Moderately Elevated Homocysteine Does Not Contribute to Thoracic Aortic Aneurysm in Mice

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Abstract

Background: Moderate hyperhomocysteinemia is an attractive target for intervention because it is present in 5–7% of the population and can be reversed by diet. This approach presupposes that hyperhomocysteinemia is directly involved in the disease process. Epidemiologic studies have indicated that moderately elevated homocysteine may contribute to thoracic aortic aneurysm (TAA) dilatation and dissection in humans. In vitro, elevated homocysteine disrupts the structure and function of extracellular matrix components, suggesting that moderate hyperhomocysteinemia may contribute to the development and/or progression of TAA.

Objective: We investigated moderately elevated homocysteine in the development and progression of TAA in a mouse model of Marfan syndrome (MFS) and in isogenic wild-type mice. The MFS mouse is a well-described model of a systemic connective tissue disorder characterized by thoracic aortic dilatation, dissection, and rupture. We used this model as a sensitized indicator system to examine the impact of homocysteine on the progression of TAA.

Methods: Murine fibrillin 1 gene (Fbn1)^{C1039G/+} MFS and C57BL/6J wild-type mice were fed a cobalamin-restricted diet to induce moderate hyperhomocysteinemia from weaning until the age of 32 wk. Homocysteine and methylmalonic acid were measured and aortic root diameter assessed with the use of echocardiography in mice aged 3, 7, 15, and 32 wk. **Results:** Cobalamin-restricted mice exhibited significantly higher homocysteine ($P < 0.0001$) and methylmalonic acid (P < 0.0001) in the blood. For both strains, no significant difference in thoracic aortic diameter was observed in mice on the cobalamin-restricted diet compared with those on the control diet.

Conclusions: Fbn1^{C1039G/+} mice are a well-characterized model of progressive aortic root dilation. Hyperhomocysteinemia in the physiologic range did not induce abnormal aortic growth in wild-type mice and did not accelerate or otherwise influence aortic root growth and pathologic progression in mice with an underlying predisposition for aortic dilatation. J Nutr 2017;147:1290–5.

Keywords: thoracic aortic aneurysm, homocysteine, methylmalonic acid, Marfan syndrome, vitamin B-12, cobalamin, mouse model

Introduction

Thoracic aortic aneurysm (TAA) is defined as a segmental fullthickness dilation of the aorta that measures \geq 50% larger than normal for a given sex, age, and body size. Various etiologies have been associated with TAA; the most common is a degenerative

Abbreviations used: Fbn1, murine fibrillin 1 gene; MFS, Marfan syndrome; MMA, methylmalonic acid; TAA, thoracic aortic aneurysm.

process associated with hypertension, age, and tobacco use. Other causes include atherosclerotic disease, genetic syndromes [e.g., Marfan syndrome (MFS)], bicuspid aortic valve, aortitis, and trauma. Overall, TAA occurs with an estimated annual incidence of 5.9–10.4 cases/100,000 patient-years (1, 2). The most severe complications of TAA include dissection, which may cause arterial occlusion and end-organ ischemia, and rupture, which is usually fatal. Approximately 50% of patients with TAA rupture die before reaching the hospital (3). Emergent surgical repair carries an additional mortality risk of $25 - 50\%$ (3).

Aneurysm size is the most important determinant of TAA rupture risk. In asymptomatic patients the risk of TAA rupture is near zero for aneurysms <5.0 cm, 1.7%/y for aneurysms

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5.0–5.9 cm, and 3.6%/y for aneurysms ≥ 6.0 cm (4). The risk of rupture, dissection, or death from all causes is 6.5%/y for 5.0– 5.9-cm aneurysms and 14.1%/y for aneurysms ≥ 6.0 cm (4). Elective preemptive surgical repair restores survival to rates similar to those in matched control individuals but carries a \geq 2.5% risk of death, even at experienced surgical centers (4). Other perioperative complications include myocardial infarction, stroke, paraplegia, paraparesis, and acute renal failure requiring dialysis. Current medical management of TAA relies on an aggressive antihypertensive treatment to slow aneurysmal growth (5). Additional interventions may include lipidlowering therapy, smoking cessation, and lifestyle modifications (5). However, even with adequate medical management, many individuals with TAA will meet the criteria for surgical intervention. Given the substantial morbidity and mortality associated with TAA repair, it is important that additional therapies be developed. An understanding of risk factors that contribute to disease severity and progression would assist in this process.

In adults, the normal range of circulating homocysteine is \sim 5–14 µmol/L and is known to vary by age and sex (6). Hyperhomocysteinemia is defined as exceeding these concentrations and can be further categorized based on homocysteine concentrations into severe (>100 µmol/L), intermediate $(>30-100 \mu \text{mol/L})$, and moderate $(15-30 \mu \text{mol/L})$ (7). Severe hyperhomocysteinemia can be caused by severe cobalamin deficiency or inborn errors of metabolism and is rare $\langle 0.02\% \rangle$; intermediate hyperhomocysteinemia caused by renal failure or moderate to severe cobalamin or folate deficiency is also not common (<1%) (8, 9). Moderate hyperhomocysteinemia can be caused by mild folate or cobalamin deficiency or impaired renal function and is much more common (6, 8). Moderately elevated homocysteine concentrations have been associated with acute thoracic aortic dissection (mean \pm SD homocysteine: 22 \pm 14 μ mol/L) and chronic aortic aneurisms (mean \pm SD homocysteine: 17 \pm 5μ mol/L) (10). Moderately elevated concentrations of homocysteine have also been observed in individuals with severe cardiovascular manifestations of MFS [median homocysteine: 13.5 µmol/L (IQR: 9, 23.1 µmol/L)] compared with those with no cardiovascular manifestations [median homocysteine: 7.5 μ mol/L (IQR: 7, 9 μ mol/L)] (11). Similarly, moderate hyperhomocysteinemia has been demonstrated in aortic dilation and dissection outside the thoracic cavity, including abdominal aortic aneurysm (12) and spontaneous cervical artery dissection [median homocysteine: 18.2 μ mol/L (IQR: 14.3, 30 μ mol/L)] (13). Taken together, these findings suggest that elevated homocysteine concentrations may influence the severity and kinetics of arterial dilation, dissection, and rupture across the vasculature. Some have theorized that hyperhomocysteinemia may contribute to this process by causing structural changes in the extracellular matrix (14). Fibrillin-1 is a major component of the 10–12-nm microfibrils and largely consists of 2 types of disulfide-rich motifs: 47 epidermal growth factor (EGF)-like domains and 7 transforming growth factor β -binding protein-like domains (15). Homocysteine has a higher acid dissociation constant for the thiol group than Cys and may alter the structural integrity, stability, and/or function of fibrillin-1 by reducing disulfide bonds in the Cys-rich protein (16). The damaging effect of hyperhomocysteinemia on fibrillin-1 is supported by experiments that have demonstrated that homocysteine treatment of a fibrillin peptide fragment containing 3 calcium-binding epidermal growth factor domains (domains 32–34) increased the risk for proteolysis (15).

Although current evidence suggests that moderate hyperhomocysteinemia may contribute to TAA pathology, a clear relation has not been established. The hyperhomocysteinemia

associated with TAA may simply be a marker for another biological change or environmental exposure. To explore the role of moderate hyperhomocysteinemia in TAA further, we used a nutritional intervention to examine the effects of elevated homocysteine on thoracic aorta diameters in C57BL/6J and MFS mice. In a recent study (17), C57BL/6J mice aged 15 wk maintained on a cobalamin-restricted diet showed significant increases in mean homocysteine concentrations (8.7 or 10.5 μ mol/L in mice on different cobalamin-restricted diets compared with 4.2μ mol/L in controls). This moderate hyperhomocysteinemia resulted from a decrease in the activity of methionine synthase, a cobalamin-dependent cytoplasmic enzyme that methylates homocysteine to Met.

MFS is a systemic connective tissue disorder characterized by ocular, skeletal, and cardiovascular system manifestations, including TAA. It is caused by the disruption of the fibrillin-1 domain structure and function with missense mutations that either create or substitute a commonly reported Cys residue (15). Because moderate hyperhomocysteinemia may also modify fibrillin-1, MFS was a particularly informative and sensitive model for our study. In addition, advances in the treatment of MFS currently guide therapy for all causes of TAA. Identifying a risk factor that contributes to TAA in MFS would assist in the development of new therapies for all underlying etiologies.

Methods

Mice. We studied 2 independent groups of mice. To establish a model of diet-induced hyperhomocysteinemia, wild-type C57BL/6J mice and their dams were maintained either on a control diet with vitamin B-12 or on a standard unpurified diet until being switched to a cobalaminrestricted diet at weaning. Distributions by sex and diet are described in Supplemental Table 1. Aortic roots were not measured in these mice because they were used to determine the long-term biochemical impact of these diets. The mice were maintained at the NIH, and the National Human Genome Research Institute Animal Care and Use Committee approved all experiments that involved dietary manipulations.

Independent parallel experiments were performed with the use of a previously described mouse line harboring the fibrillin-1 gene mutation C1039G (heterozygous Fbn1^{C1039G/+} MFS mice) (18). All MFS mice were back-crossed (>10 generations) onto the C57BL/6J background, allowing valid comparisons between litters and with wild-type C57BL/6J mice. A total of 43 mice were used in this study. Approximately 50% of the mice were maintained on a cobalamin-restricted diet to raise plasma total homocysteine from weaning (age 3 wk) until the age of 32 wk (a total of 29 wk on the diet). The remaining mice were maintained on a control diet after weaning. Distribution by genotype, sex, and diet are described in Supplemental Table 2. Homocysteine and methylmalonic acid (MMA) measurements were obtained when the mice were aged 15 wk. Aortic root diameter was assessed in vivo in mice aged 3, 7, 15, and 32 wk with the use of echocardiography. Wild-type littermates were used as controls. All experiments were approved by the Johns Hopkins University School of Medicine Animal Care and Use Committee.

Defined diets. Two soy protein-based mouse diets were custom-blended by Harlan Teklad Laboratories (Supplemental Tables 3 and 4). The cobalamin-restricted diet was derived completely from plant sources and lacked any vitamin B-12. The control diet was supplemented with 50 μ g vitamin B-12/kg unpurified diet. Distinguishing food-safe dyes were added to each formulation to ensure that the correct diet was used. Both formulations included the antibiotic succinylsulfathiazole (1% wt:vol) to limit the intestinal production of vitamin B-12 from gut flora. All unpurified diets were sterilized via γ irradiation. In all experiments, food and water were available ad libitum. Weight gain and growth during the course of the experiments were not measured, but no overtly striking differences in size were observed between the different treatment groups.

FIGURE 1 Metabolic measurements of mice aged 12 and 24 wk on the CRD or CTRL diet for 9 and 21 wk, respectively. The squares, circles, and up and down triangles indicate the values obtained from an individual mouse. (A) Homocysteine. (B) Methylmalonic acid. Homocysteine and methylmalonic acid concentrations were significantly elevated in wild-type C57BL/6J mice on the CRD. Homocysteine and methylmalonic acid concentrations in mice aged 29 wk were significantly higher than similarly aged mice on the CTRL diet. Data are presented as medians \pm IQRs. $*P < 0.05$, $*P < 0.01$, $*P < 0.001$, and $***P < 0.0001$. CRD, cobalamin-restricted diet; CTRL, control.

Blood chemistry. Blood for serum homocysteine and MMA was collected in mice aged 15 wk via the retro-orbital sinus cavity. A tetracaine ophthalmic anesthetic was used as an adjunct analgesia for retro-orbital sinus bleeding in isoflurane-anesthetized mice. All mice were deeply anesthetized with the use of isoflurane (4–5% isoflurane in 100% O_2) until the respiration rate was greatly reduced. The plane of anesthesia was confirmed by the absence of a deep pain response (i.e., withdrawal or twitching) to the pinching of the foot or toe with blunt forceps. Volumes of 0.1–0.2-mL peripheral blood were collected from the retro-orbital sinus cavity into heparinized microcapillary tubes (Sarstedt). Blood was expelled into microcentrifuge tubes, placed on ice, and centrifuged (6800 \times g; 10 min; 4°C). The upper plasma layer was immediately transferred to a new microcentrifuge tube, diluted 1:5 in

FIGURE 2 Metabolite measurements of mice aged 15 wk on the CRD or CTRL diet for 12 wk. The squares, circles, triangles, and diamonds indicate the values obtained from an individual mouse. (A) Homocysteine. (B) Methylmalonic acid. Homocysteine and methylmalonic acid concentrations were significantly elevated in mice on the CRD. No significant difference was observed in Fbn1^{C1039G/+} MFS mice compared with CTRL wild-type C57BL/6J mice in either the CRD or CTRL diet groups. For clarity, an outlying methylmalonic acid measurement (23.17 μ mol/L) was not included in the graphical representation of methylmalonic acid values. This datum is included in our statistical analysis. Data are presented as medians \pm IQRs. **P < 0.01, ***P < 0.001, and $***P < 0.0001$. CRD, cobalamin-restricted diet; CTRL, control; Fbn1, murine fibrillin 1 gene; MFS, Marfan syndrome.

deionized water, and frozen at -80° C. MMA and homocysteine were assayed in diluted plasma samples via HPLC.

Echocardiography. Echocardiographic assessment was performed in mice aged 3, 7, 15, and 32 wk. Nair hair removal cream was used on all mice the day before echocardiograms. All echocardiograms were performed on mice that were awake and unsedated with the use of the Vevo 660 V1.3.6 imaging system and an RMV603 40-MHz transducer (VisualSonics). The aorta was imaged in the parasternal long-axis view. Three separate measurements of the maximal internal dimension at the sinus of Valsalva were made from distinctly captured images and averaged. All studies were interpreted by a single echocardiographer who was blinded to the genotype and diet.

Statistical analysis. Statistical analysis was performed with the use of a 2 tailed Mann-Whitney U test. P < 0.05 was considered significant. Statistical analyses were performed with GraphPad Prism version 5.0 for MacOS X.

Results

Cobalamin restriction raised homocysteine and MMA in wild-type C57BL/6J mice. C57BL/6J mice maintained on a cobalamin-restricted diet postweaning exhibited higher homocysteine concentrations at \sim 15 and \sim 29 wk of age than similarly aged mice on the control diet (Figure 1A). The mice aged 29 wk developed moderate hyperhomocysteinemia compared with similarly aged mice on the control diet (Mann-Whitney $U = 7$; $P = 0.0136$) (Figure 1A). The mean homocysteine measurement for the C57BL/6J mice on the cobalamin-restricted diet was 14.9 mmol/L, a 2-fold increase in mean circulating homocysteine induced by dietary cobalamin restriction. Concentrations of circulating homocysteine increased with age and exposure to the cobalamin-restricted diet.

To confirm that the dietary manipulations increased homocysteine concentrations by reducing in vivo concentrations of cobalamin, we also measured plasma MMA in all mice (Figure 1B). This metabolite was elevated when the activity of the cobalamin-requiring enzyme, methylmalonyl CoA mutase, was reduced. MMA concentrations in mice aged 29 wk were significantly higher (Mann-Whitney $U = 0$; $P = 0.0016$) in those maintained on the cobalamin-restricted diet than those maintained on the control diet (medians: 222.4 and $2.9 \mu m o l / L$, respectively). Similar to the homocysteine response, MMA

concentrations were notably higher in mice on the cobalaminrestricted diet, and this effect increased with age.

Cobalamin restriction raised homocysteine and MMA in Fbn1^{C1039G/+} MFS mice. In an independent cohort of C57BL/6J and $Fbn1^{\text{C1039G}/+}$ MFS mice, the same dietary effect on homocysteine and MMA was observed (Figure 2). When these strains were analyzed together, mice aged 15 wk on the cobalamin-restricted diet had a median homocysteine measurement of 5.1 μ mol/L compared with 3.5 μ mol/L for mice on the control diet. This represented a significant difference between the 2 groups (Mann-Whitney $U = 30; P < 0.0001$) (Figure 2A).
When examined separately, the Ehra^{(C1039G/+} MES mice on the When examined separately, the $Fbn1^{\text{C1039G}/+}$ MFS mice on the cobalamin-restricted dist also had significantly bigher homocyscobalamin-restricted diet also had significantly higher homocysteine measurements than $Fbn1^{\text{C1039G}/+}$ MFS mice on the control diet (median: 5.2 and 3.45 μ mol/L, respectively) (Mann-Whitney $U = 12.5$; $P = 0.0002$). A similar dietary effect on homocysteine was observed in C57BL/6J mice (Figure 2A); an increase of homocysteine concentrations was observed in mice on the cobalamin-reduced diet regardless of genotype. No significant difference was observed in homocysteine measurements of MFS mice on the cobalamin-restricted diet compared with C57BL/6J mice on the cobalamin-restricted diet. Similarly, there was no significant difference in the homocysteine measurements of MFS mice on the control diet compared with C57BL/6 mice on the control diet.

When C57BL/6J and $Fbn1^{\text{C1039G}/+}$ MFS mice were analyzed together, MMA measurements were significantly higher in mice on the cobalamin-restricted diet (median: 2.81 µmol/L) than those on the control diet (median: 0.73 µmol/L) (Mann-Whitney $U = 0$; $P < 0.0001$) (Figure 2B). When $Fbn1^{\text{C1039G}/+}$ MFS mice were evaluated separately, significantly different MMA values were also observed in the cobalamin-restricted mice (median: 2.83μ mol/L) compared with those on the control diet (median: 0.73 μ mol/L) (Mann Whitney U = 0; P < 0.0001). No difference was noted for MMA measurements between MFS mice and C57BL/6J mice on the cobalamin-restricted diet or between MFS and C57BL/6J mice on the control diet.

Moderately elevated homocysteine did not correlate with thoracic aortic diameter. Longitudinal echocardiography was performed on mice aged 3, 7, 15, and 32 wk. $Fbn1^{\text{CI}039\text{G}/\text{i}}$ MFS mice with or without the cobalamin-restricted diet (higher homocysteine) had enlarged aortic roots (maximal dimension at

FIGURE 3 Echocardiographic measurements. The squares, circles, and up and down triangles indicate the values obtained from an individual mouse. (A) Absolute aortic root diameter measured by echocardiography of wild-type ($n = 16$) and $Fbn1^{C1039}G/+$ MFS mice ($n = 27$) aged 3 wk (baseline measurement before feeding with the diets). (B) Absolute aortic root diameter measurements of mice aged 7 wk on the diet for 4 wk. (C) Absolute aortic root diameter measurements of mice aged 15 wk on the diet for 12 wk. (D) Absolute aortic root diameter measurements of mice aged 32 wk on the diet for 29 wk. There was no significant difference in the aortic root diameter of the Fbn1^{C1039G/+} MFS mice and CTRL wild-type C57BL/6J mice between the CTRL diet cohort ($n = 14$) and CRD cohort ($n = 13$). Data are presented as medians \pm IQRs. ****P < 0.0001. CRD, cobalamin-restricted diet; CTRL, control; Fbn1, murine fibrillin 1 gene; MFS, Marfan syndrome.

the sinus of Valsalva) compared with wild-type C57BL/6J mice (aortic root size at 32 wk: 1.97 \pm 0.11 compared with 1.64 \pm 0.07 mm; P < 0.0001), but no significant difference was observed
between Fhu1^{C1039G/+} MES mice on the cobalamin-restricted diet between Fbn1^{C1039G/+} MFS mice on the cobalamin-restricted diet
(higher homocysteine) compared with MFS mice on the control (higher homocysteine) compared with MFS mice on the control diet (aortic root size at 32 wk: 1.97 \pm 0.10 compared with 1.97 \pm 0.13 mm; $P = 0.9$) (Figure 3). Likewise, no significant difference was noted in the aortic root diameter of wild-type C57BL/6 mice on the cobalamin-restricted diet compared with wild-type C57BL/6J mice on the control diet (aortic root size at 32 wk: 1.64 ± 0.09 compared with 1.64 ± 0.04 mm; $P = 0.9$).

Discussion

Epidemiologic studies and some basic laboratory investigations have suggested that hyperhomocysteinemia is positively correlated with thoracic aortic dilation, dissection, and rupture. Moderate hyperhomocysteinemia produced by genetic changes and diet is present in 5–7% of the population and in many cases may be reversed with nutritional interventions (6, 19). Therefore, it presents an attractive target for medical therapy and merits further investigation.

We used a nutritional intervention to raise homocysteine 1.5– 2-fold above the normal physiologic concentrations in mice and to examine its role in TAA. We demonstrated an elevated mean homocysteine in the moderate range $(\sim 15 \mu mol/L)$ in mice aged 29 wk when they were placed on the cobalamin-restricted diet at weaning (Figure 1A). To confirm a similar trend in both genotypes, a single measurement of homocysteine and MMA was obtained in all mice aged 15 wk (Figure 2). This nutritional intervention allowed us to measure the effect of moderately elevated homocysteine in normal mice and in mice genetically engineered to be highly susceptible to TAA. Mice heterozygous for a Cys substitution in an epidermal growth factor-like domain of fibrillin-1 ($Fbn1^{C1039G/}$) are a well-characterized model of MFS with progressive aortic root dilation starting as early as 2 wk of age. By 7 wk of age, the aortic root of $Fbn1^{\text{C}_{1039}\text{G}/+}$ mice
is significantly larger than the aortic root of wild-type mice is significantly larger than the aortic root of wild-type mice, making this an appealing model of TAA (20). In addition, in vitro studies have demonstrated that elevated homocysteine may alter the structure and function of fibrillin-1, making MFS an even more attractive system in which to study the effect of homocysteine on TAA.

We found no significant differences in the thoracic aortic diameter between the mice fed the control and cobalaminrestricted diets in either the MFS or C57BL/6J cohorts. The lack of any demonstrated effect of homocysteine on the thoracic aortic diameter of MFS mice argues against a substantial direct role for the metabolite in TAA pathology in the context of cobalamin deficiency. A possible limitation of our work was the use of an animal model to recapitulate aspects of human disease. Mice on standard lab diets have steady-state homocysteine concentrations that are lower than the normal range for humans [$3-5 \mu$ mol/L in wild-type C57BL/6J mice (21–27) and 6–10 and $8-12$ µmol/L in men and women, respectively (6)]. The diet manipulations used in this study produced homocysteine concentrations that were comparable to elevated concentrations observed in humans, i.e., \sim 15 µmol/L at the experimental endpoint. If the absolute versus relative concentrations were critical, we may not have observed a change in aortic parameters because the upper range of homocysteine concentrations observed in humans is higher than that observed in mice aged 32 wk. It is worth noting that our experimental model was

designed to mimic the physiologic concentrations commonly observed in humans rather than manipulating the system to produce supraphysiologic concentrations of homocysteine. Our observations do not rule out the possibility that greatly elevated concentrations of homocysteine would influence the progression of TAA. In the same vein, we note that although monitoring for subtle prelesion changes was beyond the scope of this study, a histology of the mice maintained on the cobalamin-restricted diet may have revealed the cellular effects observed in TAA, such as the loss of smooth muscle cell organization, loss of elastic fibers, or evidence of inflammation (28).

The most severe complications of TAA, including rupture and death, are associated with an increased aneurysmal diameter. Medical management is often insufficient in preventing progressive thoracic aortic dilatation, and surgical intervention carries a considerable risk of morbidity and mortality. Additional therapeutic strategies for TAA are needed. Animal studies that investigate promising interventions are part of this process. Equally important is the publication of negative results. Studies such as ours that use well-developed hypotheses and simple experimental design avoid duplicated scientific effort and assist in directing research toward better interventions.

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