



Case Report

Novel recessive mutations in *COQ4* cause severe infantile cardiomyopathy and encephalopathy associated with CoQ₁₀ deficiency



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ABSTRACT

Coenzyme Q₁₀ (CoQ₁₀) or ubiquinone is one of the two electron carriers in the mitochondrial respiratory chain which has an essential role in the process of oxidative phosphorylation. Defects in CoQ₁₀ synthesis are usually associated with the impaired function of CoQ₁₀-dependent complexes I, II and III. The recessively transmitted CoQ₁₀ deficiency has been associated with a number of phenotypically and genetically heterogeneous groups of disorders manifesting at variable age of onset. The infantile, multisystemic presentation is usually caused by mutations in genes directly involved in CoQ₁₀ biosynthesis. To date, mutations in *COQ1* (*PDSS1* and *PDSS2*), *COQ2*, *COQ4*, *COQ6*, *COQ7*, *COQ8A/ADCK3*, *COQ8B/ADCK4*, and *COQ9* genes have been identified in patients with primary form of CoQ₁₀ deficiency. Here we report novel mutations in the *COQ4* gene, which were identified in an infant with profound mitochondrial disease presenting with perinatal seizures, hypertrophic cardiomyopathy and severe muscle CoQ₁₀ deficiency.

1. Introduction

Coenzyme Q₁₀, also known as ubiquinone, is a lipophilic component of all cellular membranes and has multiple metabolic functions. In the mitochondrial electron-transport chain, it transfers electrons from complex I (NADH dehydrogenase-ubiquinone oxidoreductase) and complex II (succinate dehydrogenase) to complex III (ubiquinone-cytochrome *c* reductase). It also receives electrons from de novo pyrimidine biosynthesis, fatty acid, amino acid, and sulfide oxidation [1]. In addition to its central role in the mitochondrial respiratory chain as an electron carrier, CoQ₁₀ also plays a role as an antioxidant in the cell membrane [2] and may modulate apoptosis [3]. Diseases associated with CoQ₁₀ deficiency are phenotypically heterogeneous and can manifest at any age. Five major clinical phenotypes have been described: (i) an encephalomyopathic form, characterized by mitochondrial myopathy and recurrent myoglobinuria and central nervous signs [4,5]; (ii) an ataxic form dominated by cerebellar ataxia and cerebellar atrophy [6,7]; (iii) an isolated myopathic form with lipid storage and respiratory dysfunction [8,9]; (iv) a multisystemic infantile form with encephalopathy and renal disease [10,11] and (v) nephropathy [12–14].

The precise biosynthesis pathways of CoQ₁₀ in mammals are yet to

be characterized. However, in yeast, and possibly in mammals, at least 12 mitochondrial enzymes are known to participate in CoQ₁₀ biosynthesis [15]. Mutations in *PDSS1* (MIM607429) [16], *PDSS2* (MIM610564) [10], *COQ2* (MIM609825) [16,17], *COQ4* (MIM 616227) [18,19], *COQ6* (MIM614647) [20], *COQ7* (MIM616733) [21], *COQ8A/ADCK3* (MIM 612016) [22], *COQ8B/ADCK4* (MIM615573) [12], and *CoQ9* (MIM614654) [23] genes have been described in patients with early-onset multisystemic mitochondrial encephalopathy, cardiomyopathy, and renal failure associated with severe CoQ₁₀ deficiency. To date, primary CoQ₁₀ deficiency due to mutations in *COQ4* (CoQ₁₀ D7, MIM616276) has been described in twelve patients from nine unrelated families. All affected patients but one harbored either homozygous or compound heterozygous mutations in the *COQ4* gene [18,19]. A single case of CoQ₁₀ deficiency was reported to be associated with haploinsufficiency of *COQ4* [24].

Here we report novel mutations in *COQ4* identified in an infant presenting with early onset cardiomyopathy, hypotonia, hearing loss, seizures, and lactic acidosis associated with severe muscle CoQ₁₀ deficiency.

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2. Case presentation

The infant male was born at term to a 24-year-old gravida 3 para 1 mother. The pregnancy was uncomplicated with normal serology and ultrasounds. Apgar scores were 1 at 1 min and 7 at 5 min and positive pressure ventilation was required at delivery. On the second day of life, the infant had an apneic event with bradycardia and reduction in oxygen saturation. A chest X-ray was performed which noted cardiomegaly. Biventricular hypertrophy was observed by echocardiogram. Lactate levels were persistently elevated at 4–6 mM (nl < 2.4 mM) during the first several weeks of life but subsequently normalized. Seizures were noted on the fifth day of life and the patient required intubation after the initiation of anti-epileptic medications. The patient was successfully extubated at 11 days of age. Seizure management was initially acceptable, but the patient eventually became refractory to multi-drug therapy (phenobarbital, topiramate, clobazam). Additional problems included gastroesophageal reflux requiring fundoplication, delayed visual maturation without structural abnormality of the eyes, bilateral hearing loss, profound hypotonia and absence of development. There was no evidence of renal dysfunction, transaminitis or liver synthetic dysfunction.

Brain MRI studies at one week of age showed focal regions of cortical increased T1 signal and magnetic resonance spectroscopy identified enlarged lactate peaks. Subsequent MRI at 10 weeks showed microcephaly with volume loss and increasing prominence of lactate peaks.

His initial hospitalization lasted for 4 months. He was discharged on palliative measures but was readmitted six days later with profound acidosis and respiratory failure. Ventilator support was withdrawn with the agreement of his parents and he expired.

At autopsy, there was marked bi-ventricular cardiac hypertrophy. EM studies of cardiomyocytes mitochondria showed swollen mitochondria with loss of cristae and some mitochondria with semi-circular arrangements of cristae (Fig. 1). Light microscopy and EM studies of mitochondria in the kidney and liver were unremarkable.

3. Biochemical studies

CoQ₁₀ concentration in muscle extract and in skin fibroblasts were measured by reversed-phase high-performance liquid chromatography and electrochemical detection system methodology as described previously [7]. The levels of CoQ₁₀ in autopsied skeletal muscle (6.9 µg/g tissue; normal range 19.6–46.8) and in skin fibroblasts (19.1 µg/mg protein; normal range 45.4–65.7) were found to be severely reduced. Due to the tissue availability and specimen quality, we were able to determine the activities of the electron transport complexes only in skin fibroblasts using the methods described [25]. As shown in the table,

consistent with the low CoQ₁₀ level found in fibroblasts, the combined activities of CoQ₁₀-dependent complexes i.e., rotenone-sensitive NADH cytochrome *c* reductase (complex I + III) and succinate-cytochrome *c* reductase (complex II + III) were also found to be significantly reduced, whereas the activities of other complexes were essentially comparable to those found in normal controls.

4. Molecular studies

High throughput next generation sequencing of the whole mitochondrial genome as well as 319 mitochondrial nuclear genes (Mito Nuclear Gene Panel, GeneDx® Co., Gaithersburg, MD) identified three previously unreported heterozygous variants in the *COQ4* gene: a 11 base pair deletion (c.23_33del11; p.Val8AlafsX19), and two missense mutations: c.331G > T; p.Asp111Tyr and c.356C > T; p.Pro119Leu. Parental testing showed that the frameshift mutation was maternally inherited and the two point mutations were paternally inherited as shown in the family pedigree (Fig. 2A). Sequencing of mitochondrial DNA showed several uncommon variants, but none were pathogenic.

The frameshift mutation (c.23_33del11) is not present in the Exome Aggregation Consortium (ExAC) database (Cambridge, Massachusetts, USA; URL: <http://exac.broadinstitute.org>), dbSNP, or ClinVar. It results in truncation of the COQ4 protein at residue 26 and the mRNA would be subject to nonsense mediated decay. The c.311G > T mutation is present at a frequency of 1/121200 alleles in ExAC, and is also present in dbSNP (rs530213004), but has not been observed in ClinVar. In silico analysis predicts that the resulting coding mutation (p.Asp111Tyr) would be deleterious (Provean, – 7.40) and damaging (SIFT, 0.001) to protein function. The c.356C > T mutation is present in the ExAC database at a frequency of 1/121238 alleles, but is not observed in dbSNP or ClinVar. In silico analysis likewise predicts that the resulting p.Pro119Leu missense mutation would have a deleterious (Provean, – 9.32) and damaging (SIFT, 0.002) effect on protein function.

To confirm the expected effects upon the RNA and protein, we performed quantitative real-time PCR analysis (qPCR) for the COQ4 transcript and Western blotting. Western blotting shows a significant loss in the level of the COQ4 protein, consistent with a loss in immunoreactive protein encoded by the frameshift allele (Fig. 3A, B). qPCR showed a reduced level of the *COQ4* transcript, which would be anticipated due to nonsense-mediated decay of the frameshift transcript (Fig. 3C).

5. Discussion

In the inner membrane of human mitochondria, CoQ₁₀ functions primarily as a mobile electron carrier which is necessary for ATP production through the oxidative phosphorylation system. In addition,

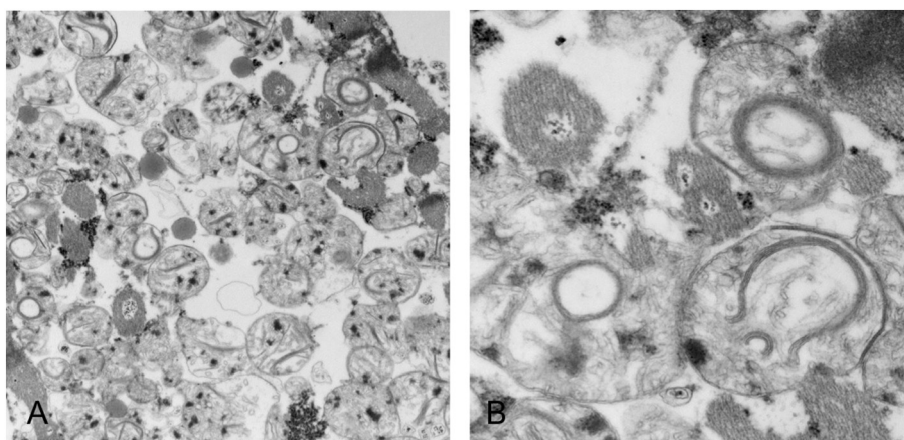


Fig. 1. Electron micrographs of cardiac muscle show strikingly abnormal mitochondrial morphology, with unusual semi-circular arrangements of crista noted in multiple myofiber mitochondria, which are non-artefactual in nature. A & B, transmission electron microscopy, A × 17,200; B × 57,500.

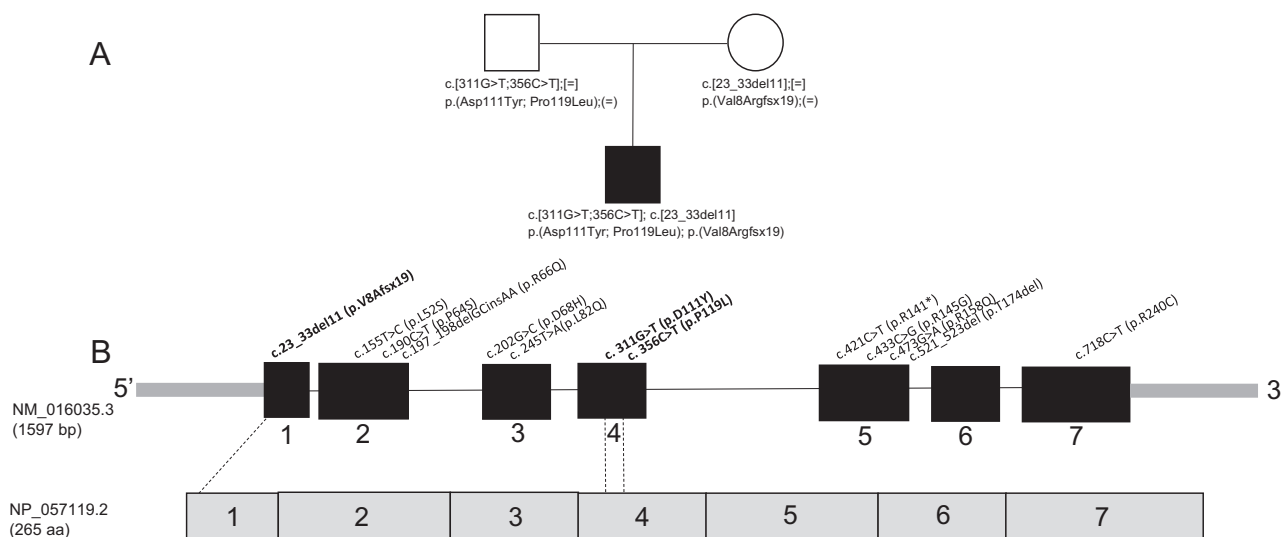


Fig. 2. Family pedigree and location of CoQ4 mutations. **A.** Family pedigree showing inheritance of mutations identified in this study. **B.** Location of mutations on CoQ4 gene (novel mutations described here are shown in bold).

CoQ₁₀ also participates in other important cellular functions such as pyrimidine biosynthesis, modulating apoptosis, and activation of mitochondrial uncoupling proteins [15]. Thus, because of its multiple cellular functions, CoQ₁₀ deficiency would be expected to cause the impairment of several different biochemical pathways and produce different clinical diseases [3]. Given that CoQ₁₀ is synthesized endogenously and dietary contribution to the cellular content is negligible, the reduction of CoQ₁₀ observed in CoQ₁₀ defects directly results from defects in the biosynthesis pathway. To date, mutations in nine genes participating in CoQ₁₀ biosynthesis have been associated with primary CoQ₁₀ deficiency. It is intriguing that although all of these gene products function in the same pathway to synthesize CoQ₁₀, their clinical presentations are quite distinct. For example, mutations in *PDS2*, encoding for one of the two subunits of COQ1 (the first committed enzyme in the CoQ₁₀ biosynthetic pathway) have been reported in an infant with severe Leigh's syndrome, nephrotic syndrome and CoQ₁₀ deficiency [10], whereas autosomal recessive mutations in *COQ2*, encoding for the *para*-hydroxybenzoate-polyprenyl transferase (the second committed enzyme in the pathway) was reported in patients with variable clinical presentation of the multisystem infantile form of CoQ₁₀ deficiency or isolated nephropathy [13,14,26–28]. It is

tempting to speculate that this clinical heterogeneity may reflect specific problem associated with defects at different steps in the CoQ₁₀ biosynthetic pathway. However, the fact that different molecular defects in the same gene (i.e. *COQ2*) cause extremely variable clinical manifestations suggests that the impact may be only a product of the overall loss of CoQ₁₀ synthesis. A correlation between the CoQ₁₀ content of yeast expressing diverse CoQ2 mutations and phenotypes in *S. cerevisiae* has recently been reported [29].

Although the complex biosynthesis of CoQ₁₀ in yeast has been extensively studied, in humans, its synthetic pathway is less well characterized. In particular, the exact function of the COQ4 protein in this pathway still remains unknown. However, based on studies in yeast and human cells, it may function as the core component in the multisubunit complex required for CoQ₁₀ biosynthesis [30,31]. To date, eleven recessive mutations in *COQ4* have been reported in patients presenting neonatal encephalopathy and severe CoQ₁₀ deficiency (Fig. 2B). Chung et al. [19] reported five recessive missense *COQ4* mutations (Arg66Glu, Asp68His, Leu82Gln, Arg158Gln, Arg240Cys) in five patients with severe, early-onset mitochondrial disease presenting encephalopathy and/or cardio-myopathy and lactic acidosis. In this study, biochemical (low muscle CoQ₁₀ content and

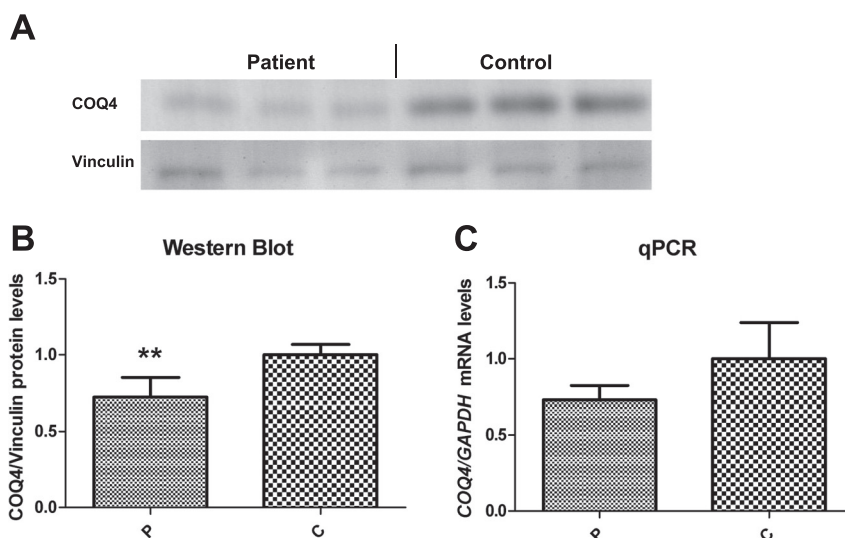


Fig. 3. **A.** Representative Western blot showing the level of CoQ4 protein in skin fibroblasts from patient (left) and control (right). **B.** Relative level of protein normalized to vinculin. **C.** Relative level of *CoQ4* transcript normalized to *GAPDH*. Error bars represent standard deviations of six experiments. Mann-Whitney *U* test. **Indicates a value of *p* > 0.01.

Table 1

Citrate synthase-normalized activities of mitochondrial electron transport chain enzyme (ETC) in the patient's skin fibroblasts.

MRC enzyme	Patient	Control (range)
Complex I/CS	0.63	0.86–1.70
Complex II/CS	1.86	1.53–1.87
Complex I + III/CS	0.11	0.55–1.30
Complex II + III/CS	0.08	0.20–0.79
Complex IV/CS	0.54	0.37–1.74
Complex V/CS	8.02	5.07–11.35
Citrate synthase (nmol/min/mg protein)	48.15	49.60–56.18

impaired respiratory chain enzyme activities) data were provided for only one patient who harbored compound heterozygous variants of Leu82Gln/Arg158Gln. Therefore, although the authors provide strong molecular evidence in support of the pathogenicity of these variants, further biochemical and functional data are required to ascertain, with high certainty, the status of Arg66Glu, Asp68His, and Arg240Cys variants (Table 1).

In the study reported by Brea-Calvo et al. [18], six pathogenic, autosomal recessive, variants were reported in five patients with early-onset mitochondrial disease and CoQ₁₀ deficiency (Fig. 2B). The authors validated the pathogenicity of these variants by conducting extensive functional studies in a recombinant yeast model. Although the clinical presentations of these patients were heterogeneous, all patients had early-onset of the disease and four out of five had early fatal outcome. The only surviving member of this cohort in the study who reached adulthood is an 18-year-old male proband in who harbored a homozygous missense (c.190C > T; p. Pro64Ser) variant in exon 2 of *COQ4* gene. Interestingly, although this variant was accompanied by a profound reduction in ETC complex I, a 70% reduction in complex II + III, and a 50% reduction in complex III, there was minimal effect on the level of CoQ₁₀ in the muscle when compared to that found in controls [18]. Therefore, from these data it becomes tempting to hypothesize that perhaps, at least in human, the c.190C > T transition in the second exon of *CoQ4* gene may have minimal functional impact on the CoQ4 protein and consequently on the CoQ₁₀ biosynthesis. The underlying cause of the profound reduction in the complex I activity detected in this patient in the cohort, was not explained. However, it is noteworthy that deficiency of complex I activity was found also in fibroblasts carrying *PDSS2* mutations [10]. Moreover, analysis by blue native gel electrophoresis in cerebellum of Coq9^{x/x} mice showed reduced level of CI, supporting the hypothesis that CoQ₁₀ deficiency causes instability of super complex I + III [32]. In Chung et al. [19], five out of five patients had hypotonia and five out of six patients presented with cardiomyopathy, yet the prevalence of hypotonia and cardiomyopathy in the cohort reported by Brea-Calvo et al. [18] was one in five and two in five respectively. Nevertheless, amongst all reported patients (including ours) harboring *COQ4* mutations, cardiomyopathy is the most prominent features in 8/12 patients.

Cardiomyopathy is a frequent manifestation of the multisystemic form of CoQ₁₀ deficiency, and has been described in association with mutations in *PDSS1* [16], *PDSS2* [33], *COQ2* [28], and *COQ9* [11,34]. Intriguingly, whereas nephropathy is a prevalent manifestation in patients harboring mutations in *COQ1* (*PDSS2* subunit) [10], *COQ2* [17], *COQ6* [20], and *COQ9* [23] genes, to date, none of the mutations of *COQ4* gene identified in patients with CoQ₁₀ deficiency were associated with renal dysfunction. This could be due to the different level of *COQ4* expression in different tissues.

6. Conclusion

In this study, we report three novel variants in the *COQ4* gene that are associated with a severe early-onset mitochondrial disease and profound CoQ₁₀ deficiency. Identification of disease-causing mutations

in genes involved in CoQ₁₀ biosynthesis is important as it may lead not only into a new insight in the pathogenesis of CoQ₁₀ deficiency, but also to early diagnosis and treatment of these disorders.

Declaration of conflict of interest

None.

Ethical approval

This retrospective case report did not require ethics committee approval at our institution.

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