The potential adverse effects of haemolysis

Francesca Rapido

Department of Anaesthesia and Critical Care, "Fondazione IRCCS Istituto Nazionale dei Tumori", Milan, Italy

Abstract

Haemolysis occurs in many haematologic and nonhaematologic diseases. Transfusion of packed red blood cells (pRBCs) can result in intravascular haemolysis, in which the RBCs are destroyed within the circulation, and extravascular haemolysis, in which RBCs are phagocytosed in the monocyte-macrophage system. This happens especially after RBCs have been stored under refrigerated conditions for long periods. The clinical implications and the relative contribution of intra- *vs* extra-vascular haemolysis are still a subject of debate. They have been associated with adverse effects in animal models, but it remains to be determined whether these may be involved in mediating adverse effects in humans.

Keywords: intravascular haemolysis, extravascular haemolysis, red blood cells, transfusion, blood storage.

Haemolysis occurs in many haematologic and nonhaematologic diseases and can be defined as the removal of senescent or damaged red blood cells (RBCs) from the circulation¹. Haemolysis also occurs after transfusion of stored blood. In particular, there is increasing evidence to suggest that increasing the storage period between blood donation and transfusion results in a decrease in RBC recovery and consequently an increase in posttransfusional haemolysis^{2,3}. However, the storage timerelated adverse effects and the potential mechanisms associated with transfusion-related toxicity remain controversial, and the relative contribution of intra- *vs* extra-vascular haemolysis is still under discussion. after RBCs have been stored under

compounds typically compartmen

ons for long periods. The clinical

such as haemoglobin and haeme,

relative contribution of intra-vs criteral and the state clinical end

colded with adv

In fact, haemolysis can be distinguished as intravascular haemolysis, in which the RBCs are destroyed within the circulation and release free haemoglobin (Hb) and RBC contents into the bloodstream, and extravascular haemolysis, in which RBCs are phagocytosed in the monocyte-macrophage system of organs such as the liver and the spleen^{4,5}. To the extent that clinically-relevant adverse effects of transfusions exist, it is likely that they are due to a combination of intra- and extra-vascular haemolysis, as shown by some animals models designed to describe the consequences of massive transfusions⁶. Of course, the complex biochemical and structural changes occurring during blood storage, and generally referred to as the Storage Lesion, can also contribute to these effects.

Intravascular haemolysis

The primary acute pathophysiological responses to extracellular Hb in plasma are increased blood pressure⁷ and pro-oxidative toxicity occurring in vascular and renal tissues^{6,8}. Pulmonary arterial pressure (PAP) was also observed to increase after exposure to free Hb9,10. During intravascular haemolysis, some toxic compounds typically compartmentalised within RBCs, such as haemoglobin and haeme, are released into the circulation. The adverse clinical effects associated with intravascular haemolysis are thought to be caused by: 1) extravascular translocation of haemoglobin and other RBC content; 2) imbalance between nitric oxide (NO) and reactive oxygen species (ROS); 3) platelet and haemostatic activation; and (4) haeme, haemoglobin and ATP-mediated activation of the innate immune system. The scavenger systems to limit the toxicity of the RBC contents include soluble plasma proteins, among which haptoglobin and haemopexin are considered to be the most important.

The first line of defence is haptoglobin, which irreversibly binds to the released haemoglobin. The resulting complex is rapidly cleared from the circulation via receptor-mediated endocytosis (CD-163 scavenger receptor $11,12$) and degraded in the liver, leading to a reduction in plasma haptoglobin. Cell-free plasma haemoglobin may overwhelm this scavenger system causing an intensified consumption of the endogenous NO and the formation of methemoglobin, which releases free haeme. Haemopexin, an acute phase protein primarily expressed in the liver, binds haeme and in addition this complex is removed by receptor-mediated endocytosis.

Various adverse effects, such as vascular dysfunction, injury, and inflammation, can be caused by the presence of free haemoglobin and free haeme in the circulation¹³. The first mechanism causing these effects is the imbalance between NO, a critical regulator of vasodilation and vascular homeostasis, and ROS. NO produced by endothelium and oxyhaemoglobin can quickly and irreversibly react, but this process is usually limited by compartmentalisation of haemoglobin inside the erythrocyte. During intravascular haemolysis, haemoglobin circulates in vessels free or in small microvesicles that can react faster with NO via the NO deoxygenation reaction and iron nitrosylation

reactions, as shown in some animal models 7,14 . In particular, it has been shown that more than 0.01 g/dL of free haemoglobin in plasma can potently inhibit NO-dependent vasodilation *in vivo*^{15,16}. The decrease in NO availability during intravascular haemolysis can also be due to other mechanisms. Free haeme can cause NO consumption and vasoconstriction by increasing adhesion molecule expression and endothelial activation, serving as a pro-inflammatory ligand of innate immune receptors (e.g., TLR4). This process also promotes inflammatory cell recruitment, platelet aggregation, and oxidation of low-density lipoprotein $17-20$. In addition, during RBC haemolysis significant concentrations of the enzyme arginase 1 are released into the circulation. Arginase 1 can metabolise L-arginine to ornithine, reducing the available L-arginine which is required for NO synthesis by the endothelial NOS (NO synthase) enzyme.

Therefore, during intravascular haemolysis, low levels of decompartmentalised or cell-free plasma Hb can impair NO signalling, reducing its bioavailability and producing vasomotor instability, endothelial dysfunction and systemic vasocontriction that clinically results in an increase in systemic vascular resistance and, as a consequence, a rising systolic, diastolic and mean arterial blood pressure with a decrease in or either unchanged cardiac output $8,9,15$, and a decreased perfusion to some organ systems, such as kidneys 21 . NO supplementation before free Hb exposure seems to attenuate these phenomena and the consequent clinical effects^{9,22}.

An increase in ROS production is also observed during haemolysis. In fact, free Hb auto-oxidises to methemoglobin and participates in a catalytic pseudoperoxidase cycle producing ROS. Haeme, which contains iron, is also responsible for the production of ROS through the Fenton reaction and by other distinct signalling pathways^{23,24}.

During intravascular haemolysis, platelet and haemostatic activation can occur. *In vitro* experiments demonstrate that NO inhibits both platelet aggregation and endothelial adhesion molecule expression. Thus, during intravascular haemolysis, the acute reduction in NO bioavailability can lead to the activation of platelets and the haemostatic system25,26. Furthermore, NO may affect coagulation by inhibiting Factor XIII, enhancing clot stability and reducing clot dissolution²⁷. Finally, RBCs contain high levels of ADP, the release of which can activate platelets via the P2Y receptors²⁸.

As mentioned before, haeme and haemoglobin can mediate the activation of the innate immune system causing macrophage and neutrophil migration to the lung and the release of DNA neutrophil extracellular traps (NETs)29-31 within the lung parenchyma. This process induces activation of inflammation and

thrombosis, through endothelial activation, RBC and activated platelet recruitment, and fibrin deposition. Haeme may also trigger pro-inflammatory and prothrombotic pathways through the stimulation of macrophage and endothelial cell toll-like receptor 4 (TLR)-4, involving Weibel-Palade body degranulation and nuclear factor-kappa B (NF- κ B) activation^{18,19}. Finally, intravascular haemolysis leads to ATP release, which can activate inflammatory pathways leading to sterile inflammation³². Therefore, intravascular release of RBC content after transfusion of older stored blood could contribute to cardiovascular and renal dysfunction, as well as inflammation, thrombosis, and enhanced susceptibility to infection, in severely ill patients.

Extravascular haemolysis

Damaged or aged RBCs accumulate over time within stored blood bags. Some degree of acute haemolysis occurs after transfusion through phagocytosis by the macrophage-monocyte system of the liver or spleen. This process is called extravascular haemolysis, and it classically occurs to eliminate senescent circulating RBCs displaying surface markers that identify them as cells requiring removal. During extravascular haemolysis, the RBC content is not found in plasma because the cell is lysed inside the macrophage. The degradation products deriving from this process are salvaged and recycled. In particular, the iron derived from haemoglobin is either stored intracellularly in ferritin deposits or returned to the plasma to be bound by transferrin and transported to the erythroid marrow for erythropoiesis and to other tissues for re-use. In circulation, iron (Fe $3+$) is carried by transferrin, which binds it with high affinity and renders it unable to react with ROS and other substances. Furthermore, at a steady state, the rate of RBC destruction is equal to the rate of red cell production, generating an equilibrium between waste production and re-use. However, this process is intensified after transfusion, when an average of up to 25% of the transfused RBCs can be cleared from the circulation according to regulatory agency criteria for blood storage³. The majority of the storage-damaged RBCs are cleared from the circulation very rapidly (within the first hour after transfusion³³), causing an excessive rate of delivery of haeme-iron to reticuloendothelial macrophages. Consequently, the rate of release of iron into the circulation can surpass the rate of uptake by transferrin, producing circulating non-transferrin-bound iron (NTBI)^{3,34}. NTBI is a heterogeneous group of iron complexes, mainly Fe³⁺⁻ citrate or albumin complexes, which is considered potentially toxic. A fraction of NTBI, known as labile plasma iron (LPI), is very loosely bound to proteins and is highly redox active, and is probably the main cause Extravelaxional MOS (NO synthase)

Entravascular haemolysis

endothelial NOS (NO synthase)

tored blood bags. Some degree of

intravascular haemolysis, low

occurs after transfusion through ph

entalised or cell-free plas

of iron-mediated oxidative damage^{35,36}. NTBI and LPI can also enter many cell types, such as liver, pancreas, endocrine glands cells and cardiomyocytes by nontransferrin dependent pathways, resulting in increased labile intracellular iron $(LIC)^{37}$, a highly reactive form of iron. LIC can generate ROS from reactive oxygen intermediates, over-riding the cell antioxidant defences and compromising cell integrity and causing organ damage and failure. Under normal conditions, NTBI and LPI should not be found in plasma. However, NTBI can be detected in the plasma as soon as transferrin becomes more saturated³⁸, and rises significantly when transferrin saturation exceeds 70-80%³⁸⁻⁴⁰.

The full implications of the increased extravascular haemolysis after transfusion of stored blood, regardless of the chronic or acute nature of the transfusions, remain to be determined. However, animal studies using mice⁴¹ and dogs⁴² suggest that there is a proinflammatory response following transfusion of older, stored RBCs. This can exacerbate an underlying systemic inflammatory response syndrome $(SIRS)^{41}$, increase alloimmunogenicity to RBC antigens⁴³, and enhances proliferation of certain pathogens^{3,44,45}. Thus, as expected, multiple observational studies have suggested an association between transfusion of RBC stored for longer durations and worse clinical outcomes⁴⁶ (e.g., increases in sepsis, pneumonia, multi-organ failure, myocardial infarction, acute renal failure, thrombosis, and mortality). However, these studies have significant flaws, mainly owing to the difficulty in disentangling the contribution of the age of the RBCs from the increased underlying disease severity in patients receiving more, and therefore older, units of RBCs. Thus, despite the completed and ongoing controlled trials designed to address these questions, the issue of whether transfusion of RBCs stored for a prolonged period is harmful is still controversial. active nature of the transfusions,

anarophage seavenger receptor fo

mariophage seavenger receptor fo

members in the absent

maripole hemoglobis in the absent

scale of the state that the ris a pro-

scale did since the

The Author declares no conflicts of interest.

References

- 1) Dhaliwal G, Cornett PA, Tierney LM Jr. Hemolytic anemia. Am Fam Physician 2004; **69**: 2599-606.
- 2) Rapido F, Bandyopadhyay S, La Carpia F, et al. Prolonged red cell storage before transfusion increases extravascular hemolysis. J Clin Invest 2017; **127**: 375-82.
- 3) Hod EA, Brittenham GM, Billote GB, et al. Transfusion of human volunteers with older, stored red blood cells produces extravascular hemolysis and circulating non-transferrin-bound iron. Blood 2011; **118**: 6675-82.
- 4) Cazzola M, Beguin Y. New tools for clinical evaluation of erythron function in man. Br J Haematol 1992; **80**: 278-84.
- 5) Bossi D, Giardina B. Red cell physiology. Mol Aspects Med 1996; **17**: 117-28.
- 6) Baek JH, D'Agnillo F, Vallelian F, et al. Hemoglobin-driven pathophysiology is an in vivo consequence of the red blood cell storage lesion that can be attenuated in guinea pigs by haptoglobin therapy. J Clin Invest 2012; **122**: 1444-58.
- 7) Rother RP, Bell L, Hillmen P, Gladwin MT. The clinical sequelae of intravascular hemolysis and extracellular plasma hemoglobin: a novel mechanism of human disease. JAMA 2005; **293**: 1653-62.
- 8) Boretti FS, Buehler PW, D'Agnillo F, et al. Sequestration of extracellular hemoglobin within a haptoglobin complex decreases its hypertensive and oxidative effects in dogs and guinea pigs. J Clin Invest 2009; **119**: 2271-80.
- 9) Minneci PC, Deans KJ, Zhi H, et al. Hemolysis-associated endothelial dysfunction mediated by accelerated NO inactivation by decompartmentalized oxyhemoglobin. J Clin Invest 2005; **115**: 3409-17.
- 10) Berra L, Pinciroli R, Stowell CP, et al. Autologous transfusion of stored red blood cells increases pulmonary artery pressure. Am J Respir Crit Care Med 2014; **190**: 800-7.
- 11) Kristiansen M, Graversen JH, Jacobsen C, et al. Identification of the haemoglobin scavenger receptor. Nature 2001; **409**: 198-201.
- 12) Schaer DJ, Schaer CA, Buehler PW, et al. CD163 is the macrophage scavenger receptor for native and chemically modified hemoglobins in the absence of haptoglobin. Blood 2006; **107**: 373-80.
- 13) Potoka KP, Gladwin MT. Vasculopathy and pulmonary hypertension in sickle cell disease. Am J Physiol Lung Cell Mol Physiol 2015; **308**: L314-24.
- 14) Jeney V, Balla G, Balla J. Red blood cell, hemoglobin and heme in the progression of atherosclerosis. Front Physiol 2014; **5**: 379.
- 15) Reiter CD, Wang X, Tanus-Santos JE, et al. Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease. Nat Med 2002; **8**: 1383-9.
- 16) Pohl U, Lamontagne D. Impaired tissue perfusion after inhibition of endothelium-derived nitric oxide. Basic Res Cardiol 1991; **86**: 97-105.
- 17) Belcher JD, Nath KA, Vercellotti GM. Vasculotoxic and proinflammatory effects of plasma heme: cell signaling and cytoprotective responses. ISRN Oxidative Med 2013; **2013**: pii: 831596.
- 18) Belcher JD, Nguyen J, Chen CS, et al. Plasma hemoglobin and heme trigger Weibel Palade body exocytosis and vasoocclusion in transgenic sickle mice. Blood 2011; **118**: 896.
- 19) Palsson-McDermott EM, O'Neill LA. Signal transduction by the lipopolysaccharide receptor, Toll-like receptor-4. Immunology 2004; **113**: 153-62.
- 20) Valentijn KM, Sadler JE, Valentijn JA, et al. Functional architecture of Weibel-Palade bodies. Blood 2011; **117**: 5033-43.
- 21) Schaer DJ, Buehler PW. Cell-free hemoglobin and its scavenger proteins: new disease models leading the way to targeted therapies. Cold Spring Harb Perspect Med 2013; **3**: 6.
- 22) Yu B, Volpato GP, Chang K, et al. Prevention of the pulmonary vasoconstrictor effects of HBOC-201 in awake lambs by continuously breathing nitric oxide. Anesthesiology 2009; **110**: 11322.
- 23) Prousek J. Fenton chemistry in biology and medicine. Pure Appl Chem 2007; **79**: 13.
- 24) Goldstein S, Meyerstein D, Czapski G. The Fenton reagents. Free Radic Biol Med 1993; **15**: 435-45.
- 25) Hu W, Jin R, Zhang J, et al. The critical roles of platelet activation and reduced NO bioavailability in fatal pulmonary arterial hypertension in a murine hemolysis model. Blood 2010; **116**: 1613-22.
- 26) Patel RP, McAndrew J, Sellak H, et al. Biological aspects of reactive nitrogen species. Biochim Biophys Acta 1999; **1411**: 385-400.
- 27) Gries A, Bode C, Peter K, et al. Inhaled nitric oxide inhibits human platelet aggregation, P-selectin expression, and fibrinogen binding in vitro and in vivo. Circulation 1998; **97**: 1481-7.
- 28) Dutra FF, Bozza MT. Heme on innate immunity and inflammation. Front Pharmacol 2014; **5**: 115.
- 29) Chen G, Zhang D, Fuchs TA, et al. Heme-induced neutrophil extracellular traps contribute to the pathogenesis of sickle cell disease. Blood 2014; **123**: 3818-27.
- 30) Graça-Souza AV, Arruda MA, de Freitas MS, et al. Neutrophil activation by heme: implications for inflammatory processes. Blood 2002; **99**: 4160-5.
- 31) Porto BN, Alves LS, Fernandez PL, et al. Heme induces neutrophil migration and reactive oxygen species generation through signaling pathways characteristic of chemotactic receptors. J Biol Chem 2007; **282**: 24430-6.
- 32) Belcher JD, Chen C, Nguyen J, et al. Heme triggers TLR4 signaling leading to endothelial cell activation and vasoocclusion in murine sickle cell disease. Blood.2014; **123**: 377-90.
- 33) Luten M, Roerdinkholder-Stoelwinder B, Schaap NP, et al. Survival of red blood cells after transfusion: a comparison between red cells concentrates of different storage periods. Transfusion 2008; **48**: 1478-85.
- 34) Stark MJ, Keir AK, Andersen CC. Does non-transferrin bound iron contribute to transfusion related immune-modulation in preterms? Archives of Disease in Childhood. Fetal and Neonatal Edition 2013; **98**: F424-9.
- 35) Esposito BP, Breuer W, Sirankapracha P, et al. Labile plasma iron in iron overload: redox activity and susceptibility to chelation. Blood 2003; **102**: 2670-7.
- 36) Cabantchik ZI, Sohn YS, Breuer W, Esposito BP. The molecular and cellular basis of iron toxicity in iron overload disorders. Diagnostic and therapeutic approaches. Thalassemia Reports 2013; **3**: 7-13.
- 37) Hider RC, Kong XL. Glutathione: a key component of the cytoplasmic labile iron pool. Biometals 2011; **24**: 1179-87.
- 38) Sahlstedt L, Ebeling F, von Bonsdorff L, et al. Non-transferrinbound iron during allogeneic stem cell transplantation. Br J Haematol 2001; **113**: 836-8.
- 39) Bradley SJ, Gosriwitana I, Srichairatanakool S, et al. Non-transferrin-bound iron induced by myeloablative chemotherapy. Br J Haematol 1997; **99**: 337-43.
- 40) Roob JM, Khoschsorur G, Tiran A, et al. Vitamin E attenuates oxidative stress induced by intravenous iron in patients on hemodialysis. J Am Soc Nephrol 2000; **11**: 539-49.
- 41) Hod EA, Zhang N, Sokol SA, et al. Transfusion of red blood cells after prolonged storage produces harmful effects that are mediated by iron and inflammation. Blood 2010; **115**: 4284-92.
- 42) Callan MB, Patel RT, Rux AH, et al. Transfusion of 28-dayold leucoreduced or non-leucoreduced stored red blood cells induces an inflammatory response in healthy dogs. Vox Sang 2013; **105**: 319-27.
- 43) Hendrickson JE, Hod EA, Spitalnik SL, et al. Storage of murine red blood cells enhances alloantibody responses to an erythroid-specific model antigen. Transfusion 2010; **50**: 642-8.
- Prestia K, Bandyopadhyay S, Slate A, et al. Transfusion of stored blood impairs host defenses against Gram-negative pathogens in mice. Transfusion 2014; **54**: 2842-51.
- 45) Solomon SB, Wang D, Sun J, et al. Mortality increases after massive exchange transfusion with older stored blood in canines with experimental pneumonia. Blood 2013; **121**: 1663-72.
- 46) Wang D, Sun J, Solomon SB, Klein HG, Natanson C. Transfusion of older stored blood and risk of death: a metaanalysis. Transfusion 2012; **52**: 1184-95.

Arrived: 8 November 2016 - Revision accepted: 13 December 2016 **Correspondence:** Francesca Rapido Dipartimento di Anestesia e Rianimazione Fondazione IRCCS Istituto Nazionale dei Tumori Via G. Veneziano 1 20133 Milano, Italy e-mail: francesca.rapido@gmail.com neutrates or contract storage periods.

Carameteric software in the system experimental preumonine with experimental preumonine in the Warg D. Sum J, Solomon SB. Klein Start and Transfission of older stored blood and

anal