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## Sensing Bad: Are Co-stimulatory CAR-Expressing $\gamma\delta$ T Cells Safer?

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Cellular immunotherapy using chimeric antigen receptor (CAR)-bearing T cells has shown promise in early clinical trials, especially for the treatment of CD19<sup>+</sup> B cell malignancies.<sup>1</sup> Despite impressive remission rates in this clinical setting, associated toxicity has been observed.<sup>2</sup> CAR T cells directed against solid tumor antigens have fared less well, with few durable clinical responses but still associated toxicity.<sup>3</sup> Toxicity results in part from the “on-target, off-tumor” effects of CAR-T cells, as the tumor-targeting ectodomain of the CAR molecule cannot discern target molecules expressed on tumors versus normal tissues. In this issue of *Molecular Therapy*, Fisher et al.<sup>4</sup> describe a unique approach using  $\gamma\delta$ -T cells as the platform for expression of a novel co-stimulatory

CAR, arguing for the potential of these uniquely engineered cells to mediate tumor-specific killing with far less off-tumor toxicity.

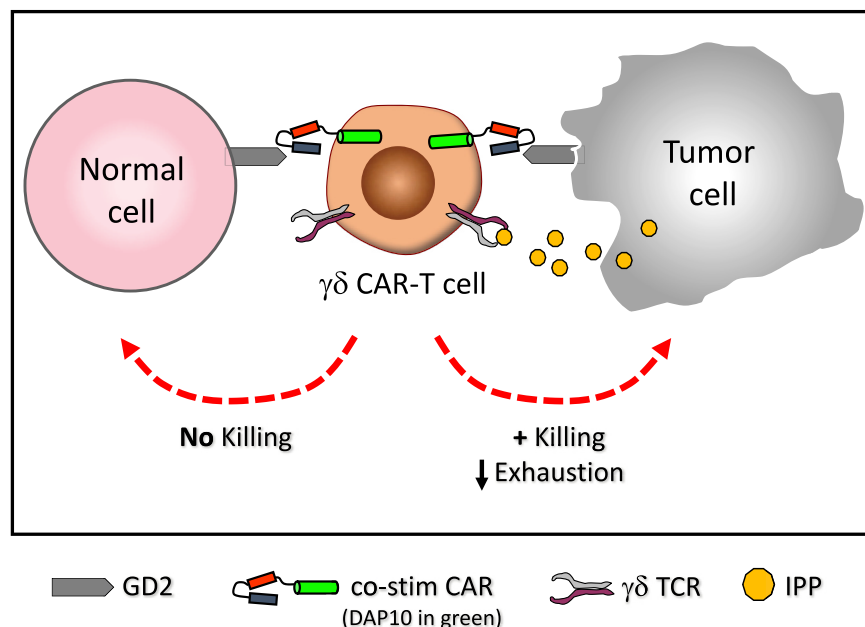
$\gamma\delta$ -T cells are a minor subset of peripheral lymphocytes in humans (<5%). Unlike  $\alpha\beta$ -T cells (the major circulating T cell subset, which are commonly utilized in creating CAR T cell immunotherapy products),  $\gamma\delta$ -T cells do not require major histocompatibility complex (MHC) class I or II molecules for recognizing antigens.<sup>5</sup>  $\gamma\delta$ -T cells respond to non-peptide phosphoantigens generated in the eukaryotic mevalonate metabolic pathway. The most abundant  $\gamma\delta$ -T cells express the V $\gamma$ 9V $\delta$ 2 T cell receptor (TCR) that recognizes isopentenyl pyro-

phosphate (IPP), which is overproduced in cancer cells.<sup>5</sup> The dysregulated mevalonate pathway in tumors leads to higher concentrations of IPP, which is sensed by  $\gamma\delta$ -TCR as a “danger signal.” Hence,  $\gamma\delta$ -T cells can discriminate tumor cells with dysregulated metabolism, which express these danger signals, from healthy cells, which do not.

$\gamma\delta$ -T cells are thought to help bridge the innate and adaptive immune systems, and, thus, functionally and phenotypically share components of both.<sup>6</sup> For example,  $\gamma\delta$ -T cells express numerous receptors typically found on innate immune effectors, such as natural killer (NK) cells, which play

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**Figure 1. “Co-stimulation Only” CAR- $\gamma\delta$  T Cell Activation after Recognition of Two Separate Tumor-Associated Molecules**

The cytotoxic capacity of CD3 $\zeta$  (signal 1) is mediated through the native  $\gamma\delta$ -TCR recognizing the tumor-associated danger signal, IPP, while co-stimulation (signal 2) is provided by a CAR recognizing the GD2 solid tumor antigen with an endodomain consisting of the innate NKG2D signaling molecule, DAP10. Normal healthy tissue does not express IPP and thus does not activate  $\gamma\delta$  T cells through their TCR.

crucial roles in anti-tumor responses. Natural killer group 2 member protein D (NKG2D) expressed on V $\gamma$ 9V $\delta$ 2 T cells binds to non-classical MHC molecules (MICA/B and the ULBPs) expressed on tumor cells. Ligand binding to NKG2D activates  $\gamma\delta$ -T cells via the intracellular signaling molecule, DAP10, with subsequent release of anti-tumor cytokines and enhanced cytotoxicity.

The unique tumor recognition function of  $\gamma\delta$ -T cells that provides broad reactivity to many different types of tumors, combined with recent success in their large-scale expansion for adoptive transfer into humans, has created a renewed interest to explore their anti-tumor therapeutic potential. The safety and efficacy of unmanipulated ex vivo-expanded  $\gamma\delta$ -T cells have been evaluated in several clinical trials.<sup>7</sup> Introduction of CAR expression on  $\gamma\delta$ -T cells has resulted in enhanced cytotoxicity in pre-clinical models,<sup>8</sup> but has not yet been evaluated in clinical trials.

Traditional CARs comprise an ectodomain that recognizes a tumor-associated antigen fused to endodomains that provide stimulatory signals. When expressed in  $\alpha\beta$ -T cells, CAR endodomains contain CD3 $\zeta$  (signal 1 for T cell activation), which provides cytotoxic capacity, in addition to one (2<sup>nd</sup> generation) or two (3<sup>rd</sup> generation) co-stimulatory endodomains, such as CD28 or 41BB (signal 2 for T cell activation), that foster expansion and viability.<sup>9</sup> Unfortunately, this all-in-one signaling moiety, which is completely independent of the native TCR, has led to unexpected toxicity, mostly owing to CAR-T cell activity in response to off-tumor antigen expression.<sup>3</sup> In order to limit this toxicity against normal tissue, Fisher et al.<sup>4</sup> devised a CAR design in which signals 1 and 2 for  $\gamma\delta$ -T cell activation are provided by separate receptors. The cytotoxic capacity of CD3 $\zeta$  (signal 1) is mediated through the native  $\gamma\delta$ -TCR recognizing the tumor-associated danger signal, IPP, while co-stimulation (signal 2) is provided by a CAR recognizing the GD2 solid tumor antigen with an endo-

domain containing the innate NKG2D signaling molecule, DAP10. In this schema, the CAR- $\gamma\delta$  T cell can be stimulated only after recognition of two separate and distinct tumor-associated molecules, whereas recognition of either alone would result in no activation (Figure 1). In this manner, CAR recognition of a tumor-associated antigen on normal (off-tumor) tissue would not result in unwanted toxicity.

The investigators compared the effects of two retroviral CAR constructs (a traditional 2<sup>nd</sup> generation CAR directed against GD2 and the  $\gamma\delta$  “co-stimulation only” CAR with anti-GD2 ectodomain and DAP10 endodomain) on  $\gamma\delta$ -T cell antitumor efficacy. Using in vitro cytokine secretion and cytotoxicity against human tumor lines as markers for  $\gamma\delta$ -T cell activation, the investigators showed that, while the traditional GD2 CAR-mediated activation of  $\gamma\delta$ -T cells in the presence of the CAR antigen alone, the “co-stimulation only” CAR-expressing  $\gamma\delta$ -T cells required both tumor-associated IPP and the CAR antigen for activation. This effect was not seen in  $\alpha\beta$ -T cells expressing the “co-stimulation only” CAR or against GD2<sup>-</sup> tumor targets, confirming the specificity of both signals in  $\gamma\delta$ -T cells. Interestingly, when the human GD2 tumor antigen was expressed in a murine tumor line that failed to engage the human  $\gamma\delta$ -TCR but would still be able to elicit a CAR response,  $\gamma\delta$ -T cells expressing the traditional CAR exhibited killing of tumor targets, whereas the “co-stimulation only” CAR-expressing  $\gamma\delta$ -T cells did not. The investigators argued that this was a method to mimic “healthy tissue” that expresses GD2 antigen to show the potential for decreased “off-tumor” toxicity. Finally, the authors compared the expression of exhaustion markers on  $\gamma\delta$ -T cells expressing the traditional CAR versus the “co-stimulation only” CAR. They showed that while “co-stimulation only” CAR expression induced upregulation of the exhaustion markers, PD-1 and TIM-3, they were far more pronounced when the traditional CAR was expressed. The authors thus provide a proof-of-principle of their concept that by using  $\gamma\delta$ -T cells expressing a “co-stimulation only” CAR, as opposed to  $\gamma\delta$ -T cells expressing a traditional CAR or  $\alpha\beta$ -T



cells expressing a “co-stimulation only” CAR, activation can occur only in the presence of two distinct tumor-associated signals.

The findings of the current study inspire a few key points that deserve additional consideration. First, the concept of “off-tumor” toxicity is dependent on normal tissue not expressing danger signals, such as IPP, which are capable of activating the native  $\gamma\delta$ -TCR. Although these danger signals are known to not be expressed in healthy adults, patients with a highly pro-inflammatory disease, such as cancer, may express these signals from “normal” tissues at various time points in the disease process. Indeed, other stress-induced danger signals, such as the NKG2D ligands MICA/B, are known to be upregulated in normal tissues in patients with cancer,<sup>10</sup> making off-tumor toxicity possible even when employing “co-stimulation only” CAR  $\gamma\delta$ -T cells. Although it has been difficult for the CAR-T cell field in general to develop animal models that mimic “off-tumor” toxicity, testing an approach such as this certainly would require either development of such a model or early implementation of phase 1 trials to test the concept in patients with cancer.

Second, as the authors themselves concede, the concept of a co-stimulatory CAR approach in which signals 1 and 2 of T cell activation are provided by separate receptors has been proposed and investigated previously in  $\alpha\beta$ -T cells. Lanitis et al.<sup>11</sup> developed a *trans* signaling CAR strategy where signal 1 (CD3 $\zeta$ ) was physically dissociated from co-stimulatory signal 2 (CD28) in two separate CARs of differing antigen specificity. These *trans*-signaling CAR-T cells showed weak cytokine secretion and cytotoxicity against target cells expressing only one antigen but showed enhanced activity against tumor cells expressing both antigens. To address the possibility that a CAR containing signal 1 could “overpower” the need for co-stimulation from the second CAR, depending on

the density and binding avidity of the tumor antigens, Kloss et al.<sup>12</sup> reported a combinatorial antigen recognition and balanced signaling approach in which the CAR containing signal 1 (CD3 $\zeta$ ) was constructed to be deliberately inefficient at killing and, only when combined with signal 2 (a separate CD28-containing CAR), would lead to optimal T cell activation. More recently, Roybal et al.<sup>13</sup> reported a combinatorially activated T cell circuit in which a synthetic Notch receptor for one tumor antigen induced the expression of a CAR for a second tumor antigen. These dual receptor “AND-gate” T cells were only armed/activated in the presence of dual antigen-expressing tumor targets. All these approaches, however, were employed in  $\alpha\beta$ -T cells in which the contributions of the native TCR were essentially ignored. The approach detailed in the current study by Fisher et al.<sup>4</sup> utilizes the endogenous signaling cascade of the native  $\gamma\delta$ -TCR, which potentially has the benefit of more robust and controlled signaling. Importantly, use of danger signals, as opposed to a tumor-associated antigen, allows the  $\gamma\delta$ -T cell approach to be utilized for myriad tumor types, some of which may not have more than one tumor-associated antigen target available. Further modeling both *in vitro* and *in vivo* is needed to clarify several points about universality and safety.

In an era when, for some specific cancer types, the clinical efficacy of CAR-T cell adoptive immunotherapy is being established, approaches to improve tumor specificity and avoidance of off-tumor toxicity are now needed. Certainly, the approach detailed in the current report by Fisher et al.<sup>4</sup> represents a unique method that offers the possibility of an alternative T cell product that can deliver dual targeted anti-tumor activity with minimal toxicity.

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