

PHARMACOKINETICS

Tolerability, pharmacokinetics and antiviral activity of rHSA/IFN α 2a for the treatment of chronic hepatitis B infection

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AIMS

A recombinant human serum albumin-interferon alpha2a fusion protein (rHSA/IFN α 2a) is expected to extend the half-life of IFN α 2a. This study aims to evaluate the tolerability, safety and efficacy of rHSA/IFN α 2a.

METHODS

This is an open, randomized, positive control, multiple-dose ascending Phase Ib study. A panel of 32 treatment naïve and non-cirrhotic chronic hepatitis B patients were divided into four cohorts, and each received 600, 750 or 900 μ g of rHSA/IFN α 2a or 180 μ g of PEG-IFN α 2a for 3 months. Tolerability, pharmacokinetics and antiviral responses were assessed.

RESULTS

Thirty-one of 32 enrolled patients completed the treatment study. The rHSA/IFN α 2a treatment was better tolerated than the PEG-IFN α 2a 180 μ g treatment, as evidenced by blood cell counts and higher serum albumin levels. Half-life ($t_{1/2}$) of rHSA/IFN α 2a was estimated to be 120–140 h, and is potentially suitable for a dosing interval of 2 weeks or longer. Pharmacokinetics of the last dose between rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g, with the exception of $t_{1/2}$, was comparable, and a similar kinetics of inhibiting HBV DNA replication was observed in both groups. Mean reductions in serum HBV DNA levels after treatment were -1.32 , -2.13 , -1.10 and -2.48 log₁₀ IU/ml in the 600, 750 and 900 μ g rHSA/IFN α 2a groups and PEG-IFN α 2a group, respectively.

CONCLUSIONS

The rHSA/IFN α 2a treatment was well tolerated and can be administered biweekly. Similar efficacy in inhibiting HBV replication was observed in both PEG-IFN α 2a and rHSA/IFN α 2a 750 μ g groups.

WHAT IS ALREADY KNOWN ABOUT THIS SUBJECT

- A recombinant human serum albumin-interferon alpha2a fusion protein (rHSA/IFN α 2a) was expressed in *Pichia pastoris* in fusion with albumin, which is expected to extend the half-life of IFN α 2a.
- Little is known about the tolerability, pharmacokinetics and antiviral activity of rHSA/IFN α 2a treatment for HBV.

WHAT THIS STUDY ADDS

- rHSA/IFN α 2a was well tolerated and effective at inhibiting HBV DNA.
- The rHSA/IFN α 2a treatment was well tolerated and can be administered biweekly.

Introduction

Chronic hepatitis B (CHB) affects 240 million people worldwide, and the disease burden is enormous in endemic regions [1, 2]. Persistent viral replication is independently linked to dismal outcomes of chronic HBV infection, including cirrhosis, hepatocellular carcinoma and severe complication-related mortality [3, 4]. Although effective antiviral therapies exist, all have specific limitations from the emergence of drug resistance to certain safety concerns associated with long-term use. To date, the recommended therapy for chronic HBV infection includes the use of either α -interferon or nucleoside analogues (i.e. lamivudine, adefovir, entecavir, telbivudine or tenofovir). The α -interferon therapy is only partially effective, is frequently limited by adverse effects, such as fatigue/asthenia, pyrexia, myalgia and headache, and is also expensive. Although HBV replication can be efficiently inhibited by nucleoside analogues, it rebounds after withdrawal. The development of drug-resistant mutants is frequently detected with early generation of nucleotide analogues [5, 6]. Furthermore, lifelong antiviral treatment is necessary for most patients, as fewer than 10% of treated patients experience clearance of chronic HBV infection, which is marked by the seroconversion of positive hepatitis B surface antigen (HBsAg) to positive anti-HBs antibody [5, 6].

IFN- α is one of the approved antivirals for treating chronic HBV infection, and the advantages of IFN- α treatment include the lack of drug resistance and a definite treatment course that usually takes 48 weeks. Nucleos(t)ide analogues (NAs) suppress viral replication, improve liver injury, reverse a certain degree of fibrosis and block the progression of chronic liver disease. However, indefinite treatment is required and chronic HBV infection rarely cured.

Different treatment strategies for using long acting immunomodulation, RNA interference and viral entry inhibition are being explored and likely advance the treatment of hepatitis B [7, 8]. rHSA/IFN α 2a is a novel fusion protein translated from genes encoding human IFN- α and albumin, which is expressed in *Pichia pastoris*. The resultant 85.7-kDa molecule is a single polypeptide that combines the antiviral property of IFN- α with the long serum half-life of albumin [9]. The fusion with albumin can delay the degradation of interferon, which is functionally similar to the pegging of IFN α 2a.

We conducted a Phase 1b trial with rHSA/IFN α 2a. The objectives of this study were to present the characteristics, pharmacokinetics (PK), pharmacodynamics and clinical and virologic outcomes of rHSA/IFN α 2a treatment, a novel therapeutic for treating chronic HBV infection.

Patients and methods

Subjects

The trial took place in our hospital, located in northeast China. Naïve chronic hepatitis B patients undergoing treatment were enrolled into this study. Inclusion criteria were: male or female subjects, aged 18–65 years; subjects with chronic HBV infection (serum HBsAg detectable for >6 months); subjects who are serum HBeAg positive with HBV DNA >20 000 IU ml⁻¹ or serum HBeAg negative with HBV DNA >2000 IU ml⁻¹; the subject's serum alanine aminotransferase (ALT) had to be >2 \times ULN, but below 10 \times ULN.

Exclusion criteria were: subjects who received steroid treatment or immunosuppression 3 months prior to entry; subjects who received interferon therapy or nucleotide analogue therapy 6 months prior to this study; subjects with existing active lung disease or history of interstitial lung disease; subjects with Hb < lower normal limits and/or absolute neutrophil count (ANC) <1.5 \times 10⁹ l⁻¹ and/or platelet count <90 \times 10⁹ l⁻¹ and/or white blood cell count (WBC) <3 \times 10⁹ l⁻¹; subjects with other concurrent severe chronic medical conditions; subjects with evidence of hepatic decompensation; subjects seropositive for HIV and HCV; subjects with past or current thyroid disease under treatment.

The study protocol and informed consent form were in conformance with the principles embodied in the Declaration of Helsinki, and this clinical trial (ClinicalTrials.gov number: NCT01671787) was approved by the Ethics Committee of the First Hospital of Jilin University. All subjects signed an informed consent prior to enrolment.

Study design

This was an open, positive control, multiple-dose ascending, Phase 1b clinical trial of rHSA/IFN α 2a. A total of 32 subjects were enrolled, and were divided into four cohorts. These cohorts received 600, 750 or 900 μ g of rHSA/IFN α 2a or 180 μ g of PEG-IFN α 2a (positive control); in which tolerability, antiviral response and PK were assessed. rHSA/IFN α 2a was administered in three escalating doses (600, 750 or 900 μ g) biweekly, and PEG-IFN α 2a was administered at 180 μ g weekly, subcutaneously. The treatment was paused for 4 weeks (washout) after the first administration to collect samples for PK analysis, and each group resumed treatment for an additional 12 weeks. Safety and antiviral responses were assessed at weeks 5, 9, 13 and 17. Study drugs were uniformly administered in the clinical research centre to assure the quality. After completion of the dosing period, subjects were required to undergo follow-up visits

at weeks 15, 16 or 17 for off-treatment safety, antiviral response and PK evaluations. Subjects treated with rHSA/IFN α 2a 600 μ g were first evaluated and observed for 8 weeks. If the subjects tolerated the initial dose, the next two groups started receiving 750 μ g of rHSA/IFN α 2a or Peg-IFN α -2a 180 μ g. If subjects in the 750 μ g group, who were also observed for 8 weeks, tolerated the dose, the last groups commenced receiving 900 μ g of rHSA/IFN α 2a. The study design is presented schematically in Figure 1.

Assessment of adverse events and tolerability

Adverse events were regularly assessed by clinical laboratory tests, vital signs, 12-lead electrocardiograms (ECGs), abdominal ultrasound, chest X-ray, physical examination, and concomitant medications throughout the study using the baseline data as control. Adverse events were coded using the Medical Dictionary for Regulatory Activities (MedDRA, version 16; MedDRA MSSO, McLean, VA). The severity of adverse events and laboratory abnormalities were graded according to the protocol of Common Terminology Criteria for Adverse Events (CTCAE) 4.0 that defines toxicity criteria.

Tolerability was assessed by physicians who participated in this trial at week 9, after each treatment group had been observed for 8 weeks. If more than 50% of the subjects had any of the following adverse events, the dose escalation was terminated: ANC $<0.5 \times 10^9 \text{ l}^{-1}$, platelet (PLT) count $<30 \times 10^9 \text{ l}^{-1}$, ALT $>10 \text{ ULN}$, persistently elevated ALT or accompanied by bilirubin elevation after dose titration, serum total bilirubin $>51.3 \mu\text{mol l}^{-1}$, development of ascites, hepatic encephalopathy, or psychiatric disorders, allergic reactions, uncontrolled thyroid disease, diabetes mellitus, as well as serious damage to the heart, kidneys, brain and lungs. The process for dose reduction consisted of two steps. If ANC or PLT count was between $0.5 \times 10^9 \text{ l}^{-1}$ and $0.75 \times 10^9 \text{ l}^{-1}$ or between $30 \times 10^9 \text{ l}^{-1}$ and $50 \times 10^9 \text{ l}^{-1}$, respectively, the drug dose was reduced. If ANC or PLT count fell below $0.5 \times 10^9 \text{ l}^{-1}$ or $30 \times 10^9 \text{ l}^{-1}$, respectively, the drug was discontinued. If ALT levels increased to $>10 \text{ ULN}$, the drug was also discontinued. Ascending of drug dose resumed once cytopenia was recovered (i.e. ANC were $\geq 0.75 \times 10^9 \text{ l}^{-1}$ or PLT count was $\geq 50 \times 10^9 \text{ l}^{-1}$).

Study drugs and administration method

Test drugs. rHSA/IFN α 2a was expressed in *Pichia pastoris* in fusion with albumin by Beijing Bio-Fortune Ltd. rHSA/IFN α 2a is 750 amino acids long, with a molecular weight of 85 694.50 and an isoelectric point of 5.845. The tested batch number was 92 13/20121001. This plasmid was created by genetic engineering technology, in which human serum albumin and IFN- α genes were seamlessly fused. Then the plasmid was integrated into the chromosome of *P. pastoris*. The *P. pastoris* can secrete the fusion protein (rHSA/IFN α 2a) into inorganic salt medium. The secreted fusion protein was purified by highly effective separation and purification technology and lyophilized. The lyophilized proteins are reconstituted in 1 ml physiological saline and administrated by subcutaneous injection. Pegasys (PEG-IFN α 2a 180 μ g) was purchased from Roche Pharmaceuticals Ltd (batch number is B1318//201302-201601). Both were stored at 4°C until use.

rHSA/IFN α 2a and PEG-IFN α 2a were subcutaneously injected at a 5-cm periumbilical area biweekly for seven doses and weekly for 13 doses, respectively. IFN α 2a concentration in 750 μ g of rHSA/IFN α 2a was equal to that in PEG-IFN α 2a 180 μ g.

Efficacy

The primary antiviral endpoint was the log change in serum HBV DNA from baseline (day 1) to week 17. Other end points included HBeAg serum conversion rate, the reduction in HBeAg level, and the normalization rate of ALT and AST. Blood samples were collected at weeks 5, 9, 13 and 17.

Blood collection for pharmacokinetic, neopterin kinetic and IFN antibody analysis

Blood samples for rHSA/IFN α 2a PK and neopterin kinetic analyses were collected pre-dose and at 2, 6, 12, 24, 48, 60, 72, 84, 96, 120, 168, 240, 336, 504 and 672 h post-dose at weeks 1 and 15 (first dose and last dose), and pre-dose at weeks 11 and 13 (the fifth and sixth dose). Blood samples for PEG-IFN α 2a PK analyses were collected at pre-dose and at 2, 6, 12, 24, 48, 60, 72, 84, 96, 120, 168, 240, 336 and 504 h post-dose at weeks 1 and 16 (first and last dose), and pre-dose at weeks 14 and 15 (the eleventh and twelfth dose). Blood samples for IFN antibody analyses were collected pre-dose on weeks 1, 5, 9, 13 and 17. Blood samples were

W1	W2	W3	W4	W5-W8	W9	W10	W11	W12	W13	W14	W15	W16	W17	W18
DOSE	Washout			Dose								Drug withdrawal		
PK							PK		PK		PK			

rHSA/IFN α 2a group (A)

W1	W2	W3	W4	W5-W8	W9	W10-W13	W14	W15	W16	W17	W18
DOSE	Washout			Dose						Drug withdrawal	
PK							PK	PK	PK		

PEG-IFN α 2a group (B)

Figure 1

Flow chart of the study

collected and placed into a vacutainer without an anti-coagulant, clotted for 30 min, and centrifuged at $3724 \times g$ for 10 min at 4°C .

Detection methods

HBV DNA level was tested using Roche's COBAS TaqMan kit. The undetectable level was 15 IU ml^{-1} . Tests were conducted at the Hepatology Department of the First Hospital of Jilin University, Changchun, China. Liver and kidney function, as well as biochemical parameters, were tested by an automatic biochemical instrument. Blood cell counts and routine urine were also tested by an automatic detection instrument. These were all performed at the Clinical Laboratory of the First Hospital of Jilin University. Concentrations of IFN alpha2a, neopterin and conjugated antibody in serum were determined and validated by enzyme-linked immunosorbent assay (ELISA) at our hospital laboratories. Serum IFN neutralizing antibodies were tested using the vesicular stomatitis virus and ELISA method at the same laboratory.

Statistical methods

Serum PK parameters including maximum observed serum concentration (C_{max}), time to maximum observed serum concentration (T_{max}), area under the concentration–time curve from time of dosing (zero hours) to the last time point with measurable serum concentration (AUC_{0-t}) prior to next dose, AUC from time of dosing (zero hours) extrapolated to infinity ($\text{AUC}_{0-\infty}$), as well as terminal elimination half-life of the drug in serum ($t_{1/2}$), clearance (CLz/F), $\text{MRT}_{0-\infty}$, volume (Vz/F) at first dose and $C_{\text{ss max}}$, $\text{AUC}_{\text{ss0-t}}$, $\text{AUC}_{\text{ss0}-\infty}$, $t_{\text{ss } 1/2}$, $T_{\text{ss max}}$, CLz/F_{ss} , $\text{MRT}_{\text{ss0}-\infty}$, Vz/F_{ss} , accumulation rate (R), degree of fluctuation (Df) and C_{avg} at last dose, were estimated based on the observed concentration–time data by the noncompartmental PK approach using WinNonlin version 6.4 (Pharsight Corporation, Mountain View, CA).

Variables analysed using Student's *t*-test or Kruskal–Wallis test or regression or correlation analysis were established using the SAS 9.1 software (USA). Results are presented as mean \pm standard deviation. *P*-values < 0.05 in two-sided tests were considered statistically significant.

Results

A total of 91 subjects were screened initially, and 32 of them were enrolled and treated in the trial. Thirty-one subjects completed all safe and anti-virus efficacy studies, and 28 of them completed the PK study. In general, demographics and disease characteristics in each treatment group were well-matched, except for age. Subjects were younger in the Peg-IFN α 2a 180 μg group than in the rHSA/IFN α 2a group. However, the difference was not statistically significant ($P > 0.05$, Table 1). The majority of subjects were males, 30/32 (93%) of subjects were Han Chinese, and 4/32 (12.5%) were HBeAg-negative.

Tolerability

rHSA/IFN α 2a was well-tolerated after over seven injection treatments, and tolerability was better than for PEG-IFN α 2a. During the study period, body weight fluctuated once, but no significant change was found. Physical examination, abdominal colour Doppler ultrasound, chest X-ray and ECG analysis did not reveal any significant change before and after treatment. There were no significant changes in urine and blood coagulation routines.

Red blood cell count, haemoglobin, WBC count, neutrophil cell count and lymphocyte absolute value, and PLT counts decreased to different extents in each treatment group after dosing. The extent of reduction was larger in the PEG-IFN α 2a group than in the rHSA/IFN α 2a treatment groups. The time for gradually stabilizing the decreased blood cell

Table 1

Demographics and disease characteristics of the four cohorts at baseline

Baseline parameter	rHSA/IFN α 2a 600 μg (<i>n</i> = 8)	rHSA/IFN α 2a 750 μg (<i>n</i> = 8)	rHSA/IFN α 2a 900 μg (<i>n</i> = 8)	Peg-IFN α 2a 180 μg (<i>n</i> = 8)	<i>P</i>
Gender (male/female)	6/2	5/3	6/2	4/4	>0.05
Ethnic (Han/other)	7/1	7/1	8/0	8/0	>0.05
Age, mean (SD) years	39.88 ± 9.43	30.88 ± 7.75	36.50 ± 10.98	25.00 ± 4.14	>0.05
Smoker (yes/no)	1/7	3/5	1/7	3/5	>0.05
Drinker (yes/no)	2/6	0/8	0/8	1/7	>0.05
BMI, mean (SD), kg m^{-2}	22.83 ± 2.41	22.86 ± 2.70	23.31 ± 2.68	21.55 ± 2.72	>0.05
HBV DNA, mean (SD) $\log_{10} \text{ IU/ml}$	7.93 ± 0.63	7.64 ± 0.79	7.28 ± 1.14	7.33 ± 0.92	>0.05
HBeAg, IU ml^{-1}	>250	>250	>250	>250	–
HBeAg, IU ml^{-1}	755.67 ± 505.23	509.81 ± 477.99	464.41 ± 564.22	805.19 ± 524.76	>0.05
ALT, mean (SD) U l^{-1}	189.13 ± 71.27	199.50 ± 99.80	141.63 ± 51.65	139.63 ± 63.86	>0.05
HBeAg negative, <i>n</i> (%)	1 (12.5)	2 (25)	0 (0)		>0.05

count was shorter in the rHSA/IFN α 2a groups (approximately 4–7 weeks) than that in the PEG-IFN α 2a group (approximately 7–9 weeks), and decreased levels of blood cell count between different rHSA/IFN α 2a groups were similar. There were no significant dose-related reductions among the rHSA/IFN α 2a groups (Figure 2 and Table 2).

The severity of adverse events was evaluated according to CTCAE version 4.0. There were 76, 96, 110 and 126 adverse event occurrences in the 600, 750 or 900 μ g rHSA/IFN α 2a or PEG-IFN α 2a 180 μ g groups, respectively. The frequency of adverse events did not significantly increase with the increased rHSA/IFN α 2a dosages. The majority of these adverse events were reductions in neutrophil absolute value, WBC counts, albumin, PLT count and haemoglobin absolute value, which were all evaluated to be drug-related.

The severity of the majority of the adverse events was grade 1–2. Adverse events at grade 3 occurred in 21 subjects, in which three, eight, two and eight of these subjects, respectively, appeared in the 600, 750, 900 μ g rHSA/IFN α 2a groups and the PEG-IFN α 2a 180 μ g group, indicating no dose-dependence. Grade 3 adverse events mainly involved the reduction in neutrophil absolute value and WBC count, as well as elevated ALT (Table 3).

At the early stage of treatment, most subjects experienced fever and headache. With the treatment time extended, most subjects appeared to adapt to interferon therapy. Furthermore, the adaptation period was shorter in the rHSA/IFN α 2a

groups (approximately 6–10 weeks) than in the PEG-IFN α 2a group (approximately 8–13 weeks). Occurrence rates of fever, body aches and headache were higher in the PEG-IFN α 2a 180 μ g group than in the rHSA/IFN α 2a 750 μ g groups (Table 4). Three subjects were administered 0.5 g of acetaminophen once for fever, and two subjects were given 4 mg of chlorpheniramine once for allergic reactions. All subjects recovered well from these adverse events.

ALT level in one subject, who received 600 μ g of rHSA/IFN α 2a, was elevated to 620 U l $^{-1}$ (>10 ULN); and the drug was discontinued after 8 weeks of treatment. A 25% reduction of dosage was applied at the sixth and eighth dosing in one subject, and at the sixth dosing in the second patient in PEG-IFN α 2a group, due to ANC reduction. All the remaining subjects completed their designated regimens. Prior to the last dosing, one subject in the PEG-IFN α 2a 180 μ g group experienced erythema nodosum, which was PEG-IFN α 2a-related and recorded as a serious adverse event (SAE) by the principal investigator. After 4 months of treatment with hydroxychloroquine, the erythema nodosum was cured without adjusting the PEG-IFN α 2a dosage. No patient death occurred during the study period.

Immunogenicity test

There was no detectable neutralizing antibody to IFN α 2a and rHSA after rHSA/IFN α 2a treatment (Table 5), and some of the subjects appeared to have detectable activity of binding

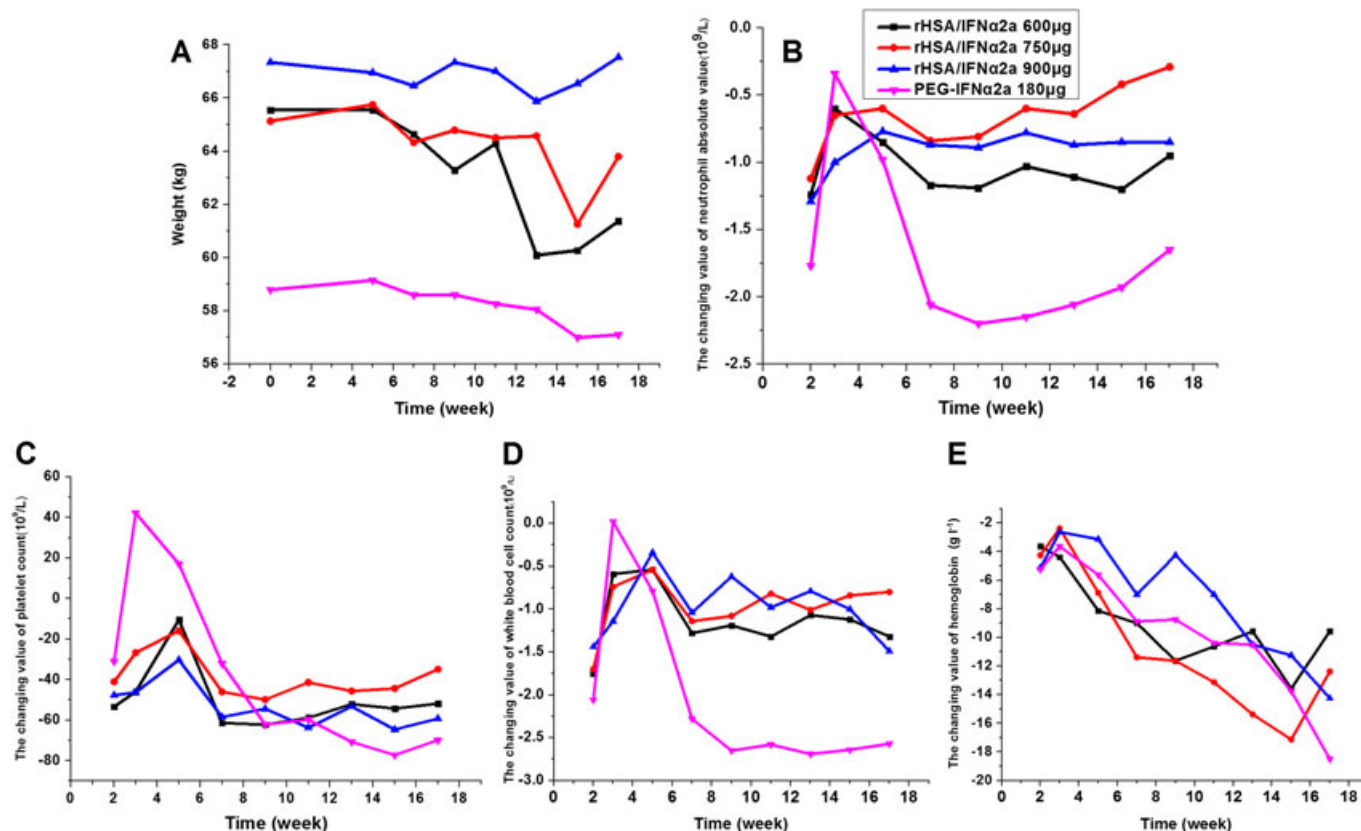


Figure 2

Reduced value of safety factors at different time points after treatment in each group (all compared with the baseline values). Body weight (A); neutrophil absolute value (B); platelet counts (C); WBC counts (D); haemoglobin absolute value (E)

Table 2

Stabilization time and lowest value of decreased blood cell counts in each treatment group

Factor	rHSA/IFN α 2a group			PEG-IFN α 2a 180 μ g group		
	Maximal reduction in parameter (Week 17)	Stabilization time	Lowest value (time)	Reduction in parameter (Week 17)	Stabilization time	Lowest value (time)
Neutrophil absolute value	$-0.95 \times 10^9 \text{ l}^{-1}$	Week 4	$0.69 \times 10^9 \text{ l}^{-1}$ (Week 2)	$-1.65 \times 10^9 \text{ l}^{-1}$	Week 7	$0.7 \times 10^9 \text{ l}^{-1}$ (Week 9)
WBC count	$-1.49 \times 10^9 \text{ l}^{-1}$	Week 7	$1.38 \times 10^9 \text{ l}^{-1}$ (Week 2)	$-2.57 \times 10^9 \text{ l}^{-1}$	Week 9	$2.15 \times 10^9 \text{ l}^{-1}$ (Week 15)
Platelet count	$-59.25 \times 10^9 \text{ l}^{-1}$	Week 7	$65 \times 10^9 \text{ l}^{-1}$ (Week 2)	$-69.88 \times 10^9 \text{ l}^{-1}$	Week 9	$77 \times 10^9 \text{ l}^{-1}$ (Week 17)
Haemoglobin Absolute value	-14.25 g l^{-1}	Week 7 or persistently decreased ^a	110 g l^{-1} (Week 5)	-18.5 g l^{-1}	Persistently decreased	93 g l^{-1} (Week 11)

^aWeek 7 (rHSA/IFN α 2a 600 μ g and 750 μ g) or persistently decreased (rHSA/IFN α 2a 900 μ g); WBC, white blood cell**Table 3**

Treatment-related adverse events in each treatment group (times)

Group	Grade 1	Grade 2	Grade 3	Grade 4/5	Total
rHSA/IFNα2a 600 μg	52 (68.42)	21 (27.63)	3 (3.95)	0 (0.00)	76
rHSA/IFNα2a 750 μg	64 (66.67)	24 (25.00)	8 (8.33)	0 (0.00)	96
rHSA/IFNα2a 900 μg	85 (77.27)	23 (20.91)	2 (1.82)	0 (0.00)	110
Peg-IFNα2a 180 μg	87 (69.05)	31 (24.60)	8 (6.35)	0 (0.00)	126

Table 4

Frequency of common adverse events in each treatment group (number of subjects)

Event, n (%)	rHSA/IFN α 2a 600 μ g (n = 8)	rHSA/IFN α 2a 750 μ g (n = 8)	rHSA/IFN α 2a 900 μ g (n = 8)	Peg-IFN α 2a 180 μ g (n = 8)
Albumin decreased	5	4	3	6
White blood cell count decreased	5	7	3	6
Haemoglobin decreased	1	3	2	5
Platelet count decreased	4	3	5	6
Neutrophil absolute value decreased	6	8	6	7
Vomit	0	0	2	0
Back pain	0	0	1	1
Fever	1	2	4	3
Fatigue	0	2	5	1
Ache all over	0	2	0	3
Headache	0	4	1	5
Dizziness	0	0	3	1
Knee-joint pain	0	1	1	0
Drowsiness	0	0	3	0

Table 5Detection of rHSA/IFN α 2a and Peg-IFN α 2a antibodies

Group	IFN alpha2a binding antibody	Recombinant human serum albumin binding antibody	Neutralizing antibody
rHSA/IFN α 2a 600 μ g	2	2	0
rHSA/IFN α 2a 750 μ g	1	1	0
rHSA/IFN α 2a 900 μ g	2	2	0
Peg-IFN α 2a 180 μ g	1	–	0

antibody. However, there was no observed effect on the efficacy of interferon *in vivo*.

Pharmacokinetic analysis of IFN α 2a

Serum IFN α 2a concentration–time profiles of the different treatment groups following the first and last dose of rHSA/IFN α 2a and PEG-IFN α 2a are shown in Figure 3. IFN α 2a PK parameters are shown in Table 6. The end-stage elimination of the IFN α 2a was estimated in accordance with the one compartment model, using mean serum IFN α 2a concentration–time profiles. Serum IFN α 2a concentration appeared to be maintained at a steady state after multiple doses, as extracted from the concentration–time profiles (Figure 4).

Following the single dose administration of rHSA/IFN α 2a on day 1, serum IFN α 2a concentrations were increased with increased doses, reaching a maximum level within 75–117 h. Consistent with the longer half-life of IFN α 2a (122.8–146.81 h), serum IFN α 2a concentrations in the rHSA/IFN α 2a groups were higher than in the PEG-IFN α 2a group at 168 h after dosing (the next administration time of PEG-IFN α 2a) (Figure 3 and Table 6).

Following the last dosing of rHSA/IFN α 2a, serum IFN α 2a concentrations increased with increased doses, reaching a maximum level within 56.57–80.57 h. The half-life of IFN α 2a (>100 h) was not changed after the last dose of rHSA/IFN α 2a. However, the half-life of PEG-IFN α 2a 180 μ g was extended to 100 h or longer. Serum IFN α 2a concentrations were similar among the rHSA/IFN α 2a 750 and 900 μ g, and PEG-IFN α 2a 180 μ g groups at 168 h following the last dose (the next administration time of PEG-IFN α 2a). Long half-life, together with the features of PK parameters and serum concentration–time profiles, are shown in Figures 3 and 4.

Analysis of the extent of fluctuation. The extent of fluctuation was similar among the rHSA/IFN α 2a 600, 750 and 900 μ g groups (1.30–1.70), which was higher than in the PEG-IFN α 2a group (0.559) (Table 6).

Analysis of the accumulation rate. The accumulation rate was similar among the rHSA/IFN α 2a 600, 750 and 900 μ g groups

(1.1–1.2), but was lower than in the PEG-IFN α 2a group (1.55) (Table 6).

Comparison of PK parameters between the first and last dose. IFN α 2a AUC and C_{\max} increased while CLz/F decreased after the last dose, compared to the first dosing in the PEG-IFN α 2a 180 μ g group ($P < 0.05$). IFN α 2a PK parameters did not change after the last dose in the rHSA/IFN α 2a 600 and 750 μ g groups, compared to the first dosing ($P < 0.05$). Furthermore, T_{\max} , $MRT_{0-\infty}$ and Vz/F of IFN α 2a decreased after the last dose in the rHSA/IFN α 2a 900 μ g group, compared to the first dosing ($P < 0.05$) (Table 6).

Comparison of PK parameters between the rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g groups. The $t_{1/2}$, AUC and $MRT_{0-\infty}$ were higher in the rHSA/IFN α 2a 750 μ g group than in the PEG-IFN α 2a 180 μ g group, and CLz/F was lower in the rHSA/IFN α 2a 750 μ g group than in the PEG-IFN α 2a group at first dosing ($P < 0.05$). The $t_{1/2}$, C_{\max} , AUC, MRT and Vz/F were similar between the rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g groups ($P < 0.05$). CLz/F of the last dose was lower in the rHSA/IFN α 2a 750 μ g group than in the PEG-IFN α 2a 180 μ g group ($P < 0.05$) (Table 7).

Analysis of linear correlations between exposure and rHSA/IFN α 2a dosage. Over the range of IFN α 2a exposures (C_{\max} and AUC) for each of the rHSA/IFN α 2a dose groups on day 1, when the slope (90% confidence interval) in the power model was less than 1, this indicated the saturation of absorption PKs at the first dose. In addition, when the slope in the power model was more than 1, this indicated the saturation of elimination PKs at the last dose (Table 8).

Kinetic analysis of neopterin

Serum neopterin concentration–time profiles in different treatment groups following the first and last dose of rHSA/IFN α 2a and PEG-IFN α 2a are shown in Figure 5. Neopterin kinetic parameters are shown in Table 9. The end-stage elimination of neopterin deviated from the mean serum neopterin concentration–time profiles (Figure 5).

Comparison of neopterin kinetic parameters between the first and last dose. Neopterin AUC and C_{\max} decreased and T_{\max} increased at the last dose in the rHSA/IFN α 2a group, compared to the first dosing, except for T_{\max} in the rHSA/IFN α 2a 600 μ g group. Neopterin C_{\max} decreased and AUC and T_{\max} increased at the last dose in the PEG-IFN α 2a group, compared to the first dosing. The P -values are listed in Table 9.

Comparison of neopterin kinetic parameters between the rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a groups. Neopterin AUC $_{0-t}$ of the first dose was higher in the rHSA/IFN α 2a 750 μ g group than in the PEG-IFN α 2a group ($P < 0.05$). Other PK parameters were similar between these two groups (Table 7).

Antiviral efficacy

HBV DNA level decreased after treatment, and mean changes in serum HBV DNA were -1.32 , -2.13 , -1.10 and -2.48 log $_{10}$

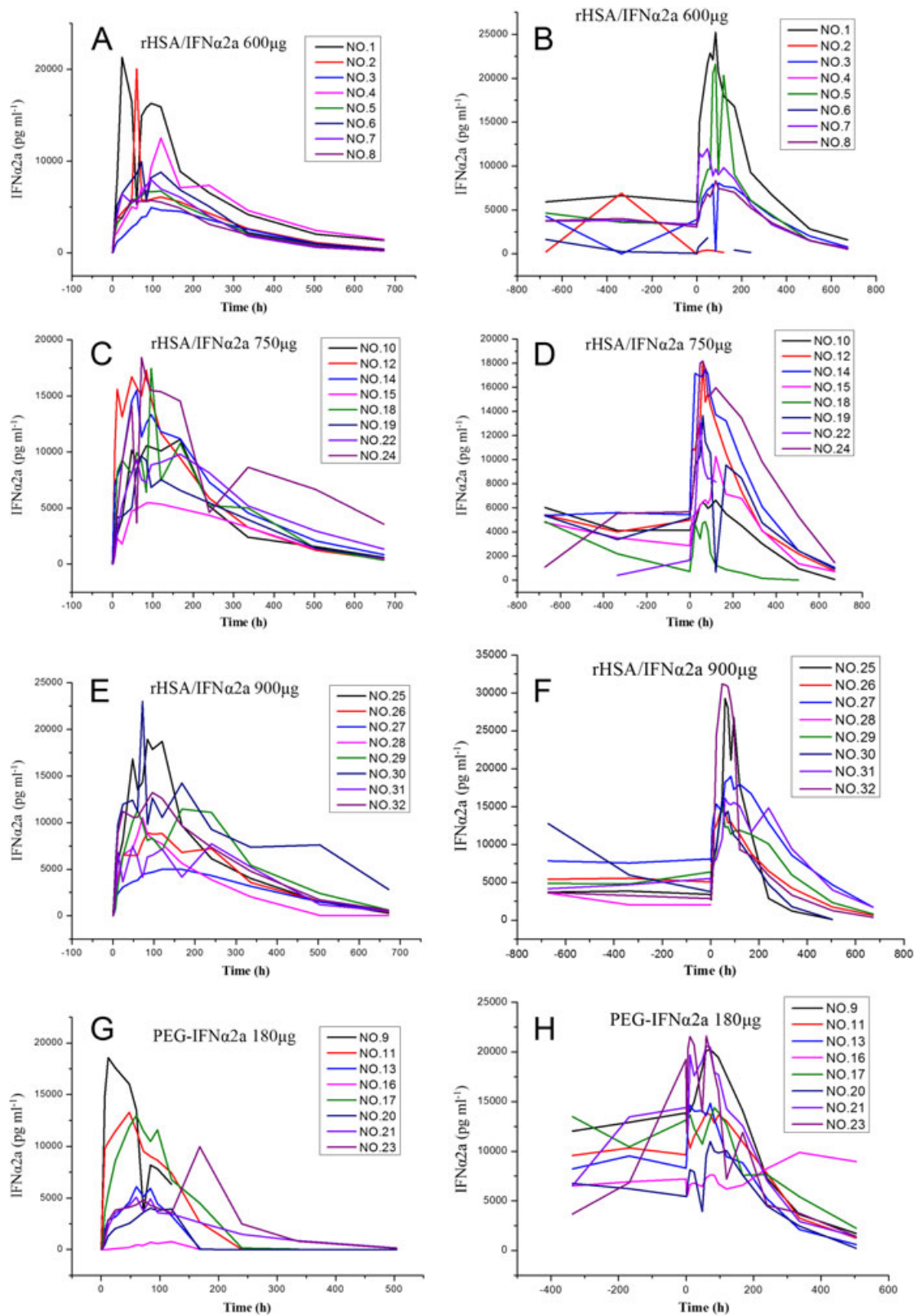


Figure 3

IFN α 2a plasma concentration–time profiles in the treatment groups, following the first and last dose of rHSA/IFN α 2a 600, 750 and 900 μ g and PEG-IFN α 2a 180 μ g. First dose of rHSA/IFN α 2a 600 μ g (A); last dose of rHSA/IFN α 2a 600 μ g (B); first dose of rHSA/IFN α 2a 750 μ g (C); last dose of rHSA/IFN α 2a 750 μ g (D); first dose of rHSA/IFN α 2a 900 μ g (E); last dose of rHSA/IFN α 2a 900 μ g (F); first dose of PEG-IFN α 2a 180 μ g (G); last dose of PEG-IFN α 2a 180 μ g (H)

Table 6
Comparison of pharmacokinetic parameters of IFN α 2a between the first and last dosing in each treatment group

Group	$t_{1/2}$ h	T_{max} h	C_{max} pg ml ⁻¹	AUC_{0-t} pg ml ⁻¹ h	$AUC_{0-\infty}$ pg ml ⁻¹ h		
rHSA/IFNα2a 600 μg	First dose	135.01 \pm 36.47	11 167.54 \pm 6341.74	2 381 878.01 \pm 879 806.14	2 518 504.60 \pm 1 022 285.89		
	Last dose	121.61 \pm 46.76	74.00 \pm 20.67	12 602.02 \pm 9248.22	3 094 334.86 \pm 1 948 097.91		
	<i>P</i>	0.92	0.72	0.35	0.35		
rHSA/IFNα2a 750 μg	First dose	146.81 \pm 53.16	102.00 \pm 42.55	13 087.84 \pm 4721.09	3 469 923.88 \pm 1 077 195.14		
	Last dose	114.02 \pm 38.72	80.57 \pm 27.46	12 723.33 \pm 5572.80	3 525 403.11 \pm 1 927 750.91		
	<i>P</i>	0.31	0.23	0.87	0.4		
rHSA/IFNα2a 900 μg	First dose	122.80 \pm 58.90	117.00 \pm 59.05	12 346.84 \pm 5962.98	3 360 387.88 \pm 1 277 834.78		
	Last dose	107.76 \pm 41.63	56.57 \pm 13.35	20 285.66 \pm 6933.92	4 540 507.10 \pm 1 288 646.44		
	<i>P</i>	0.87	0.02	0.06	0.18		
PEG-IFNα2a 180 μg	First dose	58.49 \pm 41.67	76.50 \pm 47.97	8844.80 \pm 5896.85	997 666.67 \pm 582 275.47		
	Last dose	112.40 \pm 29.14	69.00 \pm 8.49	15 510.89 \pm 4961.44	3 761 834.45 \pm 707 272.76		
	<i>P</i>	0.06	1	0.01	0.01		
Group	CLz/F L h ⁻¹	MRT _{0-\infty} h	Vz/F L	Df %	R	Cavg pg ml ⁻¹	
rHSA/IFNα2a 600 μg	First dose	0.35 \pm 0.10	255.44 \pm 55.19	64.22 \pm 14.26			
	Last dose	2.68 \pm 6.00	229.04 \pm 71.46	161.24 \pm 285.76	166.97 \pm 96.49	1.19 \pm 0.11	7331.42 \pm 4725.62
	<i>P</i>	0.35	0.92	0.35			
rHSA/IFNα2a 750 μg	First dose	0.31 \pm 0.10	294.83 \pm 94.74	63.26 \pm 21.08			
	Last dose	0.43 \pm 0.43	243.34 \pm 70.08	52.93 \pm 19.92	74.14 \pm 125.49	0.1 \pm 1.16	4156.16 \pm 6831.17
	<i>P</i>	0.74	0.31	0.31			
rHSA/IFNα2a 900 μg	First dose	0.39 \pm 0.14	279.55 \pm 85.73	65.53 \pm 30.69			
	Last dose	0.25 \pm 0.05	218.84 \pm 78.44	37.17 \pm 12.06	73.08 \pm 135.66	0.11 \pm 1.14	2267.22 \pm 10 869.59
	<i>P</i>	0.09	0.04	0.02			
PEG-IFNα2a 180 μg	First dose	2.35 \pm 3.92	130.80 \pm 67.30	93.75 \pm 98.69			
	Last dose	0.39 \pm 0.14	207.54 \pm 32.82	54.98 \pm 13.46	55.9 \pm 22.61	1.55 \pm 0.23	12 573.85 \pm 3706.2
	<i>P</i>	0.01	0.06	0.5			

Last dose: IFN α 2a serum concentration achieved a steady state after multiple doses, and pharmacokinetic parameters were C_{5max} , AUC_{50-t} , $AUC_{50-\infty}$, $t_{50 1/2}$, T_{50max} , CLz/F_{50} , $MRT_{50-\infty}$, and Vz/F_{50} respectively.

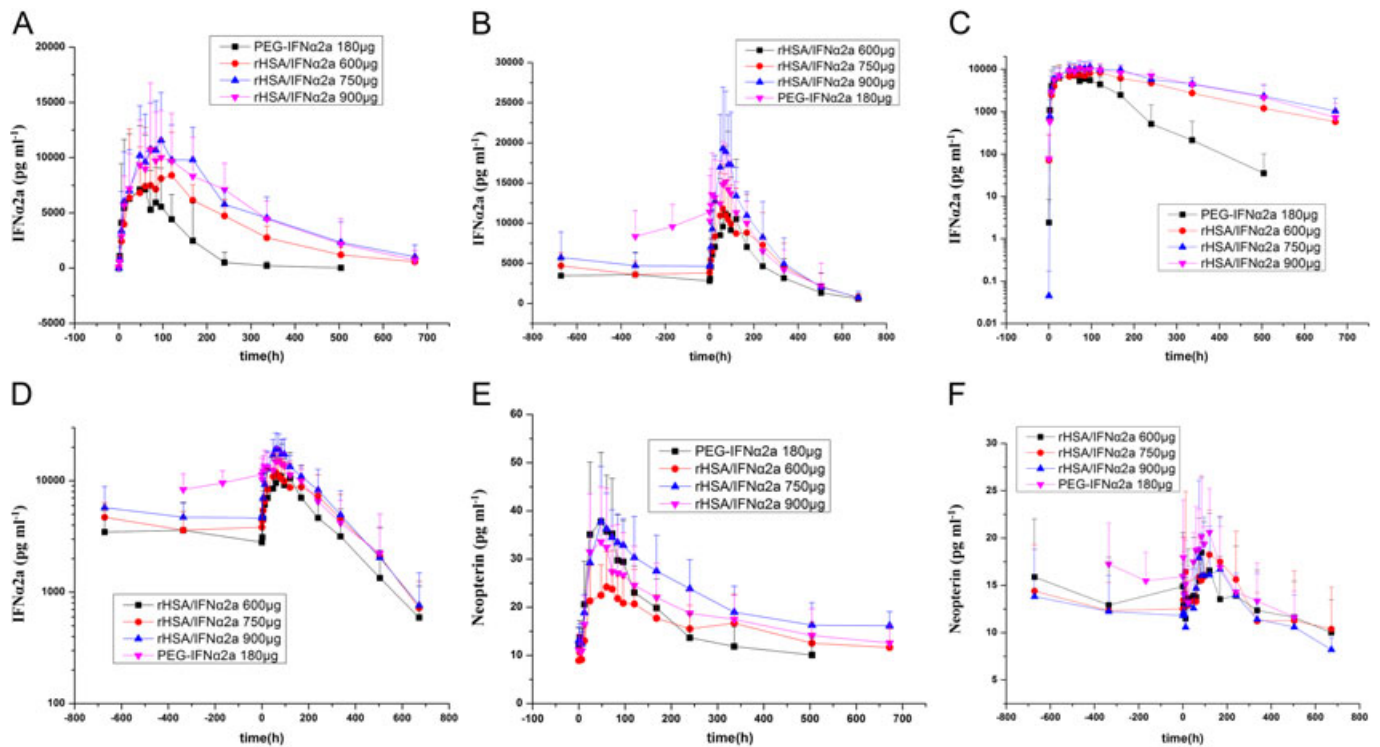


Figure 4

Mean (\pm SD) IFN α 2a or neopterin plasma concentration–time profiles in the treatment group, following the first and last dose of rHSA/IFN α 2a 600, 750 and 900 μ g, and PEG-IFN α 2a 180 μ g. Vertical error bars represent the standard deviation of the mean. IFN α 2a plasma line concentration–time profiles of the first dose (A); IFN α 2a plasma line concentration–time profiles of the last dose (B); IFN α 2a plasma log concentration–time profiles of the first dose (C); IFN α 2a plasma log concentration–time profiles of the last dose (D); neopterin plasma line concentration–time profiles of the first dose (E); neopterin plasma line concentration–time profiles of the last dose (F)

IU/ml for the 600, 750 and 900 μ g groups and the PEG-IFN α 2a group, respectively. The decreased level between the rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a groups were similar. At the end of treatment, there was no significant difference in reduced HBV DNA levels among the treated groups ($P = 0.35$). Almost all of the subjects in the rHSA/IFN α 2a 750 μ g group had HBV DNA that decreased for more than 2 \log_{10} compared to baseline after the first dosing, and in the PEG-IFN α 2a group after the fifth dosing (week 9) (Figure 6). There was no difference in the percentage with more than 2 \log_{10} reduction between groups ($P = 0.25$).

Serum HBeAg level decreased after treatment. HBeAg-negative conversion rates were 0% (0/6), 14.28% (1/7), 0% (0/6) and 0% (0/8), respectively; and HBeAg levels decreased by -488.8 ± 493.23 , -183.7 ± 292.35 , -118.2 ± 404.09 and -556.4 ± 434.87 IU ml^{-1} ($P = 0.10$) in the 600, 750 and 900 μ g groups and the PEG-IFN α 2a group, respectively. The HBeAg-negative conversion rate and normalization rate of ALT and AST were higher in the rHSA/IFN α 2a 750 μ g groups than in the PEG-IFN α 2a group (Figure 6). None of treated patients had HBsAg-negative conversion, HBeAg serum conversion, or HBsAg serum conversion.

ALT and AST levels decreased in all groups. ALT normalization rates were 14.28%, 50%, 37.5% and 37.5% ($P = 0.54$), while AST normalization rates were 14.28%, 50%, 25% and 25% ($P = 0.46$) in the 600, 750 and 900 μ g groups and in the PEG-IFN α 2a group, respectively.

Because this was a Phase Ib study, the sample size was small, resulting in insufficient power in statistics. However, the antiviral efficacy can be observed through the changed trend.

Analysis of linear correlations between the anti-viral effect and exposures of IFN α 2a and the production of neopterin in the rHSA/IFN α 2a group

First, decreased levels in serum HBV DNA and HBeAg from week 17 were determined and compared with baseline values; and AUC_{0-t} (IFN α 2a), C_{max} (IFN α 2a), AUC_{0-t} (neopterin) and C_{max} (neopterin) were computed after the last dose of rHSA/IFN α 2a. Then, a linear correlation analysis was carried out between the decreased levels of HBeAg and HBV DNA and exposures to IFN α 2a and neopterin at the last dose of rHSA/IFN α 2a. All correlation coefficients were negative. However, there were no significant differences, and the correlation was very weak, if at all (Table 10).

Discussion

In this Phase 1b study, we evaluated the safety and efficacy profiles of 600, 750 and 900 μ g of rHSA/IFN α 2a in the treatment of chronic hepatitis B patients for 17 weeks. We found

Table 7

Comparison of pharmacokinetic parameters between rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g

Group	$t_{1/2}$ h	T_{max} h	C_{max} pg ml ⁻¹	AUC_{0-t} pg ml ⁻¹ h	$AUC_{0-\infty}$ pg ml ⁻¹ h	P -value
First dose of IFNα2a						
rHSA/IFN α 2a 600 μ g	135.01 \pm 36.47	75.00 \pm 30.59	11 167.54 \pm 6341.74	2 381 878.01 \pm 879 806.14	2 518 504.60 \pm 1 022 285.89	
PEG-IFN α 2a 180 μ g	58.49 \pm 41.67	76.50 \pm 47.97	8844.80 \pm 5896.85	997 666.67 \pm 582 275.47	1 096 047.58 \pm 510 189.16	
P-value	<0.0001	0.16	0.23	<0.0001	<0.0001	
Last dose of IFNα2a						
rHSA/IFN α 2a 600 μ g	121.61 \pm 46.76	74.00 \pm 20.67	12 602.02 \pm 9248.22	3 094 334.86 \pm 1948 097.91	3 241 543.80 \pm 2 060 188.09	
PEG-IFN α 2a 180 μ g	112.40 \pm 29.14	69.00 \pm 8.49	15 510.89 \pm 4961.44	3 761 834.45 \pm 707 272.76	3 938 956.18 \pm 856 911.91	
P-value	0.96	0.9	0.78	0.28	0.78	
First dose of neopterin						
rHSA/IFN α 2a 600 μ g		78.00 \pm 45.36	29.24 \pm 7.37	10 607.02 \pm 2893.85		
PEG-IFN α 2a 180 μ g		51.00 \pm 22.90	40.70 \pm 14.29	8939.34 \pm 2368.70		
P-value		0.19	0.96	<0.0001		
Last dose of neopterin						
rHSA/IFN α 2a 600 μ g		68.33 \pm 54.59	19.71 \pm 6.71	8759.53 \pm 3026.83		
PEG-IFN α 2a 180 μ g		87.00 \pm 27.77	28.36 \pm 3.96	10 348.77 \pm 2107.19		
P-value		0.96	0.61	0.19		
Group	CLZ/F L h ⁻¹	MRT _{0-\infty} h	Vz/F L	R	Cavg pg ml ⁻¹	Df %
First dose of IFNα2a						
rHSA/IFN α 2a 600 μ g	0.35 \pm 0.10	255.44 \pm 55.19	64.22 \pm 14.26			
PEG-IFN α 2a 180 μ g	2.35 \pm 3.92	130.80 \pm 67.30	93.75 \pm 98.69			
P-value	<0.0001	0.01	0.87			
Last dose of IFNα2a						
rHSA/IFN α 2a 600 μ g	2.68 \pm 6.00	229.04 \pm 71.46	161.24 \pm 285.76	1.19 \pm 0.11	7331.42 \pm 4725.62	166.97 \pm 96.49
PEG-IFN α 2a 180 μ g	0.39 \pm 0.14	207.54 \pm 32.82	54.98 \pm 13.46	1.55 \pm 0.23	12 573.85 \pm 3706.2	55.9 \pm 22.61
P-value	<0.0001	0.34	0.07	0.71	0.71	0.07

Last dose: IFN α 2a serum concentration achieved a steady state after multiple doses, and the parameters were C_{ss0-t} , $AUC_{ss0-\infty}$, $t_{ss 1/2}$, T_{ssmax} , CLZ/F_{ss} , $MRT_{ss0-\infty}$, and Vz/F_{ss} , respectively.

Table 8Linear regression analysis between exposure of rHSA/IFN α 2a and its dosage

PK parameter	Dosing	Regression coefficient	Lower 90% CI	Upper 90% CI
C_{max}	First dose	0.35	-0.67	1.37
C_{max}	Last dose	2.33	0.27	4.38
AUC_{0-t}	First dose	0.86	0.13	1.59
AUC_{0-t}	Last dose	2.41	-0.23	5.06
$AUC_{0-\infty}$	First dose	0.85	0.04	1.66
$AUC_{0-\infty}$	Last dose	2.31	-0.28	4.91

Last dose: IFN α 2a serum concentration achieved a steady state after multiple doses, and pharmacokinetic parameters were C_{ssmax} , AUC_{ss0-t} and $AUC_{ss0-\infty}$, respectively. PK, pharmacokinetic.

that rHSA/IFN α 2a was better tolerated while delivering comparable efficacy with the 750 μ g dose in inhibiting HBV replication and normalizing ALT, compared to the 180 μ g PEG-IFN α 2a treatment.

Interferon treatment is often accompanied by adverse reactions that range from flu-like symptoms, bone marrow suppression and nervous system symptoms to gastrointestinal discomfort. The frequency and severity of these adverse reactions are related to the size and frequency of interferon dosage, as well as individual factors [10]. Fever and headache symptoms occurred more often and the reduction of blood cell count was more severe in the PEG-IFN α 2a 180 μ g group, compared with the rHSA/IFN α 2a 750 μ g group in this study. The adaptation time to IFN-related side effects was shorter in the rHSA/IFN α 2a groups than in the PEG-IFN α 2a group. Polyethylene glycol may have immunogenicity to incite immune response, while albumin is a component of human serum proteins. Different properties in carrier molecules may produce different extents of adverse events [11].

The half-life of rHSA/IFN α 2a determined in this study was approximately 140 h. A longer half-life was a result of the slow progress at the elimination phase and flat accumulation over time. The accumulation rate over time was approximately 1.1–1.2, which indicates that the elimination and accumulation of IFN were at an equilibrium state during rHSA/IFN α 2a treatment. Therefore, the rHSA/IFN α 2a was successfully constructed to improve PK profile and enable the reduction of dosing [12].

It is worth noting that the IFN α 2a level in the rHSA/IFN α 2a 750 μ g dose was equal to the PEG-IFN α 2a 180 μ g dose. Systemic exposures were similar between rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g doses after multiple dosing. These results support the conclusion that serum IFN α 2a concentration can be effectively maintained for a longer period through fusion with rHSA, which reduces dosing frequency from weekly to biweekly. Pegylation prolongs the IFN's half-life to approximately 40–80 h, and increases sustained virological response (SVR) rate among the treated chronic hepatitis C patients [13]. Since human serum albumin is a carrier protein with a half-life of 14–20 days, the fusion of IFN α 2a to albumin extends the half-life of the recombinant polypeptide to approximately 150 h, while it can maintain biological activity for 2–4 weeks, as tested in treated chronic hepatitis C patients [12, 14, 15].

There was no detectable neutralizing antibody to rHSA/IFN α 2a after rHSA/IFN α 2a treatment, which indicates that no new antigenic epitope was introduced in the fused rHSA/IFN α 2a, and the recombinant molecule would not generate immune response to compromise antiviral function [16].

Albumin-IFN alpha was originally developed to treat chronic hepatitis C [17]. In order to investigate clinical applications for treating chronic hepatitis B patients, the antiviral efficacy of rHSA/IFN α 2a was evaluated in this study. After seven doses of rHSA/IFN α 2a or 13 doses of PEG-IFN α 2a, similar patterns of reducing serum HBV DNA were observed between rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g treatments, which revealed that the efficacy of rHSA/IFN α 2a in suppressing HBV DNA replication was comparable to PEG-IFN α 2a.

Factors for predicting response to the antiviral treatment of chronic hepatitis B include high ALT level at baseline, HBV DNA < 10⁷ IU ml⁻¹, female and good treatment compliance [18–21]. A significant reduction in serum HBV DNA levels was detected in each of the rHSA/IFN α 2a treatment groups (Figure 6). At week 17, mean change in HBV DNA were approximately -1.32, -2.13 and -1.10 log₁₀ IU/ml in rHSA/IFN α 2a 600, 750 and 900 μ g, respectively. The potency of inhibiting HBV DNA replication was comparable between rHSA/IFN α 2a 750 μ g and PEG-IFN α 2a 180 μ g, and both resulted in a more than 2 log₁₀ reduction in HBV DNA level. HBeAg-negative conversion rate and ALT and AST normalization rates were higher in the rHSA/IFN α 2a 750 μ g group than in the PEG-IFN α 2a 180 μ g group, which may have resulted from the prolonged half-life of IFN α 2a that helped maintain effective drug concentration [22].

It was not clear why antiviral efficacy in the rHSA/IFN α 2a 900 μ g group was inferior to that in the rHSA/IFN α 2a 750 μ g group. Subjects in the rHSA/IFN α 2a 900 μ g group were older than subjects in the rHSA/IFN α 2a 750 μ g group, although other parameters were similar between these two groups, indicating that older age may be a factor for poor response in the rHSA/IFN α 2a 900 μ g group [23]. The saturation production of neopterin at the first and last dose indicated that rHSA/IFN α 2a 750 μ g was an effective dose, and was also better tolerated [19].

Since patient compliance to the treatment regimen is important in maximizing SVR rates, longer-acting rHSA/IFN α 2a

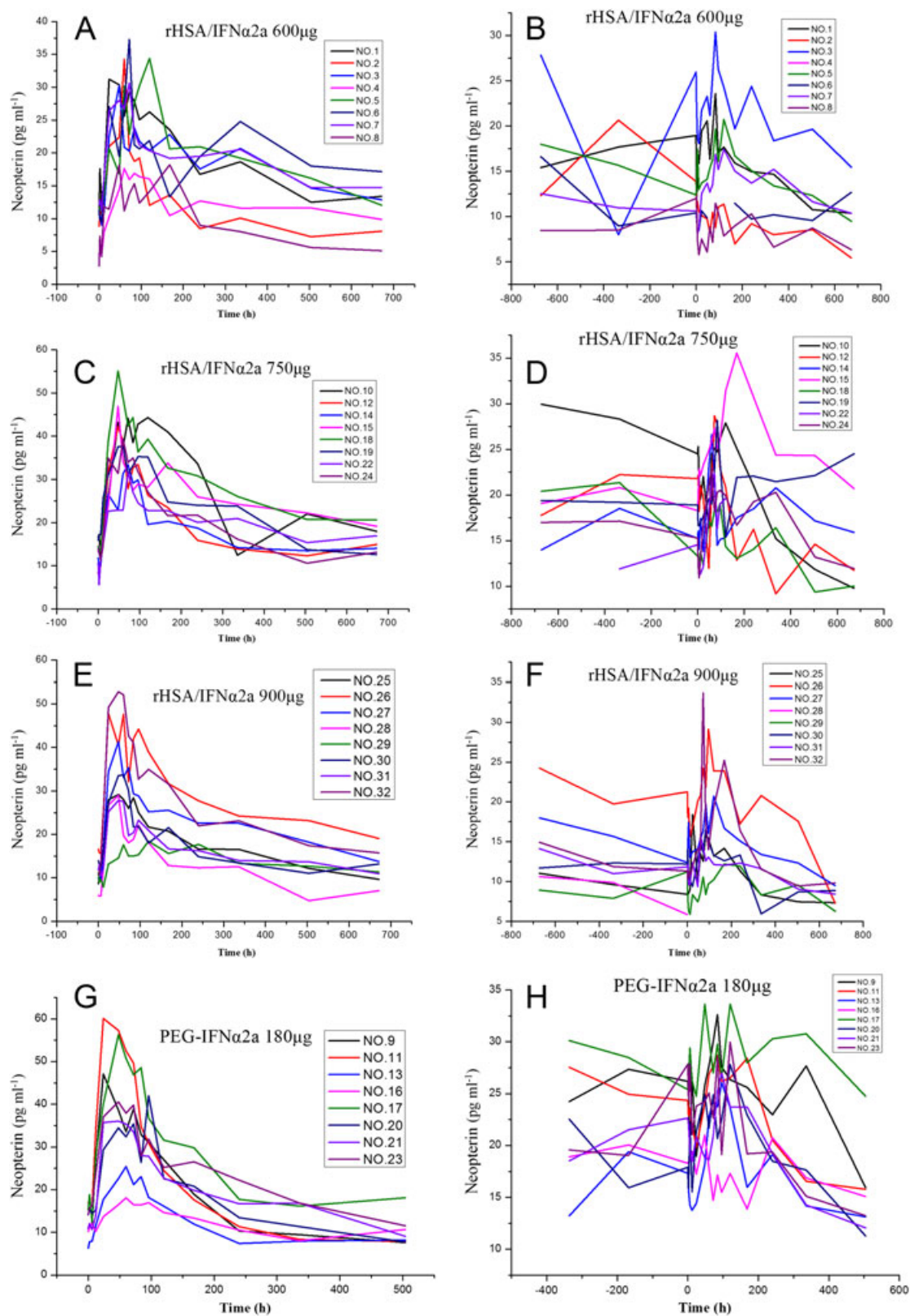


Figure 5

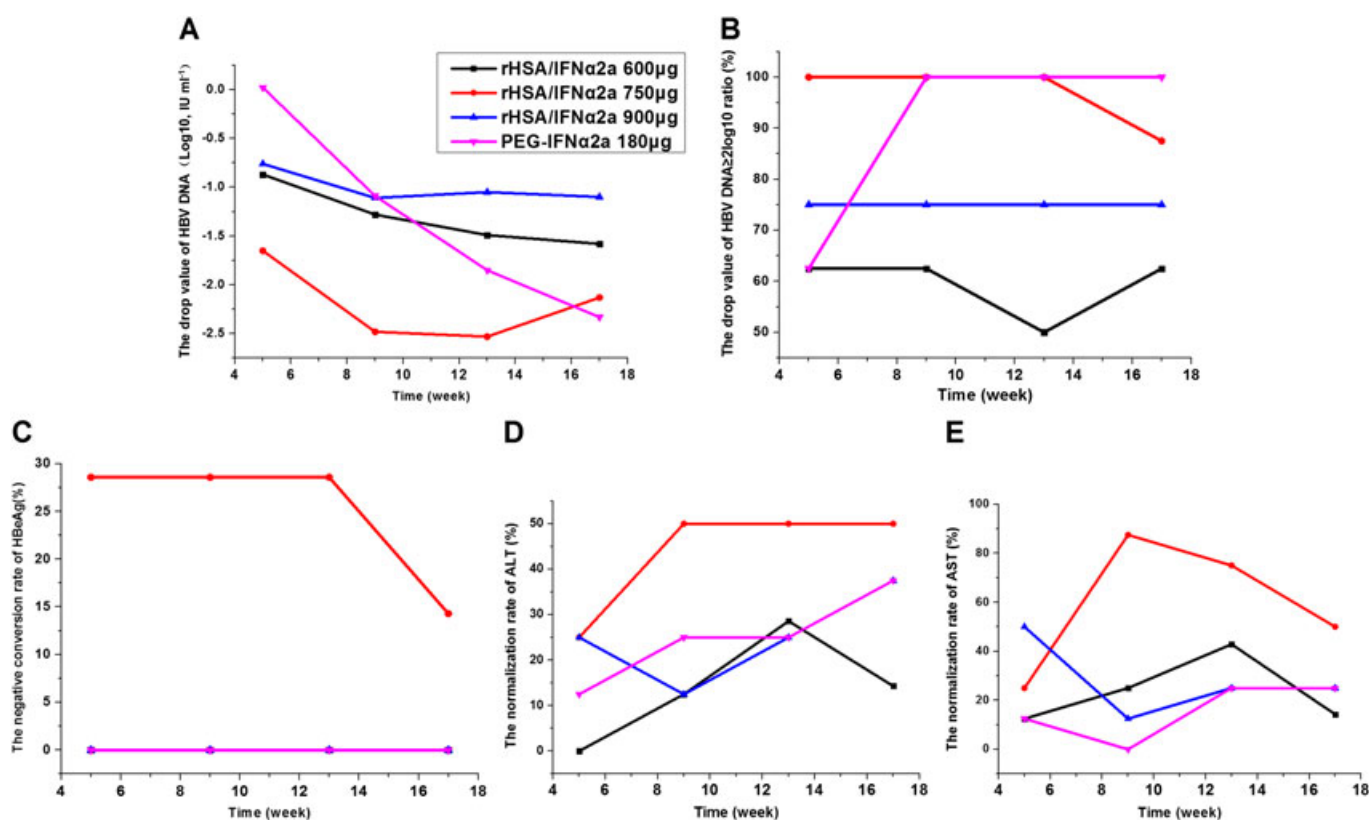
Neopterin plasma concentration–time profiles in treatment groups following the first and last doses of rHSA/IFN α 2a 600, 750 and 900 μ g, and PEG-IFN α 2a 180 μ g. First dose of rHSA/IFN α 2a 600 μ g (A); last dose of rHSA/IFN α 2a 600 μ g (B); first dose of rHSA/IFN α 2a 750 μ g (C); last dose of rHSA/IFN α 2a 750 μ g (D); first dose of rHSA/IFN α 2a 900 μ g (E); last dose of rHSA/IFN α 2a 900 μ g (F); first dose of PEG-IFN α 2a 180 μ g (G); last dose of PEG-IFN α 2a 180 μ g (H)

Table 9

Comparison of kinetics parameters of neopterin between the first and last dosing in each group

Group		T_{max} h	C_{max} Nmol l ⁻¹ ml ⁻¹	AUC_{0-t} Nmol l ⁻¹ ml ⁻¹ *h
rHSA/IFN α 2a 600 μ g	First dose	78.00 \pm 45.36	29.24 \pm 7.37	10 607.02 \pm 2893.85
	Last dose	68.33 \pm 54.59	19.71 \pm 6.71	8759.53 \pm 3026.83
	<i>P</i>	0.79	0.05	0.07
rHSA/IFN α 2a 750 μ g	First dose	66.00 \pm 24.00	41.32 \pm 7.59	14 677.07 \pm 2778.31
	Last dose	94.29 \pm 38.84	26.98 \pm 5.13	12 232.28 \pm 2969.15
	<i>P</i>	0.08	0.02	0.13
rHSA/IFN α 2a 900 μ g	First dose	55.50 \pm 27.91	35.00 \pm 11.45	12 464.27 \pm 3781.08
	Last dose	89.14 \pm 45.88	20.93 \pm 7.72	8268.78 \pm 2181.01
	<i>P</i>	0.06	0.02	0.02
PEG-IFN α 2a 180 μ g	First dose	51.00 \pm 22.90	40.70 \pm 14.29	8939.34 \pm 2368.70
	Last dose	87.00 \pm 27.77	28.36 \pm 3.96	10 348.77 \pm 2107.19
	<i>P</i>	0.03	0.04	0.07

Last dose: IFN α 2a serum concentration achieved a steady state after multiple doses, and pharmacokinetic parameters were C_{ssmax} , AUC_{ss0-t} , $AUC_{ss0-\infty}$, $t_{ss1/2}$, T_{ssmax} , CLz/F_{ss} , $MRT_{ss0-\infty}$, and Vz/F_{ss} , respectively.

**Figure 6**

Reduced levels of HBV markers at different time points after treatment in each group (all compared with the baseline values). HBV DNA (A); rate of HBV DNA reducing value $\geq 2 \log_{10}$ (B); HBeAg seroconversion (C); ALT normalization rate (D); AST normalization rate (E)

treatments can reduce dosing frequency, which also contributes to better tolerability [24–26]. Albumin can prevent the peak concentration of interferon by biweekly doses, but the

long-term steady-state of IFN level kept stimulating antiviral function, which decreases adverse effects and facilitates compliance. Unpegged IFN- α was administered three times a

Table 10Possible correlations between decreased levels of HBeAg and HBV DNA, and exposures of IFN α 2a and neopterin at last dosing of rHSA/IFN α 2a

Anti-viral factors	Parameter	AUC _{ss0-t} (IFN α 2a)	C _{ssmax} (IFN α 2a)	AUC _{ss0-t} (Neopterin)	C _{ssmax} (Neopterin)
Decreased levels of HBV DNA	r	-0.07716	0.02092	-0.27961	-0.19993
	P value	0.7465	0.9302	0.2325	0.398
	N	20	20	20	20
Decreased levels of HBeAg	r	-0.11988	-0.13495	-0.4296	-0.28997
	P value	0.6147	0.5705	0.0587	0.2149
	N	20	20	20	20

week, and serum IFN concentrations rapidly increased and decreased, which may affect the maintenance of constant antiviral efficacy.

The inconvenient administration that requires multi-injection and significant adverse effects limits the clinical application of interferon. Many patients select nucleos(t)ide analogues (NAs) as the first line of therapy because they are convenient to use and well-tolerated despite infinite treatment course. However, a finite duration of pegylated interferon is still an attractive alternative treatment because it provides higher rates of suppression of HBV replication off-therapy compared with NAs. In addition, the rates of HBeAg/HBsAg loss or seroconversion are increased over time among patients who respond to PEG-IFN therapy [27]. Here, we demonstrate that the rHSA/IFN α 2a treatment was well-tolerated and can be administered biweekly, only requiring 24 injections over a 48-week course. In addition, this new product will likely break the monopoly market of pegylated interferon and introduce the competition that will cut the price of the drug. Convenient administration, lower cost and a similar anti-HBV efficacy are the advantages of rHSA/IFN α 2a.

In conclusion, rHSA/IFN α 2a, a new long-acting IFN α 2a, was found to be better tolerated by chronic hepatitis B patients compared to PEG-IFN α 2a. The rHSA/IFN α 2a delivers a significantly longer half-life of IFN α 2a and increases IFN α 2a exposure compared to PEG-IFN α 2a. The antiviral efficacy of rHSA/IFN α 2a 750 μ g was similar to PEG-IFN α 2a 180 μ g. On the basis of these results, rHSA/IFN α 2a 750 μ g dose was selected for further evaluation in Phase II trials for treating chronic hepatitis B patients.

The study does have some limitations. The ratio of male to female patients was 21:11, so the majority of subjects were males. When the tolerability and pharmacokinetic characteristics of a drug are analysed, an ideal ratio of males to females is 1:1. However, the incidence of HBV infection is higher in the male population, which impacts the availability of female subjects.

Competing Interests

There are no competing interests to declare.

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