

RESEARCH ARTICLE

CD4 T-cell expression of IFN- γ and IL-17 in pediatric malarial anemia

Evans Raballah^{1,2,3}, Prakasha Kempaiah⁴, Zachary Karim⁴, George O. Orinda³, Michael F. Otieno⁵, Douglas J. Perkins^{1,4}, John Michael Ong'echa^{1*}

1 University of New Mexico Laboratories of Parasitic and Viral Diseases, Centre for Global Health Research, Kenya Medical Research Institute, Kisumu, Kenya, **2** Department of Medical Laboratory Sciences, Masinde Muliro University of Science and Technology, Kakamega, Kenya, **3** Department of Biochemistry and Biotechnology, Kenyatta University, Nairobi, Kenya, **4** Center for Global Health, Department of Internal Medicine, University of New Mexico, Health Sciences Centre, Albuquerque, NM, United States of America, **5** Department of Medical Laboratory Sciences, Kenyatta University, Nairobi, Kenya

* michaelongecha@yahoo.com



OPEN ACCESS

Citation: Raballah E, Kempaiah P, Karim Z, Orinda GO, Otieno MF, Perkins DJ, et al. (2017) CD4 T-cell expression of IFN- γ and IL-17 in pediatric malarial anemia. PLoS ONE 12(4): e0175864. <https://doi.org/10.1371/journal.pone.0175864>

Editor: Adrian J.F. Luty, Institut de recherche pour le developpement, FRANCE

Received: October 5, 2016

Accepted: March 31, 2017

Published: April 20, 2017

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the [Creative Commons CC0](https://creativecommons.org/licenses/by/4.0/) public domain dedication.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This study was funded by the Fogarty International Center grant R01 TW007631-03 (JMO) and National Institutes of Health grants R01 AI51305-07 (DJP) and TW05884-07 (DJP). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abstract

In *Plasmodium falciparum* holoendemic transmission regions of western Kenya, life-threatening pediatric malaria manifests primarily as severe malarial anemia (SMA, Hb \leq 6.0 g/dL with any density parasitemia). To determine the role that CD4+ T-cell-driven inflammatory responses have in the pathogenesis of SMA, peripheral CD4+ T-cell populations and their intracellular production of pro-inflammatory cytokines (IFN- γ and IL-17) were characterized in children aged 12–36 months of age stratified into two groups: non-severe malarial anemia (non-SMA, Hb \geq 6.0 g/dL, $n = 50$) and SMA ($n = 39$). In addition, circulating IFN- γ and IL-17 were measured as part of a Cytokine 25-plex Antibody Bead Kit, Human (BioSource™ International). Children with SMA had higher overall proportions of circulating lymphocytes ($P = 0.003$) and elevated proportions of lymphocytes expressing IFN- γ ($P = 0.014$) and comparable IL-17 ($P = 0.101$). In addition, SMA was characterized by decreased memory-like T-cells (CD4+CD45RA-) expressing IL-17 ($P = 0.009$) and lower mean fluorescence intensity in memory-like CD4+ T-cells for both IFN- γ ($P = 0.063$) and IL-17 ($P = 0.006$). Circulating concentrations of IFN- γ were higher in children with SMA ($P = 0.009$), while IL-17 levels were comparable between the groups ($P = 0.164$). Furthermore, circulating levels of IFN- γ were negatively correlated with IL-17 levels in both groups of children (SMA: $r = -0.610$, $P = 0.007$; and non-SMA: $r = -0.516$, $P = 0.001$), while production of both cytokines by lymphocytes were positively correlated (SMA: $r = 0.349$, $P = 0.037$; and non-SMA: $r = 0.475$, $P = 0.001$). In addition, this correlation was only maintained by the memory-like CD4+ T cells ($r = 0.365$, $P = 0.002$) but not the naïve-like CD4+ T cells. However, circulating levels of IFN- γ were only associated with naïve-like CD4+ T cells producing IFN- γ ($r = 0.547$, $P = 0.028$), while circulating levels of IL-17 were not associated with any of the cell populations. Taken together, these results suggest that enhanced severity of malarial anemia is associated with higher overall levels of circulating lymphocytes, enhanced intracellular production of IFN- γ by peripheral lymphocytes and high circulating IFN- γ levels. In addition, the observed inverse relationship between the circulating levels of IFN- γ and IL-17 together with the reduction in the levels of memory-like CD4+ T cells expressing IL-17 in children with SMA

Abbreviations: MFI, mean fluorescence intensity; SMA, severe malarial anemia; non-SMA, non-severe malarial anemia.

may suggest possible relocation of these cells in the deeper tissues for their pathological effect.

Introduction

Malaria continues to be a major public health problem, which resulted in about 214 million cases and over 438,000 deaths world-wide in 2015 [1]. The vast majority of cases (~88%) and deaths (>90%) occur in sub-Saharan Africa, largely in immune-naïve children under five years of age [1]. *Plasmodium falciparum* is responsible for over 98% of the morbidity and mortality borne by African children [2]. The two primary severe disease outcomes of malaria are cerebral malaria and severe malarial anemia (SMA) with the distribution of these severe forms being largely dependent on malaria transmission intensity [3]. Although cerebral malaria is more common in older children in regions of low-to-moderate transmission intensity, SMA is the primary manifestation seen in children with median ages of 15 months (IQR 9–25 months) that live in holoendemic transmission areas [4]. As such, in the current study area in western Kenya, severe malaria primarily manifests as SMA (hemoglobin (Hb) < 6.0g/dL [5] peaking in children of 7–24 months of age [6].

The pathogenesis of pediatric malarial anemia (MA) in holoendemic *P. falciparum* transmission areas is largely determined by the degree of red blood cell (RBC) destruction and production [7–9]. Chronic forms of *P. falciparum* resulting from persistent parasitemia and repeated infections are a primary cause of enhanced anemia severity in African children [5, 6, 10–13] with anemia often persisting even after successful clearance of parasitemia [14] due to bone-marrow suppression [15] which is characterized by dyserythropoiesis and ineffective erythropoiesis [13, 16, 17]. Suppressed and ineffective erythropoietic responses are associated with enhanced production of macrophage-derived inflammatory cytokines [18, 19] as a consequence of prolonged immune activation [20] driven, at least in part, through interactions with CD4+ T-cells [21, 22]. For example, production of IL-12 and IL-23 by macrophages/monocytes and other myeloid antigen presenting cells (APCs) induce naïve and memory CD4+ T-cells to produce IFN- γ and IL-17, respectively [21, 22]. Although IFN- γ may induce protective immune responses against parasitemia, re-infections, anemia and clinical malaria [23–26], increased IFN- γ production has also been associated with enhanced pathogenesis [27–29]. Enhanced pathogenesis, and particularly anemia, in the context of elevated IFN- γ is consistent with murine studies showing that treatment with CpG-ODN [30] and acute toxoplasmosis [31] cause increased IFN- γ levels that appear to contribute to suppression of erythropoiesis. The myriad of effects that IFN- γ has on pathophysiological processes in bone marrow was recently reviewed by de Bruin *et al.* [32].

IL-17 is a pro-inflammatory cytokine that has been implicated in the inflammation of autoimmune diseases in response to IL-23 production [33]. We have also shown that differing levels of IL-12 and IL-23 are associated with disease severity in children with malarial anemia [34–36]. However, the effect of altered IL-12 and IL-23 on conditioning naïve and memory CD4+ T-cells to produce IFN- γ and IL-17, respectively, during an acute malarial infection is unknown. Recent studies in murine models reported that chronic malaria infection was associated with an increased proportion of effector memory CD4+ T-cells that produced IFN- γ and TNF- α which were effective at delaying and reducing parasitemia and pathology [37]. Additional studies in murine models reported involvement of IFN- γ and the IL-17/IL-23 axis in malaria associated pathology [38]. However, human studies from Cameroon reported that enhanced production of IFN- γ and IL-17 were associated with protection against pregnancy-

associated malaria [39]. Recent studies in chronic inflammatory diseases reported plasticity in the ability of effector Th17 cells to produce IL-17 and/or IFN- γ following conditioning with IL-23 [40, 41]. Similarly, Perez-Mazliah and Langhorne [42] recently reviewed plasticity in CD4 T-cell sub-sets during malaria infections.

Based on previous work from our laboratory and others [34–36, 43], we investigated the hypothesis that enhanced CD4+ T-cell-derived IFN- γ and/or IL-17 promoted enhanced pathogenesis of malarial anemia. As such, we characterized CD4+ T-cell populations and their intracellular production of IFN- γ and IL-17 in children with SMA and non-SMA. Since malarial anemia severity is significantly influenced by co-infections, such as human immunodeficiency virus type 1 (HIV-1) [44], bacteremia [45, 46] and helminthic infections [47], all children with these infections were excluded from the analyses. We show that children presenting with SMA have elevated levels of lymphocytes producing IFN- γ compared to non-SMA children. In addition, we demonstrate that children with SMA had decreased levels of memory-like CD4+ T-cells expressing IL-17 and elevated circulating IFN- γ levels. Taken together, these results implicate both IFN- γ and IL-17 in the pathogenesis of SMA.

Materials and methods

Ethics statement

The study was approved by the National Ethical Review Committee of the Kenya Medical Research Institute and the Institutional Review Board of the University of New Mexico (SSC696/SSC1377). All the parents or legal guardians of the children gave written informed consent in their language of choice (Dholuo, Kiswahili or English) before enrollment into the study. The study was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice.

Study area

The current study was performed at Siaya County Referral Hospital (SCRH) in Siaya County, western Kenya. SCRH is situated in a *P. falciparum* holoendemic transmission area that reported increased pediatric malarial admissions despite recent anti-malarial interventions [48]. SMA is the primary clinical manifestation of severe malaria in children under the age of 5 years, peaking in children aged 7–24 months [5, 6]. Previous studies observed that 53% of all the malaria-related deaths in hospitalized children under the age of 3 years were due to SMA [49]. The manifestation of pediatric MA in the study population has been described in detail elsewhere [50].

Study population

Children ($n = 89$, aged 12–35 months) presenting at the SCRH were recruited into the study. Children with *P. falciparum* parasitemia (any density) were stratified based on Hb levels as follows: non-SMA (Hb > 6.0 g/dL; $n = 50$) and SMA (Hb \leq 6.0 g/dL; $n = 39$). SMA was defined with an Hb cutoff of 6.0 g/dL based on previous investigations of >14,000 longitudinal Hb distributions in children aged less than 48 months in western Kenya [5]. Since HIV-1 promotes anemia in children with falciparum malaria [44], only HIV-1 negative children were included in analyses of the present study. HIV-1 status was determined by two rapid serological antibody tests and HIV-1 proviral DNA PCR tests as previously described [44]. Similarly, bacteremic children, as well as those with hook-worm infections, were excluded from the study. All the study participants were also free from cerebral malaria, (which is a rare occurrence in this high malaria transmission area [6]), non-falciparum malaria parasites, and hypoglycemia. Children with malaria were treated according to the Ministry of Health, Kenya (MOH)

guidelines using Coartem® (artemether and lumefantrine) for uncomplicated malaria and intravenous quinine for severe malaria. Supportive care and blood transfusions were administered according to MOH guidelines. All blood samples were obtained prior to antimalarial and/or any other treatment interventions.

Parasitemia and Hb levels determination

Venous blood (<3.0mL) samples were obtained from the study participants after the parent/guardian signed an informed consent. Thick and thin peripheral blood smears were prepared from venous blood samples and stained with Giemsa reagent for malaria parasite identification and quantification by microscopy. Asexual malaria parasites were counted against 300 leukocytes, and parasite densities were determined by multiplying the parasite count by the total leukocyte counts from an automated hematology analyzer (Beckman Coulter® AcT diff2™, Beckman-Coulter Corporation, Miami, USA). Hb levels were determined using the automated hematology analyzer (Beckman Coulter® AcT diff2™, Beckman-Coulter Corporation, Miami, USA).

Characterization of CD4+ T cell populations

Approximately 500 μ L of venous blood was obtained and stained with mouse anti-human (APC CD3, BD Pharmingen™ Cat.555335; FITC CD4, BD Pharmingen™ Cat.555346; and PE-Cy™5 CD45RA, BD Pharmingen™ Cat.555490) antibodies. Briefly, venous blood was diluted with an equal volume of plain RPMI-1640 media and aliquots of 100 μ L were pipetted into snap-cap tubes. The blood-media mixture was then stained with 10 μ L of fluorochromed surface antibodies and incubated at 4°C for 30 minutes. The red blood cells (RBCs) were then lysed using cold 1X BD Pharm Lyse™ lysing buffer (Cat.555899) at room temperature for 10 minutes. The cells were washed twice in cold wash buffer (5g Bovine serum albumin (BSA) + 0.5g Sodium azide in 500mL Phosphate buffered saline (PBS)) and fixed in 1% paraformaldehyde at 4°C. Cells were then acquired using a four-color FACSCalibur within 24 hours of staining. Lymphocytes were acquired and stored on gate R1 (Fig 1A). These were further gated as CD4+ and CD4- lymphocytes as shown in Fig 1B. In order to determine the proportion of CD4+ lymphocytes that were T cells, a third gate for CD3+ was used (Fig 1C). This gate demonstrated that ~98.0% (median value) of the CD4+ lymphocytes were (CD3+) T cells.

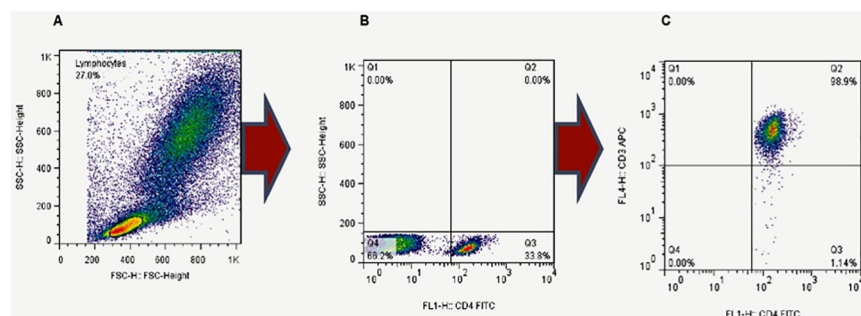


Fig 1. Characterization of CD4+ T-cell sub-populations. (A) Initial gating was performed on lymphocytes in the overall cell population during acquisition. (B) The CD4+ T-cell population was then selected by gating on CD4+ cells in the lymphocyte cell population during analyses. (C) Due to the limitation of the four-color cytometer, the populations of CD4+ cells used in subsequent analyses were CD4+ cells from the lymphocyte gate. To determine the proportion of CD4+ cells that were T-cells (CD3+CD4+), expression of CD3 by the CD4+ cells (gated in B) was determined. The median proportion of CD3+CD4+ (T-cells) among the CD4+ cells was 98.9%, suggesting that the CD4+ gating selected an enriched population of CD4+ T-cells.

<https://doi.org/10.1371/journal.pone.0175864.g001>

Intracellular cytokine stimulation assays

Since all children in the study had malaria parasitemia, measurement of *ex vivo* responses to malaria infections was determined by obtaining ~500 μ L of venous blood that was diluted with an equal volume of plain RPMI 1640 media followed by stimulation with 0.1 μ g/mL anti-CD28/CD49d (Cat.347690) in the presence of monensin (0.5 μ g/mL) for IL-17, or Brefeldin A (0.5 μ g/mL) for IFN- γ and incubated at 37°C and 5% CO₂ for six hours. After six hours of incubation cells were fixed in 1% paraformaldehyde at 4°C, permeabilized with 0.1% saponin (0.1g saponin in 100mL PBS) for thirty minutes and then stained with 10 μ L of surface antibodies (CD4 and CD45RA), and intracellular cytokine antibodies (FITC IFN- γ , BD Pharmingen™ (Cat.554551) and PE IL-17, R&D Systems, (Cat.ICP3171P)) and incubated at 4°C for thirty minutes. The cells were then processed following the steps described above and acquired using a four-color FACSCalibur immediately after staining.

Determination of circulating cytokine levels

Plasma samples were obtained from venous blood and snap-frozen at -80°C until use. Concentrations of IL-17 and IFN- γ were determined using the human Cytokine 25-plex Ab Bead Kit, (BioSource™ International) according to the manufacturer's protocol. Plates were read on a Luminex 100™ system (Luminex Corporation) and analyzed using the Bio-Plex Manager Software (Bio-Rad Laboratories), with detection limits of 4.0 pg/mL (IFN- γ) and 10.0 pg/mL (IL-17).

Data analysis

All CD4+ frequencies presented are data that resulted after compensation using unstained, single-stained and multiple-stained cells on FlowJo 7.6.3 software (TreeStar, Ashland, OR, USA). Proportions of CD4+ T-cells expressing the various surface markers and the intracellular cytokines were analyzed using FlowJo. MFIs were calculated by the geomean of the population of interest in FlowJo. Statistical analyses were performed using SPSS® software (version 19.0, IBM SPSS Inc., IL, USA). Between group comparisons were performed by Mann-Whitney U tests. Proportional differences between the clinical groups was determined using χ^2 -tests. The relationships between circulating cytokine levels and intracellular cytokine expression by different CD4+ T cell sub-populations were determined by Spearman's correlation coefficient. Statistical significance was set at $P \leq 0.050$.

Results

Characteristics of the study participants

The demographic and clinical characteristics of the study participants are summarized in [Table 1](#). Children in the two clinical categories (non-SMA and SMA) were comparable in gender ($P = 0.673$), axillary temperature ($P = 0.422$), parasite density ($P = 0.256$), prevalence of high-density parasitemia (HDP, $\geq 10,000$ parasites/ μ L) ($P = 0.259$) and carriage of sickle-cell trait ($P = 0.257$). However, children presenting with SMA were significantly younger than those with non-SMA ($P = 0.019$). In addition, children with SMA had significantly low levels of reticulocyte production index (RPI; $P = 0.007$), an indicator for erythropoietic response based on reticulocyte counts correcting for the degree of anemia [51]. Furthermore, children with SMA had elevated levels of peripheral white blood cell (WBC) counts ($P = 0.014$), proportion of circulating lymphocytes ($P = 0.003$) and reduced Hb ($P = 0.001$) relative to the non-SMA group.

Table 1. Demographic, clinical and laboratory characteristics of study participants.

Characteristic	Non-SMA	SMA	P
Participants, <i>n</i>	50	39	
Gender, female, <i>n</i> (%)	25 (50.00)	17 (43.59)	0.673 ^a
Age, months	23.00 (13)	18.00 (14)	0.019^b
Axillary temperature, °C	38.00 (1.4)	38.00 (1.2)	0.422 ^b
Hemoglobin, g/dL	9.60 (2.5)	5.00 (1.6)	<0.001^b
Reticulocyte production index	0.52 (0.34)	0.33 (0.41)	0.007^b
White blood cells, ($\times 10^9/L$)	10.60 (5)	13.30 (8)	0.014^b
Lymphocytes (%)	33.65 (24)	40.20 (13)	0.003^b
Parasite density, / μL	31,200 (131,761)	25,604 (91,327)	0.256 ^b
High-density parasitemia, <i>n</i> (%)	37 (74.00)	23 (58.97)	0.259 ^a
Sickle-cell trait, <i>n</i> (%)	10 (20.00)	4 (10.26)	0.257 ^a

Data are presented as medians (interquartile range; IQR) unless otherwise noted. Children (*n* = 89) were categorized into non-SMA (*n* = 50, Hb \geq 6.0 g/dL with any density parasitemia) and SMA (*n* = 39, Hb<6.0 g/dL with any density parasitemia). Statistical significance was set at *P*<0.05.

^aStatistical significance was determined by Chi-square analysis.

^bStatistical significance was determined by Mann-Whitney U tests.

<https://doi.org/10.1371/journal.pone.0175864.t001>

Characterization of distinct CD4+ T-cell populations

The gating strategy used in characterizing the CD4+ T-cell populations is outlined in Fig 1 (and S1–S3 Figs). The lymphocyte population was gated on to provide the overall cell populations during acquisition (Fig 1A). This gate was then dissected into CD4+ and CD4- populations (Fig 1B). The CD4+ T-cell populations reported were based on the CD4+ cells selected by this gating. However, to confirm the proportion of CD4+ cells that were T-cells (CD3+CD4+), cells expressing CD3 among the CD4+ cells (Fig 1B) were also determined. We observed that the CD4+ cells isolated (Fig 1B) were mainly CD3+ T-cells (Median (IQR) ~98.0% (3.15), Fig 1C).

Lymphocytes expressing intracellular IFN- γ and IL-17 are elevated in children with SMA

Preceding the examination of the role of CD4+ T-cells in malarial immunity, the distribution of lymphocytes and their intracellular cytokines were examined. Children with SMA had elevated lymphocytes (median (IQR) 40.00% (13.00)) relative to the non-SMA group (33.65% (25.00)); *P* = 0.003; Fig 2A). Further analyses demonstrated that the SMA group had significantly higher circulating lymphocytes expressing IFN- γ (7.78% (7.71)) and comparable IL-17 (10.30% (14.25)) relative to children with non-SMA (6.03% (4.45) and 7.80% (11.55)); *P* = 0.014 and *P* = 0.101, respectively; Fig 2B and 2C).

Memory-like CD4+ T-cells expressing IL-17 are decreased in children with SMA

Prior to characterizing naïve-like (CD4+CD45RA+) and memory-like (CD4+CD45RA-) CD4+ T-cells expressing IFN- γ and IL-17, we compared the distribution of CD4+ T-cell counts between the groups. These results demonstrated that the proportion of CD4+ T-cells was comparable between SMA and non-SMA groups (median (IQR) SMA 52.73% (11.51) and non-SMA 55.60% (16.17)); *P* = 0.394; Data not shown). In addition, naïve-like CD4+ T-cells expressing IFN- γ were non-significantly elevated in the SMA group (median (IQR) SMA

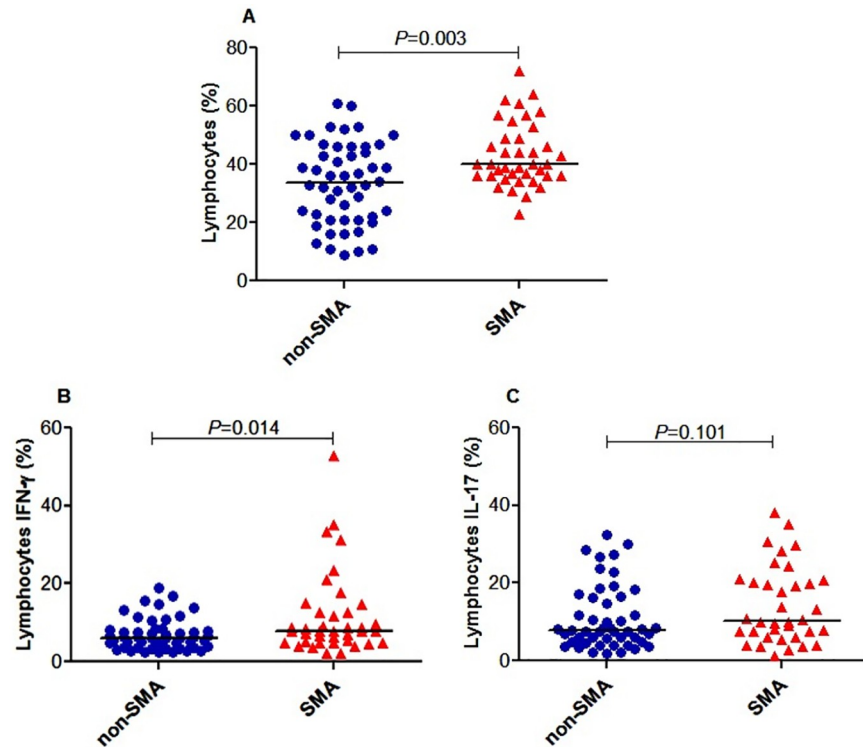


Fig 2. Lymphocytes expressing IFN- γ and IL-17 in acute malaria. Data are presented as proportions (%). The proportions were determined by flow cytometry immediately after staining. The proportions of different lymphocyte populations were determined using FlowJo Software (TreeStar, Ashland, OR, USA). The line in the middle represents the median value. Between groups comparisons were performed by Mann-Whitney U test. **(A)** The proportion of lymphocytes was higher in the SMA group relative to non-SMA group. **(B)** The proportion of lymphocytes expressing IFN- γ was higher in the SMA group relative to non-SMA group. **(C)** The proportion of lymphocytes expressing IL-17 was higher in the SMA group relative to non-SMA group.

<https://doi.org/10.1371/journal.pone.0175864.g002>

26.90% (17.50) and non-SMA 16.00% (28.28); $P = 0.250$; Fig 3A). Naïve-like CD4+ T-cells expressing IL-17 were also non-significantly higher in the SMA group (median (IQR) SMA 10.80% (16.70) and non-SMA 8.00% (12.12); $P = 0.071$; Fig 3B). Further analysis demonstrated that the memory-like CD4+ T-cells expressing IFN- γ were non-significantly reduced in the SMA group (median (IQR) SMA 1.90% (4.06) and non-SMA 3.03% (4.79); $P = 0.485$; Fig 3C). Memory-like CD4+ T-cells expressing IL-17 were significantly lower in children with SMA (median (IQR) SMA 2.86% (5.37) and non-SMA 4.85% (12.20); $P = 0.009$; Fig 3D).

Intensity of IFN- γ and IL-17 production by memory-like CD4+ T-cells is decreased in children with SMA

The intensity of T-cell response is more informative than the proportions of the cells producing the response [52], therefore we extended the above investigations by examining the intensity of IFN- γ and IL-17 in memory-like T helper cells. The strength of expression of IFN- γ and IL-17 by memory-like CD4+ T-cells (CD4+CD45RA-) was determined by calculating the mean fluorescence intensity (MFI). These results demonstrated that the MFI for IFN- γ in memory-like CD4+ T-cells (CD4+CD45RA-IFN- γ) was reduced in children with SMA (median (IQR) SMA, 18.83 (20.74) and non-SMA, 31.17 (41.93); $P = 0.063$; Fig 4A). The MFI for IL-17 in memory-like CD4+ T-cells (CD4+CD45RA-IL-17) was also significantly lower in the SMA group (median (IQR) SMA, 51.52 (35.61) and non-SMA, 83.41 (111.58); $P = 0.006$; Fig 4B).

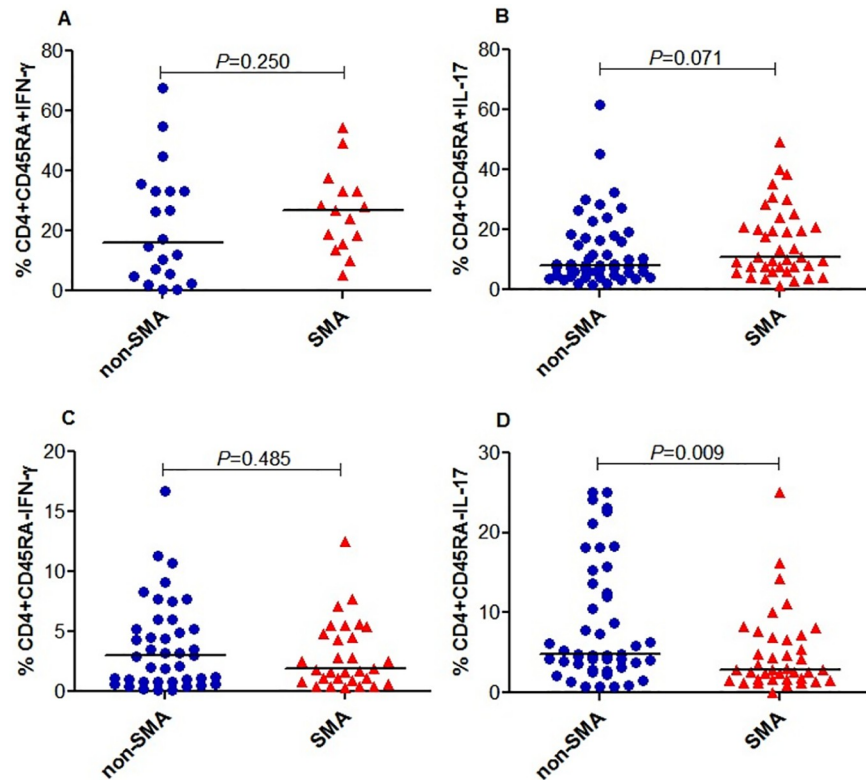


Fig 3. Memory-like CD4+ T-cells expressing IFN- γ and IL-17 in acute malaria. Data are presented as proportions (%). The proportions were determined by flow cytometry immediately after staining. The proportions of different lymphocyte populations were determined using FlowJo Software (TreeStar, Ashland, OR, USA). The line in the middle represents the median value. Between groups comparisons were performed by Mann-Whitney U test. The proportion of CD4+ T-cells was comparable between the groups (data not shown). **(A)** The proportion of naïve-like CD4+ T-cells expressing IFN- γ was comparable between the groups. **(B)** The proportion of naïve-like CD4+ T-cells expressing IL-17 was comparable between the groups. **(C)** The proportion of memory-like CD4+ T-cells expressing IFN- γ was comparable between the groups. **(D)** The proportion of memory-like CD4+ T-cells expressing IL-17 was decreased in SMA relative to non-SMA group.

<https://doi.org/10.1371/journal.pone.0175864.g003>

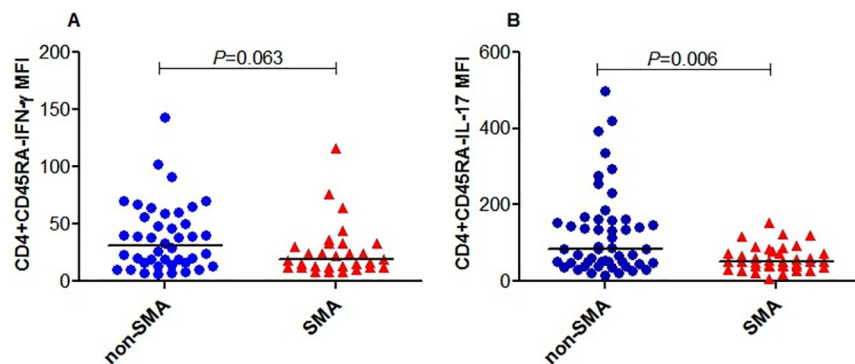


Fig 4. Intensity of IFN- γ and IL-17 production by memory-like CD4+ T-cells. Data are presented as mean fluorescence intensity (MFI). The MFI was determined by FlowJo Software (TreeStar, Ashland, OR, USA). The line in the middle represents the median value. Between groups comparisons in the clinical categories were performed by Mann-Whitney U tests. **(A)** MFI of IFN- γ in CD4+CD45RA- cells. **(B)** MFI of IL-17 in CD4+CD45RA- cells.

<https://doi.org/10.1371/journal.pone.0175864.g004>

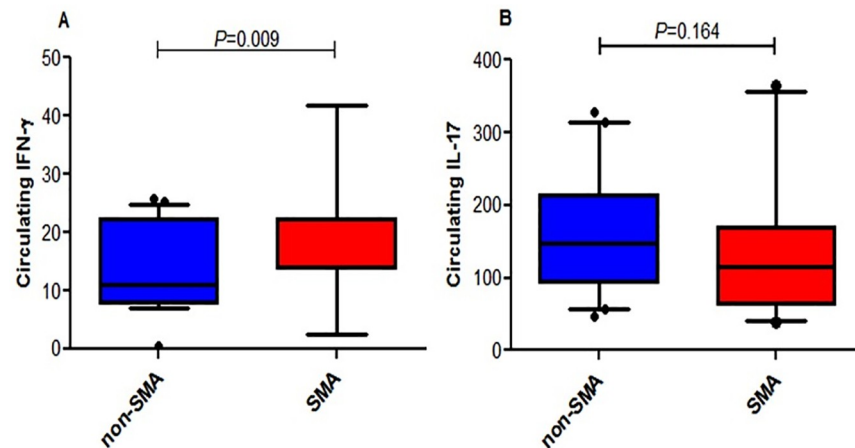


Fig 5. Circulating levels of IFN- γ and IL-17 in children with differing malarial anemia severity. Box-plots depict the data where the box represents the interquartile range, the line through the box is the median, and whiskers illustrate the 10th and 90th percentiles. Circulating IFN- γ ($n = 64$; non-SMA = 42; SMA = 22) and IL-17 ($n = 64$; non-SMA = 42; SMA = 22) concentrations were determined in plasma using the human Cytokine 25-plex Ab Bead Kit, (BioSource™ International) according to the manufacturer's protocol. Plates were read on a Luminex 100™ system (Luminex Corporation) and analyzed using the Bio-Plex Manager Software (Bio-Rad Laboratories). Between groups comparisons in the clinical categories were performed by Mann-Whitney U tests. **(A)** Circulating IFN- γ levels between non-SMA and SMA. **(B)** Circulating IL-17 levels between non-SMA and SMA.

<https://doi.org/10.1371/journal.pone.0175864.g005>

Relationship between circulating cytokines and SMA

To determine the overall production of IFN- γ and IL-17, circulating levels of the cytokines were measured in the two groups of children. Children with SMA had elevated circulating levels of IFN- γ relative to the non-SMA group (median (IQR) SMA 22.22 (8.41) and non-SMA 11.04 (14.34); $P = 0.009$; Fig 5A). Circulating IL-17 levels were comparable between the two groups (median (IQR) SMA 115.00 (105.84) and non-SMA 147.00 (120.76); $P = 0.164$; Fig 5B).

Relationship between circulating cytokine levels and intracellular cytokine expression by different CD4+ T cell sub-populations

Overall, circulating levels of IFN- γ and IL-17 were negatively correlated with each other ($r = -0.464$, $P = 0.0001$). This pattern was further observed independently in the SMA ($r = -0.610$, $P = 0.007$) and non-SMA ($r = -0.516$, $P = 0.001$) groups. However, production of both cytokines by lymphocytes was positively correlated ($r = 0.453$, $P = 0.0001$) and independently in the SMA ($r = 0.349$, $P = 0.037$) and non-SMA ($r = 0.475$, $P = 0.001$) groups. Furthermore, there was no relationship between the production of both cytokines by naïve-like CD4+ T cells ($r = 0.232$, $P = 0.425$) while there was a positive correlation in the production of both cytokines by memory-like CD4+ T cells ($r = 0.365$, $P = 0.002$). In addition, the circulating levels of IFN- γ were positively correlated with intracellular expression of IFN- γ by naïve-like CD4+ T cells ($r = 0.547$, $P = 0.028$), but not memory-like CD4+ T cells ($r = -0.214$, $P = 0.179$) nor lymphocytes ($r = 0.144$, $P = 0.298$). There was no relationship between the circulating levels of IL-17 with intracellular expression of IL-17 by naïve-like CD4+ T cells ($r = 0.252$, $P = 0.330$), memory-like CD4+ T cells ($r = 0.096$, $P = 0.492$), nor lymphocytes ($r = 0.099$, $P = 0.483$). Similarly, there was no relationship between the circulating levels of the two cytokines individually and their expression by the different cell sub-populations neither in the SMA nor the non-SMA groups ($P > 0.05$ for all comparisons).

Discussion

Understanding the mechanisms through which malaria-induced inflammatory responses modulate SMA pathogenesis will aid in the rational design of therapeutic interventions. A number of novel findings emerged in the current studies including data that suggest that naïve-like CD4+ T-cells (CD4+CD45RA+) are elevated in children with SMA relative to children with non-SMA. In addition, children with SMA had higher circulating lymphocytes expressing IFN- γ . Moreover, as previously reported by our group, children with SMA had higher concentrations of circulating IFN- γ relative to the non-SMA group [53]. In addition, we observed that circulating levels of IFN- γ were negatively correlated with circulating levels of IL-17 in both groups of children, while expression of both cytokines by lymphocytes was positively correlated. Further analysis demonstrated that circulating levels of IFN- γ were positively associated with naïve-like CD4+ T cells expressing IFN- γ while circulating levels of IL-17 were not associated with intracellular expression of IL-17 by any of the cell populations. Taken together, these results suggest that high levels of lymphocyte-derived IFN- γ may contribute to the pathogenesis of pediatric SMA. Alternatively, the observed low levels of circulating memory-like CD4+ T-cells producing IFN- γ and IL-17 in children with SMA may also suggest possible sequestration in the deeper tissues where they could induce pathogenic processes.

We hypothesized that since pediatric SMA is driven by a sustained inflammatory response following chronic plasmodial infection [5, 10–13, 54], specific populations of immune cells should contribute to the production of pro-inflammatory cytokines (e.g., IFN- γ and IL-17). In addition, recent studies by Boyle *et al.* [55] illustrated that there are malaria-specific CD4+ T-cells that produce differing cytokine responses following stimulation with *P. falciparum*-infected red blood cells (iRBCs). Therefore to test this hypothesis, we characterized the *ex vivo* ability of peripheral lymphocytes and CD4+ T-cell sub-populations to produce IFN- γ and IL-17 in acutely infected children presenting with non-SMA and SMA. In agreement with previous reports, children presenting with SMA had leukocytosis [56, 57]. In addition, our observation of elevated levels of lymphocytes expressing IFN- γ in children with SMA and memory-like CD4+ T-cells expressing IL-17 in children with non-SMA, suggests *in vivo* induction of these cells as has been reported in animal models [43, 58–60]. Furthermore, our observation that children with SMA had lymphocytosis, high IFN- γ expression by peripheral lymphocytes and the observed correlation between circulating levels of IFN- γ and naïve-like CD4+ T cells expressing IFN- γ may implicate naïve CD4+ T cells in the production of high levels of circulating IFN- γ in children with SMA. Although recent studies have been able to characterize cytokine profiles of pediatric CD4+ T-cell populations [55, 61, 62], these have related the cytokine responses with malaria protection. However, studies among Malian children investigated the cytokine profiles during the control of malaria-induced inflammation and observed a more enhanced production of anti-inflammatory cytokines during re-exposure to malaria [63].

Our observation that children with SMA were younger than those with non-SMA is consistent with previous studies among African children [50, 64]. However, the young age was not observed to affect the levels of the cell populations and cytokines investigated (see [S1 Table](#)). This observation is also consistent with recent investigations in Malawian children which observed that lymphocyte subset profiles of different forms of severe malaria were independent of age [64]. Furthermore, earlier pediatric studies in an area of high malaria transmission in western Kenya also reported no differences in these cell populations and cytokine production among children aged 0.5 to 5 years [65].

Recent studies showed that reduced protective immunity in C5aR^{-/-} mice was due to impaired function of malaria-specific CD4+ T-cells and consequent differentiation of effector

memory (CD62L^{low}CD44^{high}) and central memory (CD62L^{high}CD44^{high}) CD4+ T-cells [66]. This underscores the importance of memory CD4+ T-cells in malaria immunity. Moreover, although memory CD4+ T-cells are important in protective immunity against blood-stage parasites, they can also contribute to enhanced pathogenesis [67]. Our observation that children with SMA had decreased levels of memory-like CD4+ T-cells expressing IL-17 suggests that high levels of these cells may be required for protection against SMA pathogenesis or alternatively, they may be sequestered in the deeper tissues where they could result in SMA pathogenesis.

Previous studies in both murine models and humans have reported a strong induction of IFN- γ during acute plasmodial infections [68, 69]. Although IFN- γ can be produced by various cell types, it still remains unclear which cell types are responsible for IFN- γ -induced protection versus pathology [42]. The significantly higher proportion of IFN- γ -producing lymphocytes in children with SMA suggests that elevated lymphocytic production of IFN- γ is associated with enhanced pathogenesis. We have previously shown that children with SMA in this holoendemic region have inefficient erythropoiesis [70]. Since IFN- γ can suppress erythropoiesis [30, 32, 71, 72], enhanced lymphocytic production of IFN- γ in children with SMA may be a central process for enhanced pathogenesis in these children. Our observation that children with SMA also had low RPI may support the role of IFN- γ in suppressing erythropoiesis. However, since low RPI has also been associated with hemolysis in African children with SMA [73], the role of these cytokines in suppressing erythropoiesis needs to be investigated in the context of hemolysis and other anemia causing processes.

Initially, the IL-17/IL-23 pathway was shown to be important in the pathology of inflammation associated with autoimmune diseases [33], and in malaria pathogenesis [36, 38]. Results presented here showing decreased proportions of memory-like CD4+ T-cells expressing IL-17 in children with SMA, and comparable circulating levels of IL-17 in the two groups of children contrasts with previous investigations which, demonstrated that IL-17F levels were elevated with increased malaria severity in pediatric populations of Togo [74]. However, the finding on circulating levels of IL-17 is consistent with other studies in which IL-17 was not found to be associated with cerebral malaria (the phenotype of severe malaria in the region) and mortality in Ghanaian children [75]. Furthermore, our findings also diverge from previous findings which reported association of IL-17 with protection against pregnancy-associated malaria [39] and development of cerebral malaria in *P. berghei* infected mice [43]. These disparate findings on the role of IL-17 during malaria infections can be explained by the different pathological mechanisms proposed for the malaria disease outcomes [39, 75, 76] and by Perez-Mazliah and Langhorne's [42] review report that although Th17 cells are activated during malaria infections, their precise role is yet to be established.

In order to dissect the long term effect of IFN- γ in malaria pathogenesis, we compared the circulating IFN- γ levels between the non-SMA and SMA groups. Our data indicated that children with SMA had higher circulating levels of IFN- γ relative to the non-SMA children from the same population. This observation corroborates the findings of a previous study in western Kenya, in which the SMA phenotype had higher levels of circulating IFN- γ relative to the non-SMA children [53]. Other studies among Thai adults also showed that circulating IFN- γ levels were higher in patients with severe malaria [77]. We therefore, postulate that IFN- γ appears to be protective during early activation and becomes pathologic following chronic parasitemia. This could be due to the fact that as an inflammatory mediator, initial inflammatory responses purpose to control parasite densities. However, it has been reported that depending on the timing and magnitude of inflammatory mediators' induction, the immunological response to malaria can result to damaging host tissues [76]. Although previous studies reported that IFN- γ levels correlate with control of parasitemia [24], this study did not make such observations.

It has previously been reported that IL-12 promotes differentiation of naïve CD4+ T cells and their subsequent production of IFN- γ while IL-23 induces IL-17 production from activated CD4+ memory T cells [21, 22]. We and others have previously reported that African children with SMA have low levels of circulating IL-12 and increased levels of IL-23 [34–36]. Since IL-12 can suppress IL-23-induced IL-17 production [21, 22] our observation that circulating levels of IFN- γ and IL-17 were negatively correlated suggests that this could possibly be through the IL-12/IL-23 signaling pathway. This could also explain our observation that circulating IFN- γ levels were positively associated with naïve-like CD4+ T cells expressing IFN- γ . Although this model proposes that IL-23 induces activated CD4+ memory T cells to produce IL-17, our observation of low levels of memory-like CD4+ T cells expressing IL-17 in children with SMA may be due to that fact that differentiation of naïve CD4+ T cells to memory CD4+ T cells is accompanied by expression of receptors (CCR7 and/or CD62L) that facilitate their migration and homing to lymphoid organs or inflamed tissues [78, 79]. This pathway could also possibly explain the observed differences in the patterns seen between the naïve-like and memory-like CD4+ T cells and the low circulating IL-17 levels in children with SMA. However, additional studies are still required to confirm the source and role of IL-17 in SMA pathogenesis.

Based on the on-ground facilities in Kenya at the time of the study, we were met with several challenges. For example, we were only capable of examining two cytokines using a single parameter, whereas use of multi-parametric flow cytometry would have allowed for additional insight into the characteristics of CD4+ T-cell or CD8+ T cell populations and their role in malaria-induced pathology e.g., IFN- γ /IL-10 and IFN- γ /IL-17 co-producing cells, etc. In addition, we did not measure the levels of IL-27 that has been shown to suppress IFN- γ production by CD4+ T-cells during malaria infection [80]. Since we used peripheral blood, we do not know the distribution of these cells in different tissues, and their possible impact within the tissues.

In conclusion, we demonstrate that children with SMA have elevated expression of IFN- γ by peripheral lymphocytes. We further demonstrate that children presenting with SMA have decreased levels of memory-like CD4+ T-cells expressing IL-17 and elevated circulating IFN- γ levels. The inverse correlation observed between circulating levels of IFN- γ and IL-17 suggests that both cytokines may be implicated in modulating pathogenesis of SMA. However, future studies are warranted to determine if the responses characterized here are directly or indirectly involved in the suppression of erythropoiesis in children with malarial anemia.

Supporting information

S1 Fig. Representative flow cytometry plots of cytokine production and the gating strategy used to set the cut-offs for cytokine-positive and -negative cells. (A) Representative forward versus side scatter gating strategy for the peripheral blood stimulation for the study participants. Dot plot (B) CD4+CD45RA+ T cells, (C) CD4+CD45RA+IFN- γ + T cells, (D) CD4+CD45RA+IL-17+ T cells, (E) CD4+CD45RA-IFN- γ + T cells, (F) CD4+CD45RA-IL-17+ T cells.

(TIF)

S2 Fig. Representative flow cytometry plots showing the gating strategy using isotypic controls and matched IFN- γ antibodies to set the cut-offs for cytokine-positive and -negative cells. Isotypic controls 1–3 have their matched IFN- γ -positive cell cut-offs below them.

(TIF)

S3 Fig. Representative flow cytometry plots showing the gating strategy using isotypic controls and matched IL-17 antibodies to set the cut-offs for cytokine-positive and -negative

cells. Isotypic controls 1–3 have their matched IL-17-positive cell cut-offs below them. (TIF)

S1 Table. Characteristics of the study participants stratified by age. Data analysis performed by Mann-Whitney U tests except for gender and sickle cell trait that we compared using χ^2 tests. Data are presented as median (IQR); except for gender and sickle cell trait that are presented as n (%). (DOCX)

Acknowledgments

We are very grateful to the parents/guardians, and children from the Siaya County community, western Kenya for their participation in the study. We also thank all the University of New Mexico-KEMRI staff and the Siaya County Referral Hospital staff for their support during this study. We also wish to thank the Director of Kenya Medical Research Institute (KEMRI) for approving this manuscript for publication.

Author Contributions

Conceptualization: ER DJP JMO.

Formal analysis: ER GOO MFO DJP JMO.

Funding acquisition: DJP JMO.

Investigation: ER PK ZK JMO.

Methodology: ER PK ZK JMO.

Project administration: DJP JMO.

Resources: DJP JMO.

Supervision: DJP JMO.

Visualization: ER DJP JMO.

Writing – original draft: ER DJP JMO.

Writing – review & editing: ER PK ZK GOO MFO DJP JMO.

References

1. WHO. World Malaria Report 2015. <http://www.who.int/malaria/publications/world-malaria-report-2015/report/en/>. 2015.
2. WHO. World Malaria Report 2008. [<http://www.who.int/malaria/wmr2008>]. 2008.
3. Okiro EA, Al-Taiar A, Reyburn H, Idro R, Berkley JA, Snow RW. Age patterns of severe paediatric malaria and their relationship to *Plasmodium falciparum* transmission intensity. *Malar J*. 2009; 8:4. <https://doi.org/10.1186/1475-2875-8-4> PMID: 19128453
4. Roca-Feltrer A, Carneiro I, Smith L, Schellenberg JR, Greenwood B, Schellenberg D. The age patterns of severe malaria syndromes in sub-Saharan Africa across a range of transmission intensities and seasonality settings. *Malar J*. 2010; 9:282. <https://doi.org/10.1186/1475-2875-9-282> PMID: 20939931
5. McElroy PD, Lal AA, Hawley WA, Bloland PB, Kuile FO, Oloo AJ, et al. Analysis of repeated hemoglobin measures in full-term, normal birth weight Kenyan children between birth and four years of age. III. The Asembo Bay Cohort Project. *Am J Trop Med Hyg*. 1999 Dec; 61(6):932–40. PMID: 10674673
6. Bloland PB, Boriga DA, Ruebush TK, McCormick JB, Roberts JM, Oloo AJ, et al. Longitudinal cohort study of the epidemiology of malaria infections in an area of intense malaria transmission II. Descriptive

- epidemiology of malaria infection and disease among children. *Am J Trop Med Hyg.* 1999 Apr; 60(4):641–8. PMID: [10348241](#)
7. Chang KH, Stevenson MM. Malarial anaemia: mechanisms and implications of insufficient erythropoiesis during blood-stage malaria. *Int J Parasitol.* 2004 Dec; 34(13–14):1501–16. <https://doi.org/10.1016/j.ijpara.2004.10.008> PMID: [15582527](#)
 8. Ekvall H. Malaria and anemia. *Curr Opin Hematol.* 2003 Mar; 10(2):108–14. PMID: [12579035](#)
 9. Menendez C, Fleming AF, Alonso PL. Malaria-related anaemia. *Parasitol Today.* 2000 Nov; 16(11):469–76. PMID: [11063857](#)
 10. Ekvall H, Premji Z, Bennett S, Bjorkman A. Hemoglobin concentration in children in a malaria holoendemic area is determined by cumulated Plasmodium falciparum parasite densities. *Am J Trop Med Hyg.* 2001 Jan-Feb; 64(1–2):58–66. PMID: [11425164](#)
 11. Kitua AY, Smith TA, Alonso PL, Urassa H, Masanja H, Kimario J, et al. The role of low level Plasmodium falciparum parasitaemia in anaemia among infants living in an area of intense and perennial transmission. *Trop Med Int Health.* 1997 Apr; 2(4):325–33. PMID: [9171840](#)
 12. Luxemburger C, Nosten F, Kyle DE, Kiricharoen L, Chongsuphajaisiddhi T, White NJ. Clinical features cannot predict a diagnosis of malaria or differentiate the infecting species in children living in an area of low transmission. *Trans R Soc Trop Med Hyg.* 1998 Jan-Feb; 92(1):45–9. PMID: [9692150](#)
 13. Phillips RE, Pasvol G. Anaemia of Plasmodium falciparum malaria. *Bailliere's Best Pract Res Clin Haematol.* 1992; 5:315–30.
 14. Camacho LH, Gordeuk VR, Wilairatana P, Pootrakul P, Brittenham GM, Looareesuwan S. The course of anaemia after the treatment of acute, falciparum malaria. *Ann Trop Med Parasitol.* 1998 Jul; 92(5):525–37. PMID: [9797826](#)
 15. Helleberg M, Goka BQ, Akanmori BD, Obeng-Adjei G, Rodrigues O, Kurtzhals JA. Bone marrow suppression and severe anaemia associated with persistent Plasmodium falciparum infection in African children with microscopically undetectable parasitaemia. *Malar J.* 2005; 4(1):56.
 16. Abdalla SH, Wickramasinghe SN, Weatherall DJ. The deoxyuridine suppression test in severe anaemia following Plasmodium falciparum malaria. *Trans R Soc Trop Med Hyg.* 1984; 78(1):60–3. PMID: [6369652](#)
 17. Clark IA, Chaudhri G. Tumour necrosis factor may contribute to the anaemia of malaria by causing dyserythropoiesis and erythrophagocytosis. *Br J Haematol.* 1988 Sep; 70(1):99–103. PMID: [3179231](#)
 18. Means RT Jr., Krantz SB. Progress in understanding the pathogenesis of the anemia of chronic disease. *Blood.* 1992 Oct 1; 80(7):1639–47. PMID: [1391934](#)
 19. Wickramasinghe SN, Abdalla SH. Blood and bone marrow changes in malaria. *Baillieres Best Pract Res Clin Haematol.* 2000 Jun; 13(2):277–99. <https://doi.org/10.1053/beha.1999.0072> PMID: [10942626](#)
 20. Biemba G, Gordeuk VR, Thuma PE, Mabeza GF, Weiss G. Prolonged macrophage activation and persistent anaemia in children with complicated malaria. *Trop Med Int Health.* 1998 Jan; 3(1):60–5. PMID: [9484971](#)
 21. Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. *J Biol Chem.* 2003 Jan 17; 278(3):1910–4. <https://doi.org/10.1074/jbc.M207577200> PMID: [12417590](#)
 22. Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity.* 2000 Nov; 13(5):715–25. PMID: [11114383](#)
 23. Luty AJ, Lell B, Schmidt-Ott R, Lehman LG, Luckner D, Greve B, et al. Interferon-gamma responses are associated with resistance to reinfection with Plasmodium falciparum in young African children. *J Infect Dis.* 1999 Apr; 179(4):980–8. <https://doi.org/10.1086/314689> PMID: [10068595](#)
 24. McCall MB, Hopman J, Daou M, Maiga B, Dara V, Ploemen I, et al. Early interferon-gamma response against Plasmodium falciparum correlates with interethnic differences in susceptibility to parasitemia between sympatric Fulani and Dogon in Mali. *J Infect Dis.* 2010 Jan 1; 201(1):142–52. <https://doi.org/10.1086/648596> PMID: [19929378](#)
 25. Ong'echa JM, Lal AA, Terlouw DJ, Ter Kuile FO, Kariuki SK, Udhayakumar V, et al. Association of interferon-gamma responses to pre-erythrocytic stage vaccine candidate antigens of Plasmodium falciparum in young Kenyan children with improved hemoglobin levels: XV. Asembo Bay Cohort Project. *Am J Trop Med Hyg.* 2003 May; 68(5):590–7. PMID: [12812352](#)
 26. Robinson LJ, D'Ombrain MC, Staniscic DI, Taraika J, Bernard N, Richards JS, et al. Cellular tumor necrosis factor, gamma interferon, and interleukin-6 responses as correlates of immunity and risk of clinical Plasmodium falciparum malaria in children from Papua New Guinea. *Infect Immun.* 2009 Jul; 77(7):3033–43. <https://doi.org/10.1128/IAI.00211-09> PMID: [19380468](#)

27. Day NP, Hien TT, Schollaardt T, Loc PP, Chuong LV, Chau TT, et al. The prognostic and pathophysiologic role of pro- and antiinflammatory cytokines in severe malaria. *J Infect Dis.* 1999 Oct; 180(4):1288–97. <https://doi.org/10.1086/315016> PMID: 10479160
28. Jason J, Archibald LK, Nwanyanwu OC, Bell M, Jensen RJ, Gunter E, et al. The effects of iron deficiency on lymphocyte cytokine production and activation: preservation of hepatic iron but not at all cost. *Clin Exp Immunol.* 2001 Dec; 126(3):466–73. <https://doi.org/10.1046/j.1365-2249.2001.01707.x> PMID: 11737064
29. Perkmann T, Winkler H, Graninger W, Kremsner PG, Winkler S. Circulating levels of the interleukin (IL)-4 receptor and of IL-18 in patients with *Plasmodium falciparum* malaria. *Cytokine.* 2005 Feb 21; 29(4):153–8. <https://doi.org/10.1016/j.cyto.2004.10.008> PMID: 15652447
30. Thawani N, Tam M, Chang KH, Stevenson MM. Interferon-gamma mediates suppression of erythropoiesis but not reduced red cell survival following CpG-ODN administration in vivo. *Exp Hematol.* 2006 Nov; 34(11):1451–61. <https://doi.org/10.1016/j.exphem.2006.06.014> PMID: 17046564
31. Mullarky IK, Szaba FM, Kummer LW, Wilhelm LB, Parent MA, Johnson LL, et al. Gamma interferon suppresses erythropoiesis via interleukin-15. *Infect Immun.* 2007 May; 75(5):2630–3. <https://doi.org/10.1128/IAI.01836-06> PMID: 17307940
32. de Bruin AM, Voermans C, Nolte MA. Impact of interferon-gamma on hematopoiesis. *Blood.* 2014 Oct 16; 124(16):2479–86. <https://doi.org/10.1182/blood-2014-04-568451> PMID: 25185711
33. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med.* 2005 Jan 17; 201(2):233–40. <https://doi.org/10.1084/jem.20041257> PMID: 15657292
34. Keller CC, Yamo O, Ouma C, Ong'echa JM, Ounah D, Hittner JB, et al. Acquisition of hemozoin by monocytes down-regulates interleukin-12 p40 (IL-12p40) transcripts and circulating IL-12p70 through an IL-10-dependent mechanism: in vivo and in vitro findings in severe malarial anemia. *Infect Immun.* 2006 Sep; 74(9):5249–60. <https://doi.org/10.1128/IAI.00843-06> PMID: 16926419
35. Luty AJ, Perkins DJ, Lell B, Schmidt-Ott R, Lehman LG, Luckner D, et al. Low interleukin-12 activity in severe *Plasmodium falciparum* malaria. *Infect Immun.* 2000 Jul; 68(7):3909–15. PMID: 10858202
36. Ong'echa JM, Remo AM, Kristoff J, Hittner JB, Were T, Ouma C, et al. Increased circulating interleukin (IL)-23 in children with malarial anemia: in vivo and in vitro relationship with co-regulatory cytokines IL-12 and IL-10. *Clin Immunol.* 2008 Feb; 126(2):211–21. <https://doi.org/10.1016/j.clim.2007.08.007> PMID: 17945537
37. Stephens R, Langhorne J. Effector memory Th1 CD4 T cells are maintained in a mouse model of chronic malaria. *PLoS Pathog.* 2010; 6(11):e1001208. <https://doi.org/10.1371/journal.ppat.1001208> PMID: 21124875
38. Helmbly H. Gastrointestinal nematode infection exacerbates malaria-induced liver pathology. *J Immunol.* 2009 May 1; 182(9):5663–71. <https://doi.org/10.4049/jimmunol.0803790> PMID: 19380813
39. Megnekou R, Lissom A, Bigoga JD, Djontu JC. Effects of Pregnancy-associated Malaria on T Cell Cytokines in Cameroonian Women. *Scand J Immunol.* 2015 Jun; 81(6):508–14. <https://doi.org/10.1111/sji.12286> PMID: 25736985
40. Leavy O. T cells: Plastic TH17 cells. *Nat Rev Immunol.* 2011 Mar; 11(3):160.
41. Hirota K, Duarte JH, Veldhoen M, Hornsby E, Li Y, Cua DJ, et al. Fate mapping of IL-17-producing T cells in inflammatory responses. *Nat Immunol.* 2011 Mar; 12(3):255–63. <https://doi.org/10.1038/ni.1993> PMID: 21278737
42. Perez-Mazliah D, Langhorne J. CD4 T-cell subsets in malaria: TH1/TH2 revisited. *Front Immunol.* 2015; 5:671. <https://doi.org/10.3389/fimmu.2014.00671> PMID: 25628621
43. Ishida H, Matsuzaki-Moriya C, Imai T, Yanagisawa K, Nojima Y, Suzue K, et al. Development of experimental cerebral malaria is independent of IL-23 and IL-17. *Biochem Biophys Res Commun.* 2010 Nov 26; 402(4):790–5. <https://doi.org/10.1016/j.bbrc.2010.10.114> PMID: 21036146
44. Otieno RO, Ouma C, Ong'echa JM, Keller CC, Were T, Waindi EN, et al. Increased severe anemia in HIV-1-exposed and HIV-1-positive infants and children during acute malaria. *AIDS.* 2006 Jan 9; 20(2):275–80. <https://doi.org/10.1097/01.aids.0000200533.56490.b7> PMID: 16511422
45. Berkley J, Mwarumba S, Bramham K, Lowe B, Marsh K. Bacteraemia complicating severe malaria in children. *Trans R Soc Trop Med Hyg.* 1999 May-Jun; 93(3):283–6. PMID: 10492760
46. Were T, Davenport GC, Hittner JB, Ouma C, Vulule JM, Ong'echa JM, et al. Bacteremia in Kenyan children presenting with malaria. *J Clin Microbiol.* 2011 Feb; 49(2):671–6. <https://doi.org/10.1128/JCM.01864-10> PMID: 21106789
47. Spiegel A, Tall A, Raphenon G, Trape JF, Druilhe P. Increased frequency of malaria attacks in subjects co-infected by intestinal worms and *Plasmodium falciparum* malaria. *Trans R Soc Trop Med Hyg.* 2003 Mar-Apr; 97(2):198–9. PMID: 14584377

48. Okiro EA, Alegana VA, Noor AM, Snow RW. Changing malaria intervention coverage, transmission and hospitalization in Kenya. *Malar J.* 2010; 9:285. <https://doi.org/10.1186/1475-2875-9-285> PMID: 20946689
49. Obonyo CO, Taylor W, Ekvall H, Kaneko A, Ter Kuile F, Olliaro P, et al. Effect of artesunate plus sulfadoxine-pyrimethamine on haematological recovery and anaemia, in Kenyan children with uncomplicated, *Plasmodium falciparum* malaria. *Ann Trop Med Parasitol.* 2007 Jun; 101(4):281–95. <https://doi.org/10.1179/136485907X176337> PMID: 17524243
50. Ong'echa JM, Keller CC, Were T, Ouma C, Otieno RO, Landis-Lewis Z, et al. Parasitemia, anemia, and malarial anemia in infants and young children in a rural holoendemic *Plasmodium falciparum* transmission area. *Am J Trop Med Hyg.* 2006 Mar; 74(3):376–85. PMID: 16525094
51. Hillman RS, Flinch CA. *Red Cell Manual.* 1996; Philadelphia: F.A. Davis.
52. Seder RA, Darrah PA, Roederer M. T-cell quality in memory and protection: implications for vaccine design. *Nat Rev Immunol.* 2008 Apr; 8(4):247–58. <https://doi.org/10.1038/nri2274> PMID: 18323851
53. Ouma C, Davenport GC, Garcia S, Kempaiah P, Chaudhary A, Were T, et al. Functional haplotypes of Fc gamma (Fc γ) receptor (Fc γ RIIA and Fc γ RIIB) predict risk to repeated episodes of severe malarial anemia and mortality in Kenyan children. *Hum Genet.* 2012 Feb; 131(2):289–99. <https://doi.org/10.1007/s00439-011-1076-8> PMID: 21818580
54. Bloland PB, Lackritz EM, Kazembe PN, Were JB, Steketee R, Campbell CC. Beyond chloroquine: implications of drug resistance for evaluating malaria therapy efficacy and treatment policy in Africa. *J Infect Dis.* 1993 Apr; 167(4):932–7. PMID: 8450258
55. Boyle MJ, Jagannathan P, Farrington LA, Eccles-James I, Wamala S, McIntyre TI, et al. Decline of FoxP3+ Regulatory CD4 T Cells in Peripheral Blood of Children Heavily Exposed to Malaria. *PLoS Pathog.* 2015 Jul; 11(7):e1005041. <https://doi.org/10.1371/journal.ppat.1005041> PMID: 26182204
56. Ladhani S, Lowe B, Cole AO, Kowuondo K, Newton CR. Changes in white blood cells and platelets in children with falciparum malaria: relationship to disease outcome. *Br J Haematol.* 2002 Dec; 119(3):839–47. PMID: 12437669
57. Novelli EM, Hittner JB, Davenport GC, Ouma C, Were T, Obaro S, et al. Clinical predictors of severe malarial anaemia in a holoendemic *Plasmodium falciparum* transmission area. *Br J Haematol.* 2010 Jun; 149(5):711–21. <https://doi.org/10.1111/j.1365-2141.2010.08147.x> PMID: 20408849
58. Keswani T, Bhattacharyya A. Differential role of T regulatory and Th17 in Swiss mice infected with *Plasmodium berghei* ANKA and *Plasmodium yoelii*. *Exp Parasitol.* 2014 Jun; 141:82–92. <https://doi.org/10.1016/j.exppara.2014.03.003> PMID: 24675415
59. Meding SJ, Cheng SC, Simon-Haarhaus B, Langhorne J. Role of gamma interferon during infection with *Plasmodium chabaudi chabaudi*. *Infect Immun.* 1990 Nov; 58(11):3671–8. PMID: 1977706
60. Su Z, Stevenson MM. Central role of endogenous gamma interferon in protective immunity against blood-stage *Plasmodium chabaudi* AS infection. *Infect Immun.* 2000 Aug; 68(8):4399–406. PMID: 10899836
61. Gitau EN, Tuju J, Karanja H, Stevenson L, Requena P, Kimani E, et al. CD4+ T cell responses to the *Plasmodium falciparum* erythrocyte membrane protein 1 in children with mild malaria. *J Immunol.* 2014 Feb 15; 192(4):1753–61. <https://doi.org/10.4049/jimmunol.1200547> PMID: 24453249
62. Jagannathan P, Eccles-James I, Bowen K, Nankya F, Auma A, Wamala S, et al. IFN γ /IL-10 co-producing cells dominate the CD4 response to malaria in highly exposed children. *PLoS Pathog.* 2014 Jan; 10(1):e1003864. <https://doi.org/10.1371/journal.ppat.1003864> PMID: 24415936
63. Portugal S, Moebius J, Skinner J, Doumbo S, Doumtable D, Kone Y, et al. Exposure-dependent control of malaria-induced inflammation in children. *PLoS Pathog.* 2014 Apr; 10(4):e1004079. <https://doi.org/10.1371/journal.ppat.1004079> PMID: 24743880
64. Mandala WL, Msefula CL, Gondwe EN, Gilchrist JJ, Graham SM, Pensulo P, et al. Lymphocyte Perturbations in Malawian Children with Severe and Uncomplicated Malaria. *Clin Vaccine Immunol.* 2016 Nov 18; 23(2):95–103.
65. Chelimo K, Embury PB, Sumba PO, Vulule J, Ofulla AV, Long C, et al. Age-related differences in naturally acquired T cell memory to *Plasmodium falciparum* merozoite surface protein 1. *PLoS One.* 2011; 6(9):e24852. <https://doi.org/10.1371/journal.pone.0024852> PMID: 21935482
66. Liu T, Xu G, Guo B, Fu Y, Qiu Y, Ding Y, et al. An essential role for C5aR signaling in the optimal induction of a malaria-specific CD4+ T cell response by a whole-killed blood-stage vaccine. *J Immunol.* 2013 Jul 1; 191(1):178–86. <https://doi.org/10.4049/jimmunol.1201190> PMID: 23709683
67. Langhorne J, Ndungu FM, Sponaas AM, Marsh K. Immunity to malaria: more questions than answers. *Nat Immunol.* 2008 Jul; 9(7):725–32. <https://doi.org/10.1038/ni.f.205> PMID: 18563083
68. Langhorne J, Meding SJ, Eichmann K, Gillard SS. The response of CD4+ T cells to *Plasmodium chabaudi chabaudi*. *Immunol Rev.* 1989 Dec; 112:71–94. PMID: 2575075

69. Troye-Blomberg M, Riley EM, Perlmann H, Andersson G, Larsson A, Snow RW, et al. T and B cell responses of *Plasmodium falciparum* malaria-immune individuals to synthetic peptides corresponding to sequences in different regions of the *P. falciparum* antigen Pf155/RESA. *J Immunol*. 1989 Nov 1; 143(9):3043–8. PMID: [2478632](https://pubmed.ncbi.nlm.nih.gov/2478632/)
70. Were T, Hittner JB, Ouma C, Otieno RO, Orago AS, Ong'echa JM, et al. Suppression of RANTES in children with *Plasmodium falciparum* malaria. *Haematologica*. 2006 Oct; 91(10):1396–9. PMID: [17018392](https://pubmed.ncbi.nlm.nih.gov/17018392/)
71. Mamus SW, Beck-Schroeder S, Zanjani ED. Suppression of normal human erythropoiesis by gamma interferon in vitro. Role of monocytes and T lymphocytes. *J Clin Invest*. 1985 May; 75(5):1496–503. <https://doi.org/10.1172/JCI111853> PMID: [3923039](https://pubmed.ncbi.nlm.nih.gov/3923039/)
72. Byrd TF, Horwitz MA. Regulation of transferrin receptor expression and ferritin content in human mononuclear phagocytes. Coordinate upregulation by iron transferrin and downregulation by interferon gamma. *J Clin Invest*. 1993 Mar; 91(3):969–76. <https://doi.org/10.1172/JCI116318> PMID: [8450071](https://pubmed.ncbi.nlm.nih.gov/8450071/)
73. Fendel R, Brandts C, Rudat A, Kreidenweiss A, Steur C, Appelmann I, et al. Hemolysis is associated with low reticulocyte index and predicts blood transfusion in severe malarial anemia. *PLoS One*. 2010; 5(4):e10038. <https://doi.org/10.1371/journal.pone.0010038> PMID: [20386613](https://pubmed.ncbi.nlm.nih.gov/20386613/)
74. Ayimba E, Hegewald J, Segbena AY, Gantin RG, Lechner CJ, Agoosou A, et al. Proinflammatory and regulatory cytokines and chemokines in infants with uncomplicated and severe *Plasmodium falciparum* malaria. *Clin Exp Immunol*. 2011 Nov; 166(2):218–26. <https://doi.org/10.1111/j.1365-2249.2011.04474.x> PMID: [21985368](https://pubmed.ncbi.nlm.nih.gov/21985368/)
75. Armah HB, Wilson NO, Sarfo BY, Powell MD, Bond VC, Anderson W, et al. Cerebrospinal fluid and serum biomarkers of cerebral malaria mortality in Ghanaian children. *Malar J*. 2007; 6:147. <https://doi.org/10.1186/1475-2875-6-147> PMID: [17997848](https://pubmed.ncbi.nlm.nih.gov/17997848/)
76. Perkins DJ, Were T, Davenport GC, Kempaiah P, Hittner JB, Ong'echa JM. Severe malarial anemia: innate immunity and pathogenesis. *Int J Biol Sci*. 2011; 7(9):1427–42. PMID: [22110393](https://pubmed.ncbi.nlm.nih.gov/22110393/)
77. Phawong C, Ouma C, Tangteerawatana P, Thongshoob J, Were T, Mahakunkijcharoen Y, et al. Haplotypes of IL12B promoter polymorphisms condition susceptibility to severe malaria and functional changes in cytokine levels in Thai adults. *Immunogenetics*. 2010; 62(6):345. <https://doi.org/10.1007/s00251-010-0439-y> PMID: [20387064](https://pubmed.ncbi.nlm.nih.gov/20387064/)
78. Sallusto F, Geginat J, Lanzavecchia A. Central memory and effector memory T cell subsets: function, generation, and maintenance. *Annu Rev Immunol*. 2004; 22:745–63. <https://doi.org/10.1146/annurev.immunol.22.012703.104702> PMID: [15032595](https://pubmed.ncbi.nlm.nih.gov/15032595/)
79. Silva AL, Lacerda MV, Fujiwara RT, Bueno LL, Braga EM. *Plasmodium vivax* infection induces expansion of activated naive/memory T cells and differentiation into a central memory profile. *Microbes Infect*. 2013 Nov; 15(12):837–43. <https://doi.org/10.1016/j.micinf.2013.07.009> PMID: [23911843](https://pubmed.ncbi.nlm.nih.gov/23911843/)
80. Villegas-Mendez A, de Souza JB, Lavelle SW, Gwyer Findlay E, Shaw TN, van Rooijen N, et al. IL-27 receptor signalling restricts the formation of pathogenic, terminally differentiated Th1 cells during malaria infection by repressing IL-12 dependent signals. *PLoS Pathog*. 2013; 9(4):e1003293. <https://doi.org/10.1371/journal.ppat.1003293> PMID: [23593003](https://pubmed.ncbi.nlm.nih.gov/23593003/)