



Published in final edited form as:

*Exp Eye Res.* 2017 April ; 157: 34–37. doi:10.1016/j.exer.2016.12.005.

## Neuroprotection for Glaucoma: Requirements for Clinical Translation

Leonard A. Levin<sup>a,b,\*</sup>, Megan E. Crowe<sup>b</sup>, Harry A. Quigley<sup>c</sup>, and The Lasker/IRRF Initiative on Astrocytes and Glaucomatous Neurodegeneration Participants<sup>1</sup>

<sup>a</sup>Department of Ophthalmology, McGill University, Montreal, Quebec, Canada

<sup>b</sup>Department of Ophthalmology and Visual Sciences, University of Wisconsin School of Medicine and Public Health, Madison, Wisconsin, USA

<sup>c</sup>Wilmer Eye Institute, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

### Abstract

Within the field of glaucoma research, neuroprotection is defined as slowing the functional loss in glaucoma by a mechanism independent of lowering of intraocular pressure. There is currently a great potential for research surrounding neuroprotection as it relates to glaucoma. Anatomical targets for neuroprotection should focus on upstream rather than downstream factors, and could include any part of the retinal ganglion cell, the glia, especially astrocytes or Muller cells, and vasculature. The great number of anatomical targets is exceeded only by the number of possible biochemical pathways and potential treatments. Successful treatment may be accomplished through the targeting of one or even a combination of multiple pathways. Once a treatment is shown effective *in vitro*, it should be evaluated *in vivo* with carefully chosen animal models and studied in sufficient numbers to detect statistically and clinically significant effects. Such a drug should have few systemic side effects and its delivery should be optimized so as to encourage compliance. There are still a multitude of possible screens available to test the efficacy of a neuroprotective drug and a single gold standard is ideal for the accurate assessment and comparison of new drugs. Future studies in neuroprotection should investigate the genetic component of the disease, novel pharmaceutical agents for new or known pathways, modulations of scleral biomechanics, and relation to research of other complex disorders of the central nervous system.

---

Corresponding Author: Leonard A. Levin, MD, PhD, McGill Academic Eye Centre, 5252 de Maisonneuve West, Suite 400, Montreal, QC, H4A 3S5, Canada.

<sup>1</sup>This article summarizes the results of a targeted session on this topic at the March 2015 conference Astrocytes and Glaucomatous Neurodegeneration. This meeting was a follow-up to the 2010 meeting on the same topic, both of which were conducted as part of The Lasker/IRRF Initiative for Innovation in Vision Science. For more information about this conference, its participants and other review articles that originated from it see Tamm and Dowling, 2016. A list of the other participants of the targeted session is provided at the end of this article.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

## 1. Introduction

Although intraocular pressure (IOP) lowering has been definitively shown to decrease progressive visual loss in most patients with glaucoma (Garway-Heath et al., 2015; Heijl et al., 2002), there are some patients for whom IOP lowering is either insufficient, difficult to achieve, or associated with risks of adverse effects, e.g., in the case of some surgical therapies. This is the rationale for therapies that prevent visual loss in glaucoma through mechanisms other than IOP lowering, with the most studied being neuroprotection. A recent meeting of the Lasker Foundation and International Retina Research Foundation held a workshop on what is needed for neuroprotection to be translated to clinical use. Participants in this workshop were Leonard Levin, Harry Quigley, Megan Crowe, Francesca Cordeiro, Larry Donoso, Joyce Liao, Rick Libby, Richard Masland, Rob Nickells, Paul Sieving, and Al Sommer.

This paper summarizes the wide-ranging discussion from the workshop, and will focus on potential targets of neuroprotection, biochemical pathways that could lead to new drugs, translating animal models to human studies, and the design of human trials for a hypothetical neuroprotective drug. There are a multitude of biological targets for neuroprotection and a number of these biochemical pathways show promise (Almasieh et al., 2012; Danesh-Meyer, 2011; Nickells et al., 2012). The variety of potential targets and biochemical pathways are consistent with glaucoma being an extremely complex disease, and suggests treatment of more than one type or pathway. Once a drug is developed and shows promise *in vitro*, it may be advantageous to show efficacy in more than one animal model in order to increase the likelihood of translation to large human clinical trials (Ergorul and Levin, 2013).

## 2. A definition for neuroprotection

In terms of drug development, there are several issues in the definition of neuroprotection. The goal of glaucoma treatment is the preservation of quality of life by decreasing the loss of functional retinal ganglion cells (RGC) and their connectivity. The lowering of intraocular pressure (IOP) has been shown to slow RGC death (Cordeiro et al., 2004) and visual field damage, (Garway-Heath et al., 2015; Heijl et al., 2002) and the amount of visual field loss is closely associated with reported quality of life and functional measures of daily living (McKean-Cowdin et al., 2008). Thus, the term, neuroprotection, is now generally defined as an approach to slowing the functional loss in glaucoma by a mechanism that is independent of and in addition to IOP lowering (Levin and Peeples, 2008). The treatment should truly slow progression and not just temporarily improve function, i.e. not simply be neuroenhancement (Chang and Goldberg, 2012). Since IOP lowering is efficacious, it will continue to be used in most, if not all, glaucoma patients, along with neuroprotective therapy, unless subsequent research shows that a new neuroprotective treatment is actually superior to IOP lowering.

### 3. Potential anatomical targets

Currently, there are host of potential anatomical targets for neuroprotection in glaucoma. It is best to focus on where the damage is occurring rather than focusing on distal targets. There is considerable evidence that the initial events in glaucoma include axonal injury at the optic nerve head as a result of one or more injuries, including mechanical, vascular, biochemical, or others (Howell et al., 2007; Levin, 2001; Quigley et al., 1981; Quigley et al., 1979; Soto et al., 2008). Thus, the RGC is a primary target for neuroprotection, including its axon, cell body and dendritic tree. The RGC is contained within several milieus along its course: the retina, nerve head, optic nerve, chiasm, tract, and its central targets. Therefore, the damage process can involve interactions of RGC with glia and other neurons in the retina and targets, vascular cells in all 3 sites, and various stresses mediated by biomechanical forces at the sclera and nerve head connective tissues. Beneficial therapies may include those that improve survival mechanisms in RGCs, glia, especially astrocytes or Muller cells, and vasculature. Some studies have shown that astrocytes respond quickly to periods of increased IOP (Son et al., 2010), and it could be that these changes have both beneficial and detrimental downstream effects on RGCs (Chong and Martin, 2014; Johnson and Morrison, 2009). While secondary effects of glaucoma occur in anterograde centers, i.e., the lateral geniculate nucleus (LGN) and cortex (Gupta et al., 2009; Zhang et al., 2016), these are unlikely to have primary value as treatment targets (Danesh-Meyer and Levin, 2015). Thus, there are a variety of potential zones of interest for neuroprotective treatment. Ultimately, it may be that a drug is found for which the exact mechanism is not understood, so long as it prevents visual field deterioration and consequent decreased quality of life.

Studies that are needed include the development of rapid high-content screening techniques to identify drugs or agents targeting cells for neuroprotection. Examples include culture models of trabecular meshwork cells, RGCs, astrocytes of the optic nerve head, and scleral fibroblasts, using assays of cellular activities such as apoptosis, neurite extension, and morphological change.

### 4. Biochemical pathways and potential treatments

There are many potential biochemical pathways that could serve as targets for neuroprotection. A successful treatment regimen could target more than one of these pathways. One group of potential targets include neurotrophic factors such as NGF (Bai et al., 2010; Wang et al., 2014), BDNF (Mansour-Robaey et al., 1994; Martin et al., 2003), or CNTF (Ji et al., 2004) as pro-survival agents. Axon damage-induced apoptosis can be blocked with a variety of caspase inhibitors (Uchibayashi et al., 2011). Intracellular signaling of axonal damage can be interrupted with dual leucine zipper kinase (DLK) inhibitors (Huntwork-Rodriguez et al., 2013; Welsbie et al., 2013) or reduction in reactive oxygen species (Almasieh et al., 2011; Kanamori et al., 2010). Modulation of scleral rigidity via TGF $\beta$  signaling pathways can prevent mechanical effects on optic nerve head neurons or glia (Quigley et al., 2015). Gene therapy (Dahlmann-Noor et al., 2010; Johnson et al., 2010; Martin et al., 2003) can provide a mechanism to induce or repress any of the previously listed targets, while broader-acting interventions such as exercise can have wide-ranging effects (Roddy and ElleMBERG, 2012). Histone deacetylase inhibitors, for example, are

currently used for cancer treatment and are considered neuroprotective in CNS disease (Pelzel et al., 2010; Schmitt et al., 2015). Similarly, potential drugs which are already marketed for other uses include CoQ10 (which can serve as an electron carrier in the mitochondrial electron transport chain (Lee et al., 2014)), losartan (which is a selective angiotensin 1 receptor that can modulate scleral rigidity via the TGF $\beta$  pathway (Quigley et al., 2015)), and others. The vast number of biochemical pathways and potential treatments hint at the complexity of glaucoma as a disease and the potential need for multiple drugs in a treatment regimen, or alternatively, the tailoring of treatment to the dominant mechanism operative in each individual.

Techniques for rapid screening of drugs or other agents in animals should be developed, including the use of *in vivo* imaging or neuroimaging of RGCs, astrocytes, and the optic nerve. The development process would be helped by determination of how combining tests of efficacy in simpler optic nerve models could predict responses in the more challenging animal glaucoma models. Delineation of which glaucoma models are likely to be highly predictive of effects in human glaucoma would advance translatability to the clinic (Levin and Danesh-Meyer, 2010).

## 5. Translating animal models to humans

Once a promising drug(s) is (are) shown efficacious in *in vitro* studies, it should be tested in at least two animal models before going on to human trials. This would help in developing the dosing and delivery methods, as well as rule out species-specific effects that could make human trials unlikely to succeed. Currently, there are a number of different animal glaucoma models in use for neuroprotection, including mice, rats, beagles, pigs, and nonhuman primates. Each has advantages and disadvantages. For example, pig eyes are similar to the human, yet they are not substantially less expensive than nonhuman primates, although pigs are more available. Rodents provide a relatively inexpensive model, but are not as biologically similar to humans as nonhuman primates. Nonhuman primates are very expensive, but their eye is very similar to the human and they are critical for toxicity studies.

For clinical trials, there are important strategic choices that should be made in proposed studies before entering full-scale human phase 3 trials. An approach that would shorten the time to determine efficacy and allow human dose-response tests before a major phase 3 trial are futility designs looking for large effects (Quigley, 2012). These could narrow the search for a highly efficacious, low side-effect approach. There is rationale for proof-of-concept clinical trials of a future glaucoma drug in other optic neuropathies that might have shorter durations. Issues include obtaining baseline measurements, recruitment, and ability to randomize in a reasonable time frame. New approaches to shortening the time to see an effect on glaucomatous progression would aid in the development of neuroprotective drugs (Zhu et al., 2015; Zhu et al., 2014).

## 6. Dynamics of a hypothetical drug in human trials

As of yet, there is no gold-standard outcome measure for neuroprotection in a glaucoma clinical trial. All drugs approved by the US Food and Drug Administration (FDA) were

based on efficacy of IOP-lowering. This is irrelevant for neuroprotection, for which an objective measure of visual loss, either functional or structural, is needed. Stated opinions from FDA regarding outcome measures in glaucoma so far only include visual field testing (Weinreb and Kaufman, 2011). For recruitment to a study using visual field testing, the number of subjects can be minimized if the intrasubject variability from test-to-test is low and if the intersubject homogeneity of progression rate is high. A specific proposal for study recruitment is to identify persons likely to give reliable visual field outcomes from existing records of patients at large glaucoma centers, maximizing efficiency for a study in which visual fields are the primary outcome. Studies could use structural tests, such as optical coherence tomography, as the outcome if the selected population were at an earlier stage of glaucoma where this approach is more effective. Visual field tests have a wider assessment spectrum and would be better in that regard to allowing inclusion of a more general glaucoma population.

The proposed drug should not have serious side effects for a lifelong and slowly progressive disease such as glaucoma. Although local drug delivery such as periocular and intraocular injections decrease systemic side effects, these techniques have not been used on a large scale in glaucoma, and may impact recruitment. Injections may be the best mode of delivery as they maximize adherence. Presently, human trials of an encapsulated cell suspension producing ciliary neurotrophic factor are undergoing study (Chang and Goldberg, 2012). Phase 3 clinical trials are expensive, but the return on investment for a neuroprotective glaucoma drug is likely high. Funding for clinical trials will usually be sought from industry or government, but additional sources include private donors and voluntary consortia, such as the Glaucoma Research Network. The duration of a phase 3 trial could likely range from 18 months to three years.

## 7. Recommendations for future studies

There are several areas that should be targeted for future research to help achieve neuroprotection in glaucoma. First, further work understanding the genetics of glaucoma should help identify novel pathways or target genes that can be approached via overexpression or knockdown (Wilson and Di Polo, 2012). Second, the continued development of novel pharmacological agents for neuroprotection, or the adaptation of pre-existing drugs towards glaucoma neuroprotection, will be useful in targeting both new and known pathways of RGC degeneration and glial activation. Third, the modulation of scleral biomechanics has the potential to alter the ocular response to IOP elevation, and thereby interfere at an early level with RGC axonal injury (Nguyen et al., 2013). Finally, there are likely similarities among neuroprotection strategies that are being developed for other complex disorders of the CNS, e.g. Alzheimer disease, Parkinson disease, and amyotrophic lateral sclerosis (Dunkel et al., 2012), and that should be applied to glaucoma neuroprotection.

## Acknowledgments

This conference was supported by the Lasker Foundation and the International Retinal Research Foundation. LAL is supported in part by NIH R21EY025074, NIH P30EY016665, and the Canada Research Chair program. HAQ is supported in part by R01EY002120.

## References

- Almasieh M, Lieven CJ, Levin LA, Di Polo A. A cell-permeable phosphine-borane complex delays retinal ganglion cell death after axonal injury through activation of the pro-survival extracellular signal-regulated kinases 1/2 pathway. *J Neurochem.* 2011; 118:1075–1086. [PubMed: 21749374]
- Almasieh M, Wilson AM, Morquette B, Cueva Vargas JL, Di Polo A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res.* 2012; 31:152–181. [PubMed: 22155051]
- Bai Y, Dergham P, Nedev H, Xu J, Galan A, Rivera JC, ZhiHua S, Mehta HM, Woo SB, Sarunic MV, Neet KE, Saragovi HU. Chronic and acute models of retinal neurodegeneration TrkA activity are neuroprotective whereas p75NTR activity is neurotoxic through a paracrine mechanism. *J Biol Chem.* 2010; 285:39392–39400. [PubMed: 20943663]
- Chang EE, Goldberg JL. Glaucoma 2.0: neuroprotection, neuroregeneration, neuroenhancement. *Ophthalmology.* 2012; 119:979–986. [PubMed: 22349567]
- Chong RS, Martin KR. Glial cell interactions and glaucoma. *Curr Opin Ophthalmol.* 2014
- Cordeiro MF, Guo L, Luong V, Harding G, Wang W, Jones HE, Moss SE, Sillito AM, Fitzke FW. Real-time imaging of single nerve cell apoptosis in retinal neurodegeneration. *Proc Natl Acad Sci U S A.* 2004; 101:13352–13356. [PubMed: 15340151]
- Dahlmann-Noor A, Vijay S, Jayaram H, Limb A, Khaw PT. Current approaches and future prospects for stem cell rescue and regeneration of the retina and optic nerve. *Can J Ophthalmol.* 2010; 45:333–341. [PubMed: 20648090]
- Danesh-Meyer HV. Neuroprotection in glaucoma: recent and future directions. *Curr Opin Ophthalmol.* 2011; 22:78–86. [PubMed: 21252670]
- Danesh-Meyer HV, Levin LA. Glaucoma as a Neurodegenerative Disease. *Journal of Neuro-Ophthalmology.* 2015; 35:S22–S28. [PubMed: 26274833]
- Dunkel P, Chai CL, Sperlagh B, Huleatt PB, Matyus P. Clinical utility of neuroprotective agents in neurodegenerative diseases: current status of drug development for Alzheimer's, Parkinson's and Huntington's diseases, and amyotrophic lateral sclerosis. *Expert Opin Investig Drugs.* 2012; 21:1267–1308.
- Ergorul C, Levin LA. Solving the lost in translation problem: improving the effectiveness of translational research. *Curr Opin Pharmacol.* 2013; 13:108–114. [PubMed: 22980732]
- Garway-Heath DF, Crabb DP, Bunce C, Lascaratos G, Amalfitano F, Anand N, Azuara-Blanco A, Bourne RR, Broadway DC, Cunliffe IA, Diamond JP, Fraser SG, Ho TA, Martin KR, McNaught AI, Negi A, Patel K, Russell RA, Shah A, Spry PG, Suzuki K, White ET, Wormald RP, Xing W, Zeyen TG. Latanoprost for open-angle glaucoma (UKGTS): a randomised, multicentre, placebo-controlled trial. *Lancet.* 2015; 385:1295–1304. [PubMed: 25533656]
- Gupta N, Greenberg G, de Tilly LN, Gray B, Polemidiotis M, Yucel YH. Atrophy of the lateral geniculate nucleus in human glaucoma detected by magnetic resonance imaging. *Br J Ophthalmol.* 2009; 93:56–60. [PubMed: 18697810]
- Heijl A, Leske MC, Bengtsson B, Hyman L, Hussein M. Reduction of intraocular pressure and glaucoma progression: results from the Early Manifest Glaucoma Trial. *Arch Ophthalmol.* 2002; 120:1268–1279. [PubMed: 12365904]
- Howell GR, Libby RT, Jakobs TC, Smith RS, Phalan FC, Barter JW, Barbay JM, Marchant JK, Mahesh N, Porciatti V, Whitmore AV, Masland RH, John SW. Axons of retinal ganglion cells are insulted in the optic nerve early in DBA/2J glaucoma. *J Cell Biol.* 2007; 179:1523–1537. [PubMed: 18158332]
- Huntwork-Rodriguez S, Wang B, Watkins T, Ghosh AS, Pozniak CD, Bustos D, Newton K, Kirkpatrick DS, Lewcock JW. JNK-mediated phosphorylation of DLK suppresses its ubiquitination to promote neuronal apoptosis. *The Journal of cell biology.* 2013; 202:747–763. [PubMed: 23979718]
- Ji JZ, Elyaman W, Yip HK, Lee VW, Yick LW, Hugon J, So KF. CNTF promotes survival of retinal ganglion cells after induction of ocular hypertension in rats: the possible involvement of STAT3 pathway. *Eur J Neurosci.* 2004; 19:265–272. [PubMed: 14725620]
- Johnson EC, Morrison JC. Friend or foe? Resolving the impact of glial responses in glaucoma. *J Glaucoma.* 2009; 18:341–353. [PubMed: 19525723]

- Johnson TV, Bull ND, Hunt DP, Marina N, Tomarev SI, Martin KR. Neuroprotective effects of intravitreal mesenchymal stem cell transplantation in experimental glaucoma. *Invest Ophthalmol Vis Sci.* 2010; 51:2051–2059. [PubMed: 19933193]
- Kanamori A, Catrinescu MM, Mahammed A, Gross Z, Levin LA. Neuroprotection against superoxide anion radical by metallocorroles in cellular and murine models of optic neuropathy. *J Neurochem.* 2010; 114:488–498. [PubMed: 20456018]
- Lee D, Shim MS, Kim KY, Noh YH, Kim H, Kim SY, Weinreb RN, Ju WK. Coenzyme Q10 inhibits glutamate excitotoxicity and oxidative stress-mediated mitochondrial alteration in a mouse model of glaucoma. *Invest Ophthalmol Vis Sci.* 2014; 55:993–1005. [PubMed: 24458150]
- Levin LA. Relevance of the site of injury of glaucoma to neuroprotective strategies. *Surv Ophthalmol.* 2001; 45:S243–249. [PubMed: 11377443]
- Levin LA, Danesh-Meyer HV. Lost in translation: Bumps in the road between bench and bedside. *JAMA.* 2010; 303:1533–1534. [PubMed: 20407063]
- Levin LA, Peeples P. History of neuroprotection and rationale as a therapy for glaucoma. *Am J Manag Care.* 2008; 14:S11–14. [PubMed: 18284310]
- Mansour-Robaey S, Clarke DB, Wang YC, Bray GM, Aguayo AJ. Effects of ocular injury and administration of brain-derived neurotrophic factor on survival and regrowth of axotomized retinal ganglion cells. *Proc Natl Acad Sci USA.* 1994; 91:1632–1636. [PubMed: 8127857]
- Martin KR, Quigley HA, Zack DJ, Levkovitch-Verbin H, Kielczewski J, Valenta D, Baumrind L, Pease ME, Klein RL, Hauswirth WW. Gene therapy with brain-derived neurotrophic factor as a protection: retinal ganglion cells in a rat glaucoma model. *Invest Ophthalmol Vis Sci.* 2003; 44:4357–4365. [PubMed: 14507880]
- McKean-Cowdin R, Wang Y, Wu J, Azen SP, Varma R. Impact of visual field loss on health-related quality of life in glaucoma: the Los Angeles Latino Eye Study. *Ophthalmology.* 2008; 115:941–948. e941. [PubMed: 17997485]
- Nguyen C, Cone FE, Nguyen TD, Coudrillier B, Pease ME, Steinhart MR, Oglesby EN, Jefferys JL, Quigley HA. Studies of scleral biomechanical behavior related to susceptibility for retinal ganglion cell loss in experimental mouse glaucoma. *Invest Ophthalmol Vis Sci.* 2013; 54:1767–1780. [PubMed: 23404116]
- Nickells RW, Howell GR, Soto I, John SW. Under pressure: cellular and molecular responses during glaucoma, a common neurodegeneration with axonopathy. *Annu Rev Neurosci.* 2012; 35:153–179. [PubMed: 22524788]
- Pelzel HR, Schlamp CL, Nickells RW. Histone H4 deacetylation plays a critical role in early gene silencing during neuronal apoptosis. *BMC Neurosci.* 2010; 11:62. [PubMed: 20504333]
- Quigley HA. Clinical trials for glaucoma neuroprotection are not impossible. *Curr Opin Ophthalmol.* 2012; 23:144–154. [PubMed: 22249238]
- Quigley HA, Addicks EM, Green WR, Maumenee AE. Optic nerve damage in human glaucoma. II. The site of injury and susceptibility to damage. *Arch Ophthalmol.* 1981; 99:635–649. [PubMed: 6164357]
- Quigley HA, Guy J, Anderson DR. Blockade of rapid axonal transport. Effect of intraocular pressure elevation in primate optic nerve. *Arch Ophthalmol.* 1979; 97:525–531. [PubMed: 84662]
- Quigley HA, Pitha IF, Welsbie DS, Nguyen C, Steinhart MR, Nguyen TD, Pease ME, Oglesby EN, Berlinicke CA, Mitchell KL, Kim J, Jefferys JJ, Kimball EC. Losartan Treatment Protects Retinal Ganglion Cells and Alters Scleral Remodeling in Experimental Glaucoma. *PLoS One.* 2015; 10:e0141137. [PubMed: 26505191]
- Roddy G, Ellemberg D. Prevention of Glaucoma through Exercise: A meta-analysis. *Journal of Vision.* 2012; 12:483.
- Schmitt HM, Schlamp CL, Nickells RW. Role of HDACs in optic nerve damage-induced nuclear atrophy of retinal ganglion cells. *Neurosci Lett.* 2015
- Son JL, Soto I, Oglesby E, Lopez-Roca T, Pease ME, Quigley HA, Marsh-Armstrong N. Glaucomatous optic nerve injury involves early astrocyte reactivity and late oligodendrocyte loss. *Glia.* 2010; 58:780–789. [PubMed: 20091782]
- Soto I, Oglesby E, Buckingham BP, Son JL, Roberson ED, Steele MR, Inman DM, Vetter ML, Horner PJ, Marsh-Armstrong N. Retinal ganglion cells downregulate gene expression and lose their axons

- within the optic nerve head in a mouse glaucoma model. *J Neurosci*. 2008; 28:548–561. [PubMed: 18184797]
- Uchibayashi R, Tsuruma K, Inokuchi Y, Shimazawa M, Hara H. Involvement of Bid and caspase-2 in endoplasmic reticulum stress- and oxidative stress-induced retinal ganglion cell death. *J Neurosci Res*. 2011; 89:1783–1794. [PubMed: 21805492]
- Wang H, Wang R, Thrimawithana T, Little PJ, Xu J, Feng ZP, Zheng W. The nerve growth factor signaling and its potential as therapeutic target for glaucoma. *Biomed Res Int*. 2014; 2014:759473. [PubMed: 25250333]
- Weinreb RN, Kaufman PL. Glaucoma research community and FDA look to the future, II: NEI/FDA Glaucoma Clinical Trial Design and Endpoints Symposium: measures of structural change and visual function. *Investigative ophthalmology & visual science*. 2011; 52:7842. [PubMed: 21972262]
- Welsbie DS, Yang Z, Ge Y, Mitchell KL, Zhou X, Martin SE, Berlinicke CA, Hackler L Jr, Fuller J, Fu J, Cao LH, Han B, Auld D, Xue T, Hirai S, Germain L, Simard-Bisson C, Blouin R, Nguyen JV, Davis CH, Enke RA, Boye SL, Merbs SL, Marsh-Armstrong N, Hauswirth WW, Diantonio A, Nickells RW, Inglese J, Hanes J, Yau KW, Quigley HA, Zack DJ. Functional genomic screening identifies dual leucine zipper kinase as a key mediator of retinal ganglion cell death. *Proc Natl Acad Sci U S A*. 2013; 110:4045–4050. [PubMed: 23431148]
- Wilson AM, Di Polo A. Gene therapy for retinal ganglion cell neuroprotection in glaucoma. *Gene Ther*. 2012; 19:127–136. [PubMed: 21975466]
- Zhang P, Wen W, Sun X, He S. Selective reduction of fMRI responses to transient achromatic stimuli in the magnocellular layers of the LGN and the superficial layer of the SC of early glaucoma patients. *Hum Brain Mapp*. 2016; 37:558–569. [PubMed: 26526339]
- Zhu H, Crabb DP, Ho T, Garway-Heath DF. More Accurate Modeling of Visual Field Progression in Glaucoma: ANSWERS. *Invest Ophthalmol Vis Sci*. 2015; 56:6077–6083. [PubMed: 26393667]
- Zhu H, Russell RA, Saunders LJ, Ceccon S, Garway-Heath DF, Crabb DP. Detecting changes in retinal function: Analysis with Non-Stationary Weibull Error Regression and Spatial enhancement (ANSWERS). *PLoS One*. 2014; 9:e85654. [PubMed: 24465636]



- Neuroprotection in glaucoma slows functional loss independent of intraocular pressure
- Anatomical targets are retinal ganglion cells, glia, and vasculature
- Animal models must be able to detect statistically and clinically significant effects
- New clinical trial designs can detect significant effects more rapidly than before
- Future targets are genetics, novel drugs, sclera biomechanics, and other CNS diseases