

Microengineered hydromechanical cochlear model

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Micromachined fluid-filled variable impedance waveguides intended to mimic the mechanics of the passive mammalian cochlea have been fabricated and experimentally examined. The structures were microfabricated with dimensions similar to those of the biological system. Experimental tests demonstrate acoustically excited traveling fluid-structure waves with phase accumulations between 1.5 and 3 π radians at the location of maximum response. The resulting measured frequency-position mapping function, with similarities to that observed in the cochlea, is presented. Results for both isotropic and orthotropic membranes are reported, demonstrating that the achieved orthotropy ratio of 8:1 in tension is insufficient to produce the sharp filtering observed in animal experiments and many computational models that use higher ratios. It is also shown experimentally that high viscosity fluids must be used to provide sufficient damping to avoid the formation of a nonphysiological standing wave pattern. A mathematical model incorporating a thin-layer viscous, compressible fluid approximation coupled to an orthotropic membrane model is validated against experimental results. The work presented herein is motivated by the possibility of producing microfabricated cochlear-like filters, thus the structure is designed for production in a scalable microfabrication process.

acoustic | micro-electro-mechanical systems | sensor | viscosity

The cochlea is the organ in the inner part of the mammalian ear that is responsible for the transduction of acoustic signals into neurological signals. The typical human cochlea operates over a 3-decade frequency band, from 20 Hz to 20 kHz, covers 120 dB of dynamic range, and can distinguish tones that differ by <0.5% (1). The cochlea is also very small, occupying a volume of ≈ 1 cm³. Perhaps most importantly, the cochlea uses a mechanical process to separate audio signals into $\approx 3,500$ channels of frequency information. Thus, the cochlea is a sensitive real-time mechanical frequency analyzer.

The effectiveness of the cochlea as a time-frequency analyzer motivates our efforts to construct a hydromechanical analog that can be fabricated repeatably and in a batch fashion. Integration of sensing elements with the mechanical structure would result in a combined acoustic sensor/filter with a unique operating modality. During the design of the mechanical structure, we also learn more about some aspects of the biological cochlea.

von Békésy's (2, 3) observation of traveling waves on the basilar membrane (BM) of cadaver cochleae motivated the construction of a number of physical models of the cochlea. The first group of these models involved scaled-up versions of the cochlea. Helle's (4) model is seven times life size and exhibits tonotopic structure in the 25- to 800-Hz band. Chadwick *et al.* (5) built a 24 times life-size model and clearly show strong standing wave patterns and the resulting discrete resonances. Because of the standing wave nature of their response, a tonotopic map is not demonstrated (5). Lechner's (6) model is 14 times life size and demonstrates experimentally the improved filter sharpening and nonlinear compression that result from adding nonlinear feedback forcing of the structure.

A few microscale models with life-size dimensions have been reported. Zhou *et al.* (7) demonstrate a tonotopic map over the 300- to 15-kHz band, with phase accumulations of approximately π radians at the location of maximum response. Despite

its small size, their model was not fabricated by using scalable batch micromachining methods. Additional work in recent years by Hemmert *et al.* (8) and Lim *et al.* (9) demonstrates fluid structure traveling waves in life-size cochlear models. Both of these models use a micromachined structure for the cochlear partition but a conventionally machined housing for the fluid ducts.

The results reported in this article take the next step in the development of physical cochlear models. In this work, we demonstrate, through both theory and experiment, that a fully micromachined cochlear analog can achieve a cochlear-like phase response and a place-to-frequency map. The effects of both membrane orthotropy and fluid viscosity on the sharpness of the mechanical filter and the place-frequency map are explored experimentally and mathematically. The batch micromachining process used to fabricate the system will allow future integration of sensing elements into the structure to economically produce low-power, micromechanical, cochlear-like sensor/filters.

Cochlear Model Design

The cochlea is a spiral-shaped organ consisting mainly of ducts filled with ion-rich water-like fluids. Acoustic energy is injected from the outer and middle ears. A fluid-structure wave then travels down the length of the cochlear spiral, interacting with the microstructures in the organ of Corti and the BM. The helicotrema provides pressure equalization at the apex of the cochlea (10).

The BM changes in width, thickness, and stiffness along the length of the cochlear spiral. Because of its changing mechanical impedance, high-frequency sounds excite amplified BM motion near the base of the cochlea (close to the stapes), and low-frequency sounds excite motion near the apex of the cochlea (the termination of the cochlear spiral). In addition, the BM is orthotropic, as it is stiffer transversely than longitudinally because of transversely oriented fibers and transversely arranged supporting cells in the organ of Corti (11). This orthotropy is thought to enhance filter sharpness, particularly at high frequencies. Schematics and additional description of the physiology can be found in Dallos *et al.* (1) and Geisler (10).

The simplest mechanical analog of the cochlea is a single fluid-filled, rigid-walled duct constrained along one side by a variable width membrane or plate. A number of authors have argued that the coiling of the cochlea is a secondary effect (12, 13), and so is neglected. Recent work by Manoussaki and Chadwick (14) explores the effects of coiling in detail. Use of a single uncoiled fluid duct rather than two coiled ducts reduces the complexity of the fabrication process considerably and facilitates mechanical measurements. A single-duct physical model corresponds to a mathematical model of the cochlea where only antisymmetric pressure contributions are included (15).

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Abbreviations: BM, basilar membrane; cSt, centiStokes.

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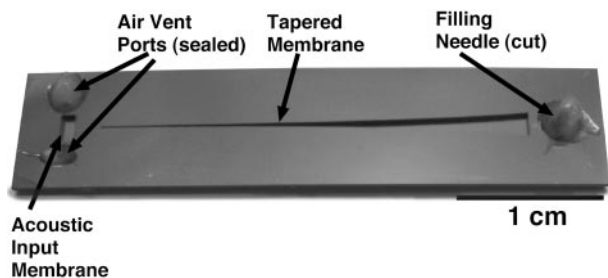


Fig. 3. Photograph of a finished device, after bonding, filling with fluid, and sealing. Hardened epoxy is visible sealing the fluid ports.

An acoustic baffle with an internally mounted electromagnetic speaker fits over the front end of the device. The baffle ensures that acoustic energy delivered by the speaker impinges primarily on the input membrane of the sample under test while enabling direct optical interrogation of the tapered membrane. A Larson-Davis (Provo, UT) 2520 0.25-in diameter microphone is mounted inside the baffle. The exact position of the microphone inside the baffle has an effect on the measured pressure level, particularly for the higher-frequency experiments. The uncertainty in the magnitude of the delivered pressure is 10 dB. Drive levels are between 85 and 105 dB re 20 μ Pa.

At each drive frequency, structure velocity is measured at 1,400 data points in a 7×200 grid over the area of the exponentially tapered membrane. A DSP lock-in amplifier [Stanford Research (Sunnyvale, CA) SR830] is used to determine the magnitude and phase, relative to the speaker drive voltage, of the structure velocity at the excitation frequency. The noise floor of the measurements is ≈ 0.01 nm.

Experimental Results

Results are presented for both isotropic and orthotropic membrane devices, filled with either 5- or 20-cSt silicone oil. Each set of results is for a unique device, but all devices come from the same wafer. Fig. 5 shows measurements of the magnitude and phase of membrane response for five driving frequencies. The phase is reported along the centerline of the membrane.

A cochlear-like frequency position map is present, as seen in the magnitude data. The best example is shown in Fig. 5c. At 4.2 kHz, the peak response occurs at 21 mm, with the peak moving to a location of 7.5 mm for a 35-kHz drive. The orthotropic cases show only slight sharpening of the response at 4.2 and 7.2 kHz, as seen by a comparison of Fig. 5c and d. For some cases, most notably 12.3 and 20 kHz in Fig. 5a and 12.3 kHz in Fig. 5d, second structural cross-modes are present. This type of response is antisymmetric with a nodal line on the

membrane center. The maximum response of this second mode occurs at a location closer to the wide end of the membrane than does the maximum first mode response.

The fluid-structure waves are predominantly traveling waves, as seen from the phase response in Fig. 5e-h. At the location of maximum response for the first symmetric structural mode, the total phase accumulation is in the range of 5 to 10 radians. For the lower viscosity cases at 4.2 and 7.2 kHz, a partial standing wave pattern is generated close to the reflecting boundary at 30 mm. This pattern can be seen from the π steps in phase appearing between 20 and 30 mm in Fig. 5e and f. Increasing the viscosity to 20 cSt causes the waves to damp out more rapidly after reaching the location of maximum response, suppressing the standing wave pattern.

One set of experiments (for the 20-cSt orthotropic case) was repeated after a 3-month interval. At four frequencies, the phase results and the shape of the magnitude response were identical. At 20 kHz, a backward-propagating, rather than forward-propagating, traveling wave was excited, perhaps caused by changes in the precise alignment of the speaker. The absolute magnitude of the response, relative to the measured excitation pressure amplitude, varied by <10 dB from the set of experiments shown here. This variation probably is caused by uncertainties in the measurement of the excitation level. At 4.2 and 7.2 kHz, additional tests were conducted with drive levels two and three times the initial drive level to test the linearity of the system. No level-dependent change was observed in the phase or relative magnitude response.

Modeling

Mathematical modeling of this system requires the solution of a coupled fluid structure problem. Many other authors have described mathematical models of the passive cochlea, both analytical and numerical. Steele and Taber (19) and de Boer and Viergever (20), among others, present 3D cochlear models by using the Wentzel-Kramer-Brillouin asymptotic technique. A number of finite element models of the cochlea have also been described, such as those by Parthasarathi *et al.* (15), Kolston and Ashmore (21), and Kagawa *et al.* (22). A direct application of these methods is not appropriate for the system considered in this study because the viscous boundary layer occupies a large fraction of the 110- μ m duct height, causing these approaches to underestimate the viscous fluid damping.

The finite element model described herein is taken from Beltman *et al.* (23). This model couples a thin, viscous, compressible acoustic medium to a structure, differing from the majority of cochlear models by focusing on viscous fluid effects. The thin gap approximations are valid for fluid film heights much less than any dimension of the response. In the case considered here, the wavelength of the response in the longitudinal direction is always >2 mm, 20 times the fluid height. The variation of the response in the transverse direction occurs over the membrane width. Hence the approximation is valid over all but the narrowest part of the membrane, where some errors may be introduced. The assumptions allow the problem to be rendered in two dimensions, with only fluid pressure and structure displacement as free variables. This process reduces the problem size, enabling a more refined solution in the x - y plane, where all of the geometric features of interest lie.

Eq. 1 models the fluid by using a modified 2D Helmholtz equation with viscous damping and structural coupling included. Viscosity enters by modifying the wave speed to a complex effective wavespeed, c_{eff} . Note that all damping terms are controlled by the dimensionless parameter δ/h_0 , the ratio of the viscous boundary layer thickness to the duct height.

P is the acoustic pressure in the fluid, w_p is the membrane displacement, h_0 is the duct height, T_x and T_y are the membrane

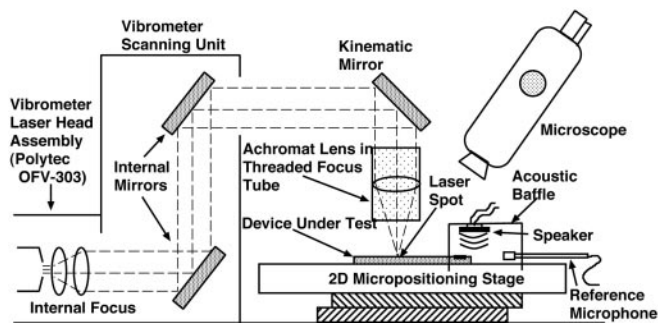


Fig. 4. Schematic of the experimental setup.

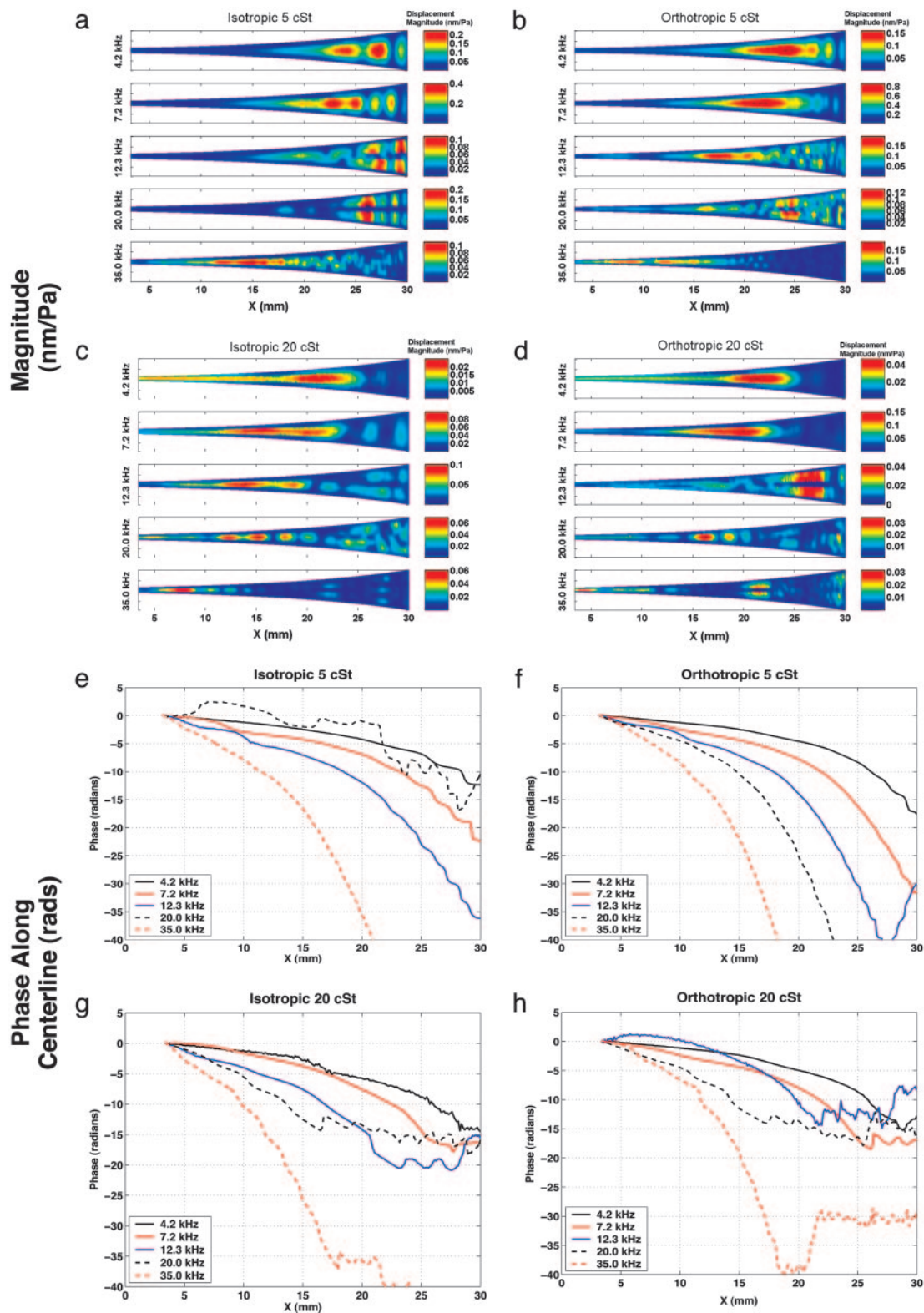


Fig. 5. Experimental results show magnitude and phase of membrane displacement response at five excitation frequencies, normalized to the input acoustic pressure, for isotropic membrane with 5-cSt fluid (a and e), orthotropic membrane with 5-cSt fluid (b and f), isotropic membrane with 20-cSt fluid (c and g), and orthotropic membrane with 20-cSt fluid (d and h). Note that because of optical constraints the entire membrane is not imaged; the data start at 4 mm from the narrow end of the device. Centerline plots of the magnitude results are available in Fig. 9, which is published as supporting information on the PNAS web site.

tensions in the x and y directions, respectively, m_a is the membrane mass per unit area, ρ_0 is the unperturbed fluid density, ω is the angular frequency, c_0 is the acoustic free wave

speed in the fluid, μ is the fluid viscosity, δ is the viscous boundary layer thickness, x, y, z are Cartesian coordinates, and j is the square root of -1 .

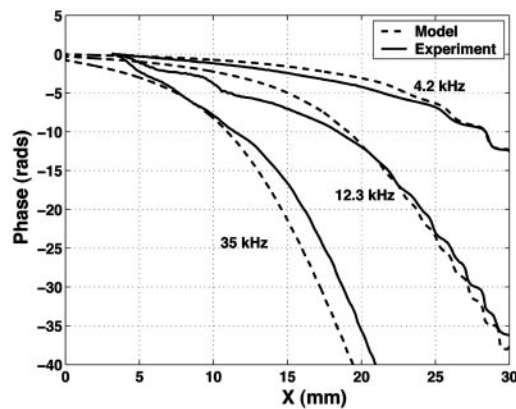


Fig. 7. Comparison of phase results between model and experiment for an isotropic membrane with 5-cSt filling fluid.

Discussion

Both the physical and mathematical models demonstrate acoustically excited fluid-structure traveling waves that produce a passive cochlear-like frequency-position map in the 4- to 35-kHz band. At the location of maximum response, the phase accumulation is $1.5\text{--}3\pi$ radians. This finding is similar to experimental data for guinea pigs, chinchillas, squirrel monkeys, and cats, which show average phase accumulations of 3π at the best frequency for a given location (24).

An examination of both experimental and theoretical results, shown in Figs. 5 and 6, indicates that to suppress the formation of nonphysiological standing waves at low frequencies 20-cSt fluid is required. However, even with this introduction of additional passive damping, the rate of cutoff of the traveling wave is sharper in animal experiments than in our model. This finding suggests that additional effects present in the cochlea, such as correctly phased active excitation by the outer hair cells, produce the sharp filtering. The helicotrema, which connects the scala at the apex of the cochlea, is not included in our model and could also provide some of the needed damping for the lowest frequencies.

Another way to increase the sharpness and gain of the mechanical filter, in theory, is to increase the orthotropy ratio. In the limit, as is done in some mathematical cochlear models, longitudinal stiffness of the BM is entirely neglected (15, 20). However, in construction of a physical model, it has proved difficult to produce membrane orthotropy ratios beyond a ratio of 8:1 in tension [resulting in a ratio of 3:1 for the transverse to longitudinal space constants, as defined by Naidu and Mountain (17)]. Experimental and mathematical results presented here show little improvement in filtering with such an orthotropy ratio. The recent experimental results by Naidu and Mountain (17) indicate that physiologically the longitudinal stiffness of the BM, while lower than transverse stiffness, is not negligible. In fact, the space constants measured by

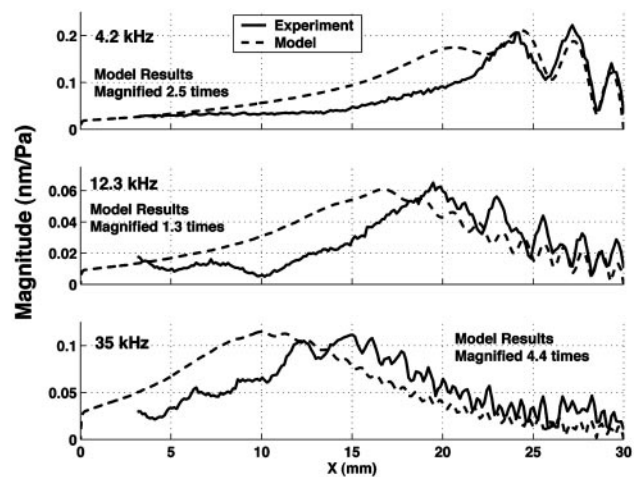


Fig. 8. Comparison of magnitude results between model and experiment for an isotropic membrane with 5-cSt filling fluid. The magnitude of the model results is magnified by the given factors to allow comparison of the shape of the response.

Naidu and Mountain that characterize longitudinal coupling in the gerbil BM are similar to those computed for the structure presented in this article. Thus, use of very high orthotropy ratios to produce sharp filtering in a model appears to not be directly physical, but might be better considered a convenient way to represent some more complex phenomenon occurring *in vivo*. Possibilities include the active processes related to outer hair cell operation or additional passive mechanical tuning resulting from interactions of the BM with the tectorial membrane. See Geisler (ref. 10, pp. 125–138) and Dallos *et al.* (ref. 1, pp. 202–203) for discussion of these mechanisms.

Comparisons of the mathematical results with measured response, such as those shown in Figs. 7 and 8, demonstrate the accuracy of the thin film fluid model. Most significantly, the effects of viscous damping are captured correctly and efficiently. To produce a good match, it is critical that the rigid boundary conditions used in the mathematics are a good approximation.

A micromachined device such as that presented here offers a unique modality for acoustic sensing. A passive device would be capable of low-power acoustic sensing and mechanical filtering, delivering multiple channels of filtered information from sensors spatially distributed along the length of the device. Both passive and active systems are made practical through the use of micro-electro-mechanical systems technology. A microfabrication process for the mechanical subsystem has been demonstrated in this work. The next step is to incorporate sensing elements and active feedback mechanisms into the structure.

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- Dallos, P., Popper, A. & Fay, R., eds. (1996) *The Cochlea*, Springer Handbook of Auditory Research (Springer, New York), Vol. 8.
- von Békésy, G. (1970) *Nature* **225**, 1207–1209.
- von Békésy, G. (1960) *Experiments in Hearing* (McGraw-Hill, New York).
- Helle, R. (1977) *J. Audiol. Technique* **16**, 138–163.
- Chadwick, R. S., Fournay, M. E. & Neiswander, P. (1980) *Hear. Res.* **2**, 475–483.
- Lechner, T. P. (1993) *Hear. Res.* **66**, 202–212.
- Zhou, G., Bintz, L., Anderson, D. Z. & Bright, K. E. (1993) *J. Acoust. Soc. Am.* **93**, 1516–1523.
- Hemmert, W., Durig, U., Despont, M., Drechsler, U., Genoelt, G., Vettiger, P. & Freeman, D. M. (2002) in *Biophysics of the Cochlea: From Molecules to Models*, ed. Gummer, A. W. (World Scientific, Teaneck, NJ), pp. 409–416.
- Lim, K.-M., Fitzgerald, A. M., Steele, C. R. & Puria, S. (1999) in *Recent Developments in Auditory Mechanics*, eds. Wada, H., Takasaka, T., Ikeda, K., Ohyama, K. & Koike, T. (World Scientific, Teaneck, NJ), pp. 223–229.
- Geisler, C. D. (1998) *From Sound to Synapse* (Oxford Univ. Press, London).

- Iurato, S. (1962) *J. Acoust. Soc. Am.* **34**, 1386–1395.
- Loh, C. H. (1984) *J. Acoust. Soc. Am.* **74**, 95–103.
- Steele, C. R. & Zais, J. G. (1985) *J. Acoust. Soc. Am.* **77**, 1849–1852.
- Manoussaki, D. & Chadwick, R. S. (2000) *SIAM J. Appl. Math.* **61**, 369–386.
- Parthasarathi, A. A., Grosh, K. & Nuttall, A. L. (2000) *J. Acoust. Soc. Am.* **107**, 474–485.
- Zwislocki, J. J. (1948) *Acta Otolaryngol.*, Suppl., **72**, 1–76.
- Naidu, R. & Mountain, D. (2001) *J. Assoc. Res. Otolaryngol.* **2**, 257–267.
- Voldrich, L. (1978) *Acta Otolaryngol.* **86**, 331–335.
- Steele, C. R. & Taber, L. A. (1979) *J. Acoust. Soc. Am.* **65**, 1007–1018.
- de Boer, E. & Viergever, M. A. (1982) *Hear. Res.* **8**, 131–155.
- Kolston, P. J. & Ashmore, J. F. (1996) *J. Acoust. Soc. Am.* **99**, 455–467.
- Kagawa, Y., Yamabuchi, T., Watanabe, N. & Mizoguchi, T. (1987) *J. Sound Vib.* **119**, 291–315.
- Beltman, W. M., vanDerHoogt, P., Spiering, R. & Tjeldeman, H. (1998) *J. Sound Vib.* **216**, 159–185.
- Robles, L. & Ruggero, M. A. (2001) *Physiol. Rev.* **81**, 1305–1352.