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Effect of different component ratio of Astragalus total saponins and Verbena total glycosides on the cerebral infarction area and serum biochemical indicators in the focal cerebral ischemia-reperfusion rat model



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ABSTRACT

Our purpose is to study the effect of different component ratio of Astragalus Total Saponins (ATS) and Verbena Total Glycosides (VTG) on the cerebral infarction area and the serum biochemical indicators in the focal cerebral ischemia-reperfusion rat model. Compared with the model group, different component ratio of ATS and VTG could significantly improve the neurological deficit scores to the focal cerebral ischemia-reperfusion rat model, and the group of 7:3, 6:4, 5:5 got the best results; it could reduce the mortality of rat model to a certain extent, and the group of 5:5 group got the best results; it can significantly reduce the cerebral infarction area, and the group of 7:3, 5:5, 4:6 got the best results; it could significantly reduce the content of TNF- α , and the group of 8:2, 6:4 got the best results; it could significantly reduce the content of NO, and the group of 7:3, 5:5 got the best results; it could significantly increase the content of SOD, and the group of 6:4, 5:5 got the best results. This indicates that different component ratio of ATS and VTG may protect the damage of focal cerebral ischemia-reperfusion rat model to a certain extent, which are compared using the comprehensive weight method and the ratio of 5:5 was proved to be the optimal active ratio.

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1. Introduction

Cerebral Ischemia Reperfusion Injury (CIRI) refers to that when blood supply is restored for a certain period of time after cerebral ischemia, its function fails to restore, or even shows more serious brain dysfunction. In modern medical research, the pathogenesis of cerebral ischemia includes energy metabolism disorders, platelet aggregation, vasoconstriction and microcirculation disorders, excitatory amino acid toxicity and so on, and the drugs clinically used for the preventing and treating cerebral ischemia are mainly antiplatelet drugs aspirin, ticlopidine and other drugs. Most cere-

bral ischemia patients need a long time of medication, but the side effect from long-term medication, drug prices, limitations of treatment course will affect the treatment of the disease (Atta et al., 2017; Halim et al., 2017). Chinese medicine believes that the occurrence of ischemic stroke relates with the Qi stagnation, Qi deficiency, phlegm turbidity, blood stasis, Yin deficiency, heat or Fire and other factors, so, clinically, some therapeutic principles like “tonifying Qi and reinforcing deficiency”, “activating blood and resolving stasis”, “clearing heat and expelling toxin” and others are adopted to prevent cerebral ischemic diseases. The components compatibility of Chinese medicine has precise clinical application, is safe and effective, and owns controllable quality. QiCao capsule is an effective preparation for clinical treatment of cerebral ischemia, with very good clinical basis. This research used the increase-decrease design of baseline geometric proportion to study the intervene effect of different component ratios of ATS and VTG, the main ingredients in QiCao capsule, on the cerebral infarction area and the serum biochemical indicators in the cerebral ischemia-reperfusion rat model (Miao and Miao, 2012; Miao et al., 2011).

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2. Material and methods

2.1. Experimental animals

SD rat, male, grade SPF, Weight 180–220 g, supplied by Hebei Experimental Animal Center, Animal permit number: 1210020.

2.2. Experimental drugs and reagents

Astragalus Total Saponins, supplied by department of chemistry, Henan University of TCM, content > 50%, batch No.: 111008; Verbena total glycosides, supplied by department of chemistry, Henan University of TCM, content = 50%, batch No.: SWT20120801; Naoluo tong capsule, produced by Guangdong Bangmin Pharmaceutical Co., Ltd., batch No.: 120109; Nimodipine tablets, produced by Shandong Xinhua Pharmaceutical Co., Ltd., batch No.: 1107044; Formaldehyde, produced by Tianjin Kernel Chemical Reagent Co., Ltd., batch No.: 20110701; Triphenyl tetrazolium chloride (TTC), produced by Shanghai Shan Pu Chemical Company, batch No.: 20110401; TNF- α kit, produced by R & D company, batch No.: 201211; SOD kit, produced by Nanjing Jiancheng Institute of Biotechnology, batch No.: 20121123; Nitric oxide (NO), produced by Nanjing Jiancheng Institute of Biotechnology, batch No.: 20121117.

2.3. Experimental instrument

UV-2000 UV-Vis spectrophotometer, UNICO (Shanghai) Instrument Co., Ltd.; TGL-16G high-speed refrigerated centrifuge, supplied by Shanghai Anting Scientific Instrument Factory; FA (N)/JA (N) series of electronic balance, Shanghai Minqiao Precision Instrument Co., Ltd; Bio-Rad-680 microplate reader, American Microplate Reader company; Adjustable pipettes, Shanghai Leibo Analysis Instrument Co. Ltd.

2.4. Experimental methods

SPF-grade SD healthy rats, male, weight 200–220 g, were taken and randomly and uniformly divided into 13 groups, and every group was 15 rats, including sham operation group (SG), model group (MG), Naoluo tong capsule group (NLT), nimodipine group (NMP), and ATS and VTG groups with different component ratios of 10:0, 8:2, 7:3, 6:4, 5:5, 4:6, 3:7, 2:8, and 0:10. The drug disposition method: the dose of the different component ratios of ATS and VTG was all 200 mg/kg. In details, the dose of 10:0 group was (ATS 200 mg and VTG 0 mg)/kg, 8:2 group (ATS 160 mg and VTG 40 mg)/kg, 7:3 group (ATS 140 mg and VTG 60 mg)/kg, 6:4 group (ATS 120 mg and VTG 80 mg)/kg, 5:5 group (ATS 100 mg and VTG 100 mg)/kg, 4:6 group (ATS 80 mg and VTG 120 mg)/kg, 3:7 group (ATS 60 mg and VTG 140 mg)/kg, 2:8 group (ATS 20 mg and VTG 160 mg)/kg, and 0:10 group (ATS 0 mg and VTG 200 mg)/kg. Besides, the dose of Naoluo tong capsule group was 500 mg/kg (equivalent to 10 times of clinical dose), and Nimodipine group 20 mg/kg (equivalent to 10 times of clinical dose). Each group of drugs were formulated with distilled water. Each treatment group was fed with corresponding drugs, and the Sham operation group and the model group were given equal volume of distilled water, in which the filling volume was 0.2 ml/10 g once a day and lasted for 7 days.

One hour after the last administration (Fasting 12 h), it's the time to build the MCAO model of each model group.

The rats were anesthetized by intraperitoneal injection with 10% chloral hydrate (0.3 ml/100 g), supine fixed, cut in mid-line and slant left of neck, layer-by-layer separated and exposed the left

common carotid artery (CCA), external carotid artery (ECA) and internal carotid artery (ICA), ligatured CCA and ECA. After using arterial clamp to clip the telecentric end of ICA, made an incision at the bifurcation of ECA and ICA and inserted smooth nylon thread with the tip coated with silicone rubber (the diameters of line body and head were 0.28 mm and 0.38 ± 0.02 mm respectively, with a mark at 20 mm from the line head) at a depth of about 20 mm, which can achieve middle cerebral artery occlusion and therefore lead to cerebral ischemia. Suture the skin at the ligation entrance with Nylon line left 1–2 cm at outside. After 2 h, gently pulled the line head that left at outside until facing a little resistance, to achieve reperfusion. During the process of brain ischemia and reperfusion, the room temperature should be kept at 23–25 °C. (For sham operation group, it only required to expose the left side of the blood vessel without inserting nylon thread). It can be deemed as a successful model when the ipsilateral Horner syndrome and that the contralaterally upper extremity was more paralyzed than lower limbs appear after the rats awake. Initially 195 rats were included in experiment, 77 rats died after the operation, and eventually 118 rats entered the final analysis.

After anesthesia awake, the rats received the neural function deficit scoring (NS) (Neuroethology criteria used the Zealanga scoring method. Score 0: No symptoms of neurological deficits, normal activities; Score 1: those who cannot fully extend the contralateral forepaw; Score 2: a circular motion to the right when crawling; Score 3: when walking, the body dumped by the side of hemiplegia; Score 4: not spontaneously walk, the loss of consciousness; Score 5: Death.). 24 h after surgery, the blood was taken to measure the contents of TNF- α , SOD and NO. Decapitated and rapidly stripped brain tissue, and cut the rest coronal region after removing the olfactory bulb, cerebellum and low brain stem into five slices. In TTC staining, after staining, the non-ischemic region was rosy red and the infarct region was white (Ishaq and Jafri, 2017). Took pictures by digital camera, and used the NIS-Elements AR 4.10.01 to calculate the areas of the total five brain slices (including 10 planes) and the infarction region, as well as the percentage of infarction region in total area.

2.5. Statistical analysis

Data analysis was made using SPSS 17.0 for windows. The measurement data was expressed by mean \pm SD. ANOVA was adopted in analysis and comparison between the groups. Those with equal variances shall be tested by the smallest used least significant difference (LSD) method, while those with heterogeneous variance

Table 1
Effect of the neurological deficit scores and the mortality ($\bar{x} \pm s$).

Group	n	Dose (g kg ⁻¹)	NS	Mortality (%)
SG	15	–	0.0 \pm 0.0 ^{**}	0.0
MG	8	–	3.1 \pm 0.4	46.7
NMP	9	0.02	1.8 \pm 0.8 ^{**}	40.0
NLT	9	0.5	1.9 \pm 0.6 [*]	40.0
10:0	8	ATS:VTG = 0.2:0	1.9 \pm 0.6 [*]	46.7
8:2	8	ATS:VTG = 0.16:0.04	1.9 \pm 0.4 [*]	46.7
7:3	9	ATS: VTG = 0.14:0.06	1.4 \pm 0.5 ^{**}	40.0
6:4	9	ATS: VTG = 0.12:0.08	1.7 \pm 0.5 ^{**}	40.0
5:5	10	ATS: VTG = 0.1:0.1	1.5 \pm 0.5 ^{**}	33.3
4:6	9	ATS: VTG = 0.08:0.12	2.0 \pm 0.7 [*]	40.0
3:7	8	ATS: VTG = 0.06:0.14	2.0 \pm 0.7 [*]	46.7
2:8	8	ATS: VTG = 0.04:0.16	1.9 \pm 0.6 [*]	46.7
0:10	8	ATS: VTG = 0:0.2	2.0 \pm 0.7 [*]	46.7

Note: Compared to the model group.

^{*} $P < 0.05$.

^{**} $P < 0.01$.

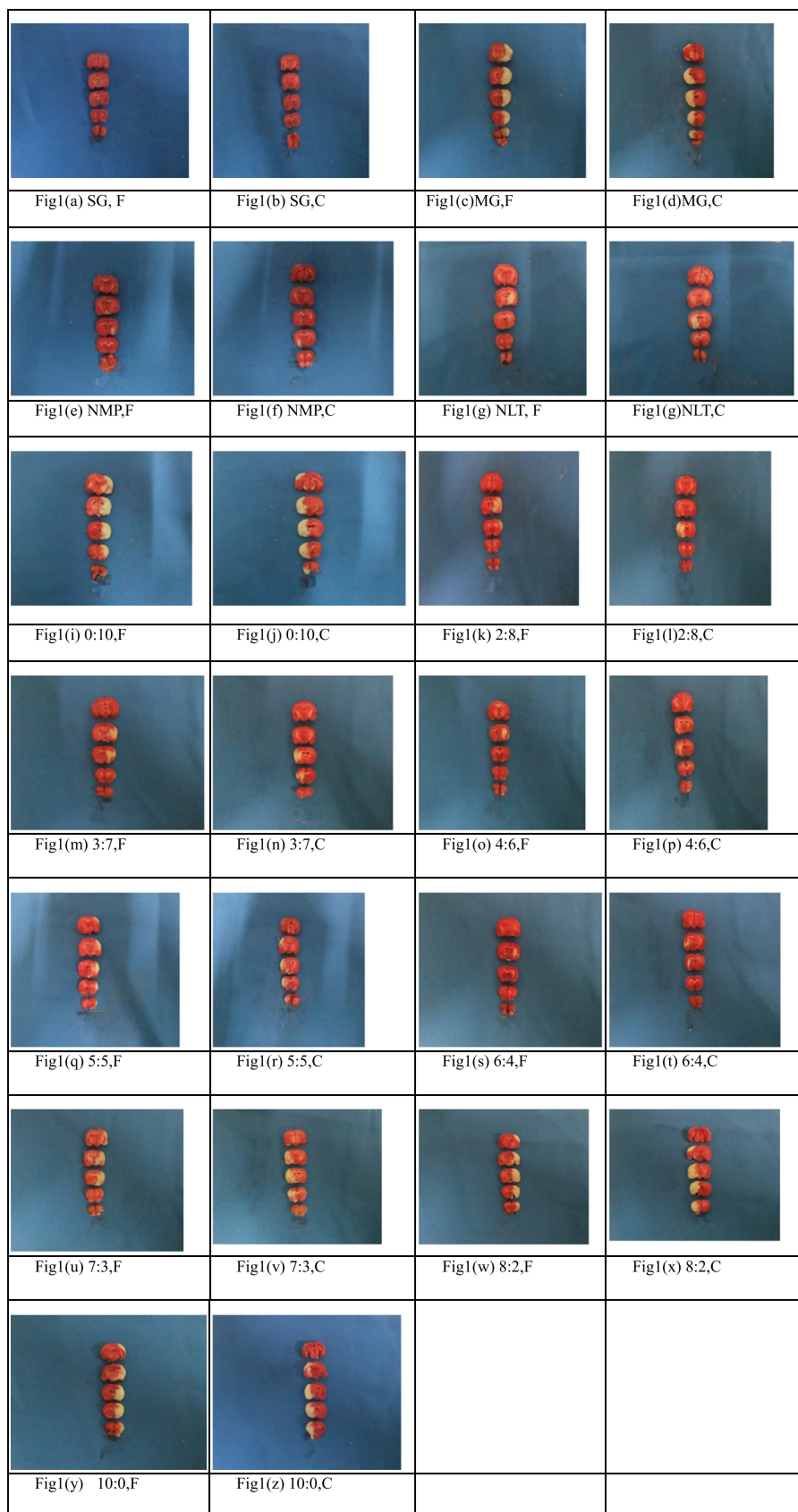


Fig. 1. The cerebral infarction area. Note: F: The front of the cerebral infarction area, C: The contrary of the cerebral infarction area.

shall be tested by Games-Howell method, and ranked data shall use Ridit test.

3. Results

3.1. Effect of the neurological deficit scores and the mortality in the cerebral ischemia-reperfusion rat model

From Table 1, compared with the SG, the neurological deficit scores of model rat is significantly increased ($P < 0.01$), which shows a successfully copied model. Compared with the MG, the groups with different component ratio of ATS:VTG can significantly reduce the neurological deficit scores ($P < 0.05$), and the groups of 7:3, 6:4 and 5:5 got the best results. And these groups can reduce the mortality to certain extent, and the group 5:5 got the best results (Muhammad et al., 2017).

3.2. Effect of the cerebral infarction area in the cerebral ischemia-reperfusion rat model

From Fig. 1 and Table 2, compared with the SG, the cerebral infarction area of model rat was significantly increased ($P < 0.01$), which shows a successfully copied model. Compared with the MG, the different component ratio of ATS:VTG can significantly reduce the cerebral infarction area ($P < 0.01$), and the groups of 7:3, 5:5 and 4:6 got the best results (see Fig. 1).

3.3. Effect of the contents of TNF- α in the cerebral ischemia-reperfusion rat model

From Table 3, compared with the SG, the content of TNF- α in model rat has been significantly increased ($P < 0.01$), which shows a successfully copied model. Compared with the MG, the different component ratio of ATS:VTG can significantly reduce the content of TNF- α ($P < 0.01$), and the groups of 8:2 and 6:4 got the best results.

3.4. Effect of the content of NO in the cerebral ischemia-reperfusion rat model

From Table 4, compared with the SG, the content of NO in model rat has been significantly increased ($P < 0.01$), which shows a successfully copied model. Compared with the MG, the different component ratio of ATS:VTG can significantly reduce the content of NO ($P < 0.01$), it shows that the different component ratio of ATS:VTG can significantly improve the content of NO in the cerebral

Table 2
Effect of the cerebral infarction area ($\bar{x} \pm s$).

Group	n	Dose (g kg ⁻¹)	CIA (%)
SG	15	–	0.0 ± 0.0 ^{**}
MG	8	–	28.05 ± 4.97
NMP	9	0.02	13.04 ± 5.46 ^{**}
NLT	9	0.5	15.46 ± 7.29 ^{**}
10:0	8	ATS:VTG = 0.2:0	16.16 ± 8.95 ^{**}
8:2	8	ATS:VTG = 0.16:0.04	17.75 ± 9.55 ^{**}
7:3	9	ATS:VTG = 0.14:0.06	12.96 ± 6.89 ^{**}
6:4	9	ATS:VTG = 0.12:0.08	15.84 ± 9.36 ^{**}
5:5	10	ATS:VTG = 0.1:0.1	15.08 ± 9.69 ^{**}
4:6	9	ATS:VTG = 0.08:0.12	15.50 ± 6.76 ^{**}
3:7	8	ATS:VTG = 0.06:0.14	16.63 ± 8.92 ^{**}
2:8	8	ATS:VTG = 0.04:0.16	17.38 ± 10.18 ^{**}
0:10	8	ATS:VTG = 0:0.2	20.05 ± 10.10 ^{**}

Note: Compared to the model group.

^{*} $P < 0.05$.

^{**} $P < 0.01$.

Table 3
Effect of the contents of TNF- α ($\bar{x} \pm s$).

Group	n	Dose (g kg ⁻¹)	TNF- α (ng/ml)
SG	15	–	1.667 ± 0.577 ^{**}
MG	8	–	2.580 ± 0.300
NMP	9	0.02	1.929 ± 0.428 ^{**}
NLT	9	0.5	1.856 ± 0.387 ^{**}
10:0	8	ATS:VTG = 0.2:0	1.947 ± 0.393 ^{**}
8:2	8	ATS:VTG = 0.16:0.04	1.869 ± 0.335 ^{**}
7:3	9	ATS:VTG = 0.14:0.06	1.977 ± 0.340 ^{**}
6:4	9	ATS:VTG = 0.12:0.08	1.895 ± 0.371 ^{**}
5:5	10	ATS:VTG = 0.1:0.1	1.925 ± 0.327 ^{**}
4:6	9	ATS:VTG = 0.08:0.12	1.970 ± 0.222 ^{**}
3:7	8	ATS:VTG = 0.06:0.14	1.903 ± 0.138 ^{**}
2:8	8	ATS:VTG = 0.04:0.16	1.960 ± 0.252 ^{**}
0:10	8	ATS:VTG = 0:0.2	1.949 ± 0.343 ^{**}

Note: Compared to the model group. ^{*} $P < 0.05$.

^{**} $P < 0.01$.

Table 4
Effect of the content of NO ($\bar{x} \pm s$).

Group	n	Dose (g kg ⁻¹)	NO (nmol/ml)
SG	15	–	25.06 ± 5.27 ^{**}
MG	8	–	46.77 ± 5.13
NMP	9	0.02	33.67 ± 9.58 ^{**}
NLT	9	0.5	33.95 ± 6.57 ^{**}
10:0	8	ATS:VTG = 0.2:0	35.86 ± 4.30 ^{**}
8:2	8	ATS:VTG = 0.16:0.04	38.12 ± 8.81 ^{**}
7:3	9	ATS:VTG = 0.14:0.06	34.33 ± 6.53 ^{**}
6:4	9	ATS:VTG = 0.12:0.08	36.85 ± 6.21 ^{**}
5:5	10	ATS:VTG = 0.1:0.1	34.01 ± 5.08 ^{**}
4:6	9	ATS:VTG = 0.08:0.12	35.46 ± 5.58 ^{**}
3:7	8	ATS:VTG = 0.06:0.14	36.00 ± 4.65 ^{**}
2:8	8	ATS:VTG = 0.04:0.16	36.42 ± 7.07 ^{**}
0:10	8	ATS:VTG = 0:0.2	35.57 ± 4.63 ^{**}

Note: Compared to the model group. ^{*} $P < 0.05$.

^{**} $P < 0.01$.

ischemia-reperfusion rat model, and the groups of 7:3 and 5:5 got the best results.

3.5. Effect of the activity of SOD in the focal cerebral ischemia-reperfusion rat model

From Table 5, compared with the SG, the activity of SOD in model rat has been significantly reduced ($P < 0.01$), which shows a successfully copied model. Compared with the MG, the different component ratio of ATS:VTG can significantly increase the activity of SOD ($P < 0.01$), and the groups of 6:4 and 5:5 got the best results.

Table 5
Effect of the activity of SOD ($\bar{x} \pm s$).

Group	n	Dose (g kg ⁻¹)	SOD (U/ml)
SG	15	–	214.63 ± 39.04 ^{**}
MG	8	–	138.10 ± 34.48
NMP	9	0.02	192.17 ± 43.02 ^{**}
NLT	9	0.5	168.87 ± 32.78 [*]
10:0	8	ATS:VTG = 0.2:0	162.47 ± 22.25
8:2	8	ATS:VTG = 0.16:0.04	164.96 ± 29.64
7:3	9	ATS:VTG = 0.14:0.06	168.87 ± 23.74 [*]
6:4	9	ATS:VTG = 0.12:0.08	172.72 ± 21.73 [*]
5:5	10	ATS:VTG = 0.1:0.1	177.98 ± 28.62 ^{**}
4:6	9	ATS:VTG = 0.08:0.12	168.48 ± 29.62 ^{**}
3:7	8	ATS:VTG = 0.06:0.14	170.70 ± 23.32 [*]
2:8	8	ATS:VTG = 0.04:0.16	168.86 ± 24.46 [*]
0:10	8	ATS:VTG = 0:0.2	161.71 ± 30.07

Note: Compared to the model group.

^{*} $P < 0.05$.

^{**} $P < 0.01$.

Table 6
The index classification results.

Grade	I level indicators	II level indicators	III level indicators
Index	CIA, NS	Mortality rate	SOD, NO, TNF- α
Weighting factor	0.5	0.3	0.2

Table 7
Index improvement rate at each classified level.

Improvement rate	>50%	50–30%	30–10%	<10%
Grade	Significant improvement	Improvement	More improvement	No improvement
Weighting factor	0.4	0.3	0.2	0.1

Table 8
Comprehensive weight assessment scores.

Rat	The value of P
10:0	0.48
8:2	0.48
7:3	0.58
6:4	0.48
5:5	0.61
4:6	0.48
3:7	0.48
2:8	0.48
0:10	0.43

3.6. Comprehensive evaluation of the effect of the different component ratio of *Astragalus total saponins* and *Verbena total glycosides* on the rat model using the comprehensive weight method

3.6.1. Classify above indicators, and determine the weight coefficient at all levels

See Table 6.

3.6.2. Based on the improvement of the symptoms and biochemical indexes of the animal model $V = (Y - M) / M * 100\%$ (Among them, Y was the index value of each drug group, and M was the corresponding index value of the model group), classify each index value and determine the weight factor of each level

See Table 7.

3.6.3. Based on the total evaluation score P value formula, calculate the comprehensive weight assessment scores of groups with different component ratios (Ma and Miao, 2011)

From Table 8, in the groups with different component ratios of ATS:VTG, the score of comprehensive weight assessment for group 5:5 is the highest, which indicates that the 5:5 component ratio of ATS:VTG can improve the cerebral ischemia-reperfusion rat model to the best extent.

4. Discussion and conclusion

This research used the increase-decrease design of the baseline geometric proportion to study the intervene effect of different component ratio of ATS and VTG on the cerebral infarction area and the serum biochemical indicators in the focal cerebral ischemia-reperfusion rat model, and used the comprehensive weight method to evaluate the efficacy of each component, in

which 5:5 group was the optimal active ratio group (Rahman et al., 2017). The components compatibility of Chinese medicine can improve the modern technology system that studies Chinese medicine substances and active ingredients, breakthrough the single mode that takes clinical experience to develop new drugs, provide theoretical basis and technical support for innovative drug research and quality control of traditional Chinese medicine, and promote the modernization of Chinese medicine development.

The experiment used the suture-occluded method to copy the focal cerebral ischemia-reperfusion rat model. This method simulates the different states of human cerebral ischemia and is the most widely used one. The neurological function changes of rats is a macro-indicator of cerebral ischemia-reperfusion model, and the behavioral changes in the animal model can reflect the abnormalities of the nervous system. Cerebral infarction is mainly cerebral blood supply disorder that causes ischemia and hypoxia, further leading to the ischemic necrosis or brain softening in the brain tissue, and formatting the infarction in the ischemic penumbra around the infarct area, so it can be used to evaluate the protective effect of drugs. SOD, as a protease, is an important antioxidants in the body (Dong et al., 2013) and an important indicators to evaluate the oxidation and anti-oxidation in a balance system (Yan et al., 2013). When cerebral ischemia occurs, the oxygen supply in the brain tissue will be reduced, thus producing a large number of free radicals (Kang and Zhang, 2010). SOD is a major free radical scavenger and can remove super-oxide anion free radicals by disproportionation reaction. TNF- α is a cytokines with extensive biological functions and an important initiation factor, which can directly promote neutrophil leukocyte to accumulate to the injury area, expand local inflammation, and increase the damage of brain tissue. After cerebral ischemia-reperfusion is injured, NO will be overproduced and lead to the death of a large number of neuronal and brain damage (Liu et al., 2016; Sarfraz et al., 2017). Therefore, the determination of NO and TNF- α can reflect the protection of cerebral neurotoxicity and the inhibition of inflammatory reaction in cerebral ischemia-reperfusion injury.

Through observing the effect of different component ratio of ATS and VTG on the neurological deficit scores, mortality, TNF- α content, NO content, and SOD activity of focal cerebral ischemia-reperfusion rat model, the experiment indicates that different component ratio of ATS:VTG can protect the focal cerebral ischemia-reperfusion rat model to a certain extent. By using the comprehensive weight method to evaluate the efficacy of each component, the results demonstrate that 5:5 group has the optimal active ratio.

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