



Published in final edited form as:

Biol Psychiatry. 2017 August 01; 82(3): 165–175. doi:10.1016/j.biopsych.2016.12.030.

Imaging Genetics and Genomics in Psychiatry: A Critical Review of Progress and Potential

Ryan Bogdan¹, Betty Jo Salmeron², Caitlin E Carey¹, Arpana Agrawal³, Vince D Calhoun⁴, Hugh Garavan⁵, Ahmad R Hariri⁶, Andreas Heinz⁷, Matthew N Hill⁸, Andrew Holmes⁹, Ned H Kalin¹⁰, and David Goldman¹¹

¹BRAIN Lab, Department of Psychological and Brain Sciences, Washington University in St. Louis, St. Louis, Missouri, USA

²Neuroimaging Research Branch, National Institute on Drug Abuse (NIDA), Intramural Research Program, Baltimore, MD, USA

³Department of Psychiatry, Washington University in St. Louis School of Medicine, St. Louis, Missouri, USA

⁴The Mind Research Network and Lovelace Biomedical and Environmental Research Institute; Department of Psychiatry and Neuroscience, University of New Mexico; Department of Electronic and Computer Engineering, University of New Mexico, Albuquerque, NM, USA

⁵Department of Psychiatry, University of Vermont, Burlington, VT, USA

⁶Laboratory of NeuroGenetics, Department of Psychology & Neuroscience, Duke University, Durham, NC, USA

⁷Department of Child and Adolescent Psychiatry, Psychosomatics, and Psychotherapy, Charité-Universitätsmedizin Berlin, Germany

⁸Hotchkiss Brain Institute, Departments of Cell Biology and Anatomy and Psychiatry, University of Calgary, Calgary, Alberta, Canada

⁹Laboratory of Behavioral and Genomic Neuroscience, National Institute on Alcoholism and Alcohol Abuse, Bethesda, United States

¹⁰Department of Psychiatry, University of Wisconsin, Madison, Wisconsin; Neuroscience Training Program (NHK, RK, PHR, DPMT, MEE), University of Wisconsin, Madison, Wisconsin; Wisconsin National Primate Research Center (NHK, MEE); Madison, Wisconsin

¹¹Laboratory of Neurogenetics, National Institute on Alcohol Abuse and Alcoholism (NIAAA), Intramural Research Program, Bethesda, MD, USA

Corresponding Author: Ryan Bogdan (rbogdan@wustl.edu, phone: 617-407-5645, address: CB 1125 Psychological and Brain Sciences Bldg Room 453B, Washington University in St. Louis, One Brookings Drive, St. Louis, MO 63130, USA).

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Disclosures:

The authors report no biomedical financial interests or potential conflicts of interest.

Abstract

Imaging genetics and genomics research has begun to provide insight into the molecular and genetic architecture of neural phenotypes and the neural mechanisms through which genetic risk for psychopathology may emerge. As it approaches its third decade, imaging genetics is confronted by many challenges including the proliferation of studies using small sample sizes and diverse designs, limited replication, problems with harmonization of neural phenotypes for meta-analysis, unclear mechanisms, and evidence that effect sizes may be more modest than originally posited, with increasing evidence of polygenicity. These concerns have encouraged the field to grow in many new directions including the development of consortia and large scale data collection projects as well as the use of novel methods (e.g., polygenic approaches, machine learning), which enhance the quality of imaging genetic studies, but also introduce new challenges. Here, we critically review progress in imaging genetics and offer suggestions and highlight potential pitfalls of novel approaches. Ultimately, the strength of imaging genetics and genomics lies in its translational and integrative potential with other research approaches (e.g., non-human animal models, psychiatric genetics, pharmacologic challenge) to elucidate brain-based pathways that give rise to the vast individual differences in behavior as well as risk for psychopathology.

Keywords

imaging; genetics; genomics; neurogenetics; mri; polygenic; candidate

By linking genetic and epigenetic variation to brain structure, function, connectivity, and chemistry via neuroimaging measures (1), imaging genetics and genomics can inform the neural mechanisms through which genetic and molecular differences impact cognition, emotion, and behavior in health and disease. Since being pioneered nearly 20 years ago by candidate gene studies of receptor ligand binding (2–6; Supplemental Material), imaging genetics has incorporated a host of allied neuroimaging techniques, most frequently, structural and functional magnetic resonance imaging (sMRI, fMRI) and has been integrated with traditional psychiatric genetics (7–9) and non-human animal models (10–13). More recently, this approach has been extended to epigenetics (14,15), and, as imaging genomics, to discovery-based (16,17) and polygenic (18,19) approaches.

Accompanying an exponential increase in publications, imaging genetics and genomics has also been confronted by several qualitative concerns including the proliferation of studies with small sample sizes, limited replication, unclear mechanisms relating genes to brain and brain to behavior, and evidence that effect sizes may be smaller than originally thought, and perhaps no larger than effects for traditional psychiatric diagnoses (9,20). Such concerns, and the desire to find new genes and pathways via genomic approaches, have led to the formation of consortia and large-scale projects to increase sample size (21–26) as well as the adoption of methodological and technological innovations in genetics (e.g., GWAS, epigenetics), neuroimaging (e.g., multimodal PET/fMRI), and psychiatric genomics (e.g., polygenic risk scores, LD score regression) (9,14,18,27–30), all of which enhance the quality of imaging genetic studies, and each of which is also subject to new potential pitfalls.

Here, we critically review the current state of imaging genetics and genomics highlighting unique strengths, considerations, and limitations of distinct approaches while considering their utility for psychiatry going forward. We suggest that some criteria to evaluate the usefulness of intermediate phenotypes according to an endophenotype conceptualization are retrograde and counterproductive when applied to imaging genetics in some instances. We argue that single variant analyses remain informative in the context of a polygenic architecture that underlies the majority of imaging phenotypes. Further, we discuss the lack of replication in imaging genetics and what has been learned, and not learned, from meta-analytic efforts. Next, we review the use of candidate and discovery-based polygenic methods that aim to better characterize the complex polygenic architecture of imaging phenotypes and consider pitfalls that these techniques may face and how they may be minimized. We highlight the potential of molecular genomic methods to verify and mechanize relationships between the dynamic genome and neural phenotypes. Finally, we consider how imaging genetics and genomics hold their greatest potential not in isolation, but as methods that can be used alongside other techniques (e.g., pharmacologic challenge), levels of analysis (e.g., the transcriptome, psychiatric genetics), and non-human animal research (e.g., genetic models) in the search for mechanistic consilience (Table 1). As imaging genetics and genomics further integrate with molecular genetics, basic neuroscience, and psychiatric genetics, and begins to accumulate not only large but also longitudinal samples, it will be able to more adequately model and test the complex interplay between genes, the brain, body, environment, and behavior and expand these pathways (Figure 1). It is hoped that such mechanistic characterization will ultimately improve the nosology, treatment, and prevention of mental illness.

Is the Endophenotype Conceptualization of Intermediate Phenotypes Useful?

Theoretically, intermediate phenotypes, such as imaging phenotypes, lie along a mechanistic pathway through which genetic variation and/or environmental experiences contribute to clinical phenotypes (Figure 1A)(31). Here, we refer to the traditional pathway from the static genome to neural intermediate phenotypes and behavior, although modern genetics regularly challenges such unidirectionality (Figure 1B). Within the theoretical discussion of intermediate phenotypes, the greatest attention has often focused on the endophenotype conceptualization, which stipulates that endophenotypes are associated with psychiatric disease and heritable, among other considerations (32).

The requirement of disease-association presupposes the research value of psychiatric nosology. This is problematic because many, if not all, psychiatric diagnoses are heterogeneous amalgamations of symptoms, with the same diagnosis having distinct putative etiologies, as is becoming more clear following RDoC (33,34). Such diagnostic heterogeneity may dilute, and even obliterate intermediate phenotype–disease association. For example, although anhedonia is a cardinal symptom of depression, it is not amongst the most common symptoms (35). As such, anhedonia-related neural circuitry may not be identified or minimized in a general patient/control study (36,37). Indeed, some reports have associated depression with blunted reward-related activity in the ventral striatum (38,39),

while others have not (40). Or consider that despite the polygenic nature of psychosis (41), some patients presenting with psychosis have a genetic variation in Huntingtin(42), or velocardiofacial syndrome(43). Thus, it is possible that distinct etiologies associated with unique presentations could be lost or minimized by a reliance on diagnosis (44). The positive results of a recent GWAS study on melancholic depression, a more severe and homogenous form of the disorder further reinforce this concern (33),but see(45). This is not to imply that understanding variability in disease associated intermediate phenotypes is not important, but rather that constraining imaging genetics research to intermediate phenotypes or genes previously linked to a disorder may stifle research on etiologic brain-based associations by generating an intellectual *file drawer problem* where only hypotheses satisfying endophenotype-disease correspondence are evaluated impeding the development of etiologically-based classification.

Heritability is on a scientific basis a more logical endophenotype criterion. Twin studies have largely focused on the heritability of morphological measures, which approach the upper end of psychiatric estimates, ranging from 60–80% (46,47). The few studies of brain function suggest more modest estimates (~40%)(46). However, *intermediate phenotypes* that are not heritable can still have genetic origins and mediate relationships between genes and behavior. For example, Williams Syndrome, which is characterized by a host of physical and personality characteristics, including excessive sociality is attributable to a microdeletion that typically occurs during the formation of germline cells in people with no history of the disorder (48,49). Common variation within the genes (e.g., *GTF2I*) spanning the microdeletion region have become candidates that are informing phenotypes related to sociability (22,50). Intermediate phenotypes can also represent stable trait differences that while not entirely heritable per se, are dependent upon experience arising as the product gene by environment interactions (e.g., *FKBP5* (20,51,52); Supplemental Material). Additionally, within genetic studies, non-heritable intermediate phenotypes may characterize individuals who are part of distinct, non-genetic subgroups, and who would otherwise be indistinguishable diagnostic phenocopies; such insight may contribute to subgroup classification, and diagnostic refinement. Finally, heritability refers to loci shared identical-by-descent representing the static genome. However, genes may traverse an imaging phenotype on their pathway to behavior, even when the intermediate phenotype is not heritable. For example, a new wave of epigenetic research (14,53–55), examining markers among discordant monozygotic twins is poised to take advantage of non-heritable intermediate phenotypes. The validity of a result that would depend on the phenotype being highly heritable (e.g., polygenic risk) would be suspect if the phenotype was not. However, the widespread application of this criterion could unintentionally impede important etiologic insight generated from genomic research on non-heritable neural phenotypes.

Single Variant Approaches

The majority of imaging genetics research has been conducted within a candidate gene framework. Most studies have focused on a limited number of functionally characterized polymorphisms (e.g., *COMT*rs4680(56), *SLC6A4* 5-HTTLPR(57) within genes coding for products that influence particular neural systems. Most of these variants have been inconsistently associated with neural phenotypes and psychopathology with both positive

and null associations reported (58–61). Recently, the unprecedented success of genomewide association studies (GWAS) has identified new candidate genes (e.g., *KTNI*(16)) and corroborated the role of prior suspects (e.g., *SIRT1*(33,62,63)). Polymorphisms discovered in psychiatric GWAS are now being investigated within a candidate framework with promising results emerging (64–66), though other evidence suggests limited overlap between polymorphisms associated with clinical and neural phenotypes (9,67).

The Controversy of Candidate Associations

As in psychiatric genetics(60,61), the intuitive mechanistic and interpretable appeal of candidate imaging genetics findings have led to many replication and extension studies, and as many contradictory findings. Several meta-analyses have concluded that effect sizes are likely smaller than originally reported, may represent false positive associations, and that publication bias may promote false confidence in the robustness and biological importance of these effects (58,68,69). However, the utility of meta-analysis for some imaging phenotypes is questionable.

Meta-analysis tends to work best under two conditions. First, when constructs are measured in a standard fashion (e.g., obesity and type 2 diabetes (70)), they estimate effects with great precision. Second, study design differences across studies can be modeled *with* a large number of studies using each design, allowing meta-analyses to examine whether design differences influence associations. Within neuroimaging, the Enhancing NeuroImaging Genetics through Meta-Analysis (ENIGMA) consortium has successfully harmonized imaging data across studies to meta-analyze structural phenotypes in a GWAS context(16). However, because many imaging genetics studies, such as those probing task-related activity, do not use standardized methodology (e.g., task and analysis), they present unique challenges.

Methodological differences across neuroimaging studies can meaningfully impact the nature of measured neural phenotypes. For instance, consider the literature on amygdala reactivity to emotional faces in autism. Early studies produced evidence of hypoactivation (71,72). However, eye tracking research has shown that children with autism typically avoid eye contact (73), which conveys important emotional information and robustly recruits amygdala activation(74). Studies directing or measuring participant eye gaze have shown elevated amygdala reactivity in autism that is correlated with eye gaze duration (75–77). A meta-analysis not considering such design differences, may produce data that autism is not associated with amygdala function (78). This is not to suggest that studies of non-imaging phenotypes are impervious to these challenges (e.g., self-report versus measured weight) or that such differences are responsible for inconsistent findings, but merely, that harmonization challenges are heightened among meta-analyses of some imaging phenotypes.

Meta-analyses have attempted to model differences across studies. For example, in a meta-analysis of the relationship between 5-HTTLPR genotype and amygdala function, Murphy and colleagues (68) examined whether a host of study characteristics influence the association. However, the small number of studies using each design and variability within study groupings may have left this approach unable to adequately model differences. For

example, studies were coded according to ethnicity and studies of German (79) and Korean participants (80), where grouped together as “not European/Mixed.” Such heterogeneous representations of study variability are inadequately powered and conceptualized leaving the conclusions of marginally significant small effects, debatable. What remains unequivocal is that data are inconclusive; whether positive or null associations better represent reality can only be addressed by further research. Overall, meta-analytic approaches have provided road maps for challenges associated with candidate studies (60,81) and identified loci conferring small effects for psychiatric and structural neuroimaging phenotypes (16,82). However, the utility of meta-analyses incorporating studies using diverse methodology when study related differences cannot be systemically evaluated, is questionable.

Much like data on complement component 4 and schizophrenia (83), some imaging genetics phenotypes may presently be better informed by convergence across modes of investigation (e.g., Table 1). For instance, the significance of 5-HTTLPR findings may be weighed alongside observations in non-human animal models, and effects of the 5-HTTLPR polymorphism on serotonin transporter expression (84). For example, work using tissue oxygen amperometry (which measures hemodynamic responses equivalent to BOLD fMRI in freely moving rodents;(85), has shown that serotonin transporter overexpression reduces amygdala responses to aversive cues in mice; a finding remarkably convergent with significant results reported in the human 5-HTTLPR BOLD fMRI amygdala literature (86,87).

Genomewide Association Study Approaches

Much like initial psychiatric GWAS, the first imaging GWAS did not identify any genomewide significant polymorphisms, likely due to inadequate power (17). While other early imaging GWAS have observed genomewide significant results these were not replicated (88). Arguably, within imaging, GWAS did not become particularly informative until the development of large consortia such as ENIGMA (21), through which investigators have pooled effect size estimates to achieve samples large enough to reliably detect loci of small effect (9,16). For example, two GWAS have linked rs7294919 genotype to hippocampal volume (89,90), with subsequent candidate replication (91).

In addition to identifying new polymorphisms, GWAS data invite speculation regarding prior assumptions. Indeed, recent evidence suggests that genetic associations for schizophrenia and subcortical brain volume are similarly small in size and non-overlapping (9). If data accumulate showing that neuroimaging measures are not associated with larger effects than clinical diagnoses, it will be important to consider factors that may contribute to this. For instance, despite being etiologically and descriptively heterogeneous, clinical diagnoses have been well studied psychometrically and have acceptable to excellent reliability, with notable exceptions (depression, generalized anxiety(92). However, other than structural neural phenotypes, which have evidence of robust reliability (93,94), the reliability of many neuroimaging phenotypes has not been rigorously investigated with inconsistent effects reported and conclusions unclear (95–99).

The success of meta-analytic psychiatric GWAS (41) has led to suggestions that GWAS may best inform psychiatry by using large samples with relatively easily assayed phenotypes

(100,101). As a consequence, imaging genetics would be most useful to understand the neural mechanisms underlying these associations. Clearly, this approach has utility, as multiple studies are beginning to demonstrate (66). However, much like the endophenotypic conceptualization, this approach presumes the value of our current conceptualization of mental illness, and further, assumes that loci linked to a particular disorder would also be linked to related neural phenotypes. However, unlike data suggesting that depression, subjective well-being, and neuroticism share substantive overlap in associated genetic variation (102), there is no overlap between genetic variation contributing to indices of subcortical brain volume and schizophrenia (9). Broadly, these results suggest that psychiatric and intermediate phenotype GWAS may provide different information that may ultimately lead to refined conceptions of mental illness decades in the future. Immediately, these data suggest that subcortical volume abnormalities observed in schizophrenia may instead arise from rare mutations (e.g., de novo), schizophrenia itself (103), its treatment (104,105), associated risk factors and potential GxE(106). By probing overlap across clinical phenotypes with neural outcomes, imaging genetics may usefully inform the origins of individual differences among psychiatrically-relevant neural phenotypes (e.g., subcortical volume schizophrenia). As larger samples accumulate ($N_s > 3000$), techniques such as LD score regression (107) may be used to estimate genetic correlations across neural and behavioral phenotypes. Imaging genomics may identify novel loci that do or do not map onto diagnostic categories, but may nonetheless contribute to our understanding of psychiatric conditions and potentially lead to refined nosology and treatment in the future.

Polygenic Approaches

With the exception of ligand-based neuroimaging techniques that target specific receptors, *in vivo* neuroimaging data provide assays of higher-order neural circuit function and structure reflective of thousands of interacting neurons and glia. As such, this resolution may be incongruous with the action of single genetic variants (20) leading imaging genetics and genomics to adopt polygenic techniques to quantify aggregate influence.

Polygenic Scores

Polygenic scoring approaches fall broadly within two categories: *polygenic risk scores (PRS)*, and *biologically-informed multilocus profile scores (BIMPS)* (20). The PRS approach summates “risk” alleles or weighted effects based upon prior GWAS summary statistics (108) and can identify neural mechanisms correlated with genetic risk for psychopathology. For example, depression PRS are associated with reduced ventromedial prefrontal cortex thickness, which is, associated with negative affect(18). For PRS studies to be maximally informative, particularly for phenotypes common in non-ascertained samples, it is important to evaluate whether associations remain after taking into account phenotypic expressions of the disease. Ideally, such relationships could be tested longitudinally to examine whether PRS-based associations with neural phenotypes precede and predict psychopathology. Notably, the PRS approach allows genomic liability to psychopathology to be evaluated among individuals without disorder expression thereby avoiding confounds of medication and disease process which plague the etiologic insight of psychiatric case-control studies (109). Further, unlike other approaches that estimate bivariate genetic correlations

(e.g., LD score) PRS allow for the degree of polygenicity to be examined and are more amenable to smaller samples, as long as the discovery cohort is sufficiently powered. Lastly, the application of Bayesian analytic approaches may have utility for imaging genetics in this context as they have improved observed effect sizes in a psychiatric phenotypes(110).

There are several limitations to the PRS approach however. It assumes additivity alone (which is supported(111)) and neglects potential epistatic effects, which while observed in imaging genetics studies (112,113) have yet to be widely replicated (114). Also, by aggregating across the genome, when used in isolation, PRS provide no insight into potential underlying molecular mechanisms. Further, this approach is constrained by the phenotypes used in the discovery-based sample, which may introduce heterogeneity (34,92) or be unrelated to the neural phenotypes under study (9). It is plausible that PRS are composed of heterogeneous gene sets contributing to distinct aspects of psychiatric diagnoses, wherein brain relationships are not observed within the full set but potentially a subset. Moreover, the predictive utility of PRS are largely based upon the sample size of discovery datasets, which arguably are just beginning to be achieved (115). Lastly, while recent developments in CRISPR/Cas techniques have facilitated multiplex genomic editing (116,117) that may eventually approximate polygenic risk, PRS approaches are not currently amenable to direct translational work in non-human animals.

The BIMPS approach summates functionally characterized polymorphisms across a given neural system to derive a composite of relative signaling within that pathway. For example, Nikolova and colleagues (28), found that BIMPS reflective of genetically-conferred elevations in dopamine signaling are predictive of elevated reward-related ventral striatum activity. Arguably, BIMPS approaches compound concerns regarding higher false discovery rates for candidate genetic association studies because they rely on priors for the genes (and loci in those genes) that constitute the system, assume how individual variants collectively contribute to overall signaling, posit that the action across these loci is additive, and provide multiple plausible profiles to be developed. For example, in light of opposing relationships between prefrontal and subcortical dopamine signaling (118,119), a dopamine BIMPS could reasonably be developed that reverse codes predominantly cortical-based genetic influence (120) as opposed to tonic dopamine regardless of region (28). As a result, it will be critical for future research to attempt replication defined as the same BIMPS and phenotype.

Notably, the integration of PRS and BIMPS may prove particularly efficacious. For example, imaging genetics could use GWAS-based results from psychiatric genetics to prioritize variants within a given system or systems. Using this framework, a recent study discovered common genetic moderators of the transcriptome response to stress hormone activation, that were also associated with depression (8). A PRS/BIMPS polygenic profile of variants associated with both stress hormone transcriptome response and depression was associated with overgeneralized amygdala responsiveness, providing a putative neural mechanism through which the transcriptome response to stress may influence depression risk (8).

System and Pathway-Level Analyses

Multiple methods have been developed to explore genetic variation at a system or pathway level, in either an *a priori* or exploratory fashion. When evidence suggests that a particular

protein or connected system contributes to a neural phenotype, yet SNP-based priors are unavailable or limited, candidate gene-level and systems-level analytic approaches may be employed, as has been more commonly done within psychiatry (7,121), but are beginning to be implemented in the context of imaging genetics (27). Clearly such approaches require adequate correction for the multiple exploratory tests conducted within and across sets to reduce Type 1 error rates; permutation-based procedures that keep genetic architecture intact while shuffling an intact phenotypic structure are particularly suited for this. Notably, how genetic variation within a gene/system-set is aggregated is controversial, with averaging being the most common (7). Nonetheless, results emerging from such analysis may prioritize particular sets and polymorphisms for further research interrogating potential function.

Using a more agnostic approach, GWAS data may also be mined to identify enrichment in known systems (122). For example, by using a full-genome pathway analysis (i.e., reducing 909,622 SNPs to 1,658 pathway), calcium responsive pathways were linked to neural activation to a face matching task in the absence of a genomewide significant locus (123). One benefit of pathway enrichment analyses is that it distills genomic data into genetic data representative of defined neural systems leading to data that may be more mechanistically interpretable and allow for greater translation with non-human animal models and pharmacologic challenge studies that can precisely target these systems. A unique concern of this approach is that it is restricted by known protein-protein interaction databases ((124) that may neglect known and unknown functional interactions among proteins.

Multivariate and Machine Learning Methods

As an alternative to univariate models, imaging genomics has begun to adopt “big data” techniques to facilitate data-driven discovery including the simultaneous modelling of genetic and imaging data to identify components with shared variance (29,125–127)). For example, parallel independent component analysis (p-ICA(29), uses genomewide and whole-brain imaging data to yield clusters of functionally related SNPs that are correlated with phenotypic components. Though traditionally performed agnostically at the whole-genome whole-brain level, modified hybrid approaches allow for the incorporation of prior information while also providing data-driven estimation (128). The multivariate fusion of imaging and genetics data allows for the identification of statistically linked genomic and neural components, which may provide insight into common mechanisms. Additionally, machine learning methods are beginning to be used in imaging genetics and genomics to predict or classify disease outcomes, which is perhaps the most direct clinical application of such methods for psychiatry. The use of these techniques in imaging genetics and genomics have typically relied upon well characterized candidate genes (e.g., (129), though data-driven analyses are also emerging (130). While in its infancy, considering clinical, neural, and genetic features in tandem for disease prediction is a promising future avenue of exploration that may have important clinical ramifications.

Despite their many benefits, multivariate techniques face a variety of unique limitations. Indeed, their use within psychiatric genetics has been controversial (131,132). For example, the high dimensionality of data frequently violates assumptions by including more features (i.e., input variables) than observations (i.e., participants). As such, dimensionality reduction

is typically required. Correspondingly, the vast number of inputs, multiple tests performed, and increased number of parameters being estimated risks overfitting the models and necessitates a heightened reliance on replication to confirm associations. However, the use of proper out-of-sample cross-validation approaches (e.g., leave-N-out), common in the machine learning literature outside of imaging genetics, can maximize the generalizability of a given study and, as such, should be universally adopted within the field.

Imaging Genetics and Genomics Going Forward: Conclusions

As imaging genetics and genomics prepares to enter its third decade, the field has exponentially expanded from its modest candidate gene investigation of ligand binding to include large scale single studies with more than 1,000 participants (perhaps unfathomable to neuroimaging researchers even 10 years ago), longitudinal designs, extensive data sharing, cross-modal investigation, and translation with non-human animal and psychiatric genetics research (21–26). Moreover, the field has begun to adopt novel methodology (e.g., the transcriptome) and analytic approaches (e.g., PRS, pathway analyses). This growth will undoubtedly enhance its ability to generate new etiologic knowledge that may ultimately enhance psychiatric nosology, treatment, and ideally, prevention. However, the same standards of skepticism, interest in replication, and insistence on biological validation apply as have arisen in candidate gene, one-locus at a time, imaging genetics. Arguably, as imaging genetics and genomics data increase in dimensionality and testing (e.g., data sharing), these concerns are only heightened.

Replication: Let's Do It When We Can but Accept When We Can't

As proposed by Carter and colleagues (133), and is applicable to research across fields (134), replication and appropriate correction for multiple testing is critical for confidence in research findings. While direct replication is the sine qua non, it is rarely done within an imaging genetics study (e.g.,(16), and we are hesitant to recommend it as a blanket criterion for publication, even when studies are small (133). Often methodological innovations are accompanied by substantive cost and going forward we could envision small samples that could yield formative insight into the genetic architecture of neural phenotypes that could not feasibly be replicated (e.g., recruitment based upon a rare variant; PET studies)(135). Ideally, we would replicate every association before it is published. However, we also must work within practical funding constraints. When replication can be tested, it undoubtedly should and null results should not be discouraged by journals (133). However, when replication cannot be attempted, perhaps it is best to take it for what it is – it might be an exceptionally innovative study that provides formative insight or a false positive, that unfortunately, may bias future research(135). The publication of null results and addressing citation biases within the literature (136), would help combat the development of such biases. When replication cannot be reasonably obtained, a compromise is using within-sample cross validation, which is feasible for small studies and would make inferences more generalizable. Further, it will be important to critically evaluate the properties of imaging phenotypes that may influence replication such as reliability and the factors that may influence this (e.g., time of day) to distill imaging phenotypes into trait-and state-related facets that enhance their research utility. Lastly, in addition to replication, we believe that

evidence of consilience should also be considered when evaluating findings, particularly in the context of increasing collaboration between imaging genetics, molecular genetics, non-human-animal models, as well as psychiatric and behavioral genetics ((84); Table 1). Evaluating convergence across methods is particularly useful when meta-analytic approaches may not yet be able to adequately model widespread between study variability. Indeed, it is precisely in the context of such convergence, that single variant candidate polymorphism investigations remain informative.

In Search of Mechanistic Understanding

Much like psychiatric and behavioral genetics, a major constraint on the utility of imaging genetics and genomics is its ability to inform molecular mechanisms which is predicated on the functional characterization of polymorphisms. With few exceptions (e.g., *FKBP5*(51)), polymorphisms have yet to be functionally detailed in a convincing manner. Pairing imaging genetics with molecular genetic and basic neuroscience research tools, holds tremendous potential. For example, in one of the most significant advances in psychiatric genetics, Sekar and colleagues (83) conducted a series of studies distilling the effects of the schizophrenia associated major histocompatibility (MHC) locus to complex variation within complement component 4 and showed that these alleles altered C4A and C4B expression in the brain, that was proportional to schizophrenia risk. Further, because C4 mediated synaptic pruning during postnatal development in mice, it is plausible that this may account for reduced synapses in schizophrenia. Indeed, it is precisely in this context that focused analyses remain relevant in our polygenic world.

In addition to better understanding molecular mechanisms using emergent technologies (e.g., RNA and methylation microarrays, RNA-Seq, bisulfite sequencing, ChIPSeq, mass spectroscopy), available databases of gene expression (BRAINEAC (137), braincloud (138), GTEx (139)) and bioinformatics tools (e.g., WUSTL epigenome browser (140)) may prove fruitful, particularly when interrogating novel and uncharacterized polymorphisms. Additionally, recent developments that allow for the imputation of the genetically-related transcriptome using GWAS data, such as TWAS (141) and PrediXcan (142), may aid in identification and confirmation of phenotype-related genes. It is also important to highlight that while integrative approaches will undoubtedly lead to greater etiologic insight, unique challenges need to be actively confronted. For example, because methylation is dynamic (143), with evidence that even proximal experiences (e.g., meal consumption), shape its landscape and measurement (144), it will be important for imaging genetics studies to collect DNA samples temporally linked to imaging data.

Summary

In conjunction with *in vitro*, *in vivo*, non-human animal research, pharmacologic manipulation, and psychiatric and behavioral genetics, imaging genetics and genomics can provide unique mechanistic insight into the genetic and experiential differences that contribute to psychiatric risk. We suggest that elements of the endophenotypic research conceptualization (e.g., disease-association) impede progress within imaging genetics and psychiatry and that some conclusions arising from meta-analyses may be premature in light

of phenotypic harmonization concerns. Further, we highlight the potential of relatively novel approaches in imaging genetics (e.g., PRS, pathway, multivariate) as well as challenges and limitations that each face while suggesting that single variant candidate gene analyses remain relevant, particularly in large samples alongside convergent evidence and anticipated small effects. Lastly, as knowledge and data continue to grow and are accompanied by methodological advances, data sharing, and prospective data collection, it becomes increasingly important to extend the traditional unidirectional model of imaging genetics (Figure 1A) to explore the complex, and testable, interplay between the genome, brain, body, and experience (Figure 1B). Presently, the most prudent manner to begin testing these pathways is through multimodal data convergence – imaging genetics is but one crucial component in the elaborate and multifaceted puzzle surrounding the interface between brain and behavior - integrating across various lines of evidence is likely to provide the most complete picture.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

RB is supported by the Klingenstein Third Generation Foundation and R01-AG045231, R01-HD083614, U01-AG052564. CEC is supported by the National Science Foundation (DGE-1143954). AA is supported by K02DA032573. VDC was supported by NIH P20GM103472, R01EB006841, R01EB005846; NSF 1539067. ARH is supported by grants R01-DA033369 and R01-AG049789 from the National Institutes of Health.

References

1. Bogdan R, Hyde LW, Hariri AR. A neurogenetics approach to understanding individual differences in brain, behavior, and risk for psychopathology. *Molecular psychiatry*. 2013; 18:288–299. [PubMed: 22614291]
2. Pohjalainen T, Rinne JO, Nagren K, Lehtikainen P, Anttila K, Syvalahti EK, et al. The A1 allele of the human D2 dopamine receptor gene predicts low D2 receptor availability in healthy volunteers. *Molecular psychiatry*. 1998; 3:256–260. [PubMed: 9672901]
3. Laruelle M, Gelernter J, Innis RB. D2 receptors binding potential is not affected by Taq1 polymorphism at the D2 receptor gene. *Molecular psychiatry*. 1998; 3:261–265. [PubMed: 9672902]
4. Heinz A, Jones DW, Mazzanti C, Goldman D, Ragan P, Hommer D, et al. A relationship between serotonin transporter genotype and in vivo protein expression and alcohol neurotoxicity. *Biological psychiatry*. 2000; 47:643–649. [PubMed: 10745057]
5. Heinz A, Goldman D, Jones DW, Palmour R, Hommer D, Gorey JG, et al. Genotype influences in vivo dopamine transporter availability in human striatum. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2000; 22:133–139. [PubMed: 10649826]
6. Heinz A, Goldman D. Genotype effects on neurodegeneration and neuroadaptation in monoaminergic neurotransmitter systems. *Neurochemistry international*. 2000; 37:425–432. [PubMed: 10871694]
7. Carey CE, Agrawal A, Zhang B, Conley ED, Degenhardt L, Heath AC, et al. Monoacylglycerol lipase (MGLL) polymorphism rs604300 interacts with childhood adversity to predict cannabis dependence symptoms and amygdala habituation: Evidence from an endocannabinoid system-level analysis. *Journal of abnormal psychology*. 2015; 124:860–877. [PubMed: 26595473]

8. Arloth J, Bogdan R, Weber P, Frishman G, Menke A, Wagner KV, et al. Genetic Differences in the Immediate Transcriptome Response to Stress Predict Risk-Related Brain Function and Psychiatric Disorders. *Neuron*. 2015; 86:1189–1202. [PubMed: 26050039]
9. Franke B, Stein JL, Ripke S, Anttila V, Hibar DP, van Hulzen KJ, et al. Genetic influences on schizophrenia and subcortical brain volumes: large-scale proof of concept. *Nature neuroscience*. 2016; 19:420–431. [PubMed: 26854805]
10. Gunduz-Cinar O, MacPherson KP, Cinar R, Gamble-George J, Sugden K, Williams B, et al. Convergent translational evidence of a role for anandamide in amygdala-mediated fear extinction, threat processing and stress-reactivity. *Molecular psychiatry*. 2013; 18:813–823. [PubMed: 22688188]
11. Pena-Oliver Y, Carvalho FM, Sanchez-Roige S, Quinlan EB, Jia T, Walker-Tilley T, et al. Mouse and Human Genetic Analyses Associate Kalirin with Ventral Striatal Activation during Impulsivity and with Alcohol Misuse. *Frontiers in genetics*. 2016; 7:52. [PubMed: 27092175]
12. Wellman CL, Camp M, Jones VM, MacPherson KP, Ihne J, Fitzgerald P, et al. Convergent effects of mouse Pet-1 deletion and human PET-1 variation on amygdala fear and threat processing. *Experimental neurology*. 2013; 250:260–269. [PubMed: 24100022]
13. Dincheva I, Drysdale AT, Hartley CA, Johnson DC, Jing D, King EC, et al. FAAH genetic variation enhances fronto-amygdala function in mouse and human. *Nature communications*. 2015; 6:6395.
14. Nikolova YS, Koenen KC, Galea S, Wang CM, Seney ML, Sibille E, et al. Beyond genotype: serotonin transporter epigenetic modification predicts human brain function. *Nature neuroscience*. 2014; 17:1153–1155. [PubMed: 25086606]
15. Ursini G, Bollati V, Fazio L, Porcelli A, Iacovelli L, Catalani A, et al. Stress-related methylation of the catechol-O-methyltransferase Val 158 allele predicts human prefrontal cognition and activity. *The Journal of neuroscience : the official journal of the Society for neuroscience*. 2011; 31:6692–6698. [PubMed: 21543598]
16. Hibar DP, Stein JL, Renteria ME, Arias-Vasquez A, Desrivieres S, Jahanshad N, et al. Common genetic variants influence human subcortical brain structures. *Nature*. 2015; 520:224–229. [PubMed: 25607358]
17. Potkin SG, Turner JA, Guffanti G, Lakatos A, Fallon JH, Nguyen DD, et al. A genome-wide association study of schizophrenia using brain activation as a quantitative phenotype. *Schizophrenia bulletin*. 2009; 35:96–108. [PubMed: 19023125]
18. Holmes AJ, Lee PH, Hollinshead MO, Bakst L, Roffman JL, Smoller JW, et al. Individual differences in amygdala-medial prefrontal anatomy link negative affect, impaired social functioning, and polygenic depression risk. *The Journal of neuroscience : the official journal of the Society for neuroscience*. 2012; 32:18087–18100. [PubMed: 23238724]
19. Terwisscha van Scheltinga AF, Bakker SC, van Haren NE, Derks EM, Buizer-Voskamp JE, Boos HB, et al. Genetic schizophrenia risk variants jointly modulate total brain and white matter volume. *Biological psychiatry*. 2013; 73:525–531. [PubMed: 23039932]
20. Bogdan R, Pagliaccio D, Baranger DA, Hariri AR. Genetic Moderation of Stress Effects on Corticolimbic Circuitry. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2016; 41:275–296. [PubMed: 26189450]
21. Thompson PM, Stein JL, Medland SE, Hibar DP, Vasquez AA, Renteria ME, et al. The ENIGMA Consortium: large-scale collaborative analyses of neuroimaging and genetic data. *Brain imaging and behavior*. 2014; 8:153–182. [PubMed: 24399358]
22. Swartz JR, Waller R, Bogdan R, Knodt AR, Sabhlok A, Hyde LW, et al. A Common Polymorphism in a Williams Syndrome Gene Predicts Amygdala Reactivity and Extraversion in Healthy Adults. *Biological psychiatry*. 2015
23. Holmes AJ, Hollinshead MO, O'Keefe TM, Petrov VI, Fariello GR, Wald LL, et al. Brain Genomics Superstruct Project initial data release with structural, functional, and behavioral measures. *Scientific data*. 2015; 2:150031. [PubMed: 26175908]
24. Schumann G, Loth E, Banaschewski T, Barbot A, Barker G, Buchel C, et al. The IMAGEN study: reinforcement-related behaviour in normal brain function and psychopathology. *Molecular psychiatry*. 2010; 15:1128–1139. [PubMed: 21102431]

25. Satterthwaite TD, Elliott MA, Ruparel K, Loughhead J, Prabhakaran K, Calkins ME, et al. Neuroimaging of the Philadelphia neurodevelopmental cohort. *Neuro Image*. 2014; 86:544–553. [PubMed: 23921101]
26. Barch DM, Burgess GC, Harms MP, Petersen SE, Schlaggar BL, Corbetta M, et al. Function in the human connectome: task-fMRI and individual differences in behavior. *Neuro Image*. 2013; 80:169–189. [PubMed: 23684877]
27. Inkster B, Nichols TE, Saemann PG, Auer DP, Holsboer F, Muglia P, et al. Pathway-based approaches to imaging genetics association studies: Wnt signaling, GSK3beta substrates and major depression. *Neuro Image*. 2010; 53:908–917. [PubMed: 20219685]
28. Nikolova YS, Ferrell RE, Manuck SB, Hariri AR. Multilocus genetic profile for dopamine signaling predicts ventral striatum reactivity. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2011; 36:1940–1947. [PubMed: 21593733]
29. Pearlson GD, Liu J, Calhoun VD. An introductory review of parallel independent component analysis (p-ICA) and a guide to applying p-ICA to genetic data and imaging phenotypes to identify disease-associated biological pathways and systems in common complex disorders. *Frontiers in genetics*. 2015; 6:276. [PubMed: 26442095]
30. Liu J, Calhoun VD. A review of multivariate analyses in imaging genetics. *Frontiers in neuroinformatics*. 2014; 8:29. [PubMed: 24723883]
31. Meyer-Lindenberg A, Weinberger DR. Intermediate phenotypes and genetic mechanisms of psychiatric disorders. *Nature reviews neuroscience*. 2006; 7:818–827. [PubMed: 16988657]
32. Gottesman II, Gould TD. The endophenotype concept in psychiatry: etymology and strategic intentions. *The American journal of psychiatry*. 2003; 160:636–645. [PubMed: 12668349]
33. CONVERGE, c. Sparse whole-genome sequencing identifies two loci for major depressive disorder. *Nature*. 2015; 523:588–591. [PubMed: 26176920]
34. Flint J, Kendler KS. The genetics of major depression. *Neuron*. 2014; 81:484–503. [PubMed: 24507187]
35. Treadway MT, Zald DH. Reconsidering anhedonia in depression: lessons from translational neuroscience. *Neuroscience and biobehavioral reviews*. 2011; 35:537–555. [PubMed: 20603146]
36. Bogdan R, Nikolova YS, Pizzagalli DA. Neurogenetics of depression: a focus on reward processing and stress sensitivity. *Neurobiology of disease*. 2013; 52:12–23. [PubMed: 22659304]
37. Wacker J, Dillon DG, Pizzagalli DA. The role of the nucleus accumbens and rostral anterior cingulate cortex in anhedonia: integration of resting EEG, fMRI, and volumetric techniques. *Neuro Image*. 2009; 46:327–337. [PubMed: 19457367]
38. Pizzagalli DA, Holmes AJ, Dillon DG, Goetz EL, Birk JL, Bogdan R, et al. Reduced caudate and nucleus accumbens response to rewards in unmedicated individuals with major depressive disorder. *The American journal of psychiatry*. 2009; 166:702–710. [PubMed: 19411368]
39. Arrondo G, Segarra N, Metastasio A, Ziauddeen H, Spencer J, Reinders NR, et al. Reduction in ventral striatal activity when anticipating a reward in depression and schizophrenia: a replicated cross-diagnostic finding. *Front Psychol*. 2015; 6:1280. [PubMed: 26379600]
40. Knutson B, Bhanji JP, Cooney RE, Atlas LY, Gotlib IH. Neural responses to monetary incentives in major depression. *Biological psychiatry*. 2008; 63:686–692. [PubMed: 17916330]
41. Schizophrenia Working Group of the Psychiatric Genomics C. Biological insights from 108 schizophrenia-associated genetic loci. *Nature*. 2014; 511:421–427. [PubMed: 25056061]
42. Shiwach R. Psychopathology in Huntington's disease patients. *Acta psychiatrica Scandinavica*. 1994; 90:241–246. [PubMed: 7831992]
43. Schneider M, Debbane M, Bassett AS, Chow EW, Fung WL, van den Bree M, et al. Psychiatric disorders from childhood to adulthood in 22q11.2 deletion syndrome: results from the International Consortium on Brain and Behavior in 22q11.2 Deletion Syndrome. *The American journal of psychiatry*. 2014; 171:627–639. [PubMed: 24577245]
44. Meyer-Lindenberg A. Intermediate or brainless phenotypes for psychiatric research? *Psychological medicine*. 2010; 40:1057–1062. [PubMed: 20540175]
45. Hyde CL, Nagle MW, Tian C, Chen X, Paciga SA, Wendland JR, et al. Identification of 15 genetic loci associated with risk of major depression in individuals of European descent. *Nature genetics*. 2016; 48:1031–1036. [PubMed: 27479909]

46. Jansen AG, Mous SE, White T, Posthuma D, Polderman TJ. What twin studies tell us about the heritability of brain development, morphology, and function: a review. *Neuropsychology review*. 2015; 25:27–46. [PubMed: 25672928]
47. Polderman TJ, Benyamin B, de Leeuw CA, Sullivan PF, van Bochoven A, Visscher PM, et al. Meta-analysis of the heritability of human traits based on fifty years of twin studies. *Nature genetics*. 2015; 47:702–709. [PubMed: 25985137]
48. Pober BR. Williams-Beuren syndrome. *The New England journal of medicine*. 2010; 362:239–252. [PubMed: 20089974]
49. Barak B, Feng G. Neurobiology of social behavior abnormalities in autism and Williams syndrome. *Nature neuroscience*. 2016; 19:647–655. [PubMed: 27116389]
50. Crespi BJ, Hurd PL. Cognitive-behavioral phenotypes of Williams syndrome are associated with genetic variation in the GTF2I gene, in a healthy population. *BMC neuroscience*. 2014; 15:127. [PubMed: 25429715]
51. Klengel T, Mehta D, Anacker C, Rex-Haffner M, Pruessner JC, Pariante CM, et al. Allele-specific FKBP5 DNA demethylation mediates gene-childhood trauma interactions. *Nature neuroscience*. 2013; 16:33–41. [PubMed: 23201972]
52. White MG, Bogdan R, Fisher PM, Munoz KE, Williamson DE, Hariri AR. FKBP5 and emotional neglect interact to predict individual differences in amygdala reactivity. *Genes, brain, and behavior*. 2012; 11:869–878.
53. Nikolova YS, Hariri AR. Can we observe epigenetic effects on human brain function? *Trends in cognitive sciences*. 2015; 19:366–373. [PubMed: 26051383]
54. Liu J, Siyahhan Julnes P, Chen J, Ehrlich S, Walton E, Calhoun VD. The association of DNA methylation and brain volume in healthy individuals and schizophrenia patients. *Schizophrenia research*. 2015; 169:447–452. [PubMed: 26381449]
55. Walton E, Hass J, Liu J, Roffman JL, Bernardoni F, Roessner V, et al. Correspondence of DNA Methylation Between Blood and Brain Tissue and Its Application to Schizophrenia Research. *Schizophrenia bulletin*. 2016; 42:406–414. [PubMed: 26056378]
56. Egan MF, Goldberg TE, Kolachana BS, Callicott JH, Mazzanti CM, Straub RE, et al. Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. *Proceedings of the National Academy of Sciences of the United States of America*. 2001; 98:6917–6922. [PubMed: 11381111]
57. Hariri AR, Mattay VS, Tessitore A, Kolachana B, Fera F, Goldman D, et al. Serotonin transporter genetic variation and the response of the human amygdala. *Science*. 2002; 297:400–403. [PubMed: 12130784]
58. Nickl-Jockschat T, Janouschek H, Eickhoff SB, Eickhoff CR. Lack of meta-analytic evidence for an impact of COMT Val158Met genotype on brain activation during working memory tasks. *Biological psychiatry*. 2015; 78:e43–e46. [PubMed: 25861704]
59. Gonzalez-Castro TB, Hernandez-Diaz Y, Juarez-Rojop IE, Lopez-Narvaez ML, Tovilla-Zarate CA, Fresan A. The Role of a Catechol-O-Methyltransferase (COMT) Val158Met Genetic Polymorphism in Schizophrenia: A Systematic Review and Updated Meta-analysis on 32,816 Subjects. *Neuromolecular medicine*. 2016
60. Duncan LE, Keller MC. A critical review of the first 10 years of candidate gene-by-environment interaction research in psychiatry. *The American journal of psychiatry*. 2011; 168:1041–1049. [PubMed: 21890791]
61. Karg K, Burmeister M, Shedden K, Sen S. The serotonin transporter promoter variant (5-HTTLPR), stress, and depression meta-analysis revisited: evidence of genetic moderation. *Archives of general psychiatry*. 2011; 68:444–454. [PubMed: 21199959]
62. Libert S, Pointer K, Bell EL, Das A, Cohen DE, Asara JM, et al. SIRT1 activates MAO-A in the brain to mediate anxiety and exploratory drive. *Cell*. 2011; 147:1459–1472. [PubMed: 22169038]
63. Kishi T, Yoshimura R, Kitajima T, Okochi T, Okumura T, Tsunoka T, et al. SIRT1 gene is associated with major depressive disorder in the Japanese population. *Journal of affective disorders*. 2010; 126:167–173. [PubMed: 20451257]
64. Zhang Q, Shen Q, Xu Z, Chen M, Cheng L, Zhai J, et al. The effects of CACNA1C gene polymorphism on spatial working memory in both healthy controls and patients with

- schizophrenia or bipolar disorder. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2012; 37:677–684. [PubMed: 22012475]
65. Paulus FM, Bedenbender J, Krach S, Pyka M, Krug A, Sommer J, et al. Association of rs1006737 in CACNA1C with alterations in prefrontal activation and fronto-hippocampal connectivity. *Human brain mapping*. 2014; 35:1190–1200. [PubMed: 23404764]
 66. Bigos KL, Mattay VS, Callicott JH, Straub RE, Vakkalanka R, Kolachana B, et al. Genetic variation in CACNA1C affects brain circuitries related to mental illness. *Archives of general psychiatry*. 2010; 67:939–945. [PubMed: 20819988]
 67. Cousijn H, Eissing M, Fernandez G, Fisher SE, Franke B, Zwiers M, et al. No effect of schizophrenia risk genes MIR137, TCF4, and ZNF804A on macroscopic brain structure. *Schizophrenia research*. 2014; 159:329–332. [PubMed: 25217366]
 68. Murphy SE, Norbury R, Godlewska BR, Cowen PJ, Mannie ZM, Harmer CJ, et al. The effect of the serotonin transporter polymorphism (5-HTTLPR) on amygdala function: a meta-analysis. *Molecular psychiatry*. 2013; 18:512–520. [PubMed: 22488255]
 69. Bastiaansen JA, Servaas MN, Marsman JB, Ormel J, Nolte IM, Riese H, et al. Filling the gap: relationship between the serotonin-transporter-linked polymorphic region and amygdala activation. *Psychological science*. 2014; 25:2058–2066. [PubMed: 25253281]
 70. Bell JA, Kivimaki M, Hamer M. Metabolically healthy obesity and risk of incident type 2 diabetes: a meta-analysis of prospective cohort studies. *Obesity reviews : an official journal of the International Association for the Study of Obesity*. 2014; 15:504–515. [PubMed: 24661566]
 71. Hadjikhani N, Joseph RM, Snyder J, Tager-Flusberg H. Abnormal activation of the social brain during face perception in autism. *Human brain mapping*. 2007; 28:441–449. [PubMed: 17133386]
 72. Baron-Cohen S, Ring HA, Wheelwright S, Bullmore ET, Brammer MJ, Simmons A, et al. Social intelligence in the normal and autistic brain: an fMRI study. *The European journal of neuroscience*. 1999; 11:1891–1898. [PubMed: 10336657]
 73. Jones W, Klin A. Attention to eyes is present but in decline in 2–6-month-old infants later diagnosed with autism. *Nature*. 2013; 504:427–431. [PubMed: 24196715]
 74. Whalen PJ, Kagan J, Cook RG, Davis FC, Kim H, Polis S, et al. Human amygdala responsivity to masked fearful eye whites. *Science*. 2004; 306:2061. [PubMed: 15604401]
 75. Tottenham N, Hertzog ME, Gillespie-Lynch K, Gilhooly T, Millner AJ, Casey BJ. Elevated amygdala response to faces and gaze aversion in autism spectrum disorder. *Social cognitive and affective neuroscience*. 2014; 9:106–117. [PubMed: 23596190]
 76. Dalton KM, Nacewicz BM, Alexander AL, Davidson RJ. Gaze-fixation, brain activation, and amygdala volume in unaffected siblings of individuals with autism. *Biological psychiatry*. 2007; 61:512–520. [PubMed: 17069771]
 77. Dalton KM, Nacewicz BM, Johnstone T, Schaefer HS, Gernsbacher MA, Goldsmith HH, et al. Gaze fixation and the neural circuitry of face processing in autism. *Nature neuroscience*. 2005; 8:519–526. [PubMed: 15750588]
 78. Sugranyes G, Kyriakopoulos M, Corrigall R, Taylor E, Frangou S. Autism spectrum disorders and schizophrenia: meta-analysis of the neural correlates of social cognition. *PloS one*. 2011; 6:e25322. [PubMed: 21998649]
 79. Dannlowski U, Konrad C, Kugel H, Zwieterlood P, Domschke K, Schoning S, et al. Emotion specific modulation of automatic amygdala responses by 5-HTTLPR genotype. *Neuro Image*. 2010; 53:893–898. [PubMed: 19962442]
 80. Lee BT, Ham BJ. Serotonergic genes and amygdala activity in response to negative affective facial stimuli in Korean women. *Genes, brain, and behavior*. 2008; 7:899–905.
 81. Flint J, Munafò MR. Candidate and non-candidate genes in behavior genetics. *Current opinion in neurobiology*. 2013; 23:57–61. [PubMed: 22878161]
 82. Cross-Disorder Group of the Psychiatric Genomics C. Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *Lancet*. 2013; 381:1371–1379. [PubMed: 23453885]
 83. Sekar A, Bialas AR, de Rivera H, Davis A, Hammond TR, Kamitaki N, et al. Schizophrenia risk from complex variation of complement component 4. *Nature*. 2016; 530:177–183. [PubMed: 26814963]

84. Caspi A, Hariri AR, Holmes A, Uher R, Moffitt TE. Genetic sensitivity to the environment: the case of the serotonin transporter gene and its implications for studying complex diseases and traits. *The American journal of psychiatry*. 2010; 167:509–527. [PubMed: 20231323]
85. Lowry JP, Griffin K, McHugh SB, Lowe AS, Tricklebank M, Sibson NR. Real-time electrochemical monitoring of brain tissue oxygen: a surrogate for functional magnetic resonance imaging in rodents. *Neuro Image*. 2010; 52:549–555. [PubMed: 20417284]
86. Barkus C, Line SJ, Huber A, Captao L, Lima J, Jennings K, et al. Variation in serotonin transporter expression modulates fear-evoked hemodynamic responses and theta-frequency neuronal oscillations in the amygdala. *Biological psychiatry*. 2014; 75:901–908. [PubMed: 24120093]
87. Bocchio M, McHugh SB, Bannerman DM, Sharp T, Capogna M. Serotonin, Amygdala and Fear: Assembling the Puzzle. *Frontiers in neural circuits*. 2016; 10:24. [PubMed: 27092057]
88. Ousdal OT, Anand Brown A, Jensen J, Nakstad PH, Melle I, Agartz I, et al. Associations between variants near a monoaminergic pathways gene (PHOX2B) and amygdala reactivity: a genome-wide functional imaging study. *Twin research and human genetics : the official journal of the International Society for Twin Studies*. 2012; 15:273–285. [PubMed: 22856363]
89. Stein JL, Medland SE, Vasquez AA, Hibar DP, Senstad RE, Winkler AM, et al. Identification of common variants associated with human hippocampal and intracranial volumes. *Nature genetics*. 2012; 44:552–561. [PubMed: 22504417]
90. Bis JC, DeCarli C, Smith AV, van der Lijn F, Crivello F, Fornage M, et al. Common variants at 12q14 and 12q24 are associated with hippocampal volume. *Nature genetics*. 2012; 44:545–551. [PubMed: 22504421]
91. Dannlowski U, Grabe HJ, Wittfeld K, Klaus J, Konrad C, Grotegerd D, et al. Multimodal imaging of a tescalcin (TESC)-regulating polymorphism (rs7294919)-specific effects on hippocampal gray matter structure. *Molecular psychiatry*. 2015; 20:398–404. [PubMed: 24776739]
92. Regier DA, Narrow WE, Clarke DE, Kraemer HC, Kuramoto SJ, Kuhl EA, et al. DSM-5 field trials in the United States and Canada, Part II: test-retest reliability of selected categorical diagnoses. *The American journal of psychiatry*. 2013; 170:59–70. [PubMed: 23111466]
93. Schnack HG, van Haren NE, Hulshoff Pol HE, Picchioni M, Weisbrod M, Sauer H, et al. Reliability of brain volumes from multicenter MRI acquisition: a calibration study. *Human brain mapping*. 2004; 22:312–320. [PubMed: 15202109]
94. Holmes AJ, Hollinshead MO, Roffman JL, Smoller JW, Buckner RL. Individual Differences in Cognitive Control Circuit Anatomy Link Sensation Seeking, Impulsivity, and Substance Use. *The Journal of neuroscience : the official journal of the Society for neuroscience*. 2016; 36:4038–4049. [PubMed: 27053210]
95. Manuck SB, Brown SM, Forbes EE, Hariri AR. Temporal stability of individual differences in amygdala reactivity. *The American journal of psychiatry*. 2007; 164:1613–1614. [PubMed: 17898358]
96. Sauder CL, Hajcak G, Angstadt M, Phan KL. Test-retest reliability of amygdala response to emotional faces. *Psychophysiology*. 2013; 50:1147–1156. [PubMed: 24128307]
97. Plichta MM, Schwarz AJ, Grimm O, Morgen K, Mier D, Haddad L, et al. Test-retest reliability of evoked BOLD signals from a cognitive-emotive fMRI test battery. *Neuro Image*. 2012; 60:1746–1758. [PubMed: 22330316]
98. Plichta MM, Grimm O, Morgen K, Mier D, Sauer C, Haddad L, et al. Amygdala habituation: a reliable fMRI phenotype. *Neuro Image*. 2014; 103:383–390. [PubMed: 25284303]
99. Bennett CM, Miller MB. How reliable are the results from functional magnetic resonance imaging? *Annals of the New York Academy of Sciences*. 2010; 1191:133–155. [PubMed: 20392279]
100. Flint J, Timpson N, Munafò M. Assessing the utility of intermediate phenotypes for genetic mapping of psychiatric disease. *Trends in neurosciences*. 2014; 37:733–741. [PubMed: 25216981]
101. Gratten J, Wray NR, Keller MC, Visscher PM. Large-scale genomics unveils the genetic architecture of psychiatric disorders. *Nature neuroscience*. 2014; 17:782–790. [PubMed: 24866044]

102. Okbay A, Baselmans BM, De Neve JE, Turley P, Nivard MG, Fontana MA, et al. Genetic variants associated with subjective well-being, depressive symptoms, and neuroticism identified through genome-wide analyses. *Nature genetics*. 2016
103. MATHALON DH, SULLIVAN EV, LIM KO, PFEFFERBAUM A. Progressive brain volume changes and the clinical course of schizophrenia in men: a longitudinal magnetic resonance imaging study. *Archives of general psychiatry*. 2001; 58:148–157. [PubMed: 11177116]
104. Tost H, Braus DF, Hakimi S, Ruf M, Vollmert C, Hohn F, et al. Acute D2 receptor blockade induces rapid, reversible remodeling in human cortical-striatal circuits. *Nature neuroscience*. 2010; 13:920–922. [PubMed: 20526332]
105. Ho BC, Andreasen NC, Ziebell S, Pierson R, Magnotta V. Long-term antipsychotic treatment and brain volumes: a longitudinal study of first-episode schizophrenia. *Archives of general psychiatry*. 2011; 68:128–137. [PubMed: 21300943]
106. Davis J, Eyre H, Jacka FN, Dodd S, Dean O, McEwen S, et al. A review of vulnerability and risks for schizophrenia: Beyond the two hit hypothesis. *Neuroscience and biobehavioral reviews*. 2016; 65:185–194. [PubMed: 27073049]
107. Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, et al. Schizophrenia Working Group of the Psychiatric Genomics C. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nature genetics*. 2015; 47:291–295. [PubMed: 25642630]
108. Plomin R, Haworth CM, Davis OS. Common disorders are quantitative traits. *Nature reviews Genetics*. 2009; 10:872–878.
109. Phillips ML, Travis MJ, Fagiolini A, Kupfer DJ. Medication effects in neuroimaging studies of bipolar disorder. *The American journal of psychiatry*. 2008; 165:313–320. [PubMed: 18245175]
110. Vilhjalmsón BJ, Yang J, Finucane HK, Gusev A, Lindstrom S, Ripke S, et al. Modeling Linkage Disequilibrium Increases Accuracy of Polygenic Risk Scores. *American journal of human genetics*. 2015; 97:576–592. [PubMed: 26430803]
111. Hill WG, Goddard ME, Visscher PM. Data and theory point to mainly additive genetic variance for complex traits. *PLoS genetics*. 2008; 4:e1000008. [PubMed: 18454194]
112. Demers CH, Drabant Conley E, Bogdan R, Hariri AR. Interactions Between Anandamide and Corticotropin-Releasing Hormone Signaling Modulate Human Amygdala Function and Risk for Anxiety Disorders: An Imaging Genetics Strategy for Modeling Molecular Interactions. *Biological psychiatry*. 2016
113. Andreasen NC, Wilcox MA, Ho BC, Epping E, Ziebell S, Zeien E, et al. Statistical epistasis and progressive brain change in schizophrenia: an approach for examining the relationships between multiple genes. *Molecular psychiatry*. 2012; 17:1093–1102. [PubMed: 21876540]
114. Hibar DP, Stein JL, Jahanshad N, Kohannim O, Hua X, Toga AW, et al. Genome-wide interaction analysis reveals replicated epistatic effects on brain structure. *Neurobiology of aging*. 2015; 36(Suppl 1):S151–S158. [PubMed: 25264344]
115. Dudbridge F. Power and predictive accuracy of polygenic risk scores. *PLoS genetics*. 2013; 9:e1003348. [PubMed: 2355274]
116. Cong L, Ran FA, Cox D, Lin S, Barretto R, Habib N, et al. Multiplex genome engineering using CRISPR/Cas systems. *Science*. 2013; 339:819–823. [PubMed: 23287718]
117. Wang H, Yang H, Shivalila CS, Dawlaty MM, Cheng AW, Zhang F, et al. One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell*. 2013; 153:910–918. [PubMed: 23643243]
118. Deutch AY. The regulation of subcortical dopamine systems by the prefrontal cortex: interactions of central dopamine systems and the pathogenesis of schizophrenia. *Journal of neural transmission Supplementum*. 1992; 36:61–89. [PubMed: 1527521]
119. Bilder RM, Volavka J, Lachman HM, Grace AA. The catechol-O-methyltransferase polymorphism: relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2004; 29:1943–1961. [PubMed: 15305167]

120. Stice E, Yokum S, Burger K, Epstein L, Smolen A. Multilocus genetic composite reflecting dopamine signaling capacity predicts reward circuitry responsivity. *J Neurosci*. 2012; 32:10093–10100. [PubMed: 22815523]
121. Perlis RH, Moorjani P, Fagerness J, Purcell S, Trivedi MH, Fava M, et al. Pharmacogenetic analysis of genes implicated in rodent models of antidepressant response: association of TREK1 and treatment resistance in the STAR(*)D study. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2008; 33:2810–2819. [PubMed: 18288090]
122. Liu JZ, McRae AF, Nyholt DR, Medland SE, Wray NR, Brown KM, et al. A versatile gene-based test for genome-wide association studies. *American journal of human genetics*. 2010; 87:139–145. [PubMed: 20598278]
123. Mattingsdal M, Brown AA, Djurovic S, Sonderby IE, Server A, Melle I, et al. Pathway analysis of genetic markers associated with a functional MRI faces paradigm implicates polymorphisms in calcium responsive pathways. *Neuro Image*. 2013; 70:143–149. [PubMed: 23274185]
124. Kanehisa M, Sato Y, Kawashima M, Furumichi M, Tanabe M. KEGG as a reference resource for gene and protein annotation. *Nucleic acids research*. 2016; 44:D457–D462. [PubMed: 26476454]
125. Chi EC, Allen GI, Zhou H, Kohannim O, Lange K, Thompson PM. Imaging Genetics Via Sparse Canonical Correlation Analysis. *Proceedings / IEEE International Symposium on Biomedical Imaging: from nano to macro IEEE International Symposium on Biomedical Imaging*. 2013; 2013:740–743.
126. Le Floch E, Guillemot V, Frouin V, Pinel P, Lalanne C, Trinchera L, et al. Significant correlation between a set of genetic polymorphisms and a functional brain network revealed by feature selection and sparse Partial Least Squares. *Neuro Image*. 2012; 63:11–24. [PubMed: 22781162]
127. Vounou M, Nichols TE, Montana G, Alzheimer's Disease Neuroimaging I. Discovering genetic associations with high-dimensional neuroimaging phenotypes: A sparse reduced-rank regression approach. *Neuro Image*. 2010; 53:1147–1159. [PubMed: 20624472]
128. Chen J, Calhoun VD, Pearlson GD, Perrone-Bizzozero N, Sui J, Turner JA, et al. Guided exploration of genomic risk for gray matter abnormalities in schizophrenia using parallel independent component analysis with reference. *Neuro Image*. 2013; 83:384–396. [PubMed: 23727316]
129. Hinrichs C, Singh V, Xu G, Johnson SC, Alzheimers Disease Neuroimaging I. Predictive markers for AD in a multi-modality framework: an analysis of MCI progression in the ADNI population. *Neuro Image*. 2011; 55:574–589. [PubMed: 21146621]
130. Yang H, Liu J, Sui J, Pearlson G, Calhoun VD. A Hybrid Machine Learning Method for Fusing fMRI and Genetic Data: Combining both Improves Classification of Schizophrenia. *Frontiers in human neuroscience*. 2010; 4:192. [PubMed: 21119772]
131. Arnedo J, Svrakic DM, Del Val C, Romero-Zaliz R, Hernandez-Cuervo H, Molecular Genetics of Schizophrenia C, et al. Uncovering the hidden risk architecture of the schizophrenias: confirmation in three independent genome-wide association studies. *The American journal of psychiatry*. 2015; 172:139–153. [PubMed: 25219520]
132. Breen, G., Bulik-Sullivan, B., Daly, M., Medland, S., Neale, B., O'Donovan, M., et al. Untitled. *PubMed Commons*. 2014. http://www.ncbi.nlm.nih.gov.libproxy.wustl.edu/pubmed/25219520#cm25219520_25216388
133. Carter CS, Bearden CE, Bullmore E, Geschwind D, Glahn DC, Gur RC, et al. Enhancing the informativeness and replicability of imaging genomics studies. *Biological psychiatry*. (In Press).
134. Ioannidis JP. Why most published research findings are false. *PLoS medicine*. 2005; 2:e124. [PubMed: 16060722]
135. Agrawal A, Bogdan R. Risky Business: Pathways to Progress in Biologically Informed Studies of Psychopathology. *Psychological inquiry*. 2015; 26:231–238. [PubMed: 27114696]
136. de Vries YA, Roest AM, Franzen M, Munafo MR, Bastiaansen JA. Citation bias and selective focus on positive findings in the literature on the serotonin transporter gene (5-HTTLPR), life stress and depression. *Psychological medicine*. 2016; 46:2971–2979. [PubMed: 27515846]
137. Hardy J, Trabzuni D, Ryten M. Whole genome expression as a quantitative trait. *Biochem Soc Trans*. 2009; 37:1276–1277. [PubMed: 19909261]

138. Colantuoni C, Lipska BK, Ye T, Hyde TM, Tao R, Leek JT, et al. Temporal dynamics and genetic control of transcription in the human prefrontal cortex. *Nature*. 2011; 478:519–523. [PubMed: 22031444]
139. Consortium GT. The Genotype-Tissue Expression (GTEx) project. *Nature genetics*. 2013; 45:580–585. [PubMed: 23715323]
140. Zhou X, Maricque B, Xie M, Li D, Sundaram V, Martin EA, et al. The Human Epigenome Browser at Washington University. *Nat Methods*. 2011; 8:989–990. [PubMed: 22127213]
141. Gusev A, Ko A, Shi H, Bhatia G, Chung W, Penninx BW, et al. Integrative approaches for large-scale transcriptome-wide association studies. *Nature genetics*. 2016; 48:245–252. [PubMed: 26854917]
142. Gamazon ER, Wheeler HE, Shah KP, Mozaffari SV, Aquino-Michaels K, Carroll RJ, et al. A gene-based association method for mapping traits using reference transcriptome data. *Nature genetics*. 2015; 47:1091–1098. [PubMed: 26258848]
143. Bjornsson HT, Sigurdsson MI, Fallin MD, Irizarry RA, Aspelund T, Cui H, et al. Intra-individual change over time in DNA methylation with familial clustering. *Jama*. 2008; 299:2877–2883. [PubMed: 18577732]
144. Rask-Andersen M, Bringeland N, Nilsson EK, Bandstein M, Olaya Bucaro M, Vogel H, et al. Postprandial alterations in whole-blood DNA methylation are mediated by changes in white blood cell composition. *The American journal of clinical nutrition*. 2016; 104:518–525. [PubMed: 27385611]
145. Swartz JR, Hariri AR, Williamson DE. An epigenetic mechanism links socioeconomic status to changes in depression-related brain function in high-risk adolescents. *Molecular psychiatry*. 2016
146. Swartz JR, Knodt AR, Radtke SR, Hariri AR. A neural biomarker of psychological vulnerability to future life stress. *Neuron*. 2015; 85:505–511. [PubMed: 25654256]
147. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhman R, et al. Human genetics shape the gut microbiome. *Cell*. 2014; 159:789–799. [PubMed: 25417156]
148. Mayer EA, Knight R, Mazmanian SK, Cryan JF, Tillisch K. Gut microbes and the brain: paradigm shift in neuroscience. *The Journal of neuroscience : the official journal of the Society for neuroscience*. 2014; 34:15490–15496. [PubMed: 25392516]
149. Chiang KP, Gerber AL, Sipe JC, Cravatt BF. Reduced cellular expression and activity of the P129T mutant of human fatty acid amide hydrolase: evidence for a link between defects in the endocannabinoid system and problem drug use. *Human molecular genetics*. 2004; 13:2113–2119. [PubMed: 15254019]
150. Boileau I, Tyndale RF, Williams B, Mansouri E, Westwood DJ, Le Foll B, et al. The fatty acid amide hydrolase C385A variant affects brain binding of the positron emission tomography tracer [¹¹C]CURB. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*. 2015; 35:1237–1240.
151. Phan KL, Angstadt M, Golden J, Onyewuenyi I, Popovska A, de Wit H. Cannabinoid modulation of amygdala reactivity to social signals of threat in humans. *The Journal of neuroscience : the official journal of the Society for neuroscience*. 2008; 28:2313–2319. [PubMed: 18322078]
152. Hariri AR, Gorka A, Hyde LW, Kimak M, Halder I, Ducci F, et al. Divergent effects of genetic variation in endocannabinoid signaling on human threat- and reward-related brain function. *Biological psychiatry*. 2009; 66:9–16. [PubMed: 19103437]
153. Morena M, Patel S, Bains JS, Hill MN. Neurobiological Interactions Between Stress and the Endocannabinoid System. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2016; 41:80–102. [PubMed: 26068727]
154. Gunduz-Cinar O, Hill MN, McEwen BS, Holmes A. Amygdala FAAH and anandamide: mediating protection and recovery from stress. *Trends Pharmacol Sci*. 2013; 34:637–644. [PubMed: 24325918]

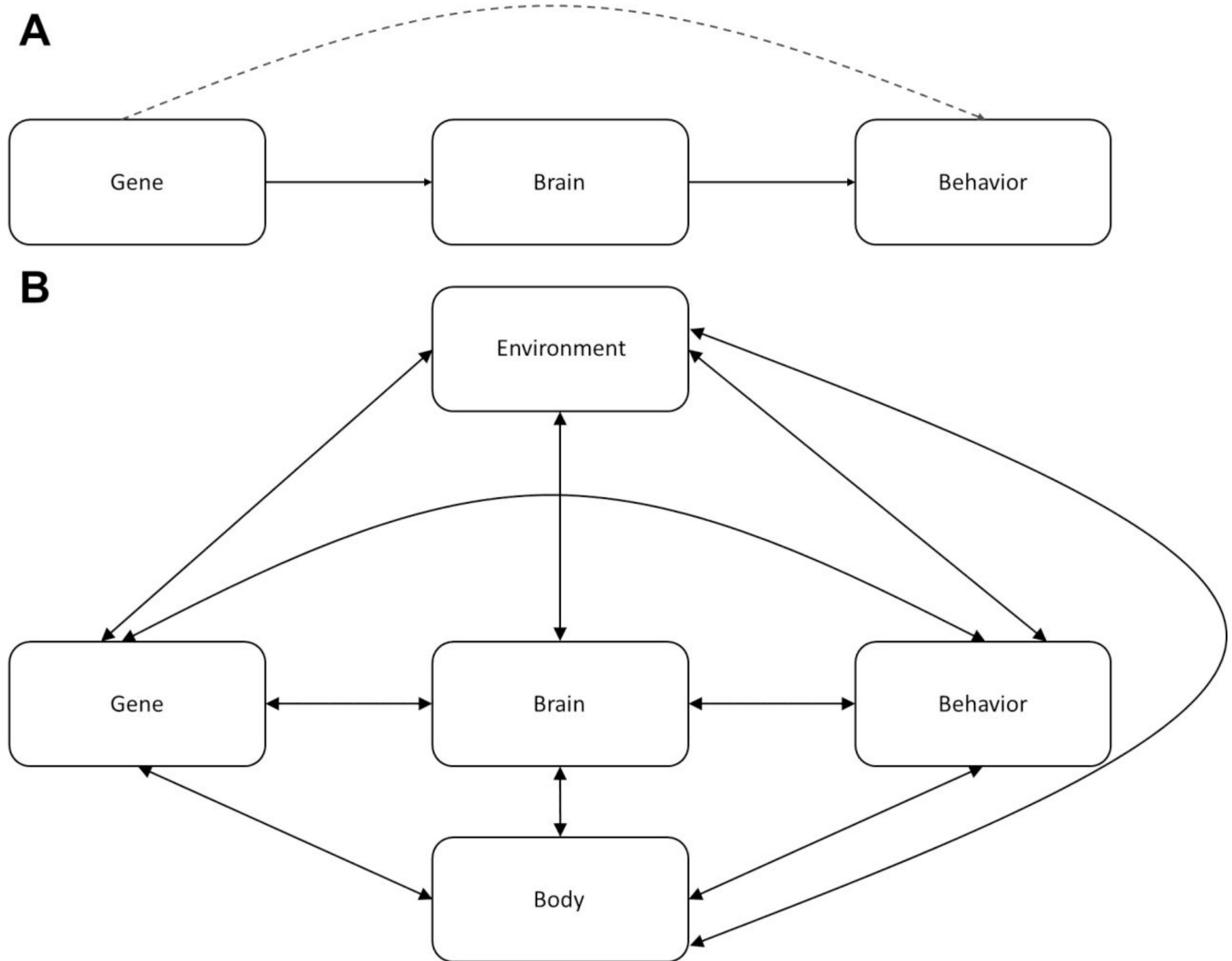


Figure 1. Imaging Genetics and Genomics Models

(A) The traditional imaging genetics and genomics model suggests that genetic variation confers risk for psychopathology indirectly through its influence on the brain. This theoretical model is well suited for traditional mediation models estimating indirect associations (demarcated with the dashed line), through which genetic background is linked to behavior through neural phenotypes. (B) *Imaging genetics and imaging genomics redux*: In the future, as imaging genetics and genomics expand to include larger and longitudinal samples it will be possible to evaluate a more complete interactive model in which bidirectional relationships between the genome, brain, and behavior may be investigated in the context of environmental experience and peripheral biological markers. For example, socioeconomic status has been associated with epigenetic modifications that are, in turn, related to psychiatrically-relevant brain function (145). Moreover, environmental experience (e.g., trauma experienced during early life) moderates genetic associations with neural phenotypes and associations between neural phenotypes and behavior (20,146). Further, genetic background influences peripheral indices such as gut microbiome (147), which in turn has been linked to neural phenotypes and psychopathology (148). As a result, a more

complete mechanistic understanding requires multiple levels of analyses in the context of longitudinal and convergent data. Currently, convergence across multiple methods and studies testing legs separately is attainable. Informed by such studies, in the future, as large multimodal longitudinal studies develop, it is plausible that more complete pathways could be tested in the framework of a single study.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 1Converging Evidence: The Example of Fatty Acid Amie Hydrolase (*FAAH*) rs324420 genotype (C/A; C385A)

Source of Evidence	Findings	Benefits	Limitations
<i>In Vitro</i> Function	A allele homozygosity is associated with less FAAH cellular expression in T-lymphocytes and transfected cells due to post-translation mechanism preceding folding (149).	Controlled functional characterization and isolation of step at which allelic variation impacts function	Unclear if similar function is observed in vivo amongst an interactive system
<i>In Vivo</i> Function	A allele carriers had lower [(11)C]CURB PET binding (FAAH binding) (150).	In vivo functional characterization	Often small samples, unclear links to behavior and other relevant phenotypes (e.g., brain function, structure)
Non-human Animal Manipulation	Knock-in mouse model: A allele associated with forebrain FAAH protein expression, hydrolytic activity, and elevated anandamide. A allele associated with increased projections from infralimbic to basolateral amygdala and enhanced fear extinction, and reduced anxiety (13).	Controlled manipulation of system using a variety of means (e.g., pharmacologic, genetic)	Unclear whether translates to humans and related conditions. Questionable phenotypic convergence across species for some phenotypes.
Human Manipulation (Pharmacologic Challenge)	Human: THC administration associated with reduced anxiety and threat-related amygdala reactivity (151).	Manipulation of a specific system allowing causal inferences to be drawn. For some substances, limitations on who can be exposed for human studies.	Temporary and chronic manipulation unclear translation to genetic risk. Uncertain whether artificial manipulations create other systematic changes.
Imaging Genetics and Genomics	A allele associated with decreased threat-related amygdala reactivity and increased amygdala habituation (152).	Provides a tractable and clinically-relevant phenotype. Offers system-level insight.	Molecular mechanisms of association unclear
Psychiatric/Behavioral Association (Candidate or GWAS)	A allele associated with enhanced fear extinction, reduced anxiety and stress sensitivity (10).	Provides clinical relevance	Unclear biological mechanisms
Treatment	Some evidence that FAAH inhibition improves anxiety in rodent models (153). Most common self-reported reason for using cannabis is anxiety reductions. THC administration reduces anxiety in clinical populations (154).	Evaluation of applicable therapeutic potential	Dependent upon other evidence, ability and safety to manipulate target. Lack of regional specificity in humans

The endocannabinoid system has been linked to stress recovery, anxiety, and substance use, across a host of models. Fatty Acid Amide Hydrolase (FAAH) is an enzymatic regulator of endocannabinoid signaling. Within the endocannabinoid system, it primarily degrades the endocannabinoid ligand anandamide.