

Tamoxifen for induction of Cre-recombination may confound fibrosis studies in female mice

Lucas L. Falke¹ · Roel Broekhuizen¹ · Alwin Huitema² · Erik Maarseveen³ · Tri Q. Nguyen¹ · Roel Goldschmeding¹

Received: 6 March 2017 / Accepted: 31 March 2017 / Published online: 11 May 2017
© The International CCN Society 2017

Abstract A variety of conditional knock-out mice relying on Tamoxifen-driven ERT2/Cre -mediated recombination are available and have been used to study involvement of specific genes in kidney disease. However, recent data suggest that Tamoxifen itself might attenuate fibrosis when administered during experimental models of kidney disease. It has remained unclear whether this still applies also if kidney damage is initiated after a wash-out period has been implemented. Here we report that the commonly applied regimen of administration of 4 alternate day doses of 1mg Tamoxifen per mouse until 14 days prior to start of the actual experiment, in this case the induction of obstructive nephropathy by Unilateral Ureteral Obstruction (UUO), still attenuated fibrosis in female obstructed mouse kidneys, whereas this effect was not seen in male obstructed kidneys. Attenuation of fibrosis was accompanied by a reduction in nuclear ER α positivity despite absence of detectable levels of the active tamoxifen metabolite endoxifen throughout the UUO experiment. In conclusion, these results indicate that the

Tamoxifen dosing regimen commonly applied in conditional gene targeting experiments might have prolonged confounding effects in female mice through attenuation of renal fibrosis independent of modulation of the expression of the targeted gene(s).

Keywords Tamoxifen · UUO · Fibrosis · Gender · ER α

Introduction

Tamoxifen is widely used for the induction of genomic recombination in mice (double-)transgenic for floxed genes and Tamoxifen specific estrogen receptors (ER) coupled to Cre-recombinase (supplemental Table 1) (Hayashi and McMahon 2002). Tamoxifen is both an antagonist and agonist of ER signaling, depending on tissue type. In kidneys of both female and male mice, ER α and β are readily detectable (Irsik et al. 2013). The kidney is highly responsive to estrogen in an ER α dependent manner and as such, it is regarded as the most estrogen-sensitive non-reproductive organ (Jelinsky et al. 2003). Of note, treatment with relatively high doses (10 mg/day) of Tamoxifen during experimental obstructive nephropathy, malignant hypertension, or diabetic nephropathy exerted an anti-fibrotic effect (Cohen and Rosenmann 1985; Dellé et al. 2012; Mao et al. 2014), in association with ER α dependent modulation of TGF β signaling (Kim et al. 2014). However, it is unclear whether also the common study designs involving pretreatment with *much lower* Tamoxifen doses for genomic recombination *prior to* the initiation of experimental kidney disease might have confounding protective effects. Therefore, we compared the development of fibrosis in obstructed kidneys of male and female mice undergoing unilateral ureteral obstruction (UUO) after a 14 day wash out

Electronic supplementary material The online version of this article (doi:10.1007/s12079-017-0390-x) contains supplementary material, which is available to authorized users.

✉ Roel Goldschmeding
R.Goldschmeding@umcutrecht.com

¹ Department of Pathology, UMC Utrecht, Heidelberglaan 100, 3584CX Utrecht, The Netherlands

² Department of Clinical Pharmacy, AVL Amsterdam, Amsterdam, Netherlands

³ Department of Clinical Pharmacy, UMC Utrecht, Utrecht, Netherlands

period following the last of 4 alternate day injections with Tamoxifen (1 mg/mouse) or vehicle-only (corn oil).

Materials & methods

Animals

Animal experiments were performed with approval of animal ethics committee of the university of Utrecht. C57Bl6/J Mice were injected 4 times every other day with either 100ul corn oil vehicle solution or corn oil Tamoxifen solution [10 mg/ml] (Sigma Aldrich), and 7 mice were injected per group. 14 days after the last injection, mice were subjected to Unilateral Ureter Obstruction (UUO) by permanent ligation of the left ureter under

general isoflurane anesthesia. 14 days after UUO mice were killed after which plasma and kidney tissue was collected.

Endoxifen measurement

For measurement of baseline endoxifen levels after two-week washout period, an additional 4 mice were killed prior to UUO. The method of Endoxifen measurement using HPLC in combination with Mass Spectrometry is extensively described elsewhere (Teunissen et al. 2011).

(Immuno)histochemistry

Formalin fixed paraffin embedded kidneys were cut into 3um sections, deparaffinized, rehydrated and stained with Sirius Red using standardized staining protocol

Fig. 1 Tamoxifen pre-treatment reduces fibrotic development in female mice upon UUO. **a**, experimental setup; **b**, representative micrographs of Sirius Red staining in male CLKs and OBKs; **c**, fibrosis quantification in male mice; **d**, representative micrographs of Sirius Red staining in female CLKs and OBKs; **e**, fibrosis quantification in female mice. 200× magnification * $p < 0.05$, *** $p < 0.005$

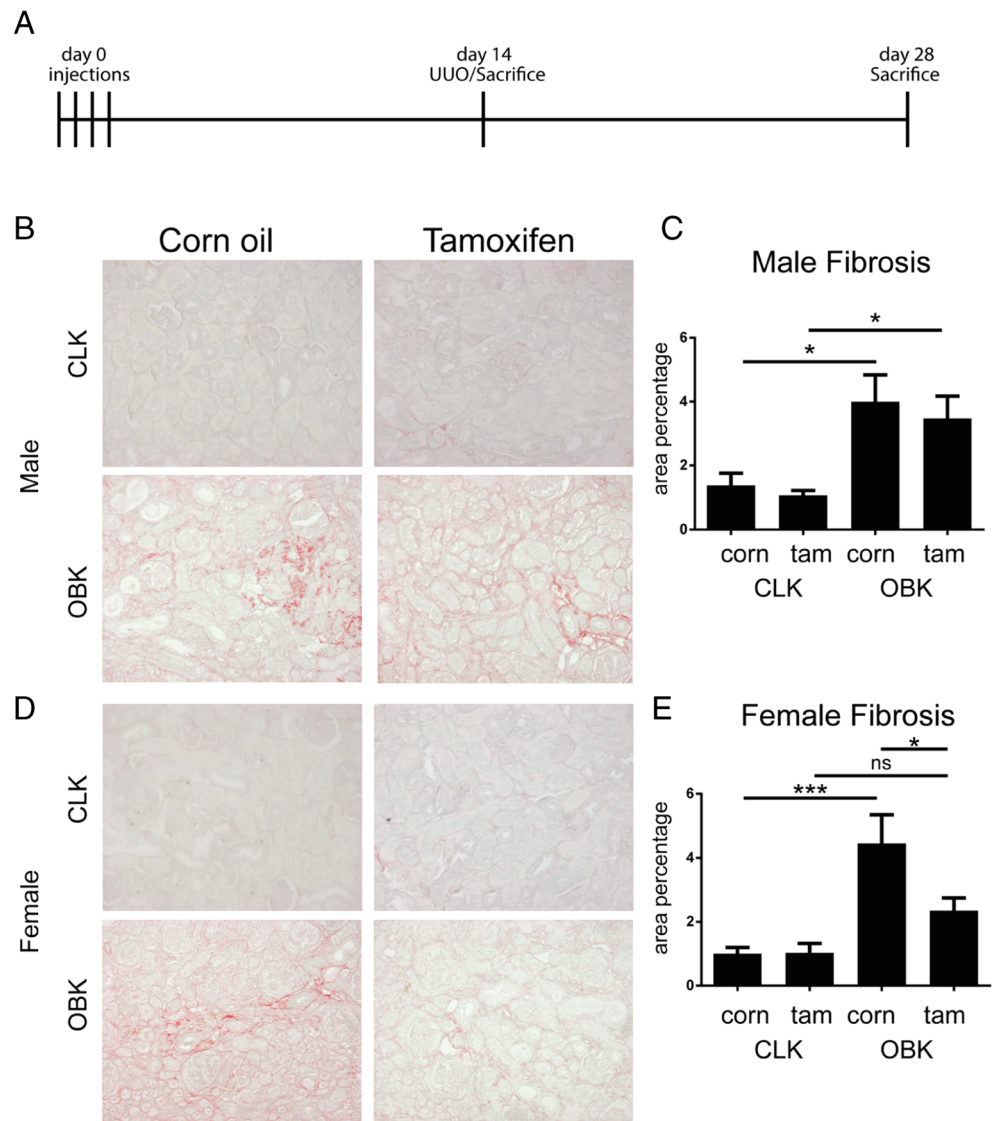
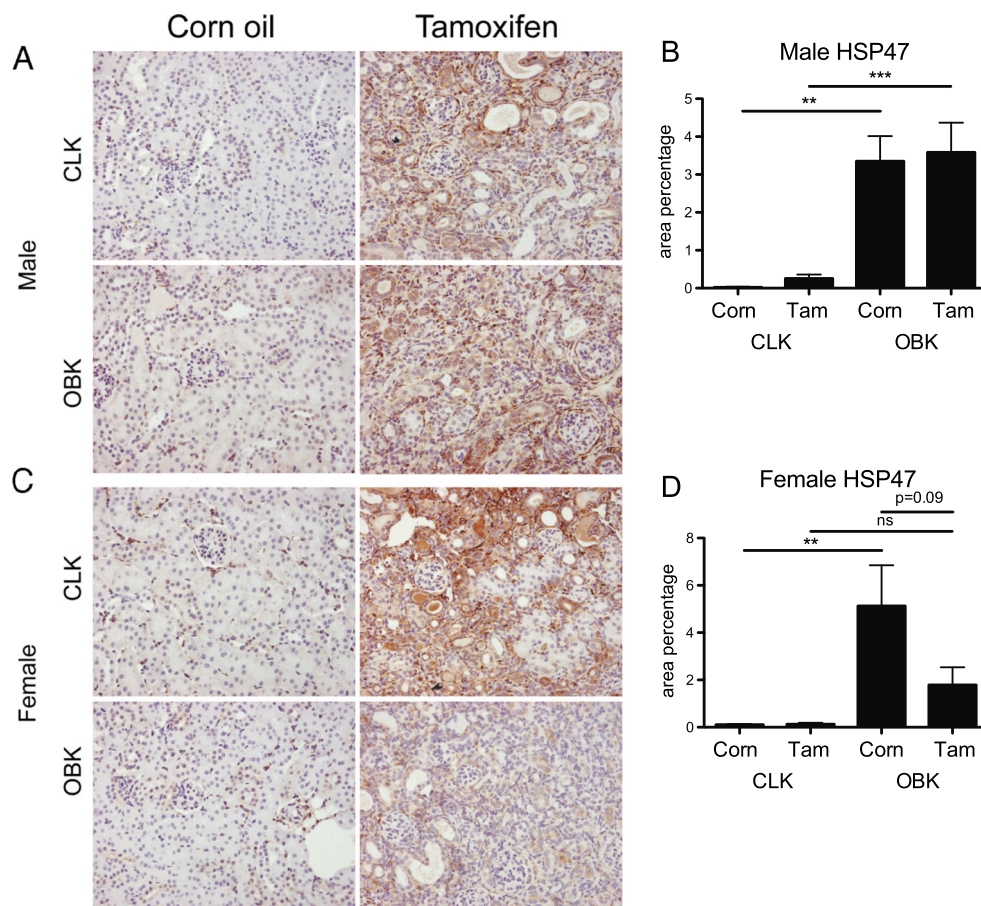


Fig. 2 Tamoxifen pre-treatment reduces de-novo collagen synthesis in female mice upon UUU. **a**, representative micrographs of Heat Shock Protein-47 staining in male CLKs and OBKs; **b**, HSP47 quantification in male mice; **c**, representative micrographs of HSP47 staining in female CLKs and OBKs; **d**, HSP47 quantification in female mice. 200× magnification. n.s. = not significant ** $p < 0.01$, *** $p < 0.005$



for diagnostics at the department of pathology of UMC Utrecht. The percentage of Sirius red positivity was determined by morphometric analysis of 10 kidney cortical fields at 200× magnification using ImageJ. For immunohistochemistry antigen retrieval (HSP47/Citrate, αSMA/EDTA, ERα/Citrate) boiling followed by endogenous peroxidase block and primary antibody incubation (HSP47: 1:1000, Enzo; αSMA: 1:200, AbCam; ERα: 1:250, Santa Cruz) was performed (for HSP47 the Animal Research Kit was used, Dako). After incubation with appropriate secondary antibody and enzymatic detection, percentage positivity using ImageJ (HSP47 & αSMA), or H-scores (ERα) were determined as described elsewhere (Wilbur et al. 1992; Lagiou et al. 2009).

Statistical analysis

All analyses were performed on blinded samples to prevent bias. ANOVA with Tukey correction for multiple testing was performed unless stated otherwise using GraphPad Prism. *P*-values below 0.05 were considered statistically significant.

Results

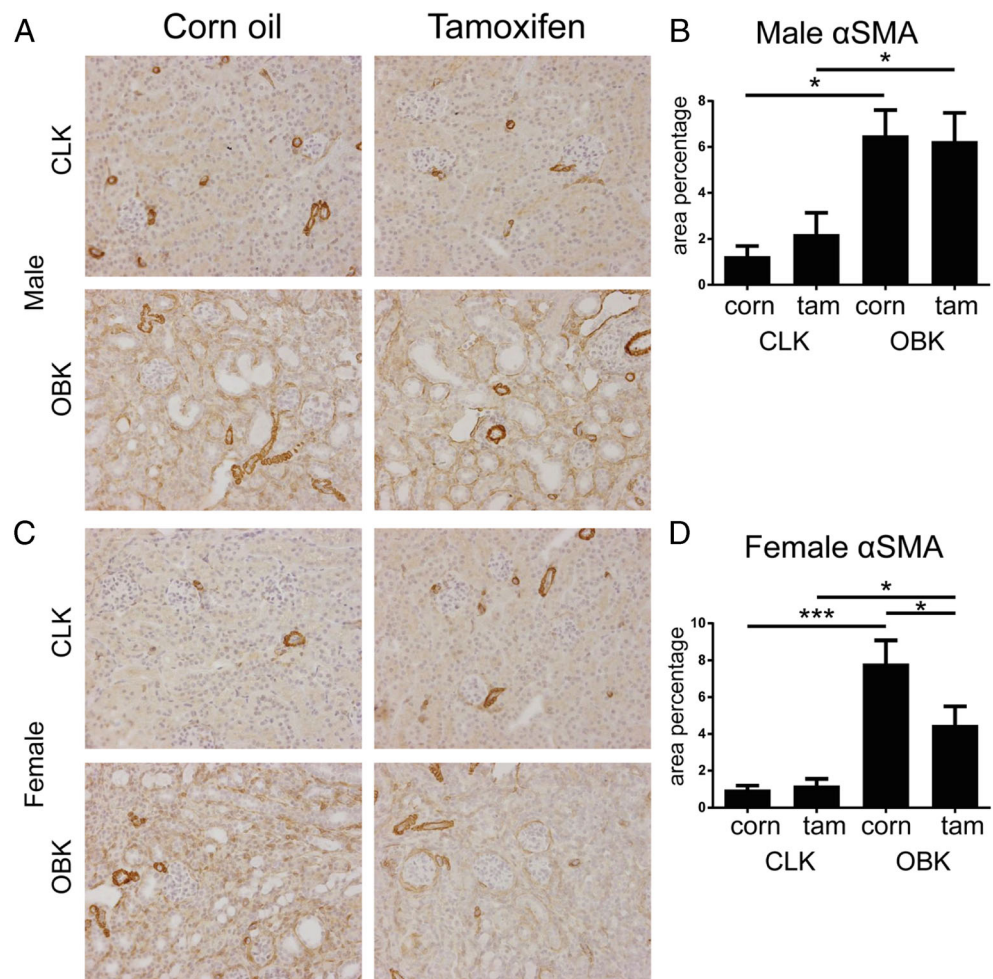
Tamoxifen/Endoxifen plasma levels are undetectable 2 and 4 weeks after initial injection

Mice were pre-treated with 4 doses of 1 mg Tamoxifen or corn oil only on alternate days until two weeks prior to unilateral ureter obstruction (UUO), and killed two weeks after UUO (Fig. 1a). Group characteristics are shown in Supplemental Table 2. First we analyzed Tamoxifen bioavailability by measuring Endoxifen (N-desmethyl-4-hydroxyTamoxifen) as a stable biologically active downstream metabolite more suitable for measurement (Teunissen et al. 2011; Ruddy et al. 2013). In our experimental setup, endoxifen levels were undetectable in plasma and kidney lysate at both 14 days (start of UUO) and 28 days (sacrifice) post injection (LLOQ < 0.1 ng/ml).

Tamoxifen reduces obstruction induced fibrosis in female but not in male mice

Analysis of Sirius red stained slides, specific for collagen deposition, showed that area positivity was not reduced in obstructed kidneys (OBKs) of Tamoxifen pre-treated male

Fig. 3 Tamoxifen pre-treatment reduces myofibroblast accumulation in female mice upon UUO. **a**, representative micrographs of α Smooth Muscle Actin staining in male CLKs and OBKs; **b**, α SMA quantification in male mice; **c**, representative micrographs of α SMA staining in female CLKs and OBKs; **d**, α SMA quantification in female mice. 200 \times magnification * $p < 0.05$, *** $p < 0.005$



mice (Fig. 1b, d). In female mice however, pre-treatment with Tamoxifen did result in reduced SR area positivity in OBKs (Fig. 1c, e).

In line with this, staining for Heat Shock Protein-47 (HSP47), a collagen chaperone protein closely related to de novo collagen production (Razzaque et al. 2005), showed that HSP47 was no longer significantly increased in Tamoxifen pre-treated female OBKs compared to CLKs (Fig. 2c, d).

Similarly, also the staining area for α -Smooth Muscle Actin (α SMA; a myofibroblast marker) was the same in Tamoxifen and corn oil injected male mice, but reduced in Tamoxifen injected female mice (Fig. 3a-d).

Tamoxifen lowers nuclear ER α in female kidneys compared to male kidneys upon UUO

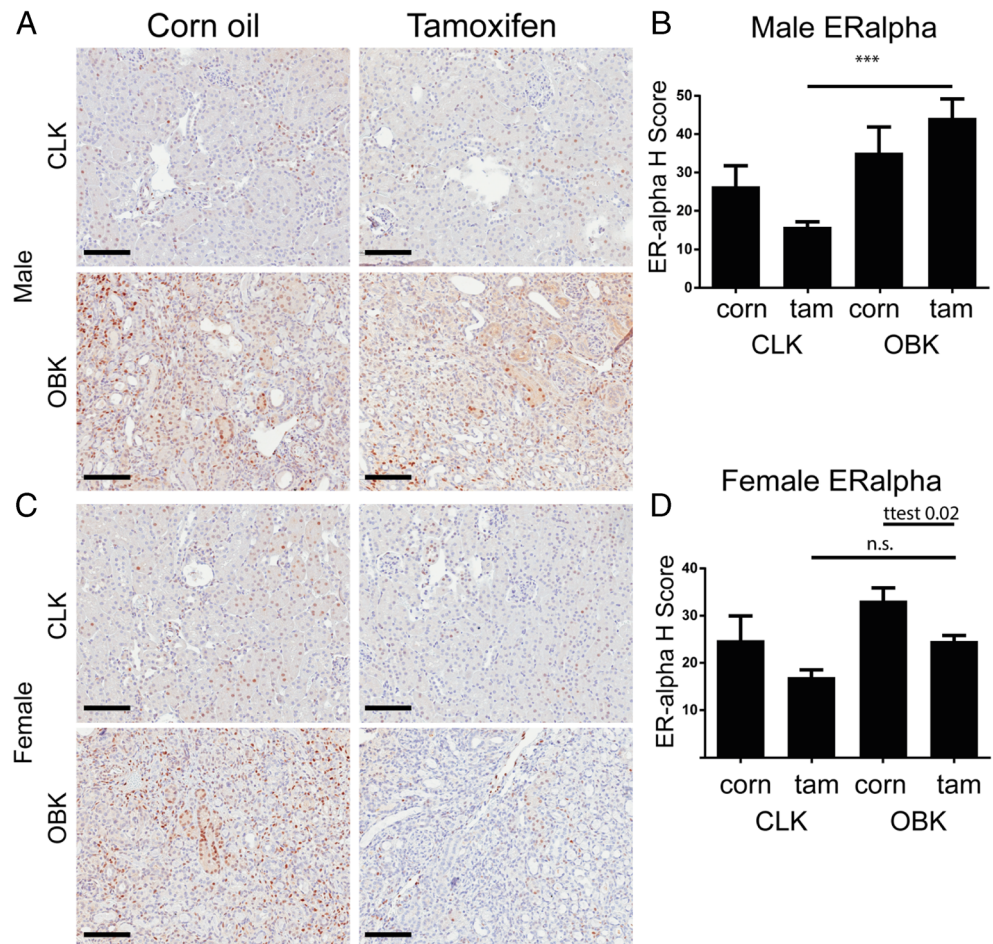
To explore a possible role of differential ER expression underlying the gender-associated difference, we analyzed nuclear estrogen receptor- α (ER α) expression by determining the H-index score, a commonly used weighted quantification method of nuclear ER α positivity (Wilbur et al. 1992; Lagiou et al. 2009). In unobstructed kidneys (CLKs),

Tamoxifen pre-treatment tended to lower nuclear ER α expression in both male and female mice but this difference was not significant ($p = 0.4$ and $p = 0.3$ resp., Fig. 4b, d). However, nuclear ER α positivity was significantly increased in OBKs of Tamoxifen pre-treated male mice (Fig. 4b), while in Tamoxifen pre-treated female mice the increase was not significant (Fig. 4d). Nuclear ER α positivity in vehicle pre-treated OBK's was similar in male and female mice ($p = 0.76$; T-test). However, nuclear ER α positivity was significantly lower in female than in male Tamoxifen pre-treated OBKs ($p = 0.0068$; T-test).

Discussion

This study shows that in male mice, the fibrogenic response upon experimental renal injury is not affected by Tamoxifen pre-treatment with the dosing regimen commonly used for modulation of floxed gene expression. Female mice pre-treated with Tamoxifen, however, showed a hampered fibrosis, despite absence of detectable endoxifen in blood and tissue throughout the UUO experiment. This is in line with

Fig. 4 Tamoxifen pre-treatment associates with reduced nuclear Estrogen Receptor α 14 days post UUU in female mice. **a**, Representative micrograph of ER α staining in male CLKs and OBKs; **b**, weighted quantification of nuclear ER α positivity in male mice; **c**, Representative micrograph of ER α staining in female CLKs and OBKs; **d**, weighted quantification of nuclear ER α positivity in female mice. 200 \times magnification *** $p < 0.005$



previous findings that kidneys of male mice are not influenced by ER α Knock Out (KO) (Lane 2008), and that the sensitivity of female mouse kidneys to acute Ischemia Reperfusion Injury was no longer reduced (compared to male kidneys) upon Tamoxifen administration or ovariectomy (Tanaka et al. 2013). Also, using a fixed dose of 4 mg per mouse might have higher impact in the 38% lighter female mice (Supplemental Table 2). However, since already prior to UUU both male and female mice had no detectable endoxifen anymore in the kidney or the blood, it seems unlikely that immediate and direct effects of residual Tamoxifen can fully explain the differential outcome, and that the possibility of sustained, possibly more indirect effects, should be taken into account. As such, the observed sustained reduction of nuclear ER α positivity only in female Tamoxifen treated mice might point to indirect mechanisms reducing ER α mediated fibrosis. In line with a sustained effect, the recurrence rate of peritoneal sclerosis was lower after discontinuation of Tamoxifen treatment, as compared to corticosteroid treatment (van der Bilt et al. 2016).

Deciphering the relative contribution of the various possible pathways is beyond the scope of this report, but a number of different mechanisms have been proposed by

which Tamoxifen can have suppressive effects on ER α mediated fibrosis is worth mentioning here. These include modulation of TGF β and EGF signaling (Britton et al. 2006; Xu et al. 2009; Goto et al. 2011; Carthy et al. 2015) and direct induction Renin expression, a protein involved in renal fibrosis (Lu et al. 2016). Since Tamoxifen treatment leads to compensatory increase of endogenous estrogen production, another pathway of possible relevance is activation of G-protein coupled receptor (GPCR; GPR30) signaling upon direct binding of estrogen (Prossnitz et al. 2008; Ignatov et al. 2011). Figure 5 summarizes a theoretical regulatory network of estrogen/ER α driven fibrosis along with tamoxifen/endoxifen intervention potentially explaining reduced fibrosis during 14 day UUU.

Prolonged anti-fibrotic effects of Tamoxifen treatment might also relate to mechanisms by which cancer cells have been noted to modulate downstream ER complex signaling through epigenetic regulation in response to Tamoxifen treatment, but it remains unclear how far such mechanisms might be operational in the non-oncological setting of kidney fibrosis (Musgrove and Sutherland 2009; Feng et al. 2014). Finally, baseline ER α

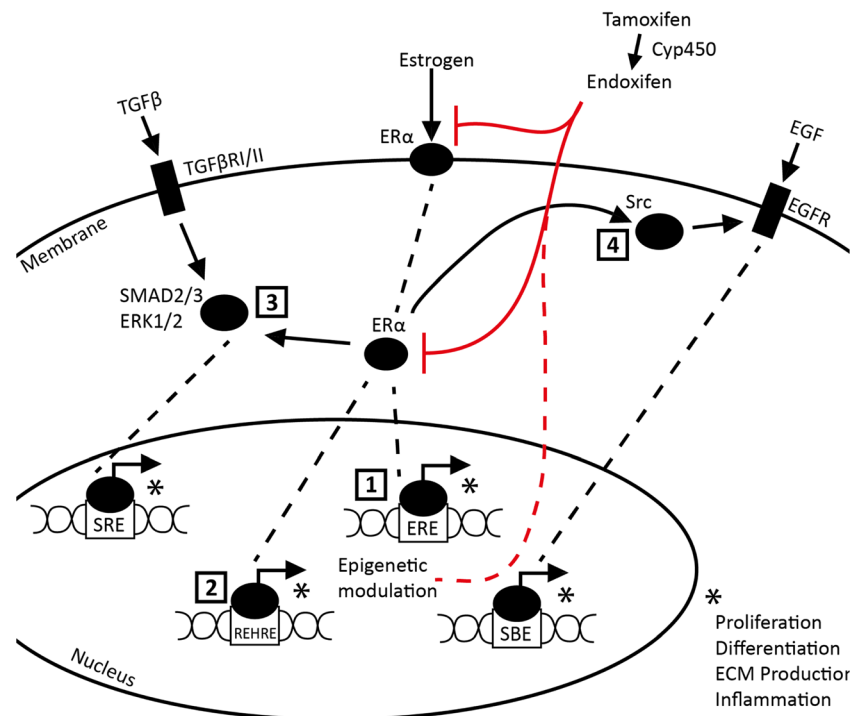


Fig. 5 Schematic overview of estrogen/Tamoxifen interaction in a fibrogenic context. 1. Direct nuclear translocation and binding of estrogen/ER α complex to the Estrogen Responsive Element (ERE). 2. Estrogen/ER α complex binding to the Renin Enhancer Hormone Response Element (REHRE). 3. Estrogen/ER α complex modulated SMAD2/3 and ERK1/2 binding to SMAD Binding Element (SBE). 4. ER α independent binding of estrogen to G-protein coupled receptor 30., 5. Estrogen/GPR30 or estrogen/ER α complex mediated modulation of

Src/EGFR interaction leading to downstream alterations in Serum Response Element (SRE) binding on MAPK. * Complex binding with ERE, REHRE, SBE or SRE respectively leads to modulation of processes involved in fibrosis (e.g. proliferation, differentiation, transcription including ECM production or infiltration/migration). Red lines indicate tamoxifen inhibition. Dashed red line indicates Tamoxifen mediated epigenetic alterations resulting in prolonged tamoxifen effects

expression increases in female but decreases in male kidneys upon ageing, indicating that studies involving Tamoxifen treatment in older mice might be even more prone to gender-related confounding than observed in the present study in young mice (Sharma and Thakur 2004).

Conclusion

We have found that for Tamoxifen-induced manipulation of gene expression in studies addressing kidney fibrosis, the commonly applied protocol with a 14 day washout period between the final dose and start of the actual experiment appears to be appropriate for studies in male mice, but it does not sufficiently prevent confounding anti-fibrotic Tamoxifen effects in female mice. Since (also in female mice) the blood and tissue endoxifen levels had fallen below the detection threshold already before the start of the experiment, a protracted indirect (e.g. epigenetic) effect might be responsible, and it remains to be seen whether longer wash-out periods could suffice to eliminate residual “off target” Tamoxifen effects (also in female mice). Until this has been resolved studies

involving Tamoxifen pre-treatment should be limited to male mice, and existing data from such studies in female mice should be interpreted with great caution.

Acknowledgements The authors would like to thank Hilde Rosingh for her technical contribution to this work. Lucas Falke was sponsored by the Netherlands Institute for Regenerative Medicine (NIRM, Grant No. FES0908).

Compliance with ethical standards

Financial/competing interests Roel Goldschmeding was employed by FibroGen Inc. in 2008/2009 and received research support and travel grants in the past.

References

- van der Bilt FE, Hendriksz TR, van der Meijden WAG et al (2016) Outcome in patients with idiopathic retroperitoneal fibrosis treated with corticosteroid or tamoxifen monotherapy. *Clin Kidney J* 9:184–191. doi:10.1093/ckj/sfv148
- Britton DJ, Hutcheson IR, Knowlden JM et al (2006) Bidirectional cross talk between ERalpha and EGFR signalling pathways regulates tamoxifen-resistant growth. *Breast Cancer Res Treat* 96:131–146. doi:10.1007/s10549-005-9070-2

- Carthy JM, Sundqvist A, Heldin A et al (2015) Tamoxifen inhibits TGF- β -mediated activation of Myofibroblasts by blocking non-Smad signaling through ERK1/2. *J Cell Physiol* 230:3084–3092. doi:10.1002/jcp.25049
- Cohen AM, Rosenmann E (1985) Effect of the estrogen antagonist, tamoxifen, on development of glomerulosclerosis in the Cohen diabetic rat. *Diabetes* 34:634–638
- Dellé H, Rocha JRC, Cavaglieri RC et al (2012) Antifibrotic effect of tamoxifen in a model of progressive renal disease. *J Am Soc Nephrol* 23:37–48. doi:10.1681/ASN.2011010046
- Feng Q, Zhang Z, Shea MJ et al (2014) An epigenomic approach to therapy for tamoxifen-resistant breast cancer. *Cell Res* 24:809–819. doi:10.1038/cr.2014.71
- Goto N, Hiyoshi H, Ito I et al (2011) Estrogen and antiestrogens alter breast cancer invasiveness by modulating the transforming growth factor- β signaling pathway. *Cancer Sci* 102:1501–1508. doi:10.1111/j.1349-7006.2011.01977.x
- Hayashi S, McMahon AP (2002) Efficient recombination in diverse tissues by a tamoxifen-inducible form of Cre: a tool for temporally regulated gene activation/inactivation in the mouse. *Dev Biol* 244:305–318. doi:10.1006/dbio.2002.0597
- Ignatov A, Ignatov T, Weissenborn C et al (2011) G-protein-coupled estrogen receptor GPR30 and tamoxifen resistance in breast cancer. *Breast Cancer Res Treat* 128:457–466. doi:10.1007/s10549-011-1584-1
- Irsik DL, Carmines PK, Lane PH (2013) Classical estrogen receptors and ER α splice variants in the mouse. *PLoS One* 8:e70926. doi:10.1371/journal.pone.0070926
- Jelinsky SA, Harris HA, Brown EL et al (2003) Global transcription profiling of estrogen activity: estrogen receptor alpha regulates gene expression in the kidney. *Endocrinology* 144:701–710. doi:10.1210/en.2002-220728
- Kim D, Lee AS, Jung YJ et al (2014) Tamoxifen ameliorates renal tubulointerstitial fibrosis by modulation of estrogen receptor α -mediated transforming growth factor- β 1/Smad signaling pathway. *Nephrol Dial Transplant* 29:2043–2053. doi:10.1093/ndt/gfu240
- Lagiou P, Georgila C, Samoli E et al (2009) Estrogen alpha and progesterone receptor expression in the normal mammary epithelium in relation to breast cancer risk. *Int J Cancer* 124:440–442. doi:10.1002/ijc.23899
- Lane PH (2008) Estrogen receptors in the kidney: lessons from genetically altered mice. *Gender Medicine* 5:S11–S18. doi:10.1016/j.genm.2008.03.003
- Lu K-T, Keen HL, Weatherford ET et al (2016) Estrogen receptor α is required for maintaining baseline Renin Expression Novelty and significance. *Hypertension* 67:992–999. doi:10.1161/HYPERTENSIONAHA.115.07082
- Mao S, Xu H, Zou L et al (2014) Estrogen preserves split renal function in a chronic complete unilateral ureteral obstruction animal model. *Exp Ther Med* 7:1555–1562. doi:10.3892/etm.2014.1663
- Musgrove EA, Sutherland RL (2009) Biological determinants of endocrine resistance in breast cancer. *Nat Rev Cancer* 9:631–643. doi:10.1038/nrc2713
- Prossnitz ER, Arterburn JB, Smith HO et al (2008) Estrogen signaling through the transmembrane G protein-coupled receptor GPR30. *Annu Rev Physiol* 70:165–190. doi:10.1146/annurev.physiol.70.113006.100518
- Razzaque MS, Le VT, Taguchi T (2005) Heat shock protein 47 and renal fibrogenesis. *Contrib Nephrol* 148:57–69. doi:10.1159/000086043
- Ruddy KJ, Desantis SD, Gelman RS et al (2013) Personalized medicine in breast cancer: tamoxifen, endoxifen, and CYP2D6 in clinical practice. *Breast Cancer Res Treat* 141:421–427. doi:10.1007/s10549-013-2700-1
- Sharma PK, Thakur MK (2004) Estrogen receptor alpha expression in mice kidney shows sex differences during aging. *Biogerontology* 5:375–381. doi:10.1007/s10522-004-3191-6
- Tanaka R, Tsutsui H, Ohkita M et al (2013) Sex differences in ischemia/reperfusion-induced acute kidney injury are dependent on the renal sympathetic nervous system. *Eur J Pharmacol* 714:397–404. doi:10.1016/j.ejphar.2013.07.008
- Teunissen SF, Jager NGL, Rosing H et al (2011) Development and validation of a quantitative assay for the determination of tamoxifen and its five main phase I metabolites in human serum using liquid chromatography coupled with tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 879:1677–1685. doi:10.1016/j.jchromb.2011.04.011
- Wilbur DC, Willis J, Mooney RA et al (1992) Estrogen and progesterone receptor detection in archival formalin-fixed, paraffin-embedded tissue from breast carcinoma: a comparison of immunohistochemistry with the dextran-coated charcoal assay. *Mod Pathol* 5:79–84
- Xu J, Wu R-C, O'Malley BW (2009) Normal and cancer-related functions of the p160 steroid receptor co-activator (SRC) family. *Nat Rev Cancer* 9:615–630. doi:10.1038/nrc2695