PROKARYOTES



Complete Genome Sequence of the Methicillin-Resistant *Staphylococcus aureus* Colonizing Strain M92

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ABSTRACT M92 is a methicillin-resistant *Staphylococcus aureus* (MRSA) colonizing strain belonging to ST239-MRSA-III. It frequently shows local nasal colonization in our hospital staff, but has never been associated with infection. We sequenced the complete genome of M92, in order to compare it to highly virulent MRSA strains to gain insight into MRSA virulence factors.

S*taphylococcus aureus* is a major human pathogen causing a wide variety of illnesses, ranging from mild skin and soft tissue infections to life-threatening illnesses such as septicemia, endocarditis, and hemorrhagic pneumonia (1). Methicillin-resistant *Staphylococcus aureus* (MRSA) continues to be a major cause of both hospital-associated (HA-MRSA) and community-associated (CA-MRSA) infection, prompting studies to characterize and understand its virulence mechanism (2, 3). MRSA colonizing strain M92 was found and isolated in our local hospitals in the Calgary, Canada area in the 1980s. This strain is frequently showing local nasal colonization in our hospital staff, but has never been associated with infection over the course of many years. M92 belongs to ST239-MRSA-III with accessory gene regulator (agr) type I. The staphylococcal protein A gene (*spa*) is truncated in M92. It has been used as an avirulence control strain in many of our infection models (4–6). We sequenced the complete genome of M92, in order to compare it to those highly virulent MRSA strains to give a more complete understanding of MRSA virulence factors.

The M92 genome was sequenced with Pacific Biosciences (PacBio) RSII sequencing technology, using one single-molecule real-time (SMRT) cell. Raw reads numbering 21,177 were generated covering a total of 239,169,848 sequenced bases, with an average read length of 11,293 bp (longest 48,331 bp). Contig assembly was done using the HGAP workflow (7): short subreads were aligned to the long subreads in a preassembly step using BLASR (8), then the corrected long reads were used to generate contigs with Celera Assembler (9). The contigs were polished by aligning raw reads on them with BLASR, then processed in the variant calling algorithm (Quiver) to generate consensus sequences. The resultant genome was determined to be 3,051,263 bp in length, with an estimated genome coverage of $77 \times$ and G+C content of 32.80%. Gene annotation was done using NCBI's Prokaryotic Genomes Annotation Pipeline, identifying a total of 3,152 genes, of which 3,009 were coding sequences (CDS), 83 were RNA genes, and 60 were pseudogenes.

Sequence analysis indicates that both the genome size and G+C content are in line with other *S. aureus* genomes, however, the size is on the higher end of what has been published. Several virulence factors common in staphylococci were located, including genes for adhesins like fibrinogen binding (*clfA*, *sdrC*, *sdrD*, *sdrE*), fibronectin binding

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(*fnbA*, *fnbB*), collagen binding (*cna*), elastin binding (*ebpS*), intercellular adhesion (*ica*), and the MHCII analogue (*map*). Common enterotoxin genes were missing (*sea*, *seb*, *sec*, *sed*, *see*, *seg*, *seh*, *sei*, *sej*), as was the toxic shock protein gene (*tst*) and immunity evasive gene (*chp*). Genes for enterotoxins *sek* and *seq* were present, as was the immunity evasive gene *scn*, and cytotoxin genes *hla*, *hld*, and *hlg*. Genes for exoenzymes such as the protease *v8*, hyaluronate lyase (*hysA*), and staphylokinase (*sak*) were also found. A more complete analysis is currently underway looking at a broader range of virulence factors, as well as comparing them to those found in more virulent strains of MRSA.

Accession number(s). The genome sequence has been deposited at DDBJ/EMBL/ GenBank under accession number CP015447.

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REFERENCES

- Diekema DJ, Pfaller MA, Schmitz FJ, Smayevsky J, Bell J, Jones RN, Beach M, SENTRY Partcipants Group. 2001. Survey of infections due to *Staphylococcus* species: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, Latin America, Europe, and the Western Pacific region for the SENTRY Antimicrobial Surveillance Program, 1997–1999. Clin Infect Dis 32:S114–S132.
- David MZ, Daum RS. 2010. Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. Clin Microbiol Rev 23:616–687. https://doi.org/10 .1128/CMR.00081-09.
- Watkins RR, David MZ, Salata RA. 2012. Current concepts on the virulence mechanisms of meticillin-resistant *Staphylococcus aureus*. J Med Microbiol 61:1179–1193. https://doi.org/10.1099/jmm.0.043513-0.
- Wu K, Conly J, McClure JA, Elsayed S, Louie T, Zhang K. 2010. Caenorhabditis elegans as a host model for community-associated methicillinresistant Staphylococcus aureus. Clin Microbiol Infect 16:245–254. https:// doi.org/10.1111/j.1469-0691.2009.02765.x.
- Wu K, Conly J, Surette M, Sibley C, Elsayed S, Zhang K. 2012. Assessment of virulence diversity of methicillin-resistant *Staphylococcus aureus* strains with a *Drosophila melanogaster* infection model. BMC Microbiol 12:274. https://doi.org/10.1186/1471-2180-12-274.

- Wu K, Simor AE, Vearncombe M, McClure JA, Zhang K. 2012. A Caenorhabditis elegans host model correlates with invasive disease caused by Staphylococcus aureus recovered during an outbreak in neonatal intensive care. Can J Infect Dis Med Microbiol 23:130–134.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/nmeth .2474.
- Chaisson MJ, Tesler G. 2012. Mapping single molecule sequencing reads using basic local alignment with successive refinement (BLASR): application and theory. BMC Bioinformatics 13:238. https://doi.org/10.1186/1471 -2105-13-238.
- Myers EW, Sutton GG, Delcher AL, Dew IM, Fasulo DP, Flanigan MJ, Kravitz SA, Mobarry CM, Reinert KHJ, Remington KA, Anson EL, Bolanos RA, Chou HH, Jordan CM, Halpern AL, Lonardi S, Beasley EM, Brandon RC, Chen L, Dunn PJ, Lai Z, Liang Y, Nusskern DR, Zhan M, Zhang Q, Zheng X, Rubin GM, Adams MD, Venter JC. 2000. A whole-genome assembly of *Drosophila*. Science 287:2196–2204. https://doi.org/10.1126/science.287.5461 .2196.