

# Patients harboring *ALK* rearrangement adenocarcinoma after acquired resistance to crizotinib and transformation to small-cell lung cancer: a case report

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**Abstract:** Anaplastic lymphoma kinase (*ALK*) rearrangement responds to *ALK* tyrosine kinase inhibitors (TKIs) in lung cancer. Many cases ultimately acquire resistance to crizotinib. Resistance, including *ALK*-dominant or *ALK* non-dominant, mechanisms have been described. Transformation to small-cell lung cancer is rare. Herein, we report a 49-year-old man diagnosed with adenocarcinoma, who was negative for *EGFR* and *ALK* genes as detected by reverse transcription polymerase chain reaction, and was treated with crizotinib. A new biopsy showed a small-cell lung cancer after disease progression. Then, next-generation sequencing (NGS) was carried out and detected a *TP53* gene mutation, an *ALK* rearrangement, and no loss of the retinoblastoma gene (*RB*). Although a regimen for small-cell lung cancer may be one treatment option, a heterogeneous tumor may exist at the time of diagnosis and manifest during the course of disease.

**Keywords:** lung cancer, *ALK*, crizotinib, small-cell lung cancer

## Introduction

Anaplastic lymphoma kinase (*ALK*) and echinoderm microtubule-associated protein-like 4 (*EML4*) gene rearrangements occur in 2%–7% of patients with non-small cell lung cancer (NSCLC).<sup>1</sup> Crizotinib is the standard treatment for advanced *ALK*-positive NSCLC and has been shown to have impressive single-agent activity in this type of patient.<sup>2,3</sup> While most patients respond to crizotinib, every patient will ultimately experience disease progression within 1–2 years.<sup>2</sup> Drug resistance has not only become the major limitation of clinical efforts, but also is the most urgent issue in need of resolution for prolonging life in patients with NSCLC. It is encouraging that molecular acquired resistance mechanisms to tyrosine kinase inhibitors (TKIs) have been identified. With in-depth research, new therapeutic strategies to overcome resistance have been used to prolong survival of patients with advanced NSCLC. Mechanisms of acquired resistance to crizotinib that have been explored include on-target genetic alterations or off-target mechanisms of resistance;<sup>4</sup> however, other complex resistance mechanisms, such as mechanisms of resistance to *EGFR*-TKIs, still exist.<sup>5</sup> Histologic transformation into small-cell lung cancer (SCLC) has been reported in some *ALK* fusion adenocarcinomas; however, there are no established treatment strategies to manage such transformations.

Here, we report a patient who developed acquired resistance to crizotinib with transformation to SCLC. Moreover, we have summarized some published reports which have improved our understanding of the transformation to SCLC in *ALK* fusion adenocarcinoma.

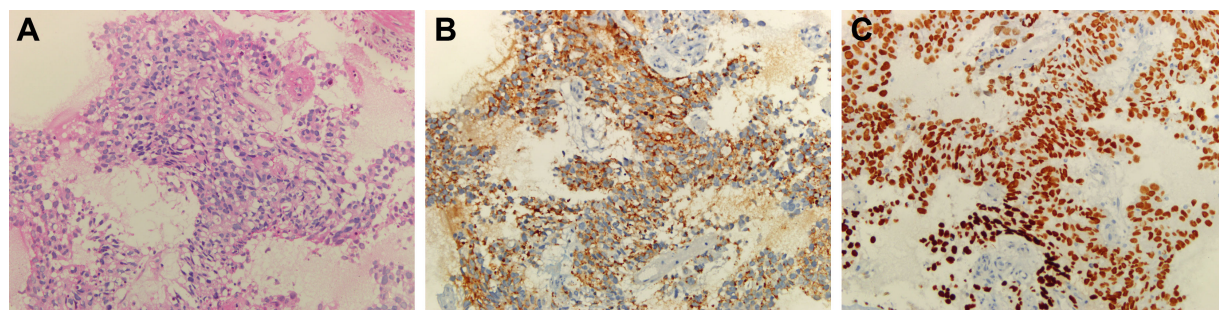
## Case report

A 49-year-old man, a never smoker, presented to our hospital with a 1-month history of cough and blood-stained sputum in October 2012. CT scans revealed a mass in the right upper lung, right hilar lymph node enlargement, and multiple nodules in the right lung. Cerebral and lumbar magnetic resonance imaging (MRI) revealed a nodule in the right cerebellum and a centrum tumor at the L<sub>2</sub> level (T<sub>4</sub>N<sub>1</sub>M<sub>1b</sub>, stage IV). A pathologic diagnosis of adenocarcinoma cells was established based on a bronchoscopic biopsy (Figure 1). Immunohistochemical (IHC) analysis demonstrated positivity for TTF-1 and Napsin A, and negativity for cytokeratin (CK) 5/6 and P63 (Figure 1 and Table 1). Tumor tissue was shown to be wild-type of epidermal growth factor receptor variants by ARMS (AmoyDx, Xiamen, People's Republic of China), and *EML4-ALK* fusion was shown by reverse transcription polymerase chain reaction (RT-PCR; AmoyDx; Figure 2). The patient received five cycles of first-line chemotherapy with cisplatin and paclitaxel from October 2012 to January 2013. The best tumor response was a partial response according to RECIST criteria, and the progression-free survival (PFS) was 7.0 months. In May 2013, the tumor progressed (right lower lobe nodules and brain metastases). The patient underwent crizotinib treatment (250 mg/bid, orally) from May 2013 to October 2014. He received whole-brain radiotherapy (2 Gy per fraction, 1 fraction per day ×20 days; total radiation dose 50 Gy). The curative effect of crizotinib treatment was stable disease (SD). After progression on crizotinib, the patient underwent multiple cycles of cytotoxic chemotherapy (gemcitabine,

docetaxel, and bevacizumab; Table 2). A secondary biopsy of the enlarging mass in the right lung was performed. Histologic examination of the biopsy specimen revealed SCLC without adenocarcinoma components (Figure 3). IHC analysis demonstrated positivity for Syn and CD56, negativity for CgA, and a Ki-67 index of 98% (Figure 3). To search for new therapeutic strategies, additional gene detection was performed on the tissue sample by next-generation sequencing (Gene plus, Beijing, People's Republic of China), which showed a *TP53* gene mutation (p.R248W) and *ALK* rearrangement accompanied by *ERBB3* p.P554Q, *NOTCH1* p.P3S, *IL7R* p.P417R, and *IL6ST* p.A896V, but no loss of the retinoblastoma gene (*RB*). The next generation sequencing (NGS) assay was performed using the HiSeq4000 (Illumina, San Diego, CA, USA). Then, the patient underwent etoposide monotherapy. After two cycles, the effect was SD. The patient remains alive at the time of writing this article. We have listed small-cell transformations following treatment with ALK-TKIs, including crizotinib/alectinib/ceritinib/lorlatinib in Table 3. The authors confirm that written informed consent for publication of case details and any accompanying images has been provided by the patient.

## Discussion

With the rapid development of molecular diagnostic technology, targeted therapy is effective for patients with advanced NSCLC and associated driver genes; however, a majority of patients will eventually acquire resistance to the targeted drug and experience disease progression. Therefore, some *ALK*-positive patients also develop acquired resistance to ALK-TKIs. Known acquired mechanisms that cause resistance to ALK-TKIs have been described, and can be divided into two types (*ALK* dominant and *ALK* non-dominant).<sup>13</sup> *ALK*-dominant mechanisms include *ALK* secondary mutations and *ALK* amplification, whereas *ALK* non-dominant



**Figure 1** Hematoxylin–eosin staining and immunohistochemistry in adenocarcinoma before crizotinib treatment.

**Notes:** (A) Hematoxylin–eosin staining revealed that tumor cells were lung adenocarcinoma (×200). (B) Immunohistochemical examination revealed that tumor cells were positive for monoclonal anti-TTF-1 antibody (×200). (C) Immunohistochemical examination revealed that tumor cells were positive for monoclonal anti-Napsin A antibody (×200).

**Table 1** Primary antibodies used for immunohistochemical staining

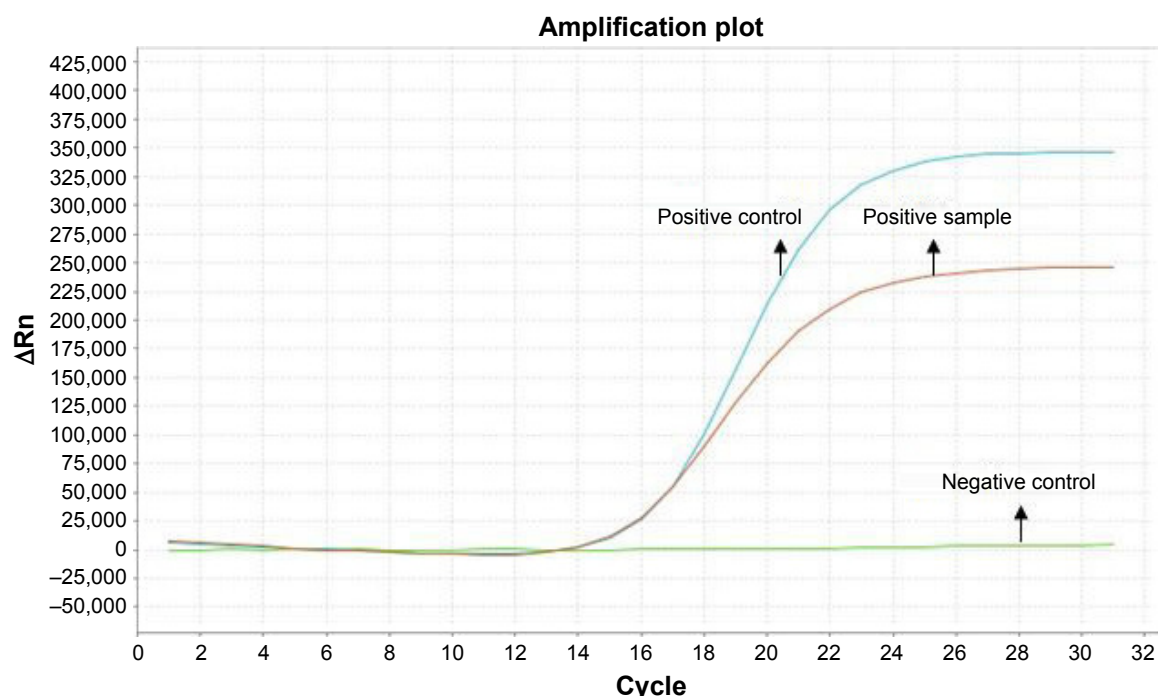
| Antibody | Clone   | Dilution | Purchased from                |
|----------|---------|----------|-------------------------------|
| TTF-1    | SPT24   | 1:100    | Zymed Laboratories, Inc.      |
| NapsinA  | OTI3E5  | 1:100    | Zymed Laboratories, Inc.      |
| P63      | UMAB4   | 1:100    | Zymed Laboratories, Inc.      |
| CK5/6    | D5/16B4 | 1:100    | Zymed Laboratories, Inc.      |
| CK8/18   | Zym5.2  | 1:100    | Zymed Laboratories, Inc.      |
| SP-A     | 32E12   | 1:100    | Zymed Laboratories, Inc.      |
| CK       | AE1/AE3 | 1:100    | Zymed Laboratories, Inc.      |
| Syn      | EPI 58  | 1:100    | Zymed Laboratories, Inc.      |
| CgA      | EP38    | 1:100    | Zymed Laboratories, Inc.      |
| CD56     | UMAB83  | 1:100    | Zymed Laboratories, Inc.      |
| Ki-67    | EP5     | 1:100    | Zymed Laboratories, Inc.      |
| PD-L1    | SPI 42  | 1:100    | Ventana Medical Systems, Inc. |

mechanisms include bypassing downstream signaling, such as the epidermal growth factor receptor (*EGFR*), *KRAS*, *KIT*, *MET*, and insulin-like growth factor 1 receptor pathways. Approximately 25% of acquired mechanisms of resistance are mediated by unknown mechanisms.<sup>14</sup> With respect to EGFR-TKIs, 3%–15% of the patients with *EGFR*-mutated lung cancer develop acquired resistance to EGFR-TKIs by histologic transformation to SCLC.<sup>15,16</sup> Similarly, a mechanism underlying SCLC transformation in *ALK*-positive tumors has been reported in some cases.<sup>6–12</sup>

In the present case in which the re-biopsied SCLC tissue was examined, *ALK* rearrangement was still detected. In addition, chemotherapy and/or anti-angiogenic drug treatment-induced change in initial tumor morphology should,

therefore, be considered. If the transformation had occurred before crizotinib treatment, a rapid growth of the primary lesion during crizotinib therapy would have been observed. Moreover, the primary lesion did not show an increase in size during albumin paclitaxel or docetaxel combination with bevacizumab therapy, and only limited agent activity on SCLC was reported with these agents.<sup>17</sup> These findings imply that the transformation to SCLC during crizotinib treatment was the main cause of acquired resistance in this case. The pathophysiologic mechanism underlying transformation to SCLC following ALK-TKI treatment is not well understood. According to the mechanism of EGFR-TKIs, two possibilities have been stated, including a phenotype switch from NSCLC to SCLC, and SCLC and adenocarcinoma may coexist at baseline, with SCLC becoming dominant during disease progression after EGFR-TKI therapy.<sup>5,18</sup> We thought the mechanism of transformation to SCLC in ALK-TKIs was similar to that with EGFR-TKIs. Although the pathologic features of the present case were re-evaluated, SCLC was not identified in the primary biopsy specimen. Because of the initial bronchial biopsy, there was limited availability of the tissue sample.

In contrast, although the cause of transformation into SCLC is unclear, inactivation of tumor suppressor genes (*RB* and *TP53*) seems important and constitutes an initial event in the tumorigenic process.<sup>16</sup> Some reports have revealed the role of *RB* gene loss and/or *TP53* mutation in *ALK*-positive

**Figure 2** Amplification plot of *EML4-ALK*-positive by amplification refractory mutation system method.

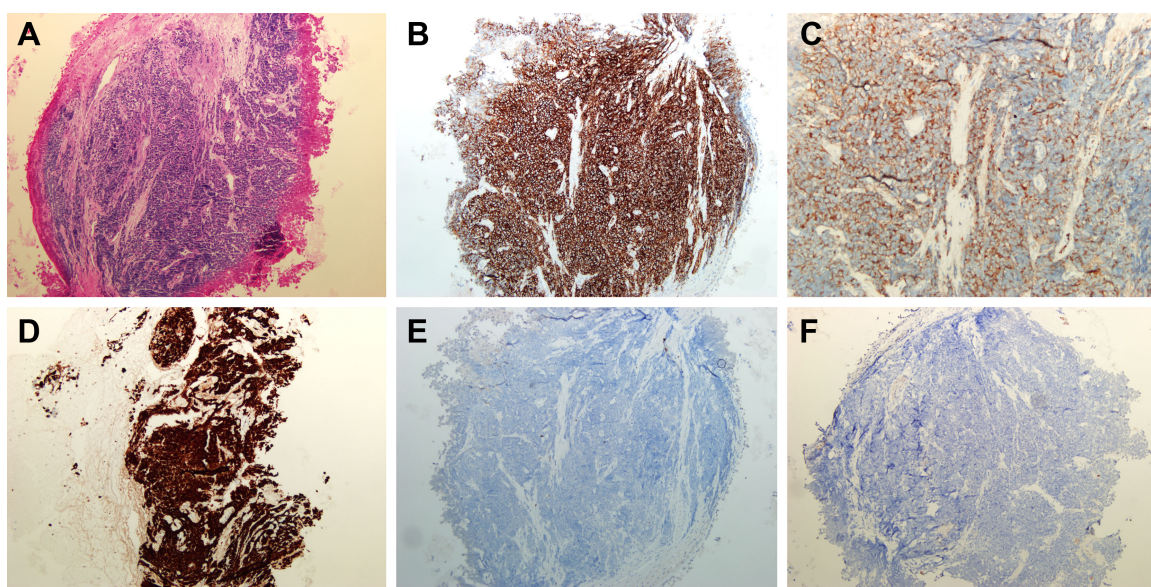
**Table 2** Details of the treatment process

| Treatment  | Time                           | Curative effect |
|--|--------------------------------|-----------------|
| Albumin paclitaxel + cisplatin, 2 cycles; albumin paclitaxel, 3 cycles | October 2014 to February 2015  | SD              |
| Anlotinib (clinical trial)   | September 2015 to June 2016    | SD              |
| Gemcitabine + Nedaplatin   | June 2016 to August 2016       | PD              |
| Docetaxel + bevacizumab, 4 cycles; bevacizumab, 2 cycles               | August 2016 to November 2016   | SD              |
| Rechallenge with crizotinib  | November 2016 to December 2016 | PD              |

**Abbreviations:** SD, stable disease; PD, progressive disease.

NSCLC transformed into SCLC<sup>11,15,16</sup>; however, in our case, there was no loss of *RB*, but a *TP53* gene mutation occurred. We reasoned that *RB* gene loss or *TP53* gene mutation could participate in the transformation of adenocarcinoma to SCLC, but the two genes were not always concurrent as the transforming event. Furthermore, we demonstrated a *NOTCH1* gene mutation, and this constituted the first reported gene involving transformation to SCLC. It has been reported that approximately 25% of patients with SCLC have a *NOTCH1* gene family abnormality identified on next-generation sequencing.<sup>19</sup> We hypothesize that, if inactivation of the *RBI* or *P53* or *NOTCH1* gene is identified in the primary tumor tissue, the tumor may be at higher risk of SCLC transformation following target drug therapy.

The present case supports attempts to re-biopsy the tumor for selection of treatment after acquired resistance. There are no established treatment strategies to manage patients with transformation to SCLC. In agreement with reported cases, Fujita et al<sup>8</sup> reported treatment with a combination of irinotecan and alectinib, resulting in an ongoing partial response to the primary lesion with a maintained partial response to the other lesions. Miyamoto et al<sup>9</sup> showed that after two cycles of cisplatin–irinotecan treatment, there was a reduction in the size of mediastinal lymph nodes, whereas there was an increase in the size and number of nodes in both lung fields. Ou et al<sup>12</sup> suggest that the patient switch to a third-generation ALK-TKI within 2 months of treatment and disease progression. The patient underwent etoposide monotherapy in our case. After two cycles, the effect was SD. Nevertheless, as the heterogeneity of response to different therapeutic strategies of the reported case suggests, the re-biopsy site only represents a partial pathology of the resistance, and the mechanism of resistance may differ from one site to another. Given the heterogeneity and the variety of acquired resistance mechanisms to target drugs, re-biopsy of only one site might not always be appropriate. We think detection of gene variations in blood samples with homogeneity in ctDNA by NGS may be a supplemental method with re-biopsy for analysis, in some cases. It is clinically challenging to choose the treatment mode when transformation to SCLC occurs. More research on optimal treatment methods are needed.



**Figure 3** Hematoxylin–eosin staining and immunohistochemistry in small-cell cancer after crizotinib treatment.

**Notes:** (A) Hematoxylin–eosin staining revealed that tumor cells were lung small-cell cancer ( $\times 100$ ). (B) Immunohistochemical examination revealed that tumor cells were positive for monoclonal anti-CD56 antibody ( $\times 100$ ). (C) Immunohistochemical examination revealed that tumor cells were positive for monoclonal anti-Syn antibody ( $\times 200$ ). (D) Immunohistochemical examination revealed that the tumor cell proliferation index was 98% for monoclonal anti-Ki-67 antibody ( $\times 100$ ). (E) Immunohistochemical examination revealed that tumor cells were negative for monoclonal anti-CgA antibody ( $\times 100$ ). (F) Immunohistochemical examination revealed that tumor cells were negative for monoclonal anti-PD-L1 antibody ( $\times 100$ ).

**Table 3** List of reported small-cell transformations resistant to crizotinib/alectinib/ceritinib/lorlatinib

| Study  | Cha et al <sup>6</sup>                         | Takegawa et al <sup>7</sup>                    | Fujita et al <sup>8</sup>                      | Miyamoto et al <sup>9</sup>                    | Caumont et al <sup>10</sup> | Levacq et al <sup>11</sup>                    |
|--|--|--|--|--|-----------------------------|---|
| Ethnicity  | Korean   | Japanese                                       | Japanese                                       | Japanese                                       | French                      | Belgian                                       |
| Age (years)  | 72   | 43   | 67   | 58   | 63                          | 53  |
| Gender   | Male   | Female   | Female   | Female   | Female                      | Female  |
| Smoking history  | 40 pack-years                                  | NA   | Never smoker                                   | Never smoker                                   | Never smoker                | Never smoker                                  |
| CNS metastasis present   | No   | Yes  | Yes  | No   | Yes                         | No  |
| Resistance to most immediate prior crizotinib/alectinib/ceritinib/lorlatinib | Crizotinib                                     | Alectinib (also crizotinib prior to alectinib) | Alectinib (also crizotinib prior to alectinib) | Alectinib (also crizotinib prior to alectinib) | Crizotinib                  | Ceritinib (also crizotinib prior to certinib) |
| Subsequent SCLC therapy  | NA   | None   | Alectinib + irinotecan (with PR)               | Cisplatin/irinotecan                           | NA                          | Cisplatin/etoposide followed by CAV           |
| Study  | <b>Ou et al<sup>12</sup></b>                   |  |  | <b>8 (present case)</b>                        |                             |   |
| Ethnicity  | American                                       |  |  | Chinese  |                             |   |
| Age (years)  | 35   |  |  | 49   |                             |   |
| Gender   | Male   |  |  | Male   |                             |   |
| Smoking history  | Never smoker                                   |  |  | Never smoker                                   |                             |   |
| CNS metastasis present   | Yes  |  |  | Yes  |                             |   |
| Resistance to most immediate prior crizotinib/alectinib/ceritinib/lorlatinib | Lorlatinib (also prior ceritinib to alectinib) |  |  | Crizotinib                                     |                             |   |
| First-line treatment assessment  | Carboplatin/etoposide + alectinib              |  |  | Etoposide                                      |                             |   |

**Abbreviations:** CAV, chemotherapy regimen (cyclophosphamide + doxorubicin + vincristine); CNS, central nervous system; PR, partial response; SCLC, small-cell lung cancer; NA, not available.

## Conclusion

Oncologists should realize the possibility of transformation to SCLC after patients acquire resistance to ALK-TKI therapy. A re-biopsy should be performed to facilitate histologic and molecular analysis. Transformation to SCLC is an important consideration for choosing appropriate therapy due to the potential efficacy of standard SCLC treatments or a combination of next-generation AKL-TKIs.

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## Disclosure

The authors report no conflicts of interest in this work.

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