

Correspondence

Extracellular ATP and P₂X₇ receptor exert context-specific immunogenic effects after immunogenic cancer cell death

AD Garg¹, DV Krysko^{2,3}, P Vandenabeele^{2,3} and P Agostinis^{*,1}*Cell Death and Disease* (2016) 7, e2097; doi:10.1038/cddis.2015.411; published online 18 February 2016

Dear Editor,

Immunogenic cell death (ICD) facilitates danger signalling-driven trafficking of damage-associated molecular patterns (DAMPs) like extracellular ATP (eATP).^{1,2} The binding of eATP to P₂X₇ receptor triggers immunogenic signalling,³ which (along with other factors) converts the dying cancer cells into an effective anticancer vaccine.³

Endoplasmic reticulum (ER) stress is central to ICD,¹ on the basis of which ICD inducers are subdivided into two types,¹ that is, Type I (e.g., some chemotherapies), which elicit danger signalling through 'collateral' non-lethal ER stress,¹ and Type II (e.g., hypericin-photodynamic therapy (Hyp-PDT)), which elicit danger signalling via 'focused' lethal ER stress.^{1,4} Type II and Type I ICD inducers differ on several levels, for example, plasticity of danger signalling and the trafficking mechanisms of DAMPs.⁴ In fact, eATP was found to be absent during Newcastle disease virus (NDV)-induced Type II ICD despite the induction of macroautophagy (a Type I ICD-associated, eATP-trafficking mechanism).^{2,5} Moreover, we have established that Hyp-PDT-induced eATP is PERK and secretory pathway-dependent,⁶ while being independent of macroautophagy⁷ or chaperone-mediated autophagy.⁸ This raised an important question – like in the case of NDV-induced ICD, could eATP be dispensable or a partial immunogenic signal for Hyp-PDT-induced ICD?

To this end, we decided to gain further insights into the eATP-trafficking mechanism and its immunogenic potential following Hyp-PDT. To address the contribution of the pannexin/connexin-caspase axes² that elicits eATP secretion (in response to Type I ICD inducers but remains enigmatic in the Type II settings), we utilized the pan-pannexin/connexin inhibitor, carbenoxolone (CBX). In CT26 cells treated with Hyp-PDT, CBX pretreatment failed to reduce eATP (Figure 1a), thereby suggesting the dispensability of pannexins/connexins. Next, we addressed the role of caspase activity using the pan-inhibitor, zVAD-fmk. Interestingly, zVAD-fmk significantly reduced Hyp-PDT-induced eATP

(Figure 1a). Considering the previously demonstrated role of casp-8 in ICD^{1,6} we wondered whether this caspase was mediating eATP secretion. Interestingly, CT26 cells expressing caspase-8 shRNA (casp-8 shRNA) also exhibited significantly reduced eATP following Hyp-PDT (Figure 1a).

The regulation of eATP secretion by casp-8 was unexpected, as our previous study found casp-8 to be dispensable for Hyp-PDT-induced ICD, *in vivo*.⁶ This suggested that eATP secretion may not be crucial for Hyp-PDT-induced ICD, *in vivo*. To resolve this, we utilized the CT26-BALB/c mice prophylactic vaccination model. Immunogenic effects of eATP were blocked using either Apyrase or Apy (an ATP-degrading enzyme, Figure 1b) or a 2,3-dialdehyde derivative of ATP, that is, oxidized-ATP (Oxi-ATP, a P₂X₇ receptor antagonist) or a combination of both (i.e., Apy+Oxi-ATP).³ Approximately 70% of the mice immunized with Hyp-PDT-based vaccine efficiently rejected the formation of CT26 tumours at the challenge site (Figure 1c). Interestingly, eATP degradation or blockade of P₂X₇ receptor, alone, failed to strongly reduce the tumour-rejecting immunity (Figure 1c). On the other hand, only the combination of Apy+Oxi-ATP significantly reduced the vaccine's tumour-rejecting capacity (Figure 1c). Thus, eATP, despite being ubiquitously secreted after Hyp-PDT,^{6,7,8} only acts as a partial immunogenic signal, and thus singular blockade of either eATP or its P₂X₇ receptor is unable to reduce the immunogenic potential of the vaccine.

These results are unprecedented because eATP and P₂X₇ receptor had been shown to act in a synergistic manner.^{1,2,3} Here, we rather observed a potentiating effect, that is, blockade of either eATP or P₂X₇ receptor did not, but combined blockade significantly reduced ICD's immunogenic potential. Thus, our results suggest that the mere presence of eATP does not ensure the presence of corresponding immunogenic activity in all contexts. Moreover, a certain degree of redundancy exists on the level of purinergic receptor agonists, and thus these results

¹Cell Death Research & Therapy (CDRT) Unit, Department of Cellular and Molecular Medicine, Faculty of Medicine, KU Leuven University of Leuven, Leuven, Belgium;
²Molecular Signaling and Cell Death Unit, Inflammation Research Center, VIB, Ghent, Belgium and ³Department of Biomedical Molecular Biology, Ghent University, Ghent, Belgium

*Corresponding author: P Agostinis, Cell Death Research & Therapy (CDRT) Unit, Department of Cellular and Molecular Medicine, Faculty of Medicine, KU Leuven University of Leuven, Campus Gasthuisberg O&N1, Herestraat 49, Box 802, Leuven 3000, Belgium. Tel: +32 16 345715; Fax: +32 16 34 5991; E-mail: patrizia.agostinis@med.kuleuven.be

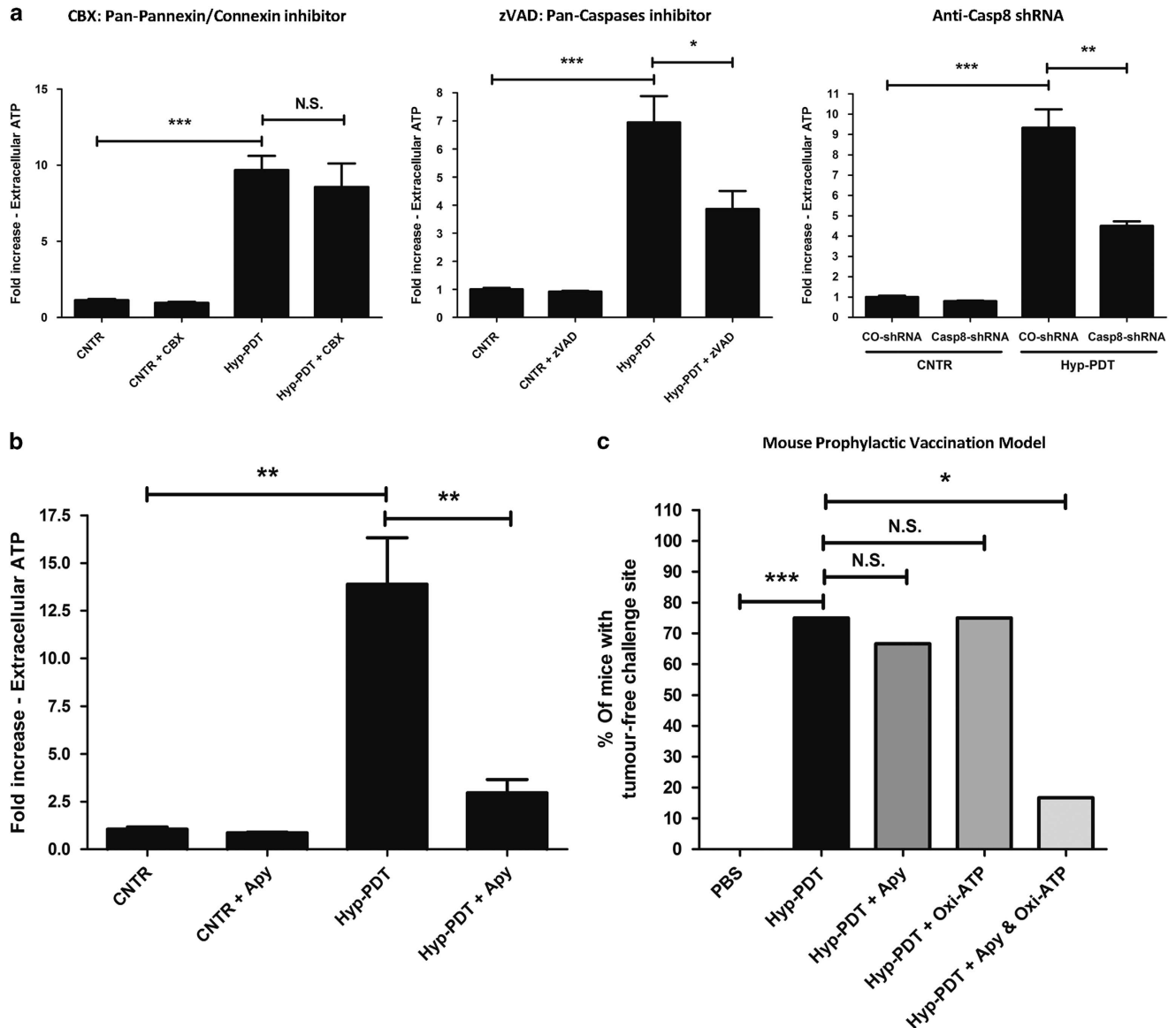


Figure 1 Extracellular ATP and P_2X_7 receptor together potentiate ICD in cancer. (a) CT26 cells were treated with Hyp-PDT (dosage: 150 nM Hypericin preincubation for 16 h followed by light irradiation with a total fluence of 2.70 J/cm²) as described previously⁶ and recovered for eATP analysis 1 h post treatment. Depending on the settings (as indicated in the legends above the graphs), the cells were preincubated with CBX (100 μ M for 1 h) or zVAD-fmk (25 μ M for 30 min). Alternatively, CT26 cells expressing control shRNA (CO-shRNA) or casp-8 shRNA were utilized as described previously.⁶ Extracellular ATP was detected using the standard luciferin-luciferase bioluminescence assay.⁷ Here, $n = 3-4$, mean \pm S.E.M., Student's *t*-test, ** $P < 0.01$ and *** $P < 0.001$, NS, non-significant; CNTR, untreated controls. (b) In another case, CT26 cells were treated with Hyp-PDT as described above and incubated for 15 min post recovery with Apyrase (Apy; 10 U/ml); eATP was then analyzed as described above. (c) For testing of immunogenicity, the CT26-BALB/c mice model was utilized.⁶ Here, the CT26 cells were treated with Hyp-PDT followed by 'vaccine' preparation as described previously.⁶ In certain cases, the vaccines were mixed/co-injected with either Apy (10 U/ml for 15 min) or Oxi-ATP (4 mg/kg per mouse) or both (Apy+Oxi-ATP). These respective vaccines were given twice with an interval of 7-8 days between vaccinations in one of the flanks of the syngenic BALB/c mice. About 8-10 days following the vaccination regimen, the vaccinated mice were challenged on the contra-lateral flank with live CT26 cells. Thereafter, the mice were monitored for the occurrence of CT26 tumours at the challenge site. Here, $n = 10$ for PBS, $n = 12$ for Hyp-PDT, $n = 12$ for Hyp-PDT+Apy, $n = 12$ for Hyp-PDT+Oxi-ATP and $n = 6$ for Hyp-PDT+Apy+Oxi-ATP, Fisher's exact test; * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$; NS, non-significant

may also point to the release of such (as-yet-uncharacterized) agonists from dying cells. Lastly, these observations are based on the heterotopic (subcutaneous) tumour model; it would be crucial to reanalyze the role of eATP in an orthotopic tumour model to overcome immunological variations stemming from incompatibility between the transplanted cancer type and the surrounding tissue.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements. ADG is supported by the FWO postdoctoral fellowship 2013 from the Research Foundation Flanders (FWO-Vlaanderen). This work is supported by FWO (G0584.12N and K202313N to PA; G.0607.13N to PA, PV/DVK; G.0A54.13N to DVK) and C16/15/073 grant of the KU Leuven to PA and BOF14/GOA/019 of Ghent University to DVK. PV holds a Methusalem grant (BOF09/01M00709) from

the Flemish Government. This paper represents research results of the IAP7/32 funded by the Interuniversity Attraction Poles Programme, initiated by the Belgian State.

1. Garg AD *et al.* *Front Immunol* 2015; **6**: 588.
2. Martins I *et al.* *Cell Death Differ* 2014; **21**: 79–91.
3. Ghiringhelli F *et al.* *Nat Med* 2009; **15**: 1170–1178.
4. Garg AD *et al.* *Cell Death Differ* 2014; **21**: 26–38.
5. Koks CA *et al.* *Int J Cancer* 2015; **136**: E313–E325.
6. Garg AD *et al.* *EMBO J* 2012; **31**: 1062–1079.
7. Garg AD *et al.* *Autophagy* 2013; **9**: 1292–1307.
8. Garg AD, Dudek AM, Agostinis P. *Cell Death Dis* 2013; **4**: e826.



Cell Death and Disease is an open-access journal published by *Nature Publishing Group*. This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>