

Novel cyanine-AMP conjugates for efficient 5' RNA fluorescent labeling by one-step transcription and replacement of [γ - 32 P]ATP in RNA structural investigation

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ABSTRACT

Two novel fluorescent cyanine-AMP conjugates, F550/570 and F650/670, have been synthesized to serve as transcription initiators under the T7 ϕ 2.5 promoter. Efficient fluorophore labeling of 5' RNA is achieved in a single transcription step by including F550/570 and F650/670 in the transcription solution. The current work makes fluorescently labeled RNA readily available for broad applications in biochemistry, molecular biology, structural biology and biomedicine. In particular, site-specifically fluorophore-labeled large RNAs prepared by the current method may be used to investigate RNA structure, folding and mechanism by various fluorescence techniques. In addition, F550/570 and F650/670 may replace [γ - 32 P]ATP to prepare 5' labeled RNA for RNA structural and functional investigation, thereby eliminating the need for the unstable and radio-hazardous [γ - 32 P]ATP.

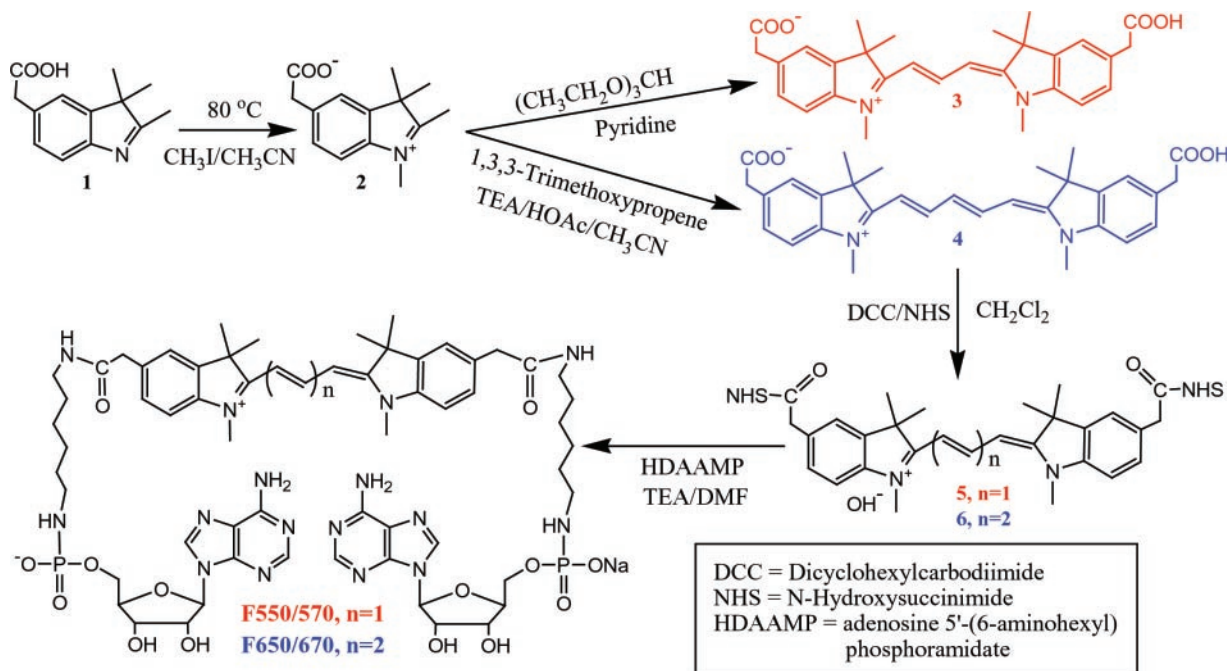
INTRODUCTION

Site-specific fluorescent labeling of RNA has many applications in biochemistry, molecular biology, structural biology and biomedicine (1–15). Fluorophores may be attached to RNA either by phosphoramidite chemistry during RNA synthesis (1–3,16), post-transcriptional fluorophore coupling with enzymatically prepared RNA (16,17), or direct fluorophore labeling during transcription (18,19). Phosphoramidite chemistry-based fluorescent labeling is commonly used to synthesize relatively small RNA molecules (<50–60 nt). When the size of RNA increases, such fluorescent-labeling procedures become impractical due to low yields of full-length RNA, high levels of impurities with short RNA

fragments and high costs of chemical synthesis. On the other hand, transcription-based RNA synthesis and fluorescent labeling has no apparent RNA size restrictions; both purities and costs are essentially independent of RNA sizes. Post-transcriptional fluorescent labeling involves two essential steps: preparation of amino- or thio-derivatized RNA by transcription followed by fluorophore coupling via fluorophore-*N*-hydroxysuccinimide esters (NHS) or fluorophore-maleimides or fluorophore-bromides (16,17). The limitations of this method lie in (i) the scarcity of available amino- and thio-nucleotide precursors for the preparation of amino- and thio-derivatized RNA, respectively; (ii) the hydrolytic lability of fluorophore-NHS; (iii) high concentrations of fluorophore-NHS, fluorophore-maleimides or fluorophore-bromides that are required to achieve efficient fluorescent labeling of RNA; and (iv) multiple steps of manipulation of RNA samples, leading to laborious sample preparation and low RNA yields.

Direct RNA labeling at the 5' end using the T7 ϕ 2.5 promoter developed in our laboratory (18,19) only requires appropriate label-linker-AMP conjugates (adenosine 5'-monophosphate, AMP) to serve as transcription initiators. The newly developed *in vitro* transcription system recognizes the adenosine moiety and labels the 5' end of RNA with high efficiency through transcription initiation (18,19). The only RNA sequence requirement is the 5' AG. We have previously reported fluorescein-HDA-AMP (1,6-hexanediamine, HDA) for direct RNA labeling (19). For this new RNA-labeling method to be widely applicable, however, diverse fluorophores with better spectroscopic properties, such as the commonly used cyanine dye family, are highly desirable. Here, we describe the synthesis of two novel cyanine-AMP conjugates, F550/570 and F650/670, and their use to fluorescently label 5' RNA in a single transcription step. Furthermore, we demonstrate one utility, among others, of F550/570 to replace the commonly used [γ - 32 P]ATP for 5' RNA labeling in RNA structure/function/mechanism investigation.

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Scheme 1. Synthesis of cyanine-AMP conjugates.

MATERIALS AND METHODS

Synthesis of cyanine-AMP conjugates

All reagents and chemicals were purchased from Aldrich and used as received. Synthetic procedures are shown in Scheme 1. Starting from 2,3,3-trimethyl-3H-indole-5-acetic acid **1** (20), the common intermediate 1,2,3,3-tetramethyl-indoleninium-5-acetate (**2**) was synthesized by methylation of **1** (20). Condensation of two molecules of **2** with one molecule of triethyl orthoformate (**21**) afforded the symmetrical red cyanine dye **3** with two free carboxyl groups that can be used for subsequent conjugation with AMP via a linker. Separately, condensation of one molecule of 1,3,3-trimethoxypropene (**21**) with two molecules of **2** produced another symmetrical blue cyanine dye **4** with two free carboxyl groups. The carboxyl groups of **3** and **4** were then activated by *N,N'*-dicyclohexylcarbodiimide (DCC) to form their corresponding NHS esters, **5** and **6**. Finally, the intermediates **5** and **6** were individually coupled with 5'-(6-aminohexyl) adenosine phosphoramidate (HDAAMP) (**19**) to afford a pair of novel symmetrical cyanine-AMP conjugates, **F550/570**, **n=1** and **F650/670**, **n=2**. Detailed description of the syntheses of **2–6**, **F550/570** and **F650/670** are given below.

Synthesis of compound 2. To a 25 ml Schlenk tube was added 0.5 g (2.3 mmol) of compound **1**, prepared from the published procedure (20), 1.2 ml (19.3 mmol) of iodomethane and 10 ml of acetonitrile. The reaction mixture was degassed with argon for 30 min and the tube was sealed with a cap and heated in an oil bath at 80°C for 1 h. After cooling, the reaction mixture was transferred into a flask and concentrated under vacuum to give 0.78 g (94%) of product **2**.

Synthesis of compound 3. The literature procedures (20,21) were used to prepare compound **3**. To 1.0 g (2.8 mmol) of

compound **2**, 20 ml of dry pyridine was added. While the reaction mixture was refluxing, 1.4 ml (8.4 mmol) of triethyl orthoformate was added slowly (0.4 ml per 15 min). After completion of addition, the reaction mixture was refluxed for another 2 h. Solvent was removed and the red residue was dissolved in 40 ml of methanol, followed by adding 200 ml of ethyl acetate. After concentrating to ~50 ml, another 100 ml of ethyl acetate was added, and concentrated to ~50 ml. To this suspension, 100 ml of ethyl acetate was added. The top solvent was decanted and the red residue was dried over under vacuum to give 0.81 g (96%) of the compound **3**. Mass spectrometry (MS) analysis gave the following results: $C_{29}H_{33}N_2O_4^+$, calcd, 473.24, found 473.2 (M^+).

Synthesis of compound 4. The literature procedure (21) was used to prepare compound **4**. To 1.0 g (2.8 mmol) of compound **2**, 28 ml of dry acetonitrile, 0.6 ml of triethylamine (TEA) and 0.2 ml of acetic acid were added. While the reaction mixture was refluxing, a solution of 1.0 g (7.6 mmol) of 1,3,3-trimethoxypropene in 4.0 ml of acetonitrile was slowly added (0.5 ml per 15 min). After completion of the addition, the reaction mixture was refluxed for another 2 h. Solvent was removed and the blue/purple residue was dissolved in 40 ml of methanol, followed by adding 200 ml of ethyl acetate. After concentrating to ~50 ml, another 100 ml of ethyl acetate was added, and concentrated to ~50 ml. To this suspension, another 100 ml of ethyl acetate was added, the top solvent was decanted and the red residue was dried over a high vacuum to give 0.84 g (96%) of the compound **4**. The molecular peak found by MS, 499.2 (M^+), is consistent with the expected formula $C_{31}H_{35}N_2O_4^+$, 499.26.

Synthesis of 5 and 6. To 100 mg (0.16 mmol) of compound **3** or **4**, 90 ml of CH_2Cl_2 , 100 mg (0.48 mmol) of DCC, 55 mg (0.48 mmol) of NHS and 50 mg of 4-(dimethylamino)pyridine

were added. The reaction mixture was stirred for 2 h. The urea-derivative precipitate was formed and filtered off. The filtrate was concentrated to dryness and used for the next step of coupling reaction without further purification. MS analysis gave the following results: **5**— $C_{37}H_{39}N_4O_8^+$, calcd, 667.28, found 667.2 (M^+); **6**— $C_{39}H_{41}N_4O_8^+$, calcd, 693.29, found 693.2 (M^+).

Synthesis and purification of F550/570 and F650/670. To a 300 μ l aqueous solution of 370 mM HDAAMP (19), 150 μ l TEA, 750 μ l *N,N*-dimethylformide (DMF) and 20 mg of **5** or **6**, which were dissolved separately in 150 μ l DMF, were added. After 30 min of reaction, 1.2 ml of water was added to the sample. The resulting solution was filtered with a 0.2 μ m syringe filter. Purification of **F550/570** and

F650/670 was achieved by semi-preparative reverse phase high-performance liquid chromatography (HPLC) (Figures 1 and 2). Isolated yields for **F550/570** and **F650/670** were \sim 20%. High-resolution MS analysis gave excellent results: **F550/570**— $C_{61}H_{85}N_{16}O_{14}P_2^+$, calcd, 1327.5901, found 1327.5869 (M^+); **F650/670**— $C_{63}H_{87}N_{16}O_{14}P_2^+$, calcd, 1353.6057, found 1353.5998 (M^+).

Spectroscopic properties of F550/570 and F650/670

UV absorbance spectra and molar extinction coefficients of **F550/570** and **F650/670** were obtained from a JASCO spectrometer (V-530) in 20 mM phosphate, pH 7.0. Fluorescence emission spectra were measured with an ISS PC1 fluorometer (Champaign, IL) in 20 mM phosphate, pH 7.0, under the

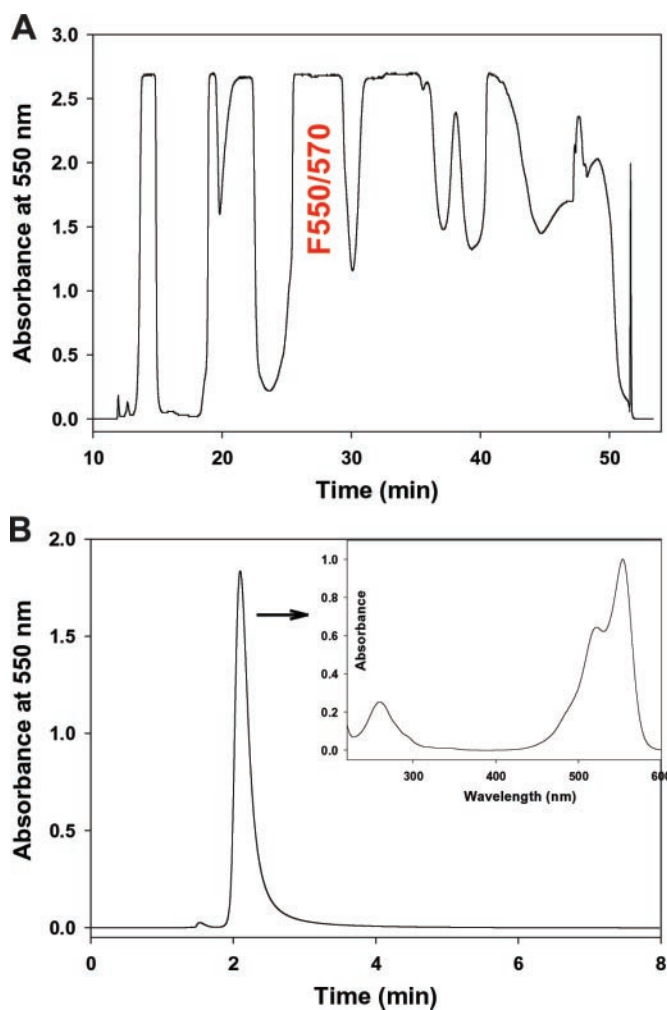


Figure 1. Purification and subsequent analysis of **F550/570** by HPLC. (A) **F550/570** sample was injected onto a Delta Pak C18 column, $7.8 \times 300 \text{ mm}^2$, pre-equilibrated in 20 mM phosphate buffer, pH 7.0, flow rate 5 ml/min. The mobile phase was manually changed in the following order: 100% water at 7 min \rightarrow 20% MeOH/80% water at 9 min \rightarrow 30% MeOH/70% water at 16 min \rightarrow 40% MeOH/60% water at 23 min \rightarrow 50% MeOH/50% water at 30 min \rightarrow 60% MeOH/40% water at 38 min \rightarrow 100% MeOH at 45 min. The 25–30 min fraction (marked as **F550/570**) has the desired UV spectrum from an online photodiode array detector. (B) Analysis of the 25–30 min fraction by an Econosphere C18 column, $4.6 \times 50 \text{ mm}^2$, in 60% MeOH/40% 20 mM phosphate, pH 7.0, flow rate 0.5 ml/min. The insert shows the UV–visible spectrum of the peak.

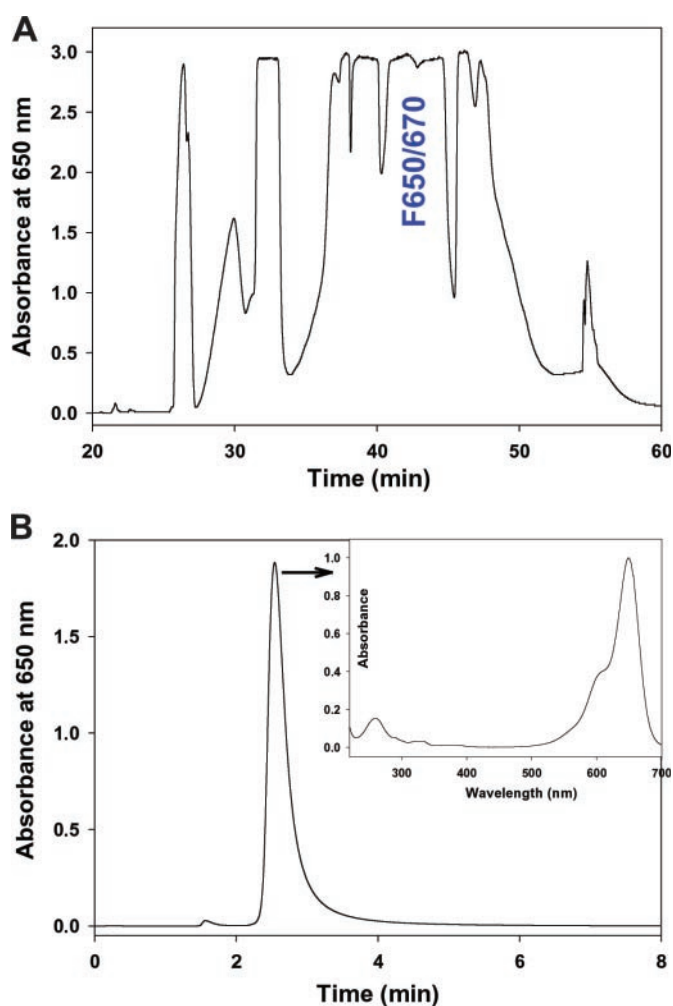


Figure 2. Purification and subsequent analysis of **F650/670** by HPLC. (A) **F650/670** sample was injected onto a Delta Pak C18 column, $7.8 \times 300 \text{ mm}^2$, pre-equilibrated in 20 mM phosphate buffer, pH 7.0, flow rate 5 ml/min. The mobile phase was manually changed in the following order: 100% water at 11 min \rightarrow 20% MeOH/80% water at 18 min \rightarrow 30% MeOH/70% water at 23 min \rightarrow 40% MeOH/60% water at 29 min \rightarrow 50% MeOH/50% water at 35 min \rightarrow 60% MeOH/40% water at 43 min \rightarrow 100% MeOH at 52 min. The 41–45 min fraction (marked as **F650/670**) has the desired UV spectrum from an online photodiode array detector. (B) Analysis of the 41–45 min fraction by an Econosphere C18 column, $4.6 \times 50 \text{ mm}^2$, in 60% MeOH/40% 20 mM phosphate, pH 7.0, flow rate 0.5 ml/min. The insert shows the UV–visible spectrum of the peak.

excitation of 510 and 610 nm for **F550/570** and **F650/670**, respectively. Quantum yields (Φ) of **3**, **4**, **F550/570** and **F650/670** were determined by using reference fluorophores according to the following equation:

$$\Phi_X = \Phi_R(SL_X/SL_R)(\eta_X/\eta_R)^2, \quad 1$$

where X and R stand for the sample and the reference, respectively. SL is the slope of integrated fluorescence area versus absorbance and η is the refractive index of the solvent. The reference for **3** and **F550/570** was rhodamine 101 ($\Phi_R = 1$; Fluka) (22). Zinc phthalocyanine ($\Phi_R = 0.3$; Aldrich) (23) was used as the reference for quantum-yield measurements of **4** and **F650/670**. Fluorescence was measured at the excitation of 490 nm (for **3** and **F550/570**) or 600 nm (for **4** and **F650/670**).

RNA 5' labeling by **F550/570** and **F650/670**

Fluorescent labeling of 5' RNA by **F550/570** and **F650/670** was performed under normal *in vitro* transcription conditions (18,19) with slight modifications: changing [ATP] from 1 to 0.25 mM and adding 2 mM of **F550/570** or **F650/670** to the transcription solution. The final transcription solution contained 40 mM Tris-HCl, pH 8.0, 5 mM dithiothreitol, 6 mM MgCl₂, 2 mM spermidine, 0.01% Triton X-100, 0.25 mM ATP, 1 mM each of UTP, GTP and CTP, 2 mM **F550/570** or **F650/670**, 0.05–0.5 μ M dsDNA containing the T7 ϕ 2.5 promoter (18,19), 500 U of T7 RNA polymerase per 100 μ l reaction and 10–20 U of RNase inhibitor per 100 μ l reaction. Also included was 2 μ l of [α -³²P]ATP per 100 μ l reaction as a tracer for RNA analysis. The labeling reaction was carried out at 37°C for 2 h before analysis by denaturing PAGE. Product detection and quantification were achieved by phosphorimaging/fluorimaging under the excitation by ³²P, a 532 nm green laser, and a 633 nm red laser (Typhoon 9400; Amersham Biosciences). For Figure 4, the RNA was a 92-nt ribozyme (TES33) with the sequence of AGGGAAGUGCUACCACACUUGCUGGUGUACGCGCCCCUUGCGUACUCUGCCCUUCCGCGUCUCCCCGUCCAACGGGCAUGCGGCCA-GCCA, which was previously isolated in our laboratory (24). In addition, to test transcription-labeling yields and PAGE separation of varying RNA, we prepared five different RNA sizes of 100, 200, 300, 400 and 500 nt using self-constructed DNA templates from Epicentre's control DNA for AmpliScribe™ system. The sequences of transcribed RNAs are AGAAUUCUAAGCGGAGAUCCGCUAGUGAUUUUAA-ACUAUUGCUGGCAGCAUUCUUGAGUCCAAUAUAA-AAGUAUUGUGUACCUUUUUGCUGGGUCAGGUUGUUCUUUAGGAGAGUAAAAGGAUCAAAUUGCACUAAA-CGAAACUGAAACAAGCGAUCGAAAAUAUCCCUUU-GGGAUUCUUGACUCGAUAAGUCUAUUUUUUCAG-AGAAAAAAUAUUCAUUGUUUUCUGGGUUGGUGAU-UGCACCAAUCAUUCUCAAUCAAUUGUUGUUUUA-CCACACCAUUCGCCCCGAUAAAAGCAUGAAUGU-UCGUGCUGGGCAUAGAAUUAACCGUCACCUCAAA-AGGUAUAGUUAAAUCACUGAAUCCGGGAGCACUU-UUUCUAUUAAAUGAAAAGUGGAAUUCUGACAAUU-CUGGCAAACCAUUUAACACACGUGCGAACUGUCC-AUGAAUUUCUGAAAGAGUUACCCUCUAAGUAAU-GAGGUGUUUAGGACGCUUUUUAUUU, where each bold-face nucleotide represents the 3' end of an RNA sequence from the beginning A.

Replacement of [γ -³²P]ATP by **F550/570** in RNA reaction site mapping

F550/570-labeled RNA (prepared as described above) was used to investigate the reaction site of an *in vitro* isolated ribozyme ACT3 that catalyzes self-aminoacylation from biocytinyl CoA (biocytinCoA) (25). Gel-purified dye-labeled ACT3 RNA was reacted for 10 min at 25°C with 1 mM biocytinCoA in the selection buffer containing 20 mM HEPES (pH 7.4), 200 mM KCl, 100 mM NaCl, 30 mM MgCl₂ and 10 mM CaCl₂ (25). After purification by membrane filtration using a microcon M30 (Millipore), the RNA was partially hydrolyzed by lead under the following conditions: 2 mM lead acetate in 7 M urea and 50 mM HEPES, pH 5.5, 40 s at 80°C, followed by quenching with 50 mM EDTA. The resultant RNA fragments were loaded onto a Neutravidin column (Pierce). Following washing with 4 M NaCl, 3 M sodium acetate and water, RNA fragments were eluted by 20 mM biotin and 5 M urea at 95°C for 10 min. The eluted RNA was EtOH-precipitated and the recovered RNA was analyzed by 8% denaturing PAGE, followed by phosphorimaging under the excitation of a 532 nm green laser.

An RNase T1 ladder of the **F550/570**-labeled RNA was prepared by digestion (10 min at 50°C) of **F550/570**-labeled RNA with 2 μ g of carrier tRNA, 1 U of RNase T1 in 7 M urea, 2 mM EDTA and 0.1 M sodium acetate, pH 5.2. The ladder served as RNA size markers in RNA fragment analysis by PAGE (Figure 6B).

RESULTS

Synthesis of **F550/570** and **F650/670**

Cyanine dyes (26) display some excellent fluorescent properties, including their high molar extinction coefficients and improved resistance to photobleaching (in contrast to the commonly used fluorescein). In addition, the color and solubility of cyanine dyes can be modulated by the number of double bonds between the two indole rings and by changing the attached chemical groups. Having successfully demonstrated 5' fluorescein labeling of RNA by direct transcription (19), we sought to develop similar methods to site-specifically label 5' RNA by cyanine dyes with the intention of bringing this simple and efficient RNA-labeling technique to broad applications in biosciences and biomedicine.

After two synthetic steps (Scheme 1), the two fluorescent cyanine dyes **3** and **4** were obtained in high yield (91%). After activation by DCC and NHS, the NHS esters **5** and **6** were readily coupled with the amino group of HDAAMP (19). Purification by reverse phase HPLC (Figures 1A and 2A) yielded pure (>96% purity) fluorescent dyes **F550/570** and **F650/670** (with ~20% total yields) as shown in Figures 1B and 2B. In addition to **F550/570** and **F650/670**, there were other peaks containing cyanine dyes (Figures 1A and 2A). Their identities were not further investigated.

UV absorbance and fluorescent properties of **F550/570** and **F650/670**

Ultraviolet and visible spectra of **F550/570** and **F650/670** (Figure 3, Ab lines) show additive contribution from both the adenosines and the cyanine cores. For both cyanine-AMP conjugates, the absorbance within 220–300 nm is

contributed mainly by the adenosine moieties. The absorbance between 450 and 580 nm of **F550/570**, with λ_{\max} at 550 nm, is purely due to the cyanine dye **3** core. Similarly, for **F650/670**, the absorbance between 580 and 700 nm ($\lambda_{\max} = 650$ nm) originates from the cyanine dye **4** core. Molar extinction coefficients measured in 20 mM phosphate buffer, pH 7.0, are $\epsilon_{550} = \sim 130\,000\text{ M}^{-1}\text{ cm}^{-1}$ and $\epsilon_{650} = \sim 210\,000\text{ M}^{-1}\text{ cm}^{-1}$ for **F550/570** and **F650/670**, respectively. Fluorescence emission spectra of **F550/570** and **F650/670** are marked as Em curves in Figure 3. **F550/570** fluoresces between 550 and 700 nm, with emission $\lambda_{\max} = 570$ nm. Under excitation, **F650/670** emits fluorescence within 640–780 nm ($\lambda_{\max} = 670$ nm). Within the visible range (440–780 nm), both the absorption spectra and fluorescence emission spectra of **F550/570** and **F650/670** are similar to those of common cyanine dyes, Cy3 and Cy5, respectively (26). The quantum yields (from three sets of measurements) of **F550/570** and **F650/670** are 0.16 ± 0.03 and 0.47 ± 0.06 , respectively, compared with 0.11 ± 0.03 and 0.27 ± 0.04 for their corresponding precursors **3** and **4**. Therefore, the attachment of two adenosines and HDA linkers at the opposite ends of the cyanine cores increases their quantum yields, but has no apparent effects on other spectroscopic properties of the cyanine dye within the visible range.

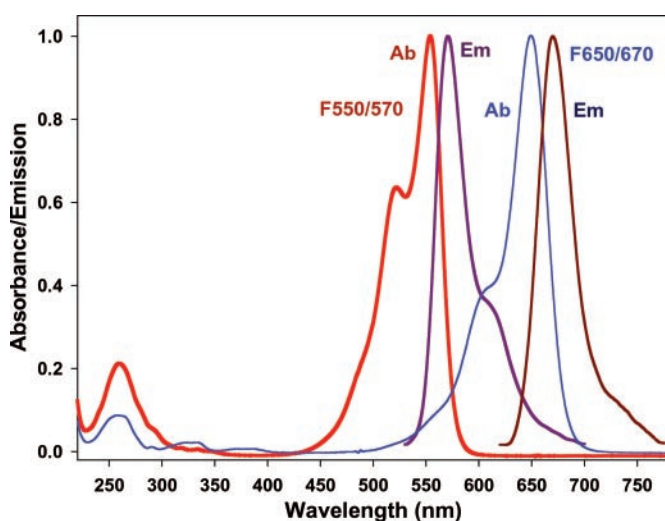


Figure 3. UV-visible absorption spectra (Ab) and fluorescence emission spectra (Em) of **F550/570** and **F650/670**. The spectra were measured in 20 mM phosphate buffer, pH 7.0. All spectra were normalized to 1 at their λ_{\max} . The λ_{\max} difference between excitation and emission is 20 nm for both **F550/570** and **F650/670**.

Fluorescent labeling of 5' RNA by **F550/570** and **F650/670**

Fluorescent labeling of 5' RNA is achieved by simply including **F550/570** or **F650/670** in transcription solutions under the T7 class II promoter $\phi 2.5$ (18,19). One of the two adenosines within **F550/570** or **F650/670** initiates transcription, resulting in 5' RNA labeling by the cyanine dyes. Although there are two identical adenosines within **F550/570** or **F650/670**, the probability of both adenosines initiating transcription to produce head-to-head joined RNA via **F550/570** or **F650/670** is low due to the high concentration ratios of **F550/570** (or **F650/670**) over transcribed RNA molecules (i.e. mM versus μM). To confirm the prediction, purified **F550/570**-RNA (TES33, ^{32}P -labeled) was added to the transcription solution in the absence of $[\alpha\text{-}^{32}\text{P}]\text{ATP}$ and **F550/570**. No head-to-head joined RNA dimer was observed by phosphorimaging after PAGE. Because neither **F550/570** nor **F650/670** contains a nucleoside 5'-triphosphate, the cyanine dyes cannot be incorporated into internal RNA positions by T7 RNA polymerase. Figure 4 shows 5' RNA (TES33) labeling by **F550/570** and **F650/670**. Three parallel transcription experiments (with $[\alpha\text{-}^{32}\text{P}]\text{ATP}$ as the internal radiolabel) were carried out in the absence of the cyanine dyes (lane 1) or in the presence of **F550/570** (lane 2) or **F650/670** (lane 3). Phosphorimaging based on ^{32}P (Figure 4A) revealed an additional slower RNA band in lanes 2 and 3. Fluorescence scanning of the same gel under the excitation with the 532 nm green laser (Figure 4B) shows only a single RNA band in lane 2, whose location overlaps with that of the upper band of lane 2 in Figure 4A. Under the excitation of a 633 nm red laser (Figure 4C), scanning of the same gel displays another single RNA band in lane 3, whose location superimposes with that of the upper band of lane 3 in Figure 4A. Taken together, the three different scanings of the same gel based on excitation by ^{32}P (Figure 4A), 532 nm photons (Figure 4B) and 633 nm photons (Figure 4C) indicate fluorescent labeling of RNA by **F550/570** (lane 2) and **F650/670** (lane 3) during transcription. The labeling yields were $60 \pm 5\%$ and $35 \pm 5\%$ for **F550/570** and **F650/670**, respectively. Total RNA yields for **F550/570**- and **F650/670**-labeled RNA were $110 \pm 30\%$ and $90 \pm 30\%$ of the control RNA (in the absence of dye-AMP). In a different set of labeling experiments with varying RNA sizes (100–500 nt), 15 independent transcriptions gave labeling yields of $78 \pm 3\%$ and $55 \pm 3\%$ for **F550/570** and **F650/670**, respectively. The respective total RNA yields relative to the control RNA were $150 \pm 30\%$ and $110 \pm 30\%$. Therefore, **F550/570** and **F650/670** appear to stimulate transcription initiation under the transcription

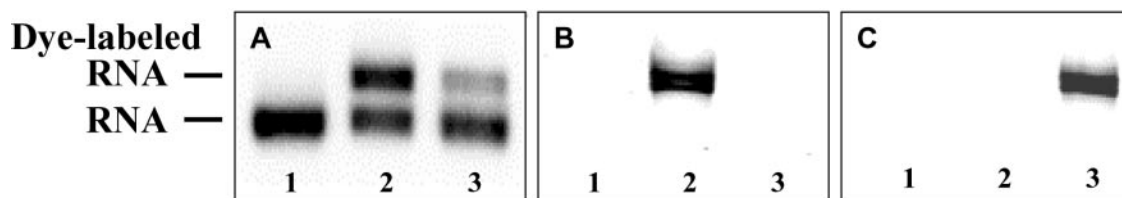


Figure 4. RNA fluorescent labeling by **F550/570** and **F650/670** under the T7 $\phi 2.5$ promoter (18,19). All RNA was also internally ^{32}P -labeled by $[\alpha\text{-}^{32}\text{P}]\text{ATP}$. After transcription, RNA samples were fractionated by PAGE. Lane 1, normal transcription; lanes 2 and 3, transcription in the presence of **F550/570** and **F650/670**, respectively. (A) ^{32}P -phosphorimaging reveals total RNA bands in different transcription experiments. (B) Scanning of the same gel under the excitation of a 532 nm laser shows only **F550/570**-labeled RNA. (C) Under excitation with a 633 nm laser, only **F650/670**-labeled RNA is visible. The RNA sequence was that of a thioester-synthesizing ribozyme TES33, 92 nt (24). RNAs from $\sim 5\ \mu\text{l}$ transcription were used for the gel.

conditions. In a typical experiment, ~ 20 μg of dye-labeled RNA can be prepared from 100 μl transcription.

Since no transcription-based methods would produce 100% labeled RNA, isolation of labeled RNA from unlabeled (normal) RNA may be required for its applications. Owing to their relatively large sizes, **F550/570**- and **F650/670**-labeled RNA displays significant migration retardation (5–6 nt difference) by PAGE. This added property of **F550/570**- and **F650/670**-labeled RNA may be exploited to achieve high purity levels of fluorescent RNA by PAGE. To establish the RNA size-PAGE resolution relationship, five different sizes of RNA (100, 200, 300, 400 and 500 nt) were labeled by both ^{32}P and **F550/570** (**F650/670**) and run 30 cm on a large sequencing gel ($42 \times 35 \times 0.08 \text{ cm}^3$). As shown in Figure 5A, gel-running time varied from ~ 4 h for an RNA of 100 nt to ~ 18 h for the 500 nt RNA. Separation between dye-labeled RNA and unlabeled RNA ranged from 12 to 3 mm when RNA increased from 100 to 500 nt (Figure 5B). Therefore, **F550/570**- and **F650/670**-labeled RNA (up to 500 nt, depending on the skill of the experimenter) can be separated from unlabeled RNA by 8% PAGE. In one experiment, we purified an **F550/570**-labeled 120-nt RNA to $>95\%$ purity using 8% denaturing PAGE ($42 \times 35 \times 0.08 \text{ cm}^3$) after running for 4 h at 45 V/cm.

Replacement of $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ by **F550/570** in RNA reaction site mapping

Investigation of RNA structure/function/mechanism frequently requires the use of $[5'\text{-}^{32}\text{P}]\text{RNA}$ (27–29). However, it may be difficult to phosphorylate some RNA by $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ and polynucleotide kinase (PNK) due to the structure of RNA. As an example, a recently isolated ribozyme (ACT3) from our laboratory poses such a problem. ACT3 is a ribozyme that catalyzes the formation of aminoacylated RNA from aminoacyl thioesters of CoA (25). Attempts to label the 5' HO-ACT3 by $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ and PNK gave very poor ($<2\%$) labeling yields due to the 5' recessed structure (Figure 6A). Here, we demonstrate that 5' **F550/570**-labeled RNA can be used to map the reaction site within the ribozyme.

BiocytinCoA-reacted RNA, which was 5' end-labeled by **F550/570**, was first treated with lead acetate to generate all possible single-cut RNA fragments. The resulting RNA sample contained both biocytin-tagged RNA fragments and normal RNA fragments. After Neutravidin affinity chromatography, only biocytin-tagged RNA fragments would be retained on the column. Analysis of the eluted RNA sample by PAGE and **F550/570**-based fluorimaging should reveal the location of the reactive site within the ribozyme. Figure 6B shows the fluorescence scanning image of RNA fragments eluted from the Neutravidin column. Figure 6C is the profile of **F550/570** fluorescence intensity from Figure 6B. The gel indicates that RNA fragments shorter than C51 (from the 5' end) were completely missing from the affinity column, while all the fragments longer than C51 were present. The result is schematically interpreted in Figure 6D. The C51 RNA fragment (with 5' **F550/570** as well) thus contained the reaction site at a 2' OH group. Since lead-induced RNA hydrolysis requires the presence of a free 2' OH group, the actual reaction site is the 2' OH group of U50, which agrees with the results obtained by reverse transcriptase-catalyzed primer extension, HPLC analysis and MS analysis (25).

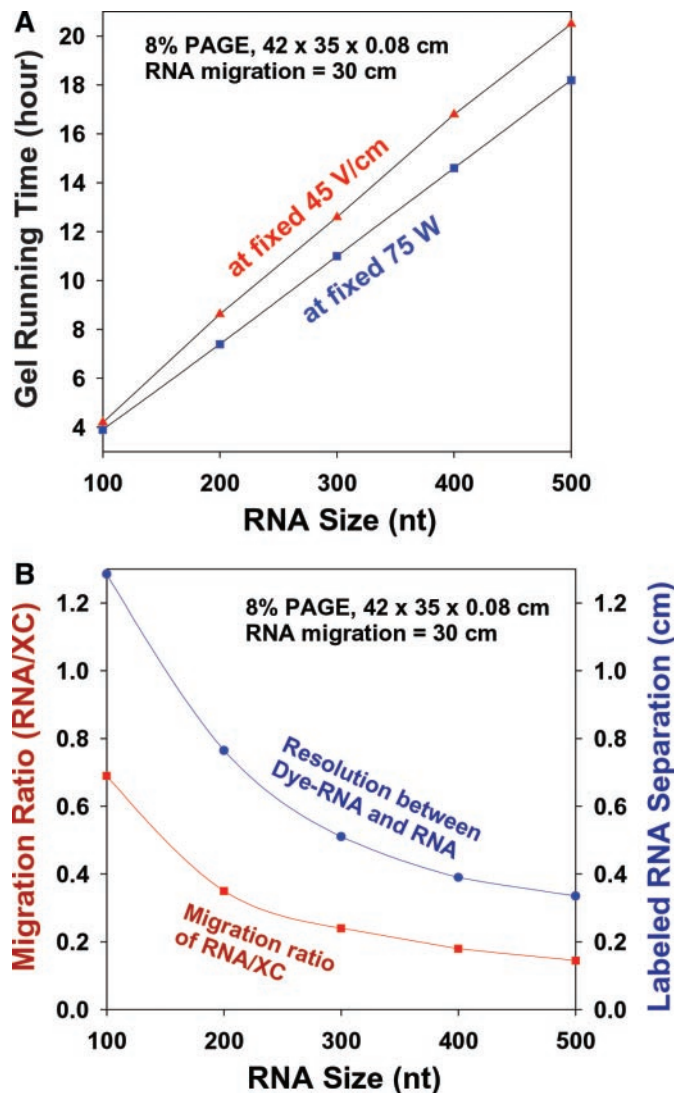


Figure 5. Resolution of dye-labeled RNA of different sizes by 8% PAGE ($42 \times 35 \times 0.08 \text{ cm}^3$). Each RNA was run 30 cm from the origin under either constant voltage or constant wattage. (A) The relationship of RNA size and gel-running time under two different conditions. (B) Relative electrophoretic mobility of different RNA sizes to xylene cyanol (XC) and resolution between dye-labeled RNA and unlabeled RNA.

DISCUSSION

Our approach for direct fluorescent labeling of RNA by **F550/570** and **F650/670** during transcription offers the following distinct advantages over the phosphoramidite chemistry method and post-transcriptional fluorescent labeling. First, site-specific RNA fluorescent labeling is achieved in a single step of transcription, greatly reducing the RNA sample handling time and RNA loss. Typically, fluorophore-labeled RNA can be made available in common laboratories within 2–4 h. Second, there is no apparent RNA size restriction; fluorescently labeled small and large RNAs can be prepared by the same transcription method with similar yields, purities and costs. The commonly used *in vitro* transcription under the T7 $\phi 6.5$ promoter produces G-initiated RNAs. The same RNA sequences can be used for fluorescence labeling under

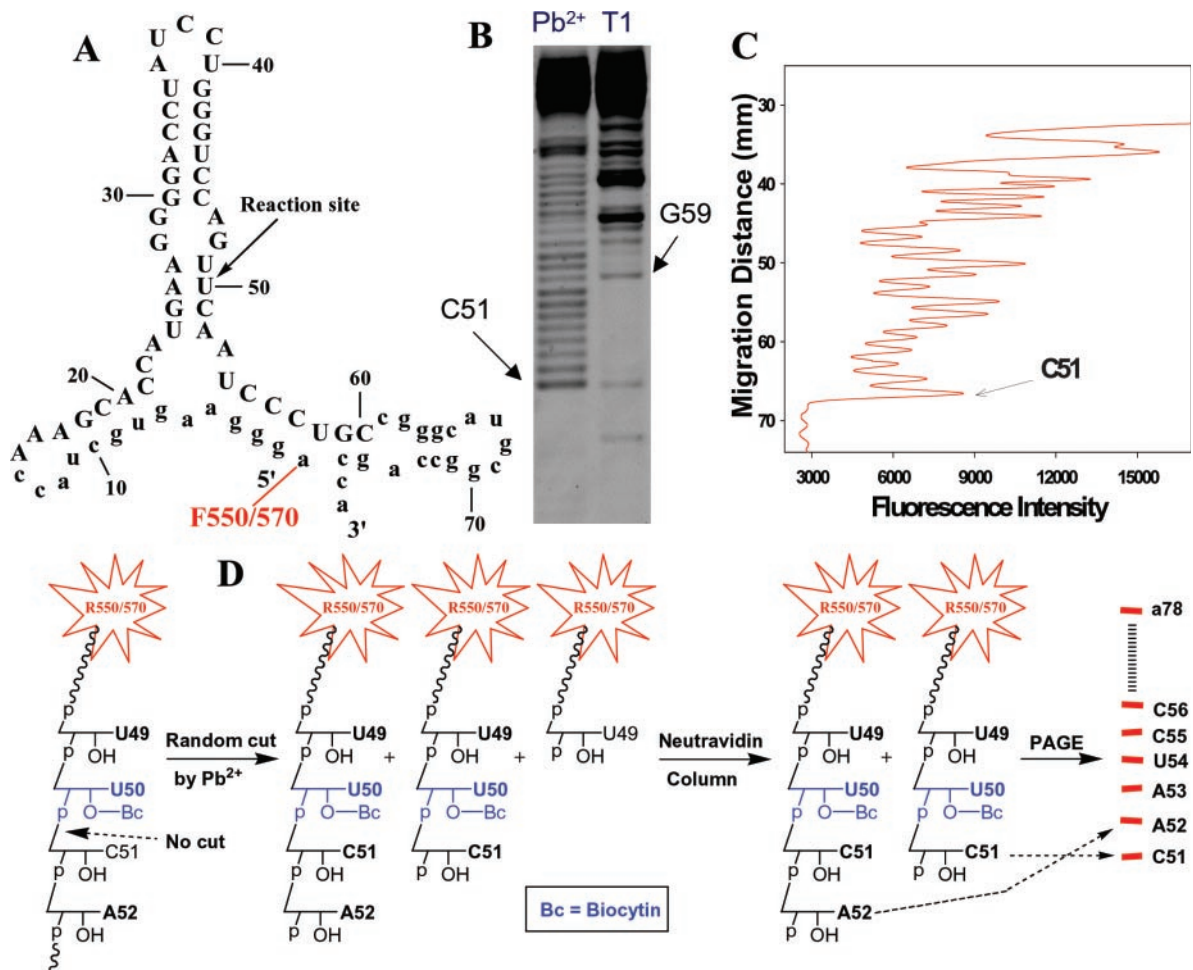


Figure 6. Ribozyme reaction site mapping by **F550/570**-labeled RNA. RNA from 10 μ l transcription was used for the analysis. (A) The secondary structure of ACT3 (25). It is difficult to label the 5' end by [γ - 32 P]ATP and PNK due to its recessed 5' end. (B) PAGE analysis of Neutravidin column-eluted RNA fragments (lane 1), along with an RNase T1-digested ladder of the same **F550/570**-labeled RNA. RNA fragments were generated by partial lead hydrolysis of biocytinCoA-reacted RNA (**F550/570**-labeled). After electrophoresis, the gel was scanned directly by an Amersham Typhoon phosphorimager under the excitation of a 532 nm laser. (C) Fluorescence intensity profile of lane 1 in (B). (D) An illustration of the reaction site mapping. Controlled lead-induced RNA hydrolysis randomly cuts the RNA to all possible fragments, except for the reactive site where the 2' OH is blocked by biocytin. The eluted RNA sample from Neutravidin chromatography contains all biocytin-tagged fragments, C51 and above. None of the untagged RNA fragments (U49 and below) is retained by the column. Therefore, PAGE analysis and **F550/570**-based phosphorimaging can detect all RNA fragments equal to or longer than C51. The reaction site is one nucleotide below C51, i.e. U50.

the T7 ϕ 2.5 promoter by adding an extra A before the first G. Third, **F550/570** and **F650/670** are chemically stable, allowing laboratories to maintain steady stocks without constant purchase of fresh supplies. The current finding will make site-specifically fluorophore-labeled RNA readily available for a variety of applications in biochemistry, structural biology and nucleic acids-based clinical diagnostics. In particular, fluorophore-labeled large RNAs prepared by the current method may be appealing to biochemists and structural biologists to investigate RNA structure, folding and mechanism by various fluorescence techniques such as fluorescence resonance energy transfer (6–9,13) and single-molecule kinetics (10,15,30). Without an apparent restriction on RNA sequence (except for the 5' AG) and size, the described fluorescent labeling by **F550/570** and **F650/670** can be easily applied to large RNAs such as the group I and group II introns, the RNase P RNA, spliceosomal RNAs, ribosomal RNAs and artificially selected functional RNAs (aptamers and ribozymes).

PNK-catalyzed phosphorylation by [γ - 32 P]ATP is a common procedure to prepare [γ - 32 P]RNA that is required in various studies of structure/function/mechanism (27–29). However, such a 32 P-based RNA labeling method may present a series of problems for the experimenter. First, 32 P radioactivity quickly diminishes with time due to its short half-life. In addition, RNA labeling efficiency by [γ - 32 P]ATP actually decreases much faster than the radiodecay of 32 P, probably due to ATP damage caused by strong 32 P radioactivity. Therefore, usable [γ - 32 P]ATP has an even shorter half-life (<14 days). Frequent supply of fresh [γ - 32 P]ATP is necessary for efficient 5' RNA labeling by 32 P and PNK. Unless it is used by several experimenters in a sizable laboratory or shared among different laboratories, a substantial amount of [γ - 32 P]ATP is wasted due to its fast radiodecay. Second, [γ - 32 P]RNA may slowly lose its function during storage as a result of structural damage by its 32 P radioactivity. Accordingly, it may be necessary to frequently prepare freshly 32 P-labeled RNA before a previous preparation is fully

consumed. Third, some RNA (such as the one shown in Figure 6A) may present difficulties for efficient labeling by PNK and [γ - 32 P]ATP. Finally, 32 P is a source of radio-hazard. Its use requires user training and strict workplace safety regulations.

Our newly developed **F550/570** and **F650/670** offer solutions to the above [γ - 32 P]ATP-based RNA labeling problems. Regardless of RNA structure, the 5' end of RNA can be readily labeled by **F550/570** and **F650/670**, if the RNA has AG at its 5' end. If the subject RNA does not possess a 5' AG sequence, AG can be added to the 5' end for the purpose of facilitating 5' fluorescent labeling by **F550/570** or **F650/670**. With phosphorimagers/fluorimagers becoming widely accessible, the RNA-labeling advantages offered by **F550/570** and **F650/670** over traditional 32 P-based methods may be fully realized.

The cyanine dye-AMP conjugates **F550/570** and **F650/670** described in this report have been made available by AdeGenix Inc. (Monrovia, CA). Detailed application notes can be found at www.adeenix.com.

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