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4-PBA and metformin decrease sensitivity to PTZ-induced seizures in a malin knockout model of Lafora disease

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Abstract

Lafora disease (LD) is a rare adolescent-onset progressive myoclonic epilepsy caused by loss-of-function mutations either in the *EPM2A* gene encoding laforin or in the *EPM2B* gene encoding malin. Mouse models with deletion in the *Epm2a* or *Epm2b* gene display intracellular aggregates of polyglucosans (Lafora bodies) and neurological complications that resemble those seen in patients with LD. In the absence of laforin or malin expression, mice also exhibit different degrees of hyperexcitability, as reflected by an enhanced response to the convulsant drug pentylenetetrazol (PTZ). Malin knockout mice treated with 4-phenylbutyric acid (4-PBA) and metformin showed decreased amounts of Lafora bodies and polyubiquitin protein aggregates in the brain, diminished neurodegeneration, and amelioration of some neurological conditions. In this study, we analyzed the action of 4-PBA and metformin treatments on response to PTZ in a malin knockout model of LD. Both treatments decreased seizure susceptibility, bringing about a reduction in both seizure number and length, and eliminated the mortality induced by PTZ. These results show a neuroprotective role of 4-PBA and metformin and extend the beneficial effects reported in the malin knockout model of LD.

Keywords

Lafora disease; epilepsy; oxidative stress; autophagy; PTZ; malin knockout mouse

Introduction

Progressive myoclonus epilepsy of Lafora, or Lafora disease (LD) (OMIM 254780; ORPHA501) is a rare autosomal recessive disease that presents in adolescence with generalized seizures, myoclonic, absence, and visual seizures or cognitive decline. Rapid neurologic deterioration with progressive ataxia, dementia, dysarthria, amaurosis, and respiratory failure leads to death within 5 to 10 years of disease onset¹. Patients with LD present Lafora bodies, which are intracellular inclusions of polyglucosan, a long, linear and

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poorly branched glycogen. Lafora bodies accumulate in brain, skin, heart, and other tissues. At present, there is no effective treatment for this disease. LD is caused by recessive mutations either in the *EPM2A* gene, which encodes the dual-specificity phosphatase laforin (OMIM 607566)²⁻⁴, or in the *EPM2B* gene, encoding the E3 ubiquitin ligase malin (OMIM 608072)^{5,6}.

A number of mouse models of LD with targeted deletions of either the *Epm2a*⁷ or the *Epm2b*⁸⁻¹⁰ genes have been generated. In the absence of laforin or malin expression, *Epm2a*^{-/-} and *Epm2b*^{-/-} mice develop Lafora bodies and neurological complications that resemble those seen in patients with LD. Thus, both *Epm2a*^{-/-} and *Epm2b*^{-/-} mice, manifest with dyskinesia, impaired motor coordination and activity, deficits in episodic memory, and distinct extents of spontaneous epileptic activity¹¹. Additionally, they also exhibit different degrees of hyperexcitability, as reflected by an enhanced response to the convulsant agent pentylenetetrazol (PTZ)¹², an antagonist of the GABA_A receptor. Laforin and malin knockout mice phenotypically express with impaired macroautophagy and altered ubiquitin-proteasome system, resulting in defects in protein clearance mechanisms^{10,13}. Both LD mouse models also display increased oxidative stress and impaired antioxidant response in the brain¹⁴. In addition to polyglucosan aggregates, Lafora bodies also present ubiquitinated proteins, advanced glycation-end products, chaperones, autophagy components, and proteasome subunits.

A previous report from Sanz's and our group analyzed the effects of treatments with 4-PBA and with metformin in malin mutant mice¹⁵. 4-PBA is a chemical chaperone that sequesters misfolded and aggregated proteins associated with several human neurodegenerative diseases while metformin promotes autophagy through the activation of the AMP-activated protein kinase (AMPK) and acts as a neuroprotective agent in different neurodegenerative diseases¹⁶. Both treatments decreased the number of Lafora bodies and polyubiquitin protein aggregates in the brain, diminished neurodegeneration, and ameliorated neurological tests in mice lacking the malin protein¹⁵.

In order to explore the effects of 4-PBA and metformin on the epileptic activity of these mice, we analyzed the susceptibility to PTZ-induced seizures in malin-deficient mice following treatment with these two drugs.

Materials and Methods

Animals and treatments

Malin-deficient mice were used for our study. Generation of malin knockout mice was performed by targeted deletion of the single exon encoding malin, as described in Criado et al¹⁰. 4-PBA at 20 mM and metformin at 12 mM (Sigma Chemicals, MO, USA) were dissolved in drinking water and administered *ad libitum* in malin knockout male mice at 3 months of age. 4-PBA and metformin treatments were administered for 2 months and animals were then tested for their sensitivity to PTZ. Four groups of 16 homozygous adult male mice were analyzed per condition: wild type mice, malin knockout mice, malin knockout mice with 4-PBA treatment, and malin knockout mice with metformin treatment.

The mouse colonies were bred at the IIS-Fundación Jiménez Díaz Animal Facility and were maintained on a 12:12-hour light/dark cycle under constant temperature (23°C), with access to food and water *ad libitum*. The experiments were conducted in accordance with the Declaration of Helsinki principles and the guidelines of the Institutional Animal Care and Use Committee, and were approved by the IIS-Fundación Jiménez Díaz ethical review board.

PTZ treatment

PTZ (Sigma Chemicals, MO, USA) was administered intraperitoneally as a single injection at 50 mg/kg. After administration of a convulsive dose of PTZ, mice displayed intervals of hyperactivity, twitching, and hyperextension of the limbs that at times progressed to generalized tonic-clonic seizures, and occasionally to death, usually within the first 20 min after injection. The percentage of mice showing PTZ-induced generalized seizures, and the lethality were monitored over a period of 45 min. The time interval between drug administration and development of generalized tonic-clonic seizures (seizure latency) and the length of the seizures were also analyzed. The appearance of additional PTZ-induced seizures for periods of up to 2 h after PTZ administration was also evaluated in 8 animals per condition, although no concomitant episodes were observed.

Statistical analysis

Generalized seizures and lethality values are given as percentages of animals that respond to PTZ treatment. The chi-square test was used to perform the pairwise comparison. Latency time and seizure length values are given as means \pm standard error of means (SEM), and differences between groups were analyzed by one-way ANOVA followed-up by Student's t-test for pairwise comparison. Statistical significance was considered to be reached at * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ (Graph-PadPrism2.0) (n=16 per condition).

Results

After 2 months of treatment with 4-PBA or metformin, a convulsive dose of PTZ was injected intraperitoneally in wild type, malin knockout, malin knockout mice after treatment with 4-PBA, and malin knockout mice after treatment with metformin. The percentage of animals showing generalized seizures, mortality, seizure latency, and seizure length were analyzed. Following injection of PTZ, mice displayed freezing and convulsive activity, which later progressed to generalized tonic-clonic seizures, sometimes associated with death. PTZ treatment induced seizures in 50% of wild type mice, whereas in mice lacking malin the percentage increased to 78% ($p < 0.05$). Both 4-PBA and metformin treatments decreased the percentage of *Epm2b*^{-/-} mice developing seizures, reaching the wild type values after 4-PBA treatment, and decreasing this percentage below wild type levels after metformin treatment (Fig. 1A). PTZ-induced mortality had a tendency to be lower in 4-PBA-treated malin knockout mice (6.25%) when compared with wild type mice (25%), although the differences were not statistically significant. After metformin treatment, PTZ-induced mortality decreased to 0% ($p < 0.05$) (Fig. 1B). Seizure latency values were lower for malin mutant mice when compared to wild type mice. Both 4-PBA and metformin treatments increased the latency for PTZ-induced seizure onset in malin knockout mice,

eliminating the statistical significance between wild type and malin knockout mice (Fig. 1C). The length of PTZ-induced seizures in the malin knockout model was significantly increased when compared to wild type mice ($p < 0.05$). After 4-PBA treatment, no significant differences were observed between malin knockout and wild type mice, while seizure lengths were even shorter after metformin treatment ($p < 0.001$) (Fig. 1D). Thus, 4-PBA and metformin decrease PTZ-induced seizures, mortality, and seizure lengths, ameliorating the hyperexcitability detected in mice lacking the malin protein.

Discussion

We have previously described the effects of 4-PBA and metformin treatments in *Epm2b*^{-/-} mice on decreasing the number of Lafora bodies, reducing neurodegeneration and gliosis, and improving motor behavior and memory¹⁵. In this study, we tested the effects of 4-PBA and metformin on the increased sensitivity of this model to PTZ. We show that both treatments ameliorate the sensitivity of malin-deficient mice to this epileptogenic agent. The mechanisms involved in the antiepileptic action of 4-PBA and metformin are unknown, as are the mechanisms involved in the generation of epileptic seizures in LD. Studies addressing how LBs or other biological alterations result in the epilepsy of LD are lacking. Laforin and malin have been involved in the regulation of glycogen metabolism¹⁷, and they have been also implicated in alternative physiological pathways, such as endoplasmic reticulum stress response and protein clearance^{10,15}. In our previous report¹⁵, we also described that 4-PBA increased the levels of the chaperone BIP/Grp78, involved in the amelioration of proteostasis dysfunction, and that metformin induced the activation of the AMPK complex. However, as we mentioned above, the way these abnormalities induce epileptic discharges and seizures is unknown. The positive effects of 4-PBA and metformin on the hypersensitivity of malin-deficient mice to PTZ may be attributed to the reduction of Lafora bodies in GABAergic neurons, since a decreased amount of LBs was observed after treatment with 4-PBA and metformin, or could also be a consequence of the amelioration of selective neuronal cell death and gliosis¹⁵.

Thus, treatments with 4-PBA and metformin show further beneficial results in our malin knockout LD mouse model. As these compounds are already approved for clinical practices in different pathologies, we argue that they could be tested in clinical trials for their potential capacity to improve some symptoms that present in patients with Lafora disease.

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References

1. Van Heycop Ten Ham MW, De Jager H. Progressive Myoclonus Epilepsy with Lafora Bodies. *Clinical-Pathological Features Epilepsia*. 1963; 4(1–4):95–119. [PubMed: 14092647]
2. Minassian BA, Lee JR, Herbrick JA, Huizenga J, Soder S, Mungall AJ, et al. Mutations in a gene encoding a novel protein tyrosine phosphatase cause progressive myoclonus epilepsy. *Nat Genet*. 1998; 20(2):171–174. [PubMed: 9771710]
3. Serratosa JM, Gomez-Garre P, Gallardo ME, Anta B, de Bernabe DB, Lindhout D, et al. A novel protein tyrosine phosphatase gene is mutated in progressive myoclonus epilepsy of the Lafora type (EPM2). *Hum Mol Genet*. 1999; 8(2):345–352. [PubMed: 9931343]
4. Ganesh S, Agarwala KL, Ueda K, Akagi T, Shoda K, Usui T, et al. Laforin, defective in the progressive myoclonus epilepsy of Lafora type, is a dual-specificity phosphatase associated with polyribosomes. *Hum Mol Genet*. 2000; 9(15):2251–2261. [PubMed: 11001928]
5. Chan EM, Young EJ, Ianzano L, Munteanu I, Zhao X, Christopoulos CC, et al. Mutations in NHLRC1 cause progressive myoclonus epilepsy. *Nat Genet*. 2003; 35(2):125–127. [PubMed: 12958597]
6. Gentry MS, Worby CA, Dixon JE. Insights into Lafora disease: malin is an E3 ubiquitin ligase that ubiquitinates and promotes the degradation of laforin. *Proc Natl Acad Sci U S A*. 2005; 102(24):8501–8506. [PubMed: 15930137]
7. Ganesh S, Delgado-Escueta AV, Sakamoto T, Avila MR, Machado-Salas J, Hoshii Y, et al. Targeted disruption of the Epm2a gene causes formation of Lafora inclusion bodies, neurodegeneration, ataxia, myoclonus epilepsy and impaired behavioral response in mice. *Hum Mol Genet*. 2002; 11(11):1251–1262. [PubMed: 12019206]
8. DePaoli-Roach AA, Tagliabracci VS, Segvich DM, Meyer CM, Irimia JM, Roach PJ. Genetic depletion of the malin E3 ubiquitin ligase in mice leads to lafora bodies and the accumulation of insoluble laforin. *J Biol Chem*. 2010; 285(33):25372–25381. [PubMed: 20538597]
9. Valles-Ortega J, Duran J, Garcia-Rocha M, Bosch C, Saez I, Pujadas L, et al. Neurodegeneration and functional impairments associated with glycogen synthase accumulation in a mouse model of Lafora disease. *EMBO Mol Med*. 2011; 3(11):667–681. [PubMed: 21882344]
10. Criado O, Aguado C, Gayarre J, Duran-Trio L, Garcia-Cabrero AM, Vernia S, et al. Lafora bodies and neurological defects in malin-deficient mice correlate with impaired autophagy. *Hum Mol Genet*. 2012; 21(7):1521–1533. [PubMed: 22186026]
11. Garcia-Cabrero AM, Marinas A, Guerrero R, de Cordoba SR, Serratosa JM, Sanchez MP. Laforin and malin deletions in mice produce similar neurologic impairments. *J Neuropathol Exp Neurol*. 2012; 71(5):413–421. [PubMed: 22487859]
12. Garcia-Cabrero AM, Sanchez-Elexpuru G, Serratosa JM, Sanchez MP. Enhanced sensitivity of laforin- and malin-deficient mice to the convulsant agent pentylentetrazole. *Front Neurosci*. 2014; 8:291. [PubMed: 25309313]
13. Puri R, Suzuki T, Yamakawa K, Ganesh S. Dysfunctions in endosomal-lysosomal and autophagy pathways underlie neuropathology in a mouse model for Lafora disease. *Hum Mol Genet*. 2012; 21(1):175–184. [PubMed: 21965301]
14. Roma-Mateo C, Aguado C, Garcia-Gimenez JL, Ibanez-Cabellos JS, Seco-Cervera M, Pallardo FV, et al. Increased oxidative stress and impaired antioxidant response in Lafora disease. *Mol Neurobiol*. 2015; 51(3):932–946. [PubMed: 24838580]
15. Berthier A, Paya M, Garcia-Cabrero AM, Ballester MI, Heredia M, Serratosa JM, et al. Pharmacological Interventions to Ameliorate Neuropathological Symptoms in a Mouse Model of Lafora Disease. *Mol Neurobiol*. 2016; 53(2):1296–1309. [PubMed: 25627694]
16. Wiley JC, Meabon JS, Frankowski H, Smith EA, Schecterson LC, Bothwell M, et al. Phenylbutyric acid rescues endoplasmic reticulum stress-induced suppression of APP proteolysis and prevents apoptosis in neuronal cells. *PLoS One*. 2010; 5(2):e9135. [PubMed: 20161760]
17. Tagliabracci VS, Girard JM, Segvich D, Meyer C, Turnbull J, Zhao X, et al. Abnormal metabolism of glycogen phosphate as a cause for Lafora disease. *J Biol Chem*. 2008; 283(49):33816–33825. [PubMed: 18852261]

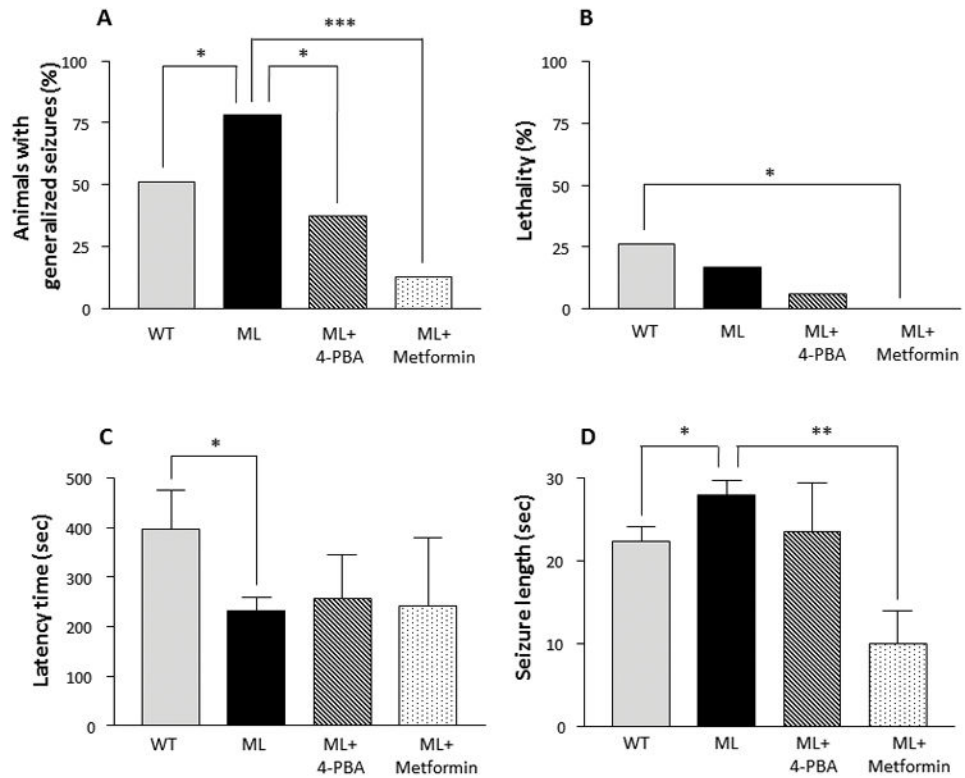


Figure 1. Sensitivity of *Epm2b*^{-/-} mice to PTZ after treatment with 4-PBA and metformin
(A) 4-PBA and metformin decreased the percentage of mice with generalized seizures after i.p. PTZ injection. The chi-square test yields $\chi^2 = 15.38$ on 3 degrees of freedom with a p-value of 0.0015. **(B)** Reduced mortality induced by PTZ after 4-PBA administration, and lack of PTZ lethal effects after metformin treatment. The chi-square test yields $\chi^2 = 7.823$ on 3 degrees of freedom with a p-value of 0.0498. **(C)** After 4-PBA and metformin treatments the differences between wild type and *Epm2b*^{-/-} in latency time (with a p-value of 0.0431 in Student's t-test) are no longer present. **(D)** Seizure length was lowered by 4-PBA, and more notoriously by metformin. The one-way ANOVA showed a p-value of 0.0294 and an F-ratio of 3.483 ML: malin knockout mice; WT: wild type mice.