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Estrogen receptor- α gene haplotype is associated with primary knee osteoarthritis in Korean population

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Abstract

Estrogen and estrogen receptors (ERs) are known to play important roles in the pathophysiology of osteoarthritis (OA). To investigate ER- α gene polymorphisms for its associations with primary knee OA, we conducted a case-control association study in patients with primary knee OA ($n = 151$) and healthy individuals ($n = 397$) in the Korean population. Haplotyping analysis was used to determine the relationship between three polymorphisms in the ER- α gene (intron 1 T/C, intron 1 A/G and exon 8 G/A) and primary knee OA. Genotypes of the ER- α gene polymorphism were determined by PCR followed by restriction enzyme digestion (*PvuII* for intron 1 T/C, *XbaI* for intron 1 A/G, and *BtgI* for exon 8 G/A polymorphism). There was no significant difference between primary knee OA patients and

healthy control individuals in the distribution of any of the genotypes evaluated. However, we found that the allele frequency for the exon 8 G/A *BtgI* polymorphism (codon 594) was significantly different between primary knee OA patients and control individuals (odds ratio = 1.38, 95% confidence interval = 1.01–1.88; $P = 0.044$). In haplotype frequency estimation analysis, there was a significant difference between primary knee OA patients and control individuals (degrees of freedom = 7, $\chi^2 = 21.48$; $P = 0.003$). Although the number OA patients studied is small, the present study shows that ER- α gene haplotype may be associated with primary knee OA, and genetic variations in the ER- α gene may be involved in OA.

Keywords: estrogen receptor, haplotype, knee osteoarthritis, polymorphism

Introduction

Osteoarthritis (OA) is a common disorder among the elderly. It is multifactorial disorder, with predisposing factors included advanced age, and genetic, hormonal and mechanical factors [1,2]. Furthermore, recent studies have revealed a role for the inflammatory process in the pathogenesis of OA [3-5]. It has been reported that the development of OA may be influenced by multiple genes [6]. A number of candidate genes have been suggested to mediate susceptibility to OA, including collagen genes (*COL1A1*, *COL2A1*, *COL9A1*, *COL11A2*), and the genes encoding cartilage matrix protein 1 (*CMP1*), vitamin D receptor (*VDR*), insulin-like growth factor-1 (*IGF1*), transforming growth factor- β_1 (*TGF β 1*), aggrecan-1 (*AGC1*), tis-

sue inhibitor of metalloproteinase 3 (*TIMP3*), interleukin-1 receptor (*IL1R*), and the estrogen receptor [6].

The human estrogen receptor (ER) has two isoforms: ER- α and ER- β . The former isoform is a ligand-activated transcription factor composed of several important domains for hormone binding, DNA binding, and activation of transcription [7]. ER- α is an important mediator in the signal transduction pathway [7]. The ER is a member of the steroid/thyroid hormone superfamily of nuclear receptors [8]. The ER- α gene is greater than 140 kilobases, contains eight exons, and is located on chromosome 6q25 [9]. The coding region has a length of 1785 nucleotides, and it is translated into a protein of 595 amino acids and 66 kDa [10].

Several variations in the DNA sequence of the ER- α gene have been reported [11,12]. A few reports have examined the association between ER- α gene polymorphisms and OA; the findings are controversial. Ushiyama and co-workers [13] reported associations between a genotype of *PvuII* and *XbaI* polymorphisms in intron 1 of the ER- α gene and generalized OA. Bergink and co-workers [14] found that an ER- α haplotype of *PvuII* and *XbaI* polymorphisms was associated with radiographic OA of the knee. However, Loughlin and co-workers [15] found no association between ER- α gene polymorphisms and idiopathic OA. Some studies of *BtgI* polymorphisms have been conducted in other diseases. Cancel-Tassin and co-workers [16] reported that a *BtgI* (594 A/G) polymorphism was not associated with risk for prostate cancer. Tanaka and colleagues [17] reported similar findings in prostate cancer patients. In another study, the *BtgI* (594 A/G) polymorphism was not associated with renal cell carcinoma [18]. Curran and co-workers [19] reported an association of A/G polymorphism in exon 8 of the ER gene with sporadic breast cancer. However, no findings regarding the relationship between exon 8 A/G polymorphism and OA have been reported.

In the present study, we first analyzed the association between the exon 8 G/A *BtgI* polymorphism (database-single nucleotide polymorphism number rs2228480; codon 594) of the ER- α gene and primary knee OA in patients from the Korean population. We also investigated the association between haplotypes of three polymorphisms (*PvuII* in intron 1, *XbaI* in intron 1, and *BtgI* in exon 8) and primary knee OA.

Materials and methods

Study subjects

A total of 151 patients with primary knee OA (98 women and 53 men) from the Korean population were included in

the study. Patients were examined at the Rheumatology Clinic in Kyung Hee University Medical Center, Seoul, Korea. In total, 397 control individuals (207 women and 190 men) underwent the 2003 health examination. OA was diagnosed according to American College of Rheumatology criteria, which include primary OA with any symptom and/or sign of OA, positive finding on radiographs according to the Kellgren-Lawrence grading [20], and no evidence of arthritis due to other disease. The study was approved by the ethics review committee of the Medical Research Institute, Kyung Hee University Medical Center, Seoul, Korea.

Clinical information and classification of primary knee osteoarthritis The age at onset of OA is important clinically because it is associated with long-term prognosis. During the study, we were able to identify the current age and age at onset of disease in the 151 knee OA patients by individual interview conducted at our outpatient clinic. The current age of OA patients (mean \pm standard deviation) was 58.8 \pm 9.6 years, and the age at onset of OA was 52.0 \pm 9.5 years. We therefore stratified clinical data according to mean onset age of disease: older or younger than 52.0 years. The group with disease onset at age <52.0 years was defined as the 'early' onset, and the group of disease onset at age >52.0 years was defined as the 'late' onset group.

The Kellgren-Lawrence grade represents disease severity, as reflected on radiographs, and Lequesne's functional index represents functional or symptomatic status of patients [21]. Radiographic findings of OA were classified into mild (Kellgren-Lawrence grade 1 or 2) or severe (Kellgren-Lawrence grade 3 or 4). The functional or symptomatic status of OA patients was classified as functionally or symptomatically good (Lequesne's functional index = 10) or poor (Lequesne's functional index > 10; Table 1).

Table 1

Clinical characteristics of the patients with primary knee osteoarthritis patients

Characteristic	Findings
Age (years)	58.8 \pm 9.6
Number women/men (n)	98/53
Age at onset age (years)	52.0 \pm 9.5
Body mass index (kg/m ²)	25.2 \pm 2.9
Duration of osteoarthritis (years)	6.8 \pm 5.9
Kellgren-Lawrence grade (n): 1/2/3/4	6/84/56/5
Lequesne's index	10.2 \pm 2.5

A total of 151 patients with primary knee osteoarthritis were included. Values are expressed as mean \pm standard deviation or as numbers.

Table 2**Sequences of primers used for estrogen receptor- α genotyping**

Polymorphism site	Primer sets	Annealing temperature (°C)	Restriction enzyme	Allele size (bp)
Intron 1 T/C	5'-ctgccaccctatctgtatcttttctattctcc-3' 5'-tctttctctgccaccctggcgtcgattatctga-3'	64	<i>PvuII</i>	T: 936 + 438 C: 1374
Intron 1 A/G	5'-ctgccaccctatctgtatcttttctattctcc-3' 5'-tctttctctgccaccctggcgtcgattatctga-3'	64	<i>XbaI</i>	A: 981 + 393 G: 1374
Exon 8 G/A	5'-gaggagacggaccaagccac-3' 5'-gccattggtgttgatgatg-3'	63	<i>BtgI</i>	G: 129 + 98 A: 227

bp, base pairs.

Analysis of estrogen receptor- α gene polymorphisms

Genomic DNA was prepared from whole blood samples using the NucleoSpin DNA isolation kit (Macherey-Nagel GmbH & Co., Düren, Germany). PCR amplifications were performed using 50 ng genomic DNA in a 30 μ l reaction volume containing 0.5 μ l 10 pmol sense primer, 0.5 μ l 10 pmol antisense primer, 0.5 μ l 2.5 mmol/l dNTP (Takara, Shiga, Japan), 1 U Taq DNA polymerase (Neurotics Inc., Seoul, Korea), and in buffer containing 25 mmol/l MgCl₂, 750 mmol/l Tris-HCl (pH 9.0), 150 mmol/l ammonium sulfate, and 1 mg/ml bovine serum albumin. Samples were subjected to 35 cycles of amplification in GeneAmp PCR system 2700 (Applied Biosystems, Foster, CA, USA). PCR primers used in the study are listed in Table 2 [19,22]. The PCR products were digested under the conditions specified by the enzyme supplier (New England Biolabs Inc, Beverly, MA, USA). Restriction fragments were separated by agarose gel electrophoresis and ethidium bromide staining.

Statistical analysis

For the case-control association study, the significance of differences in allelic and genotypic frequencies between OA patients and control populations was determined using standard χ^2 tests. We used the EH program [23] to investigate the relative risks associated with haplotypes. $P < 0.05$ was considered statistically significant.

Results**Distribution of estrogen receptor- α genotypes in osteoarthritis patients and control individuals**

As shown in Table 3, three single nucleotide polymorphisms were identified in ER- α intron 1 (T/C, A/G) and exon 8 (G/A). When the allele frequency of the exon 8 G/A *BtgI* polymorphism was compared between OA patients and control individuals, a significant difference was observed (odds ratio [OR] = 1.38, 95% confidence interval [CI] = 1.01–1.88; $P = 0.044$). However, the genotype distribution did not exhibit a significant difference between OA patients and control individuals ($P = 0.13$). Observed genotype and allele frequencies for the intron 1 T/C *PvuII* and the intron 1 A/G *XbaI* polymorphisms were not significantly different between OA patients and control individuals (Table 3). There was no evidence of deviation from Hardy-

Weinberg equilibrium in healthy control individuals (for intron 1 T/C *PvuII* polymorphism, $P = 0.85$; for intron 1 A/G *XbaI* polymorphism, $P = 0.99$; and for exon 8 G/A *BtgI* polymorphism, $P = 0.77$).

Estrogen receptor- α gene polymorphisms and sex of osteoarthritis patients

As shown in Table 4, the observed genotype distribution ($P = 0.04$) and allele frequency (OR = 1.89, 95% CI = 1.15–3.11; $P = 0.01$) for the exon 8 G/A *BtgI* polymorphism were significantly different between male OA patients and male control individuals, whereas those in females exhibited no significant difference. We compared genotype distributions and allele frequencies for the intron 1 T/C *PvuII* and the intron 1 A/G *XbaI* polymorphisms. There were no significant differences in the polymorphisms between OA patients and controls of the same sex.

Estrogen receptor- α gene polymorphisms and risk for late onset osteoarthritis

Late onset OA patients were defined as those with disease onset at age above 52 years, whereas early onset OA patients had their disease onset at age under 52 years. When comparing allele frequency of the exon 8 G/A *BtgI* polymorphism between late onset OA patients and control individuals, a significant difference was observed (OR = 1.62, 95% CI = 1.07–2.46; $P = 0.021$). However, the genotype distribution did not exhibit a significant difference between OA patients and control individuals ($P = 0.06$). The genotype distributions and allele frequencies of the intron 1 T/C *PvuII* and the intron 1 A/G *XbaI* polymorphisms were not significantly different between late onset OA patients and control individuals. When comparing genotype distributions and allele frequencies of the intron 1 T/C *PvuII*, the intron 1 A/G *XbaI*, and the exon 8 G/A *BtgI* polymorphisms between early onset OA patients and control individuals, no significant difference was observed (Table 4).

Estrogen receptor- α gene polymorphisms and risk for radiographically severe osteoarthritis

Patients with radiographically severe OA were defined as those whose Kellgren-Lawrence grade was 3 or 4, whereas

Table 3**Genotype distribution and allele frequency of estrogen receptor- α gene polymorphisms in patients with osteoarthritis and control individuals**

Groups	ER- α genotypes			ER- α alleles					
		OA (%)	Control (%)	P^a	OA (%)	Control (%)	P^b	OR (95% CI)	
<i>PvuII</i> (T/C)	TT	61 (40.4)	152 (38.3)	0.89	T	190 (62.9)	487 (61.3)	0.63	0.93 (0.71–1.23)
	CT	68 (45.0)	183 (46.1)		C	112 (37.1)	307 (38.7)		
	CC	22 (14.6)	62 (15.6)						
<i>XbaI</i> (A/G)	AA	98 (64.9)	256 (64.5)	0.81	A	245 (81.1)	638 (80.3)	0.77	0.95 (0.68–1.33)
	AG	49 (32.4)	126 (31.7)		G	57 (18.9)	156 (19.7)		
	GG	4 (2.70)	15 (3.80)						
<i>BtgI</i> (G/A)	GG	84 (55.6)	257 (64.7)	0.13	G	225 (74.5)	636 (80.1)	0.044	1.38 (1.01–1.88)
	GA	57 (37.8)	122 (30.7)		A	77 (25.5)	158 (19.9)		
	AA	10 (6.60)	18 (4.60)						

A total of 151 patients with osteoarthritis (OA) and 397 control individuals were included in the study. ^aControl individuals versus patients using the χ^2 test with 3×2 contingency table. ^bControl individuals versus patients using the χ^2 test with 2×2 contingency table. CI, confidence interval; ER, estrogen receptor; OR, odds ratio.

radiographically mild OA was defined as Kellgren-Lawrence grade 1 or 2. When comparing genotype distributions and allele frequencies of the intron 1 T/C *PvuII*, the intron 1 A/G *XbaI*, and the exon 8 G/A *BtgI* polymorphisms between patients with mild OA and control individuals, no significant difference was observed (Table 4). The genotype distributions and allele frequencies of the intron 1 T/C *PvuII*, the intron 1 A/G *XbaI*, and the exon 8 G/A *BtgI* polymorphisms did not exhibit significant differences between patients with severe OA and control individuals (Table 4).

Estrogen receptor- α gene polymorphisms and risk for functionally poor osteoarthritis

OA patients who were functionally or symptomatically poor (poor index) were defined as those who with a Lequesne's functional index score over 10, whereas those who were functionally or symptomatically good (good index) had a Lequesne's functional index score less than or equal to 10. Genotype distributions and allele frequencies of the intron 1 T/C *PvuII*, the intron 1 A/G *XbaI*, and the exon 8 G/A *BtgI* polymorphisms between OA patients with a good and those with a poor index, and control individuals were not significantly different (Table 4).

Estrogen receptor- α haplotype analysis in patients with primary knee osteoarthritis

Table 5 shows the frequency of each haplotype. The difference was significant between all OA patients combined and control individuals (degrees of freedom [df] = 7, $\chi^2 =$

21.48; $P = 0.003$). There was no significant difference between female patients and female control individuals (df = 7, $\chi^2 = 8.22$; $P = 0.31$). However, there was a significant difference between male OA patients and male control individuals (df = 7, $\chi^2 = 16.76$; $P = 0.019$; Table 5).

The late onset, radiographically severe, and poor index subgroups of OA patients exhibited significant differences in haplotype distribution (Table 6). There was a significant difference between patients with late onset OA and control individuals (df = 7, $\chi^2 = 21.96$; $P = 0.002$) but not between patients with early onset OA and control individuals (df = 7, $\chi^2 = 7.42$; $P = 0.390$). When comparing patients with radiographically severe OA and control individuals, a significant difference was observed (df = 7, $\chi^2 = 23.96$; $P = 0.001$), but this was not the case in patients with radiographically mild OA (df = 7, $\chi^2 = 13.60$; $P = 0.059$). There was a significant difference between OA patients with a poor index and control individuals (df = 7, $\chi^2 = 14.66$; $P = 0.041$), but this was not the case for OA patients with a good index (df = 7, $\chi^2 = 10.96$; $P = 0.140$; Table 6).

Discussion

We report here, for the first time, on the associations of exon 8 G/A *BtgI* polymorphism (codon 594) in the ER- α gene and ER- α haplotypes of three polymorphisms (*PvuII* in intron 1, *XbaI* in intron 1, and *BtgI* in exon 8) with primary knee OA in the Korean population. Several reports [13–15,24–26] have indicated that estrogen and its receptor

Table 4**Comparison of estrogen receptor- α gene polymorphisms in subtypes of osteoarthritis patients and control individuals**

Clinical subtypes	Genotype distributions ^a									Allele frequencies ^b					
	<i>PvuII</i>			<i>XbaI</i>			<i>BtgI</i>			<i>PvuII</i>		<i>XbaI</i>		<i>BtgI</i>	
	TT	TC	CC	AA	AG	GG	GG	GA	AA	T	C	A	G	G	A
Women (n = 98)	43 (43.9)	42 (42.8)	13 (13.3)	64 (65.3)	33 (33.7)	1 (1.00)	57 (58.2)	35 (35.7)	6 (6.10)	128 (65.3)	68 (34.7)	161 (82.1)	35 (17.9)	149 (76.0)	47 (24.0)
	$\chi^2 = 3.84; P = 0.15$			$\chi^2 = 1.05; P = 0.59$			$\chi^2 = 0.23; P = 0.89$			1.15 (0.80–1.64); $P = 0.45$		0.91 (0.58–1.41); $P = 0.67$		1.10 (0.74–1.65); $P = 0.63$	
Men (n = 53)	18 (34.0)	26 (49.0)	9 (17.0)	34 (64.1)	16 (30.2)	3 (5.70)	27 (75.5)	22 (20.7)	4 (3.80)	62 (58.5)	44 (41.5)	84 (79.2)	22 (20.8)	76 (71.78)	30 (28.3)
	$\chi^2 = 1.69; P = 0.43$			$\chi^2 = 0.07; P = 0.96$			$\chi^2 = 6.28; P = 0.043$			0.82 (0.53–1.27); $P = 0.38$		1.04 (0.62–1.78); $P = 0.86$		1.89 (1.15–3.11); $P = 0.011$	
Early onset ^c (n = 85)	34 (40.0)	41 (48.2)	10 (11.8)	52 (61.2)	31 (36.5)	2 (2.30)	51 (60.0)	29 (34.1)	5 (5.90)	109 (64.1)	61 (35.9)	135 (79.4)	35 (20.6)	131 (77.1)	39 (22.9)
	$\chi^2 = 0.82; P = 0.66$			$\chi^2 = 1.00; P = 0.60$			$\chi^2 = 0.77; P = 0.68$			0.893 (0.63–1.25); $P = 0.50$		1.06 (0.70–1.60); $P = 0.78$		1.20 (0.80–1.78); $P = 0.37$	
Late onset ^c (n = 66)	27 (40.9)	27 (40.9)	12 (18.2)	46 (69.7)	18 (27.3)	2 (3.00)	33 (50.0)	28 (42.4)	5 (7.60)	81 (61.4)	51 (38.6)	110 (83.3)	22 (16.7)	94 (71.2)	38 (28.8)
	$\chi^2 = 0.67; P = 0.71$			$\chi^2 = 0.68; P = 0.71$			$\chi^2 = 5.40; P = 0.07$			1.00 (0.68–1.46); $P = 0.99$		1.82 (0.50–1.33); $P = 0.42$		1.63 (1.07–2.46); $P = 0.021$	
Mild ^d (n = 90)	44 (48.9)	33 (36.7)	13 (14.4)	67 (74.5)	21 (23.3)	2 (2.20)	50 (55.6)	33 (36.7)	7 (7.80)	121 (67.2)	59 (32.8)	155 (86.1)	25 (13.9)	133 (73.9)	47 (26.1)
	$\chi^2 = 3.58; P = 0.17$			$\chi^2 = 3.32; P = 0.19$			$\chi^2 = 0.30; P = 0.19$			0.77 (0.55–1.09); $P = 0.14$		0.66 (0.42–1.04); $P = 0.07$		1.42 (0.98–2.07); $P = 0.06$	
Severe ^d (n = 61)	17 (27.9)	35 (57.4)	9 (14.7)	31 (50.8)	28 (45.9)	2 (3.30)	34 (55.7)	24 (39.3)	3 (4.90)	69 (56.6)	53 (43.4)	90 (73.8)	32 (26.2)	92 (75.4)	30 (24.6)
	$\chi^2 = 2.99; P = 0.22$			$\chi^2 = 4.76; P = 0.09$			$\chi^2 = 1.92; P = 0.38$			1.22 (0.83–1.79); $P = 0.31$		1.45 (0.94–2.26); $P = 0.09$		1.31 (0.84–2.05); $P = 0.23$	
Good index ^e (n = 82)	32 (39.0)	35 (42.7)	15 (18.3)	52 (63.4)	28 (34.2)	2 (2.40)	45 (54.9)	31 (37.8)	6 (7.30)	99 (60.4)	65 (39.6)	132 (80.5)	32 (19.5)	121 (73.8)	43 (26.2)
	$\chi^2 = 0.68; P = 0.78$			$\chi^2 = 0.48; P = 0.78$			$\chi^2 = 3.16; P = 0.21$			1.04 (0.74–1.47); $P = 0.82$		0.99 (0.65–1.51); $P = 0.97$		1.43 (0.97–2.11); $P = 0.07$	
Poor index ^e (n = 69)	29 (42.0)	33 (47.8)	7 (10.2)	46 (66.7)	21 (30.4)	2 (2.90)	39 (56.5)	26 (37.7)	4 (5.80)	91 (65.9)	47 (34.1)	113 (81.9)	25 (18.1)	104 (75.4)	34 (24.6)
	$\chi^2 = 1.44; P = 0.49$			$\chi^2 = 0.20; P = 0.90$			$\chi^2 = 1.72; P = 0.42$			0.82 (0.56–1.20); $P = 0.30$		0.90 (0.57–1.44); $P = 0.67$		1.32 (0.86–2.01); $P = 0.20$	

^aControl versus patients using the χ^2 test with 3×2 contingency table. ^bControl versus patients using the χ^2 test with 2×2 contingency table. ^cEarly onset osteoarthritis OA patients were defined as those with disease onset at age under 52 years, whereas late onset OA patients were those with onset at age over 52 years. ^dBased on radiographic findings, OA patients were classified into mild (Kellgren-Lawrence grade 1 or 2) or severe (Kellgren-Lawrence grade 3 or 4). ^eBased on Lequesne's functional index score, poor OA patients were defined as those with an index score over 10, whereas good OA patients had an index score less than or equal to 10.

might be involved in the etiology of OA. Until now three reports on the relationship between ER- α polymorphisms and OA had been published. Two studies [13,14] reported that a ER- α polymorphism was associated with OA. Ushiyama and co-workers [13] found an association between a genotype of *PvuII* and *XbaI* polymorphisms in intron 1 and generalized OA with severe radiographic changes in the Japanese population (65 OA patients and 318 control individuals). In a population-based study conducted in a Caucasian population (1483 subjects), Bergink and co-workers [14] reported that ER- α haplotype of *PvuII* and *XbaI* polymorphisms was associated with radiographic OA of the knee. One study showed no relationship between

ER- α gene polymorphisms and OA in a Caucasian population (371 OA patients and 369 control individuals) [15].

Our study showed that the genotype distributions for the intron 1 T/C *PvuII* and the intron 1 A/G *XbaI* polymorphisms were not associated with OA, a finding similar to that reported by Loughlin and co-workers [15]. However, the allele frequency for the *BtgI* polymorphism was significantly different between OA and control individuals. The difference in the allele frequency was more marked in OA patients with late onset of disease and in male patients (Table 4). The haplotype of three polymorphisms was associated with OA. We conducted further analysis of OA

Table 5

Comparison of estrogen receptor- α gene haplotypes in osteoarthritis patients and control individuals

Haplotype	Controls			OA patients		
	Total	Women	Men	Total	Women	Men
TAG	0.41	0.44	0.37	0.41	0.42	0.40
TAA	0.09	0.12	0.05	0.17	0.18	0.14
TGG	0.09	0.09	0.08	0.05	0.05	0.05
TGA	0.03	0.03	0.02	<0.01	<0.01	<0.01
CAG	0.25	0.18	0.31	0.18	0.18	0.18
CAA	0.06	0.06	0.06	0.05	0.04	0.07
CGG	0.06	0.06	0.08	0.10	0.10	0.09
CGA	0.02	0.01	0.02	0.04	0.02	0.07
				df = 7, $\chi^2 = 21.48$; $P^a = 0.003$	df = 7, $\chi^2 = 8.22$; $P^a = 0.314$	df = 7, $\chi^2 = 16.76$; $P^a = 0.019$

A total of 397 control individuals and 151 osteoarthritis (OA) patients were included in the study. Values given are haplotype frequencies. ^a*P* values for overall difference in haplotype distribution between OA patients and control individuals were calculated using the EH program [23]. df, degrees of freedom.

Table 6

Comparison of estrogen receptor- α gene haplotypes in subtypes of osteoarthritis patients and control individuals

Haplotypes	Control	Early onset ^a	Late onset ^a	Mild ^b	Severe ^b	Good ^c	Poor ^c
TAG	0.41	0.41	0.41	0.43	0.39	0.39	0.44
TAA	0.09	0.16	0.18	0.18	0.15	0.16	0.17
TGG	0.09	0.07	0.02	0.06	0.02	0.05	0.05
TGA	0.03	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
CAG	0.25	0.19	0.17	0.19	0.16	0.19	0.17
CAA	0.06	0.04	0.07	0.06	0.04	0.06	0.04
CGG	0.06	0.10	0.11	0.05	0.18	0.11	0.10
CGA	0.02	0.04	0.04	0.02	0.06	0.04	0.03
		df = 7, $\chi^2 = 7.42$; $P^d = 0.390$	df = 7, $\chi^2 = 21.96$; $P^d = 0.002$	df = 7, $\chi^2 = 13.60$; $P^d = 0.059$	df = 7, $\chi^2 = 23.96$; $P^d = 0.001$	df = 7, $\chi^2 = 10.96$; $P^d = 0.14$	df = 7, $\chi^2 = 14.66$; $P^d = 0.041$

Values given are haplotype frequencies. ^aThe mean age at onset in the osteoarthritis (OA) patients was 52.0 years, and so early onset OA patients were defined as those with disease onset at age under 52 years, whereas late onset OA patients were those with onset at age over 52 years. ^bBased on radiographic findings, OA patients were classified as mild (Kellgren-Lawrence grade 1 or 2) or severe (Kellgren-Lawrence grade 3 or 4). ^cBased on Lequesne's functional index score, poor OA patients were defined as those with an index score over 10, whereas good OA patients were those with an index score less than or equal to 10. ^d*P* values for overall difference in haplotype distribution between OA patients and control individuals were calculated using the EH program [23]. df, degrees of freedom.

patients subdivided by clinical parameters and found that the haplotype exhibited a strong association with late onset, radiographically severe, and functionally poor OA of the knee, and in particular with male sex (Tables 5 and 6). However, the TAA and CAG haplotypes exhibited differences between female and male control individuals (Table 5). Haplotype TAA had frequencies of 0.12 and 0.05 in female and male control individuals, but its frequencies in male and female OA patients were 0.18 and 0.14, respectively. Haplotype CAG had a frequency of 0.18 in female control individuals, 0.18 in female patients and 0.18 in male

patients, but its frequency in the male control individuals was 0.31. This discrepancy may be due to the small numbers of male control individuals (*n* = 190) and male OA patients (*n* = 53).

The genotype distribution of the *PvuII* and *XbaI* polymorphisms in the ER- α gene was reported to differ between racial and ethnic groups [13-15]. In the present study, genotype and allele frequencies in control individuals were similar to previously reported distributions in the Chinese population [27]. Interestingly, the exon 8 G/A *BtgI* polymor-

phism and haplotype of three polymorphisms (*PvuII* in intron 1, *XbaI* in intron 1, and *BtgI* in exon 8) were associated with OA in men but not in women.

Our sample size was relatively small and our data were subjected to a number of uncorrected tests, and therefore our positive results may represent false-positive findings. To confirm the association between ER- α polymorphisms and OA, additional studies are required.

Conclusion

In conclusion, we found that ER- α gene haplotype may be associated with primary knee OA in the Korean population, and that genetic variations in the ER- α gene might play a role in susceptibility to OA.

Competing interests

None declared.

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References

- Ghosh P, Cheras PA: **Vascular mechanisms in osteoarthritis.** *Best Pract Res Clin Rheumatol* 2001, **15**:693-709.
- Reginato AM, Olsen BR: **The role of structural genes in the pathogenesis of osteoarthritic disorders.** *Arthritis Res* 2002, **4**:337-345.
- Pelletier JP, Johanne MP, Abramson SB: **Osteoarthritis, an inflammatory disease: potential implication for the selection of new therapeutic targets.** *Arthritis Rheum* 2001, **44**:1237-1247.
- Yuan GH, Kayo MH, Kato T, Nishioka K: **Immunologic intervention in the pathogenesis of osteoarthritis.** *Arthritis Rheum* 2003, **48**:602-611.
- Sandell LJ, Aigner T: **Articular cartilage and changes in arthritis. An introduction: cell biology of osteoarthritis.** *Arthritis Res* 2001, **3**:107-113.
- Aigner T, Dudhia J: **Genomics of osteoarthritis.** *Curr Opin Rheumatol* 2003, **15**:634-640.
- Shupnik MA, Pitt LK, Soh AY, Anderson A, Lopes MB, Laws ER Jr: **Selective expression of estrogen receptor alpha and beta isoforms in human pituitary tumors.** *J Clin Endocrinol Metab* 1998, **83**:3965-3972.
- Evans RM: **The steroid and thyroid hormone receptor superfamily.** *Science* 1988, **240**:889-895.
- Menasce LP, White GR, Harrison CJ, Boyle JM: **Localization of the estrogen receptor locus (ESR) to chromosome 6q25.1 by FISH and a simple post-FISH banding technique.** *Genomics* 1993, **17**:263-265.
- Green S, Walter P, Kumar V, Krust A, Bornert JM, Argos P, Chambon P: **Human oestrogen receptor cDNA: sequence, expression and homology to v-erb-A.** *Nature* 1986, **320**:134-139.
- Schubert EL, Lee MK, Newman B, King KC: **Single nucleotide polymorphisms (SNPs) in the estrogen receptor gene and breast cancer susceptibility.** *J Steroid Biochem Mol Biol* 1999, **71**:21-27.
- Sasaki M, Tanaka Y, Sakuragi N, Dahiya R: **Six polymorphisms on estrogen receptor 1 gene in Japanese, American and German populations.** *Eur J Clin Pharmacol* 2003, **59**:389-393.
- Ushiyama T, Ueyama H, Inoue K, Nishioka J, Ohkubo I, Hukuda S: **Estrogen receptor gene polymorphism and generalized osteoarthritis.** *J Rheumatol* 1998, **25**:134-137.
- Bergink AP, van Meurs JB, Loughlin J, Arp PP, Fang Y, Hofman A, van Leeuwen JP, van Duijn CM, Uitterlinden AG, Pols HA: **Estrogen receptor alpha gene haplotype is associated with radiographic osteoarthritis of the knee in elderly men and women.** *Arthritis Rheum* 2003, **48**:1913-1922.
- Loughlin J, Sinsheimer JS, Mustafa Z, Carr AJ, Clipsham K, Bloomfield VA, Chitnavis J, Bailey A, Sykes B, Chapman K: **Association analysis of the vitamin D receptor gene, the type I collagen gene COL1A1, and the estrogen receptor gene in idiopathic osteoarthritis.** *J Rheumatol* 2000, **27**:779-784.
- Cancel-Tassin G, Latil A, Rousseau F, Mangin P, Bottius E, Escary JL, Berthon P, Cussenot O: **Association study of polymorphisms in the human estrogen receptor alpha gene and prostate cancer risk.** *Eur Urol* 2003, **44**:487-490.
- Tanaka Y, Sasaki M, Kaneuchi M, Shiina H, Igawa M, Dahiya R: **Polymorphisms of estrogen receptor alpha in prostate cancer.** *Mol Carcinog* 2003, **37**:202-208.
- Tanaka Y, Sasaki M, Kaneuchi M, Fujimoto S, Dahiya R: **Single nucleotide polymorphisms of estrogen receptor α in human renal cell carcinoma.** *Biochem Biophys Res Commun* 2002, **296**:1200-1206.
- Curran JE, Lea RA, Rutherford S, Weinstein SR, Griffiths LR: **Association of estrogen receptor and glucocorticoid receptor gene polymorphisms with sporadic breast cancer.** *Int J Cancer* 2001, **95**:271-275.
- Kellgren JK, Lawrence JS: **Radiological assessment of osteoarthritis.** *Ann Rheum Dis* 1957, **16**:494-501.
- Lequesne M, Méry C, Samson M, Gérard P: **Indexes of severity for osteoarthritis of the hip and knee. Validation-value in comparison with other assessment tests.** *Scand J Rheumatol* 1987, **Suppl 65**:85-89.
- Tsai SJ, Wang YC, Hong CJ, Chiu HJ: **Association study of oestrogen receptor alpha gene polymorphism and suicidal behaviours in major depressive disorder.** *Psychiatr Genet* 2003, **13**:19-22.
- Terwilliger JD, Ott J: **Linkage disequilibrium between alleles at marker loci.** In *Handbook of Human Genetic Linkage* 1st edition. Baltimore: Johns Hopkins University; 1994:188-198.
- Ushiyama T, Ueyama H, Inoue K, Ohkubo I, Hukuda S: **Expression of genes for estrogen receptors alpha and beta in human articular chondrocytes.** *Osteoarthritis Cartilage* 1999, **7**:560-566.
- Tsai CL, Liu TK, Chen TJ: **Estrogen and osteoarthritis: a study of synovial estradiol and estradiol receptor binding in human osteoarthritic knees.** *Biochem Biophys Res Commun* 1992, **183**:1287-1291.
- Spector TD, Campion GD: **Generalized osteoarthritis: a hormonally mediated disease.** *Ann Rheum Dis* 1989, **48**:523-527.
- Cai Q, Gao YT, Wen W, Shu XO, Jin F, Smith JR, Zheng W: **Genetic polymorphisms in the estrogen receptor α gene and risk of breast cancer: results from the Shanghai Breast cancer study.** *Cancer Epidemiol Biomarkers Prev* 2003, **12**:853-859.