Identification of Better Pharmacokinetic Benzothiazinone Derivatives as New Antitubercular Agents

Kai Lv,† Xuefu You,† Bin Wang,‡ Zengquan Wei,§ Yun Chai,† Bo Wang,† Apeng Wang,† Guocheng Huang, \overline{y} Mingliang Liu, \overline{x} , \overline{z} and Yu Lu^{*, \overline{x}}

† Institute of Medicinal Biotechnology, Chi[nese A](#page-3-0)cademy of M[edi](#page-3-0)cal Sciences and Peking Union Medical College, Beijing 100050, China

‡ Beijing Key Laboratory of Drug Resistance Tuberculosis Research, Department of Pharmacology, Beijing Tuberculosis and Thoracic Tumor Research Institute, Beijing ChestHospital, Capital Medical University, Beijing 101149, China

§ Tianjin Chase Sun Pharmaceutical Co. Ltd., Tianjin 301700, China

S Supporting Information

[AB](#page-3-0)STRACT: [A series of ne](#page-3-0)w 8-nitro-6-(trifluoromethyl)-1,3 benzothiazin-4-one(BTZ) derivatives containing a C-2 nitrogen spiro-heterocycle moiety based on the structures of BTZ candidates BTZ043 and PBTZ169 were designed and synthesized as new antitubercular agents. Many of them were found to have excellent in vitro activity (MIC < $0.15 \mu M$) against the drug susceptive Mycobacterium tuberculosis H37Rv strain and two clinically isolated multidrug-resistant strains. Compounds 11l and 11m display acceptable safety, greater aqueous solubility, and better pharmacokinetic profiles than PBTZ169, suggesting their promising potential to be lead compounds for future antitubercular drug discovery.

KEYWORDS: Antitubercular agents, benzothiazinones, spiro-heterocycles, structure−activity relationships, pharmacokinetics

Tuberculosis (TB), one of the leading causes of death from
an infectious disease ranking above HIV/AIDS, is caused
which has Muscle training tuberwheir (MTD)¹ The Muscle mainly by Mycobacterium tuberculosis (MTB).¹ The World Health Organization (WHO) estimated that approximately 10.4 million people were infected and 1.4 millio[n d](#page-4-0)ied from TB worldwide in $2015²$ Recently, the spread of multidrug-resistant (MDR) TB and the emergence of extensively drug-resistant (XDR) TB have r[ev](#page-4-0)italized drug discovery efforts in search of new drugs.^{3−5} Although Bedaquiline (inhibition of mitochondrial ATP synthase) and Delamanid (inhibition of mycolic acid biosynthesi[s](#page-4-0)) [w](#page-4-0)ere approved for the treatment of MDR-TB, over a huge gap of over 40 years, $6,7$ both of them have pronounced issues, including hERG toxicity concerns, as well as multiple ADME issues due to t[heir](#page-4-0) high lipophilicity.³ Moreover, there are only three new chemical entities TBA-354 (nitroimidazole), PBTZ169 (benzothiazinone), and Q20[3](#page-4-0) $(imidazopyridine)⁸$ currently in Phase 1 clinical trials. Therefore, there is a significant unmet medical need for safer, more effective TB drug[s](#page-4-0) with novel mechanisms.

Decaprenyl phosphoryl- β -D-ribose 2'-epimerase (DprE1), an essential aspect of MTB survival and a novel mechanism of antitubercular activity, has been identified as a potential target for developing potent and safer anti-TB agents. $9-11$ Some new chemical entities (NCE) were found to have excellent activity against MDR/XDR-MTB as covalent or nonco[va](#page-4-0)l[en](#page-4-0)t inhibitors of the DprE1 enzyme.12−¹⁷ As the most advanced scaffold

among these NCEs, nitrobenzothiazinones (BTZs) have garnered great interest recently, and many series of BTZ derivatives were reported.18−²⁶ The structure−activity relationship (SAR) and mechanistic studies suggest that the $-NO₂$ group at position 8 and t[he](#page-4-0) s[ulf](#page-5-0)ur atom at position 1 are critical for activity, that the $-CF_3$ at position 6 plays an important role in maintaining activity, $18,19,27,28$ and that structural modifications at position 2 are the most successful and efficient. For example, BTZ043 (Fig[ure](#page-4-0) [1\),](#page-5-0) [a 2](#page-5-0)-spiroketal BTZ nearing phase I clinical trials, displays nanomolar bactericidal activity both in vitro and in ex vivo [models o](#page-1-0)f TB.^{18,24} PBTZ169(Figure 1), a 2piperazino BTZ with slightly worse pharmacokinetic (PK) properties relative to BTZ043, [ha](#page-4-0)[s i](#page-5-0)mproved i[n vivo](#page-1-0) potency because of its greater solubility through protonation of the tertiary amino nitrogen on the piperazine.²⁰ According to the latest report, a Phase I trial of PBTZ169 was completed in Russia, and a second Phase I trial wil[l b](#page-5-0)e undertaken in Switzerland in $2017²$

In our previous study, the spiroketal moiety of BTZ043 was replaced by various [c](#page-4-0)yclic ketoximes or the terminal nitrogen on the piperazine ring of BPTZ169 was shifted outside the ring, and some of the resulting compounds were found to have

Received: March 10, 2017 Accepted: May 10, 2017 Published: May 10, 2017

Figure 1. Design of the BTZ derivatives.

improved activity and aqueous solubility, or acceptable PK profiles.²⁶ In this study, we intended to integrate the structural features of BTZ043 and PBTZ169, and construct structurally unique [co](#page-5-0)mpounds, nitrogen spiro-heterocycles as side chains at the 2-position of BTZs (Figure 1). Clearly, these side chains combine the characteristics of both the spiro-cycle of BTZ043 and the alkaline tertiary-amine of BTZ169. The antimycobacterial activity, solubility, toxicity, and PK properties of these target compounds were evaluated, aiming to identify alternative groups at position 2 of BTZs and find optimized potent anti-TB drug candidates with improved drug-like properties through SAR study.

Detailed synthetic pathways to target compounds 11−13 are outlined in Scheme 1. Introduction of various R groups to Bocprotected spiro-heterocycles 1−3 by reductive amination with aldehydes, 29 nucleophilic substitution with bromides, or amidation with carboxylic acid in the presence of EDC and HOBt³⁰ g[ave](#page-5-0) compounds 4–6, which upon removal of the Boc protecting group afforded the desired side chain compounds 7−9 [by](#page-5-0) TFA in DCM in good yields. Additionally, the Boc group of 3 was also reduced by LiALH₄ to produce the methide 9. ³¹ The target compounds 11−13 were conveniently obtained from 10^{20} by nucleophilic substitution with spiro-heterocycle d[eri](#page-5-0)vatives 7−9 or 1−3.

The t[arg](#page-5-0)et compounds 11a−i, 12a−d, and 13a−c bearing different kinds of substituent groups to ensure side chain flexibility and structure diversity were first synthesized (Table 1). They were preliminarily screened for in vitro activity against MTB H37Rv ATCC27294 strain, using the Microplate [Alamar](#page-2-0) [B](#page-2-0)lue Assay (MABA). 32 The minimum inhibitory concentration (MIC) is defined as the lowest concentration effecting a reduction in fluores[cen](#page-5-0)ce of >90% relative to the mean of replicate bacterium-only controls. The MIC values of the compounds along with PBTZ169, isoniazid (INH), and rifampicin (RFP) for comparison were obtained from three independent experiments and presented in μ M in Table 1.

The data reveal that with a few exceptions (11e−h, 12d, 13c), all the BTZ derivatives show considerable act[ivity aga](#page-2-0)inst this strain (MIC < 1 μ M). Among them, compounds 12b and 12c (MIC = 0.062 and 0.060 μ M) are more active than INH (MIC = 0.284 μ M) and RFP (MIC = 0.092 μ M), and slightly less active than PBTZ169 (MIC < 0.035 μ M). The potency of the BTZ derivatives is related to the sizes of both cycles A and B in the spiro-heterocycle moiety, as well as the nature of substituent groups. When ring A is piperidine (11−13), the contribution of ring B to the activity seems to be as follows:

Scheme 1. Synthesis of Compounds $11-13^a$

 a_{Regents} and conditions: (a) (i) RCHO, MeOH, AcOH, NaCNBH₃, rt−50 °C, 60–85%; or (ii) RBr, MeCN, K₂CO₃, rt, 55–65%; or (iii) RCOOH, EDC, HOBt, Et₃N, DCM, rt, 67%; (b) TFA, DCM, rt, 80− 95%; (c) LiAlH4, THF, 60 °C, 65%; (d) MeOH, rt−50 °C, 43−70%.

pyrrolidine > azetidine > piperidine (11a vs 12a vs 13a; 11b vs 12b vs 13b); whereas, when ring B is piperidine, the contribution of ring A is in this order: piperidine > azetidine > pyrrolidine (11i vs 12d vs 13c). However, replacement of the cyclohexylmethyl group derived from PBTZ169 with benzyl or acetate moieties leads to increased activity (11a vs 11b−d; 12a vs 12b−c; 13a vs 13b). Removal of the cyclohexyl group (11e) or replacement of the cyclohexylmethyl with benzoyl (11h) abolishes the potency against this strain. Moreover, Nacetamide was also found to be much less active than the corresponding N-acetate (11b vs 11g; 11d vs 11f). Consequently, this set of modifications suggests that the spiro-heterocycle (piperidine as ring A) with N-benzylic group might be important for antimycobacterial activity and worthy of further optimizations, considering the probable instability in vivo of the N-acetate moiety.

Next, the optimizations were focused on R group (Table 2) of the benzene ring. An additional set of N-benzylated analogues listed in Table 2 were designed and sy[nthesized](#page-2-0). Considering the commercial availability of the spiro-heterocycles, compounds [with pipe](#page-2-0)ridine as B ring synthesized from the cheapest starting material 1 were selected for the preliminary SAR study. Our results indicate that most of them show potent activity against MTB H37Rv strain (MIC < 0.15 μ M) and that the most active compounds, 11m, 11o and 12f, were found to have the same MIC values of <0.035 μ M as PBTZ169. Exploration of SAR at the para-position of phenyl group with piperidine as ring B was first conducted by substitution of various functional groups (11j−m). Compared

Table 1. Structures and Activities of 11a−i, 12a−d, and 13a− c against MTB H37Rv

with the lead compound 11c, introduction of an electronwithdrawing group (F, Br, CF_3) offers more potency $(11k-m)$, whereas the presence of an electron-donating one (CH_3O)

results in slightly decreased activity (11j). Moreover, a metafluoro atom leads to increased activity (11k vs 11n), and an additional meta-fluoro one is even more favorable (11n vs 11o). The SAR applies equally to the analogues with pyrrolidine ring as ring B. For example, introduction of an electron-donating group (CH_3O) is detrimental to the potency (12c vs 12e; 12c vs $12g$). Conversely, an electron-withdrawing one (F) causes improved activity (12c vs 12f). Additionally, a compound with azetidine ring as ring $B(13c)$ was also synthesized to investigate the potential impact of the size of ring B on activity. It is shown that azetidine \approx pyrrolidine \gg piperidine (11j vs 12e vs 13d), suggesting that BTZs with azetidine or pyrrolidine as ring B appears to be more active than piperidine. Inspired by this, more N-benzylated analogues with azetidine or pyrrolidine as B ring are now being synthesized in our lab, and the results will be reported in due course.

It was reported there is a strong correlation between MIC and lipophilicity of N-alkylpiperazyl-BTZs.²⁰ The logP values of the N-benzylated spiro-heterocycle series were calculated by Chemdraw 15.0 and listed in Table 2. [H](#page-5-0)owever, no clear correlation between logP and antimycobacterial activity was found in this study.

Encouraged by their strong potency against the drug sensitive MTB H37Rv strain (MIC < $0.15 \mu M$), compounds 11l−o, 12b-c, 12e−f, and 13d were further evaluated against two clinical isolated MTB-MDR (12525 and14231) strains resistant to both INH and RFP. As shown in Table 3, all of them exhibit excellent activity (MIC < $0.035 - 0.123 \mu M$) comparable to or slightly less potent than PBTZ[169, sugg](#page-3-0)esting their promising potential for both drug-sensitive and resistant MTB strains (Tables 1−3).

Considering acidic gastrointestinal environments, compounds 11l−o, 12b−c, 12e−f, and 13d were evaluated for their water solubility at [pH](#page-3-0) 2 (0.01 M HCl solution) by using an HPLC-UV method. 33 As expected, the compounds containing a basic nitrogen spiro-heterocycle moiety display good solubility (0.67−2.0[4](#page-5-0) mg/mL). All of them are, with the exception of 11o, more water-soluble than PBTZ169 (0.90 mg/ mL). Compound 12b with an additional acetate moiety was

Table 3. Activity, Solubility, Cytotoxicity and Acute Toxicity of Selected Compounds

	MIC (μM)				
compd.	MDR- MTB 12525	MDR- MTB 14231	water solubility ^a (mg/mL)	CC_{50}^{b} Vero cells (μM)	acute toxicity ^c
111	0.092	0.086	1.17	1677.85	5/5
11m	< 0.035	< 0.035	1.24	145.35	5/5
11n	<0.035	<0.035	1.13	1398.13	5/5
11 ₀	< 0.035	< 0.035	0.67	1169.75	5/5
12 _b	0.080	0.060	2.04	266.34	5/5
12c	< 0.035	< 0.035	1.56	289.26	3/5
12e	0.056	0.095	1.40	86.20	NT
12f	< 0.035	0.034	1.12	810.61	4/5
13d	0.119	0.123	1.20	78.86	NT
PBTZ- 169	< 0.035	< 0.035	0.90	1402.82	NT
INH	>291.7	34.48			
RFP	27.24	>48.60			

a The water solubility was tested in 0.01 M HCl solution (approximate pH 2.0). ^bThe 50% cytotoxic concentration. ^cNo. of animals that survived/total no. of animals; NT, not tested.

found to have the highest solubility (2.04 mg/mL), which is more than two times that of PBTZ169 (Table 3).

Compounds 11l−o, 12b-c, 12e-f, and 13d were examined for cytotoxicity $(CC₅₀)$ in a mammalian Vero cell line by MTT assay, 34 and the results are reported in Table 3. A comparison demonstrates that only three compounds 11l, 11n, and 11o (CC₅₀: 1169.75−1677 μM) display comparable cytotoxic to PBTZ169 (CC₅₀: 1402.82 μ M), and compounds 12e and 13d containing a N-(4-methoxyl)benzyl group are the most cytotoxic (CC₅₀: 86.20 and 78.86 μ M). Moreover, all but one (11m) of the derivatives with piperidine as ring B have higher CC_{50} than those with pyrrolidine or azetidine as ring B.

Based on the measured activity levels against all of the tested strains and cytotoxicity, compounds 11l−o, 12b−c, and 12f were further tested for in vivo tolerability by recording the number of survivors after a single oral dose in mice of 500 mg/ kg, followed by a 7-day observation. As shown in Table 3, all of the compounds with piperidine as ring B (11l−o) and compound 12b with an acetate moiety display the lowest oral acute lethal toxicity, followed by pyrrolidine (ring B) containing derivatives 12f and then 12c.

The in vivo PK profiles of compounds 11l−o, 12b, and 12f were evaluated in mice after a single oral administration of 50 mg/kg. As shown in Table 4, all the tested compounds display similar or significantly longer $T_{1/2}$ (3.11−10.66 h) and MRT (4.00−15.9 h) than PBTZ169 (2.87 and 3.73 h). Among the compounds with piperidine as ring B (11l−o), compounds 11l

and 11m with a substituent at the para-position on benzene ring display the best PK profiles, their $T_{1/2}$, C_{max} , AU $C_{0 \text{-inf}}$, and MRT are comparable to or significantly longer/higher than PBTZ169, whereas the C_{max} and $\text{AUC}_{0\text{-inf}}$ of compounds 11n– o with a substituent at the meta-position are similar to or significantly lower than PBTZ169. The results suggest that the presence of a substituent at the para-position on benzene ring is more favorable for the PK profiles than the meta-position. In addition, compound 12f with a para-fluoro atom on benzene ring has similar PK profiles to PBTZ169. However, compound 12b has very poor drug exposures as expected; its C_{max} (17.6) ng/mL) and $AUC_{0\text{-inf}}$ (41.8 ng/mL) are significantly lower than PBTZ169, and the hydrolysis of the acetate moiety in acidic conditions may account for this.

In summary, a series of new BTZ derivatives with a nitrogen spiro-heterocycle moiety at position 2 were designed as new TB agents through combining the structural features of both BTZ043 and PBTZ169. Many of them exhibit excellent in vitro inhibitory activity against both drug-sensitive MTB strain H37Rv and drug-resistant clinical isolates (MIC< $0.15 \mu M$). Compounds 11l and 11m, with $N-(4$ -bromobenzyl)- and $N-(4$ trifuorobenzyl)-3,9-diazaspiro[5,5]undecane moieties, respectively, display acceptable safety, higher aqueous solubility and better PK properties than PBTZ169, and both of them may serve as new and promising lead compounds for further antitubercular drug discovery. Studies to determine the in vivo efficacy of 11l and 11m are currently underway.

■ ASSOCIATED CONTENT

6 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsmedchemlett.7b00106.

[Experimental procedu](http://pubs.acs.org)res, analytical data, ¹H NMR and ¹³C NMR copies for compounds 11−13, and HPLC copies of compound 11o for solubility determination (PDF)

■ A[UTHO](http://pubs.acs.org/doi/suppl/10.1021/acsmedchemlett.7b00106/suppl_file/ml7b00106_si_001.pdf)R INFORMATION

Corresponding Authors

*E-mail: lmllyx@126.com. Phone: 86-010-63030965 (M.L.) *E-mail: luyu4876@hotmail.com. Phone: 86-010-89509357 $(Y.L.).$

ORCID[®]

Minglian[g](mailto:luyu4876@hotmail.com) [Liu:](mailto:luyu4876@hotmail.com) 0000-0001-8491-8115

Author Contributions

M.L.L., Y.L., X.[F.Y., and H.Y.G. co](http://orcid.org/0000-0001-8491-8115)nceived and designed the project. K.L., B.W., Z.W., Y.C., B.W., and A.W. conducted the experiments, K.L. and M.L. analyzed the data and prepared the

Table 4. PK Profiles of 11k−n and 12b, 12e Dosed Orally in Mice at 50 mg/kg $(n = 3)^a$

"Note: noncompartmental calculations gave these parameter estimates. $*P < 0.05$. $*P < 0.01$, one-tail t test compared with PBTZ169.

manuscript. All authors read and approved the manuscript. All authors have given approval to the final version of the manuscript.

Funding

We gratefully acknowledge the financial support by the National S&T Major Special Project on Major New Drug Innovations (2015ZX09102007-008), NSFC (81502923, 81373267, 21502237), CAMS Innovation Fund for Medical Sciences (CIFMS2016-I2M-1-010), and Beijing Municipal Administration of Hospitals Clinical Medicine Development of Special Funding Support (ZYLX201304).

Notes

All animal experiments were carried out in accordance with the guidelines of the Chinese Association for Laboratory Animal Sciences and approved by the institutional ethical committee (IEC) of Peking Union Medical College.

The authors declare no competing financial interest.

■ ABBREVIATIONS

ADME, absorption, distribution, metabolism, excretion, toxicity; hERG, human ether-a-go-go related gene; PK, pharmacokinetic; TB, tuberculosis; HIV, human immunodeficiency virus; AIDS, acquired immune deficiency syndrome; MTB, Mycobacterium tuberculosis; MDR, multidrug-resistant; XDR, extensively drug-resistant; WHO, World Health Organization; ATP, adenosine triphosphate; DprE1, decaprenyl phosphoryl- β -D-ribose 2'-epimerase; NCE, new chemical entities; BTZs, nitrobenzothiazinones; SAR, structure−activity relationship; EDC, 1-ethyl-3-(3-(dimethylamino)propyl)carbodiimide; THF, tetrahydrofuran; HOBt, hydroxybenzotriazole; TFA, trifluoroacetic acid; DCM, methylene chloride; MIC, minimum inhibitory concentration; MABA, microplate alamar blue assay; INH, isoniazid; RFP, rifampicin; logP, lipophilicity; HPLC, high-performance liquid chromatography; UV, ultraviolet; CC, cytotoxicity; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; $T_{1/2}$, half-life; C_{max} , maximum serum concentration; T_{max} time at which the C_{max} is observed; AUC, area under the curve; MRT, mean residence time

■ REFERENCES

(1) Nusrath Unissa, A.; Hanna, L. E.; Swaminathan, S. A Note on Derivatives of Isoniazid, Rifampicin, and Pyrazinamide Showing Activity Against Resistant Mycobacterium tuberculosis. Chem. Biol. Drug Des. 2016, 87 (4), 537−50.

(2) World Health Organization. Global Tuberculosis Report 2016. http://www.who.int/tb/publications/global_report/en/.

(3) Hoagland, D. T.; Liu, J.; Lee, R. B.; Lee, R. E. New agents for the [treatment of drug-resistant Mycobacterium tuberculos](http://www.who.int/tb/publications/global_report/en/)is. Adv. Drug Delivery Rev. 2016, 102, 55−72.

(4) Jones, D. Tuberculosis success. Nat. Rev. Drug Discovery 2013, 12 (3) , 175−6.

(5) Schraufnagel, D.; Abubaker, J. Global action against multidrugresistant tuberculosis. JAMA, J. Am. Med. Assoc. 2000, 283 (1), 54−5.

(6) Cohen, J. Infectious disease. Approval of novel TB drug celebrated–with restraint. Science 2013, 339 (6116), 130.

(7) Ryan, N. J.; Lo, J. H. Delamanid: first global approval. Drugs 2014, 74 (9), 1041−5.

(8) Pethe, K.; Bifani, P.; Jang, J.; Kang, S.; Park, S.; Ahn, S.; Jiricek, J.; Jung, J.; Jeon, H. K.; Cechetto, J.; Christophe, T.; Lee, H.; Kempf, M.; Jackson, M.; Lenaerts, A. J.; Pham, H.; Jones, V.; Seo, M. J.; Kim, Y. M.; Seo, M.; Seo, J. J.; Park, D.; Ko, Y.; Choi, I.; Kim, R.; Kim, S. Y.; Lim, S.; Yim, S. A.; Nam, J.; Kang, H.; Kwon, H.; Oh, C. T.; Cho, Y.; Jang, Y.; Kim, J.; Chua, A.; Tan, B. H.; Nanjundappa, M. B.; Rao, S. P.; Barnes, W. S.; Wintjens, R.; Walker, J. R.; Alonso, S.; Lee, S.; Kim, J.; Oh, S.; Oh, T.; Nehrbass, U.; Han, S. J.; No, Z.; Lee, J.; Brodin, P.; Cho, S. N.; Nam, K.; Kim, J. Discovery of Q203, a potent clinical candidate for the treatment of tuberculosis. Nat. Med. 2013, 19 (9), 1157−60.

(9) Manina, G.; Pasca, M. R.; Buroni, S.; De Rossi, E.; Riccardi, G. Decaprenylphosphoryl-beta-D-ribose 2′-epimerase from Mycobacterium tuberculosis is a magic drug target. Curr. Med. Chem. 2010, 17 (27), 3099−108.

(10) Riccardi, G.; Pasca, M. R.; Chiarelli, L. R.; Manina, G.; Mattevi, A.; Binda, C. The DprE1 enzyme, one of the most vulnerable targets of Mycobacterium tuberculosis. Appl. Microbiol. Biotechnol. 2013, 97 (20), 8841−48.

(11) Brecik, M.; Centarova, I.; Mukherjee, R.; Kolly, G. S.; Huszar, S.; Bobovska, A.; Kilacskova, E.; Mokosova, V.; Svetlikova, Z.; Sarkan, M.; Neres, J.; Kordulakova, J.; Cole, S. T.; Mikusova, K. DprE1 Is a Vulnerable Tuberculosis Drug Target Due to Its Cell Wall Localization. ACS Chem. Biol. 2015, 10 (7), 1631−36.

(12) Crellin, P. K.; Brammananth, R.; Coppel, R. L. Decaprenylphosphoryl-beta-D-Ribose 2 ′-Epimerase, the Target of Benzothiazinones and Dinitrobenzamides, Is an Essential Enzyme in Mycobacterium smegmatis. PLoS One 2011, 6 (2), e16869.

(13) Stanley, S. A.; Grant, S. S.; Kawate, T.; Iwase, N.; Shimizu, M.; Wivagg, C.; Silvis, M.; Kazyanskaya, E.; Aquadro, J.; Golas, A.; Fitzgerald, M.; Dai, H. Q.; Zhang, L. X.; Hung, D. T. Identification of Novel Inhibitors of M. tuberculosis Growth Using Whole Cell Based High-Throughput Screening. ACS Chem. Biol. 2012, 7 (8), 1377−84. (14) Wang, F.; Sambandan, D.; Halder, R.; Wang, J. N.; Batt, S. M.; Weinrick, B.; Ahmad, I.; Yang, P. Y.; Zhang, Y.; Kim, J.; Hassani, M.; Huszar, S.; Trefzer, C.; Ma, Z. K.; Kaneko, T.; Mdluli, K. E.; Franzblau, S.; Chatterjee, A. K.; Johnson, K.; Mikusova, K.; Besra, G. S.; Futterer, K.; Jacobs, W. R.; Schultz, P. G. Identification of a small molecule with activity against drug-resistant and persistent tuberculosis. Proc. Natl. Acad. Sci. U. S. A. 2013, 110 (27), 2510−17.

(15) Naik, M.; Humnabadkar, V.; Tantry, S. J.; Panda, M.; Narayan, A.; Guptha, S.; Panduga, V.; Manjrekar, P.; Jena, L. K.; Koushik, K.; Shanbhag, G.; Jatheendranath, S.; Manjunatha, M. R.; Gorai, G.; Bathula, C.; Rudrapatna, S.; Achar, V.; Sharma, S.; Ambady, A.; Hegde, N.; Mahadevaswamy, J.; Kaur, P.; Sambandamurthy, V. K.; Awasthy, D.; Narayan, C.; Ravishankar, S.; Madhavapeddi, P.; Reddy, J.; Prabhakar, K. R.; Saralaya, R.; Chatterji, M.; Whiteaker, J.; McLaughlin, B.; Chiarelli, L. R.; Riccardi, G.; Pasca, M. R.; Binda, C.; Neres, J.; Dhar, N.; Signorino-Gelo, F.; McKinney, J. D.; Ramachandran, V.; Shandil, R.; Tommasi, R.; Iyer, P. S.; Narayanan, S.; Hosagrahara, V.; Kavanagh, S.; Dinesh, N.; Ghorpade, S. R. 4- Aminoquinolone Piperidine Amides: Noncovalent Inhibitors of DprE1 with Long Residence Time and Potent Antimycobacterial Activity. J. Med. Chem. 2014, 57 (12), 5419−34.

(16) Panda, M.; Ramachandran, S.; Ramachandran, V.; Shirude, P. S.; Humnabadkar, V.; Nagalapur, K.; Sharma, S.; Kaur, P.; Guptha, S.; Narayan, A.; Mahadevaswamy, J.; Ambady, A.; Hegde, N.; Rudrapatna, S. S.; Hosagrahara, V. P.; Sambandamurthy, V. K.; Raichurkar, A. Discovery of Pyrazolopyridones as a Novel Class of Noncovalent DprE1 Inhibitor with Potent Anti-Mycobacterial Activity. J. Med. Chem. 2014, 57 (11), 4761−71.

(17) Shirude, P. S.; Shandil, R.; Sadler, C.; Naik, M.; Hosagrahara, V.; Hameed, S.; Shinde, V.; Bathula, C.; Humnabadkar, V.; Kumar, N.; Reddy, J.; Panduga, V.; Sharma, S.; Ambady, A.; Hegde, N.; Whiteaker, J.; McLaughlin, R. E.; Gardner, H.; Madhavapeddi, P.; Ramachandran, V.; Kaur, P.; Narayan, A.; Guptha, S.; Awasthy, D.; Narayan, C.; Mahadevaswamy, J.; Vishwas, K. G.; Ahuja, V.; Srivastava, A.; Prabhakar, K. R.; Bharath, S.; Kale, R.; Ramaiah, M.; Choudhury, N. R.; Sambandamurthy, V. K.; Solapure, S.; Iyer, P. S.; Narayanan, S.; Chatterji, M. Azaindoles: Noncovalent DprE1 Inhibitors from Scaffold Morphing Efforts, Kill Mycobacterium tuberculosis and Are Efficacious in Vivo. J. Med. Chem. 2013, 56 (23), 9701−8.

(18) Neres, J.; Pojer, F.; Molteni, E.; Chiarelli, L. R.; Dhar, N.; Boy-Rottger, S.; Buroni, S.; Fullam, E.; Degiacomi, G.; Lucarelli, A. P.; Read, R. J.; Zanoni, G.; Edmondson, D. E.; De Rossi, E.; Pasca, M. R.; McKinney, J. D.; Dyson, P. J.; Riccardi, G.; Mattevi, A.; Cole, S. T.;

Binda, C. Structural basis for benzothiazinone-mediated killing of Mycobacterium tuberculosis. Sci. Transl. Med. 2012, 4 (150), 150ra121.

(19) Makarov, V.; Manina, G.; Mikusova, K.; Mollmann, U.; Ryabova, O.; Saint-Joanis, B.; Dhar, N.; Pasca, M. R.; Buroni, S.; Lucarelli, A. P.; Milano, A.; De Rossi, E.; Belanova, M.; Bobovska, A.; Dianiskova, P.; Kordulakova, J.; Sala, C.; Fullam, E.; Schneider, P.; McKinney, J. D.; Brodin, P.; Christophe, T.; Waddell, S.; Butcher, P.; Albrethsen, J.; Rosenkrands, I.; Brosch, R.; Nandi, V.; Bharath, S.; Gaonkar, S.; Shandil, R. K.; Balasubramanian, V.; Balganesh, T.; Tyagi, S.; Grosset, J.; Riccardi, G.; Cole, S. T. Benzothiazinones kill Mycobacterium tuberculosis by blocking arabinan synthesis. Science 2009, 324 (5928), 801−4.

(20) Makarov, V.; Lechartier, B.; Zhang, M.; Neres, J.; van der Sar, A. M.; Raadsen, S. A.; Hartkoorn, R. C.; Ryabova, O. B.; Vocat, A.; Decosterd, L. A.; Widmer, N.; Buclin, T.; Bitter, W.; Andries, K.; Pojer, F.; Dyson, P. J.; Cole, S. T. Towards a new combination therapy for tuberculosis with next generation benzothiazinones. EMBO Mol. Med. 2014, 6 (3), 372−83.

(21) Trefzer, C.; Rengifo-Gonzalez, M.; Hinner, M. J.; Schneider, P.; Makarov, V.; Cole, S. T.; Johnsson, K. Benzothiazinones: Prodrugs That Covalently Modify the Decaprenylphosphoryl-beta-D-ribose 2 ′-epimerase DprE1 of Mycobacterium tuberculosis. J. Am. Chem. Soc. 2010, 132 (39), 13663−65.

(22) de Jesus Lopes Ribeiro, A. L.; Degiacomi, G.; Ewann, F.; Buroni, S.; Incandela, M. L.; Chiarelli, L. R.; Mori, G.; Kim, J.; Contreras-Dominguez, M.; Park, Y. S.; Han, S. J.; Brodin, P.; Valentini, G.; Rizzi, M.; Riccardi, G.; Pasca, M. R. Analogous Mechanisms of Resistance to Benzothiazinones and Dinitrobenzamides in Mycobacterium smegmatis. PLoS One 2011, 6 (11), e26675.

(23) Karoli, T.; Becker, B.; Zuegg, J.; Mollmann, U.; Ramu, S.; Huang, J. X.; Cooper, M. A. Identification of Antitubercular Benzothiazinone Compounds by Ligand-Based Design. J. Med. Chem. 2012, 55 (17), 7940−7944.

(24) Pasca, M. R.; Degiacomi, G.; Ribeiro, A. L.; Zara, F.; De Mori, P.; Heym, B.; Mirrione, M.; Brerra, R.; Pagani, L.; Pucillo, L.; Troupioti, P.; Makarov, V.; Cole, S. T.; Riccardi, G. Clinical isolates of Mycobacterium tuberculosis in four European hospitals are uniformly susceptible to benzothiazinones. Antimicrob. Agents Chemother. 2010, 54 (4), 1616−8.

(25) Batt, S. M.; Jabeen, T.; Bhowruth, V.; Quill, L.; Lund, P. A.; Eggeling, L.; Alderwick, L. J.; Futterer, K.; Besra, G. S. Structural basis of inhibition of Mycobacterium tuberculosis DprE1 by benzothiazinone inhibitors. Proc. Natl. Acad. Sci. U. S. A. 2012, 109 (28), 11354− 59.

(26) Zhang, R.; Lv, K.; Wang, B.; Li, L. H.; Wang, B.; Liu, M. L.; Guo, H. Y.; Wang, A. P.; Lu, Y. Design, synthesis and antitubercular evaluation of benzothiazinones containing an oximido or amino nitrogen heterocycle moiety. RSC Adv. 2017, 7 (3), 1480−83.

(27) Kloss, F.; Krchnak, V.; Krchnakova, A.; Schieferdecker, S.; Dreisbach, J.; Krone, V.; Mollmann, U.; Hoelscher, M.; Miller, M. J. In Vivo Dearomatization of the Potent Antituberculosis Agent BTZ043 via Meisenheimer Complex Formation. Angew. Chem., Int. Ed. 2017, 56 (8), 2187−91.

(28) Tiwari, R.; Miller, P. A.; Chiarelli, L. R.; Mori, G.; Sarkan, M.; Centarova, I.; Cho, S. H.; Mikusova, K.; Franzblau, S. G.; Oliver, A. G.; Miller, M. J. Design, Syntheses, and Anti-TB Activity of 1,3- Benzothiazinone Azide and Click Chemistry Products Inspired by BTZ043. ACS Med. Chem. Lett. 2016, 7 (3), 266−70.

(29) Baxter, E. W. Reductive aminations of carbonyl compounds with borohydride and borane reducing agents. Organic Reactions 2001, 59, 1−714.

(30) Mahmoud, K. A.; Long, Y. T.; Schatte, G.; Kraatz, H. B. Rearrangement of the active ester intermediate during HOBt/EDC amide coupling. Eur. J. Inorg. Chem. 2005, 1, 173−180.

(31) Corruble, A.; Davoust, D.; Desjardins, S.; Fressigne, C.; Giessner-Prettre, C.; Harrison-Marchand, A.; Houte, H.; Lasne, M. C.; Maddaluno, J.; Oulyadi, H.; Valnot, J. Y. A NMR and theoretical study of the aggregates between alkyllithium and chiral lithium amides:

control of the topology through a single asymmetric center. J. Am. Chem. Soc. 2002, 124 (51), 15267−79.

(32) Lu, Y.; Zheng, M.; Wang, B.; Fu, L.; Zhao, W.; Li, P.; Xu, J.; Zhu, H.; Jin, H.; Yin, D.; Huang, H.; Upton, A. M.; Ma, Z. Clofazimine analogs with efficacy against experimental tuberculosis and reduced potential for accumulation. Antimicrob. Agents Chemother. 2011, 55 (11), 5185−93.

(33) Sun, L. Q.; Zhu, L.; Qian, K.; Qin, B.; Huang, L.; Chen, C. H.; Lee, K. H.; Xie, L. Design, synthesis, and preclinical evaluations of novel 4-substituted 1,5-diarylanilines as potent HIV-1 non-nucleoside reverse transcriptase inhibitor (NNRTI) drug candidates. J. Med. Chem. 2012, 55 (16), 7219−29.

(34) Chai, Y.; Wan, Z. L.; Wang, B.; Guo, H. Y.; Liu, M. L. Synthesis and in vitro antibacterial activity of 7-(4-alkoxyimino-3-amino-3 methylpiperidin-1-yl)fluoroquinolone derivatives. Eur. J. Med. Chem. 2009, 44 (10), 4063−9.

■ NOTE ADDED AFTER ASAP PUBLICATION

This paper was published ASAP on May 12, 2017 with errors in the abstract graphic, figure 1, and table 2. The corrected version was reposted on May 15, 2017.