## PROKARYOTES



# Complete Genome Sequence of the Uropathogenic *Escherichia coli* Strain NU14

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**ABSTRACT** *Escherichia coli* is the most common bacterium causing urinary tract infections in humans. We report here the complete genome sequence of the uropathogenic *Escherichia coli* strain NU14, a clinical pyelonephritis isolate used for studying pathogenesis.

rinary tract infections (UTIs) are common infections of humans that primarily affect women. The majority of UTIs are caused by Escherichia coli, hence the term uropathogenic E. coli (UPEC). While antibiotic therapy is generally successful, UTI recurs in 24 to 44% of patients within 6 to 12 months, often with a strain identical to that causing the initial infection (1). A fraction (2 to 5%) of patients further suffer from frequent recurrences ( $\geq$ 3 per year) (2). In a mouse model of UTI (3, 4), human UTI isolates not only infect the lumen of the bladder but also form intracellular collections within the epithelial cells of the bladder, which are known as intracellular bacterial communities (IBCs) and quiescent intracellular reservoirs (QIRs) (5, 6). IBCs and QIRs contribute to phenotypic antibiotic resistance, host immune evasion, and recurrent infection (7). Furthermore, IBC-like structures have been identified in the urine of adult and pediatric UTI patients (8–10). Thus, intracellular infection is a leading hypothesis for why humans suffer from recurrent UTI. The UPEC strain NU14 was isolated from a UTI patient (11). It was the first strain to be observed infecting epithelial cells in a mouse UTI model (12), and it was among several strains initially demonstrated to form IBCs (6). The initial genetic identification of the type 1 pilus adhesin (FimH) was also performed for NU14 (13), which led to its use in vaccine studies for UTIs (14).

NU14 genomic DNA was sheared to a size of approximately 10 kb using a g-Tube (Covaris). A SMRTbell library was prepared according to manufacturer's instructions, loaded with a MagBead-bound library protocol, and sequenced on three SMRTCells using P5-C3 chemistry on the PacBio RSII instrument (Pacific Biosciences) with a 180-min movie time. *De novo* assembly was performed with the Hierarchical Genome Assembly Process (HGAP2) in the SMRT Analysis suite version 2.3 using all default parameters, except that the assembly target coverage parameter was changed to 20 (15). In addition, inspection of the assembled sequence and of the results from the BridgeMapper protocol led to manual resolution of an inverted sequence of ~315 bp, which corresponded to the invertible *fimS* promoter element. This was set to the "OFF" orientation, corresponding to the orientation in most other *E. coli* genomes. The sequence was then polished with data from the same SMRTCells using the Resequencing protocol in the SMRT Analysis suite. In total, there were 162,908 reads and 1,607,174,902 nucleotides that passed filtering, representing an approximate coverage of 300× (based on the final assembly) and a preassembly mean read length of 9,865 bp.

The NU14 genome consists of a single circular chromosome of 5,076,615 bp with a G+C content of 50.6%. Annotation of the NU14 genome was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (16). The NU14 chromosome contains 4,908 protein-coding sequences, 22 rRNAs, and 89 tRNAs. The finished genome se-

Received 13 March 2017 Accepted 15 March 2017 Published 4 May 2017

Citation Mehershahi KS, Chen SL. 2017. Complete genome sequence of the uropathogenic *Escherichia coli* strain NU14. Genome Announc 5:e00306-17. https://doi .org/10.1128/genomeA.00306-17.

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quence of UPEC strain NU14 will facilitate genetic manipulation and dissection of the virulence mechanisms of UPEC.

Accession number(s). The complete sequence of the UPEC strain NU14 chromosome has been submitted to GenBank under the accession number CP019777 (Bio-Project PRJNA373796).

#### ACKNOWLEDGMENTS

This work was supported by the National Research Foundation, Prime Minister's Office, Singapore Grant number NRF-RF2010-10; the National Medical Research Council, Ministry of Health, Singapore grant number NMRC/CIRG/1358/2013; and the Genome Institute of Singapore (GIS)/Agency for Science, Technology and Research (A\*STAR).

## REFERENCES

- Foxman B. 2002. Epidemiology of urinary tract infections: incidence, morbidity, and economic costs. Am J Med 113(suppl 1A):5S–13S. https:// doi.org/10.1016/S0002-9343(02)01054-9.
- Foxman B. 2010. The epidemiology of urinary tract infection. Nat Rev Urol 7:653–660. https://doi.org/10.1038/nrurol.2010.190.
- Connell I, Agace W, Klemm P, Schembri M, Mărild S, Svanborg C. 1996. Type 1 fimbrial expression enhances *Escherichia coli* virulence for the urinary tract. Proc Natl Acad Sci U S A 93:9827–9832. https://doi.org/10 .1073/pnas.93.18.9827.
- Hung CS, Dodson KW, Hultgren SJ. 2009. A murine model of urinary tract infection. Nat Protoc 4:1230–1243. https://doi.org/10.1038/nprot.2009 .116.
- Mysorekar IU, Hultgren SJ. 2006. Mechanisms of uropathogenic *Escherichia coli* persistence and eradication from the urinary tract. Proc Natl Acad Sci U S A 103:14170–14175. https://doi.org/10.1073/pnas.0602136103.
- Anderson GG, Palermo JJ, Schilling JD, Roth R, Heuser J, Hultgren SJ. 2003. Intracellular bacterial biofilm-like pods in urinary tract infections. Science 301:105–107. https://doi.org/10.1126/science.1084550.
- Hunstad DA, Justice SS. 2010. Intracellular lifestyles and immune evasion strategies of uropathogenic *Escherichia coli*. Annu Rev Microbiol 64: 203–221. https://doi.org/10.1146/annurev.micro.112408.134258.
- Rosen DA, Hooton TM, Stamm WE, Humphrey PA, Hultgren SJ. 2007. Detection of intracellular bacterial communities in human urinary tract infection. PLoS Med 4:e329. https://doi.org/10.1371/journal.pmed .0040329.
- Robino L, Scavone P, Araujo L, Algorta G, Zunino P, Pírez MC, Vignoli R. 2014. Intracellular bacteria in the pathogenesis of *Escherichia coli* urinary tract infection in children. Clin Infect Dis 59:e158–e164. https://doi.org/ 10.1093/cid/ciu634.

- Robino L, Scavone P, Araujo L, Algorta G, Zunino P, Vignoli R. 2013. Detection of intracellular bacterial communities in a child with *Escherichia coli* recurrent urinary tract infections. Pathog Dis 68:78–81. https://doi.org/10.1111/2049-632X.12047.
- Hultgren SJ, Schwan WR, Schaeffer AJ, Duncan JL. 1986. Regulation of production of type 1 pili among urinary tract isolates of *Escherichia coli*. Infect Immun 54:613–620.
- Mulvey MA, Lopez-Boado YS, Wilson CL, Roth R, Parks WC, Heuser J, Hultgren SJ. 1998. Induction and evasion of host defenses by type 1-piliated uropathogenic *Escherichia coli*. Science 282:1494–1497. https://doi.org/10.1126/science.282.5393.1494.
- Hultgren SJ, Duncan JL, Schaeffer AJ, Amundsen SK. 1990. Mannosesensitive haemagglutination in the absence of piliation in *Escherichia coli*. Mol Microbiol 4:1311–1318. https://doi.org/10.1111/j.1365-2958 .1990.tb00710.x.
- Langermann S, Palaszynski S, Barnhart M, Auguste G, Pinkner JS, Burlein J, Barren P, Koenig S, Leath S, Jones CH, Hultgren SJ. 1997. Prevention of mucosal *Escherichia coli* infection by FimH-adhesin-based systemic vaccination. Science 276:607–611. https://doi.org/10.1126/science.276.5312 .607.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/ nmeth.2474.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta)genomic annotation. OMICS 12:137–141. https://doi .org/10.1089/omi.2008.0017.