

# SCIENTIFIC REPORTS



OPEN

## Transglycosylation by a chitinase from *Enterobacter cloacae* subsp. *cloacae* generates longer chitin oligosaccharides

Mohan Krishna Mallakuntla, Papa Rao Vaikuntapu, Bhoopal Bhuvanachandra, Subha Narayan Das & Appa Rao Podile

Humans have exploited natural resources for a variety of applications. Chitin and its derivative chitin oligosaccharides (CHOS) have potential biomedical and agricultural applications. Availability of CHOS with the desired length has been a major limitation in the optimum use of such natural resources. Here, we report a single domain hyper-transglycosylating chitinase, which generates longer CHOS, from *Enterobacter cloacae* subsp. *cloacae* 13047 (*EcChi1*). *EcChi1* was optimally active at pH 5.0 and 40 °C with a  $K_m$  of 15.2 mg ml<sup>-1</sup>, and  $k_{cat}/K_m$  of  $0.011 \times 10^2$  mg<sup>-1</sup> ml min<sup>-1</sup> on colloidal chitin. The profile of the hydrolytic products, major product being chitobiose, released from CHOS indicated that *EcChi1* was an endo-acting enzyme. Transglycosylation (TG) by *EcChi1* on trimeric to hexameric CHOS resulted in the formation of longer CHOS for a prolonged duration. *EcChi1* showed both chitobiase and TG activities, in addition to hydrolytic activity. The TG by *EcChi1* was dependent, to some extent, on the length of the CHOS substrate and concentration of the enzyme. Homology modeling and docking with CHOS suggested that *EcChi1* has a deep substrate-binding groove lined with aromatic amino acids, which is a characteristic feature of a processive enzyme.

Chitin [(C<sub>8</sub>H<sub>13</sub>O<sub>5</sub>N)<sub>n-1</sub>], is a linear homopolymer of *N*-acetyl glucosamine (GlcNAc) units linked through β (1 → 4) glycosidic bonds. It is an abundant renewable natural resource next to cellulose in the biosphere. Chitin is a primary structural component of the fungal cell wall, insects, molluscs, squid, internal shells of cephalopods and the exoskeletons of arthropod, exists in three biological forms i.e. α-chitin, β-chitin and γ-chitin. The presence of acetamide group (NH-CO-CH<sub>3</sub>) allows increased hydrogen bonding between adjacent polymers, giving increased strength of chitin – polymer matrix. Chitin oligosaccharides (CHOS) generated from polymeric chitin with specific composition and length have potential applications. The remarkable properties of CHOS i.e. bio-degradability, bio-compatibility, and non-toxicity suit wider industrial application<sup>1,2</sup>. Production of CHOS by chemical methods has been challenging due to disadvantages like non-specific random hydrolysis, and difficulty to remove the acidity of the CHOS oligomers<sup>3</sup>. Chitinases could be possible alternatives for production of long chain CHOS, especially for biological applications.

Chitinases (EC 3.2.1.14) hydrolyze chitin to chitin monomer (DP1; Degree of polymerization 1), chitin dimer (DP2) and CHOS of shorter length. Chitinases, classified into two glycoside hydrolase (GH) families i.e. GH18 and GH19 could be exochitinases that act terminally and endochitinases that cleave randomly at internal sites of the chitin, eventually producing a variety of low molecular mass or short length CHOS<sup>4</sup>. Chitinases that act on crystalline polysaccharides need to associate and disrupt the polymer from packing and also direct the travelling of single polymer chain into the catalytic center through a catalytic groove, a mechanism called processivity<sup>5,6</sup>. The aromatic amino acids lining the substrate-binding site, facilitate processivity by functioning as a flexible and hydrophobic sheet and allow the polymer chain to slide<sup>7,8</sup>.

A few of the GH18 chitinases exhibit an uncommon transglycosylation (TG), in addition to chitin hydrolysis, by introducing new glycosidic bonds between donor and acceptor saccharides<sup>9</sup>. The GH18 chitinases gained special interest due to potential applications for the enzymatic production of CHOS by TG from chitin<sup>10</sup>. Bacterial

Department of Plant Sciences, School of Life Sciences, University of Hyderabad, Gachibowli, Hyderabad, 50046, Telangana, India. Correspondence and requests for materials should be addressed to A.R.P. (email: [podilerao@uohyd.ac.in](mailto:podilerao@uohyd.ac.in))

chitinases like chitinase-D from *Serratia proteamaculans* (*SpChiD*) exhibited hyper-TG with chitotriose to chitohexaose (DP3–6) substrates generating products longer than chitohexaose (DP7–13)<sup>11</sup>. Similarly, chitinase A from *Stenotrophomonas maltophilia*<sup>12</sup> and ChiCW from *Bacillus cereus* 28–9 synthesized chitohexaose (DP6) from chitotetraose (DP4)<sup>13</sup>. A close relationship between bacterial chitinases and their pathogenicity may exist<sup>14,15</sup>, and the molecular targets of chitinases in chitinous or non-chitinous hosts remain unclear. Hence, detailed understanding of chitinases at the biochemical or functional level may help in understanding their role in pathogenicity.

*Enterobacter cloacae* subsp. *cloacae* (*Ec*), a nosocomial pathogen, was originally isolated from the human cerebrospinal fluid by Edwin Oakes Jordan in 1890<sup>16</sup>. Analysis of the genome sequence of *E. cloacae* subsp. *cloacae* revealed that four chitinases and two *N*-acetyl-glucosaminidases could be involved in chitin degradation. In addition, *E. cloacae* genome has genes that code for one CBM-33 lytic polysaccharide monooxygenase (LPMO-GenBank: ADF60226.1) and one polysaccharide deacetylase (PDA-GenBank: ADF62202.1), which may have a crucial role in depolymerization of chitin. Of the four chitinases, two belong to family GH18 (*EcChi1*-GenBank: ADF62010.1 and *EcChi2* (GenBank: ADF62328.1) and other two belong to GH19 (*EcChi3*-GenBank: ADF62326.1 and *EcChi4*-GenBank: ADF61237.1). Here, the *EcChi1* with only the catalytic GH18 domain and no auxiliary domains displayed an unprecedented hyper-TG. We show that *EcChi1* has structural features of a hyper-TG chitinase similar to *SpChiD* of *Serratia proteamaculans*.

## Results

**Cloning, heterologous expression, purification and dot blot assay.** Full-length DNA sequence coding for *EcChi1* consisted of 1257 nucleotides and an ORF of 418 amino acid residues with a predicted isoelectric point of 6.15. A 1.2-kb segment of *EcChi1* gene was amplified with specific primers using gDNA of *E. cloacae* subsp. *cloacae* as a template, cloned into pET-28a (+), and transformed *E. coli* Rosetta-gami for expression. Extracellular protein was isolated from the induced *E. coli* Rosetta-gami culture pellet and purified *EcChi1* through Ni-NTA affinity chromatography. The molecular mass of *EcChi1* (42.5 kDa) calculated from amino acid sequence without the signal peptide was in agreement with experimentally observed molecular weight as obtained by 12% SDS-PAGE (Supplementary Fig. S1A). Dot - blot assay revealed that, *EcChi1* exhibited clear zone on water-soluble glycol chitin containing gel confirming the chitinase activity (Supplementary Fig. S1B).

**Steady state kinetics of *EcChi1* on colloidal chitin and chitobiose.** The *EcChi1* was active in mildly acidic to a neutral range of pH 5.0 to 7.0 with the optimum activity in 50 mM sodium citrate pH 5.0 (Fig. 1A). However, the activity decreased sharply below pH 5.0 and above 7.0, with diminished or no activity below pH 4.0 and above 9.0. *EcChi1* was optimally active at 40 °C (Fig. 1B) and retained 60% activity at 50 °C, that was decreased with increase of temp up to 80 °C, while 70% activity was retained at 30 °C. The effect of metal ions on the activity of *EcChi1* was also evaluated using colloidal chitin as substrate. The activity of *EcChi1* in presence of Cu<sup>+2</sup> was similar to control but was partially inhibited in the presence of urea, Ca<sup>+2</sup>, Mg<sup>+2</sup>, Zn<sup>+2</sup>, Mn<sup>+2</sup>, EDTA, and Fe<sup>+2</sup>. The presence of Hg<sup>+2</sup> and SDS inhibited the activity by 80% and 90%, respectively. There was no increase in the activity of *EcChi1* in the presence of test cations. The kinetic parameters of *EcChi1* were determined with colloidal chitin and chitobiose as the substrate. The velocity measurements fitted to Michaelis-Menten kinetics revealed that the  $K_m$ ,  $k_{cat}$ , and  $k_{cat}/K_m$  values were 15.2 mg ml<sup>-1</sup>, 0.16 × 10<sup>2</sup> min<sup>-1</sup> and 0.011 × 10<sup>2</sup> mg<sup>-1</sup> ml min<sup>-1</sup> for colloidal chitin (Fig. 1C) and 213.2 μM, 1.41 min<sup>-1</sup> and 0.6 × 10<sup>-2</sup> for chitobiose (Supplementary Fig. S2).

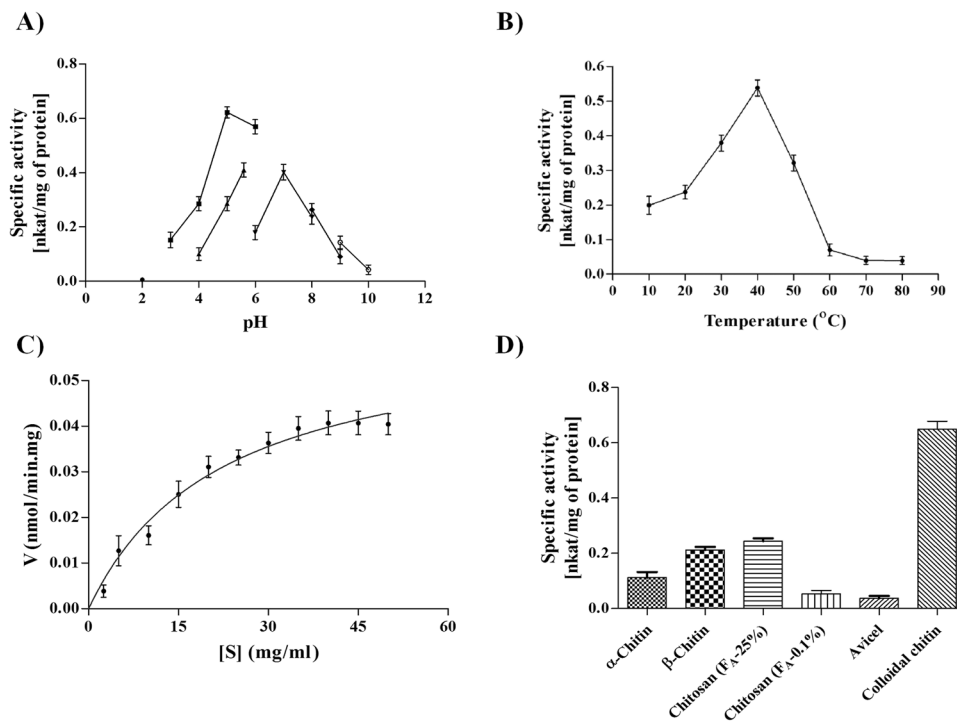
**Activity of *EcChi1* on different insoluble substrates.** *EcChi1* exhibited maximum activity on colloidal chitin followed by chitosan, β-chitin, α-chitin and avicel. The activity of *EcChi1* was 41% on chitosan ( $F_A$ -25%), 35% on β-chitin, 17% on α-chitin, 8% on chitosan ( $F_A$ -0.1%), and 5.7% on avicel.

**Binding study with soluble substrates.** Binding of *EcChi1* to the soluble substrates was analysed using native PAGE with or without polysaccharides embedded in the gel. In presence of laminarin, CM-cellulose and glycol chitin there was a decrease in electrophoretic mobility of *EcChi1* (Fig. 2). Among the three soluble substrates, the retardation in the mobility of *EcChi1* was more by soluble glycol chitin than CM-cellulose, and laminarin suggesting specificity of *EcChi1* towards chitinous substrates.

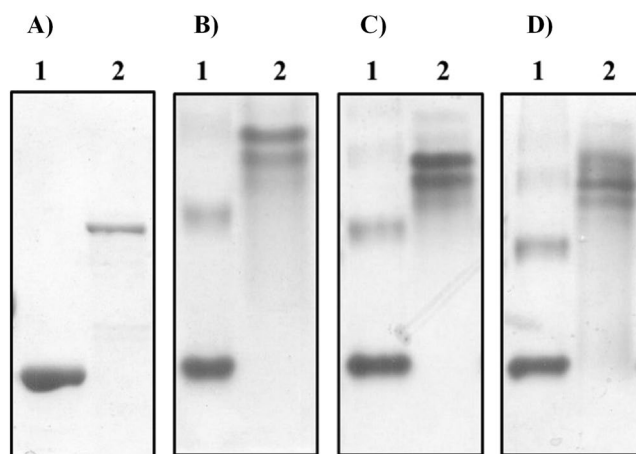
**Time-course of colloidal chitin and oligosaccharide degradation.** *EcChi1* released DP1 to DP4 hydrolytic products on polymeric colloidal chitin substrate with DP2 and DP1 as the major products (Fig. 3A and B). The concentration of DP1 increased with the extension of incubation time. *EcChi1* hydrolyzed DP2 into DP1 after 240 min, and complete hydrolysis occurred by 420 min (Supplementary Fig. S3). Chitobiose was the major hydrolytic product from both DP3 and DP4 substrate (Fig. 4A and B). With DP5 substrate, a very little DP1 (1.4%) formed from 120 min. After prolonged incubation, DP1 was the dominant product with 37.5% followed by DP2 with 28% at 720 min (Fig. 5A and C). The hydrolysis of DP6 by *EcChi1* was rapid with only 2.2% of DP6 at 120 min (Fig. 5B and D).

*EcChi1* exhibited high TG activity on chitotriose (DP3), chitotetraose (DP4), chitopentaose (DP5) and chitohexaose (DP6) substrates. With DP3 as starting substrate, TG products DP4–DP7 were detectable. DP4 and DP5 appeared from 1 min to 90 min, while DP6 and DP7 products appeared from 3 min to 60 min (Fig. 4A and C). Chitobiose was the major hydrolytic product from DP3 substrate. Use of DP4, DP5, and DP6 as starting substrates showed the formation of longer TG products (Figs 4B and 5A,B). With DP4 substrate, DP5–DP8 products formed due to TG, whereas DP9 was detectable as the longest TG product from DP5 and DP6 substrates (Figs 5B and 6). MALDI-TOF MS confirmed the products ≥DP6 of *EcChi1*. The products ranging from DP5 to DP9 and DP4 to DP9 were detectable with DP5 and DP6 substrates, respectively (Fig. 6A and B).

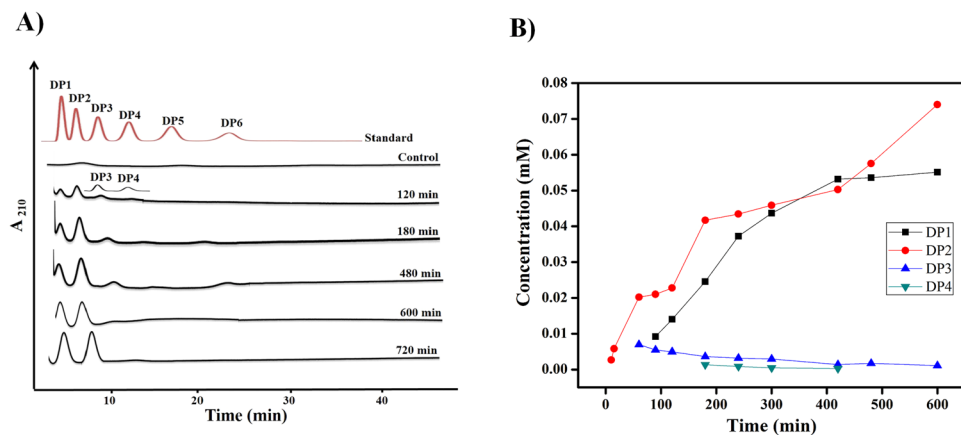
Reaction with DP4 substrate accumulated TG products from 1 min and lasted for 480 min, while DP8 product was detectable until 240 min. The TG products DP5 and DP6 reached a maximum quantity of 5.3% and 4.4% at 60



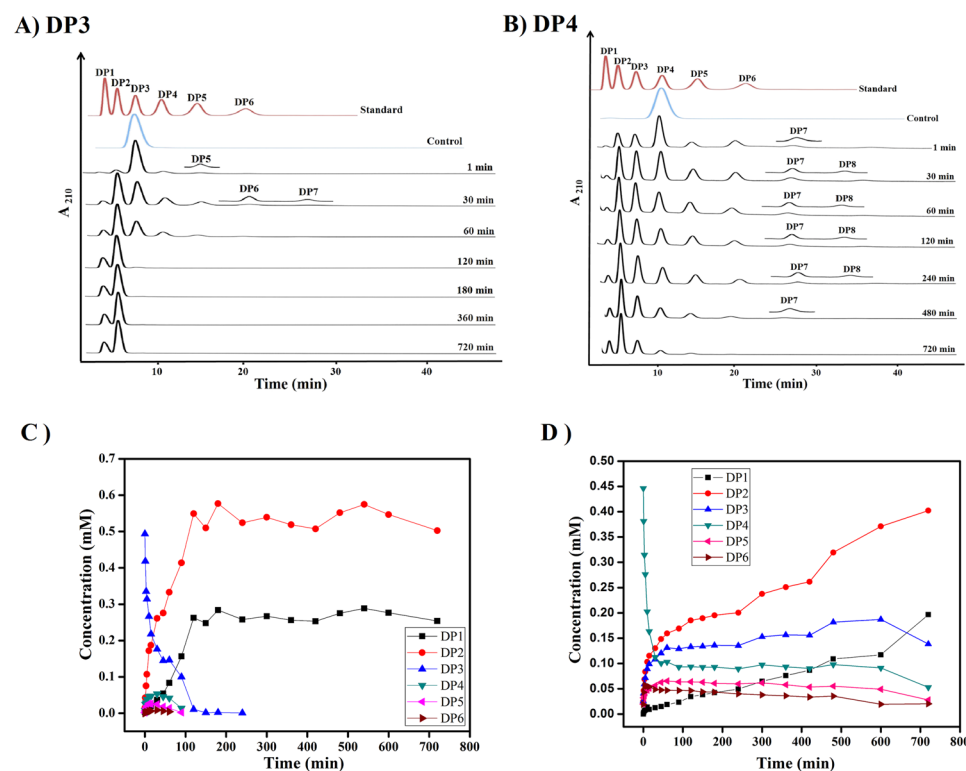
**Figure 1.** pH, temperature optima, kinetic parameters and activity on different insoluble substrates of *EcChi1*. The pH and temperature optima were determined by incubating 30 mg/ml colloidal chitin and 30  $\mu$ g of *EcChi1* for 1 h at 40 °C in various buffers at different pH values, as follows: (A) 50 mM glycine-HCl (pH 2.0,  $\blacktriangledown$ ), 50 mM sodium citrate (pH 3.0–6.0,  $\blacksquare$ ), 50 mM sodium acetate (pH 4.0–6.0,  $\blacktriangle$ ), 50 mM sodium phosphate (pH 6.0–8.00,  $\bullet$ ), 50 mM Tris-HCl (pH 8.0–9.0,  $\blacklozenge$ ) and 50 mM glycine-NaOH (pH 9.0–10.0,  $\circ$ ) and at 10 °C to 80 °C (B), Different concentrations of colloidal chitin (2.5–60 mg/ml) and 30  $\mu$ g of *EcChi1* in 50 mM sodium citrate buffer, pH 5.0. The average of the triplicate data was fitted to the Michaelis-Menten equation by a nonlinear regression function of graphpad prism version 6.0 (C), The reaction mixture containing 30  $\mu$ g *EcChi1* and 1 mg ml<sup>-1</sup> of one of the polymeric substrates, was incubated at pH 5.0 and 40 °C for 1 h. The reaction mixture was centrifuged, and products were quantified using a reducing group assay (D). The error bars represent the standard deviations from three individual experiments.



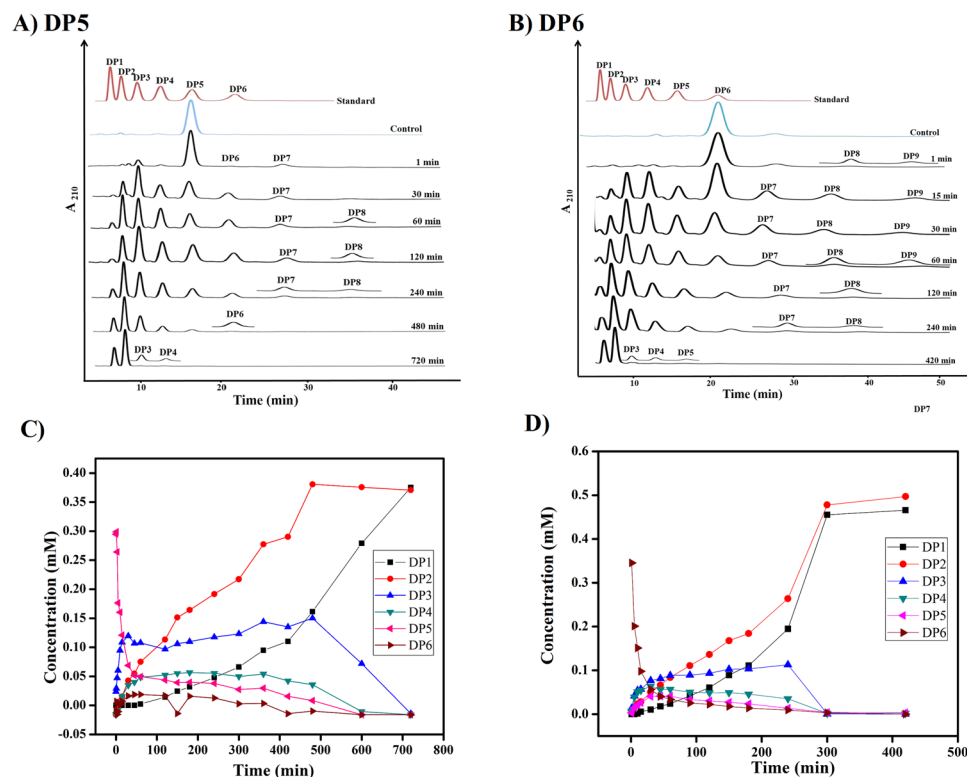
**Figure 2.** Binding of *EcChi1* with the soluble polymeric substrates. Affinity non-denaturing electrophoresis was performed in 8% native PAGE gels by preparing with or without substrate. (A) Proteins (*EcChi1* or non-interacting BSA) without substrate, while (B) with 0.1% (wt/vol) substrate glycol chitin, (C) CM-cellulose and (D) laminarin incorporated in gels. Proteins were visualized by Coomassie blue G-250 staining after electrophoresis. Lane 1. BSA, Lane 2. *EcChi1*.



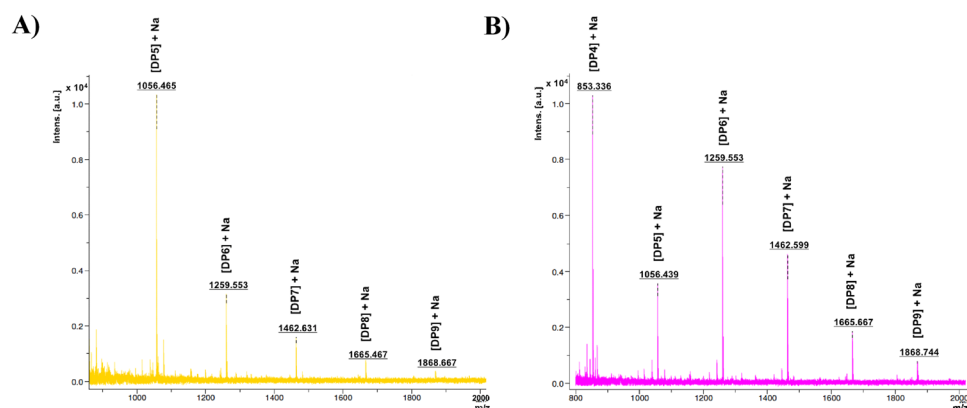
**Figure 3.** Hydrolysis of colloidal chitin by *EcChi1*. The *EcChi1* ( $0.8 \mu\text{M}$ ) was incubated with  $30 \text{ mg/ml}$  of the substrate in  $900 \mu\text{L}$  reaction mixture for different time periods from 0 to 720 min at  $40^\circ\text{C}$ . The reaction products were analyzed by binary gradient HPLC. (A) The topmost profile shows a standard mixture of CHOS ranging from DP1 to DP6. The remaining profiles are the reactions at different incubation times. Control represents the substrate without *EcChi1* while standard represents CHOS ranging from DP1 to DP6. (B) Overview of the quantifiable CHOS products generated during hydrolysis. Products were quantified from respective peak areas using standard calibration curves of CHOS ranging from DP1 to DP4.



**Figure 4.** Time course hydrolysis and TG catalyzed by *EcChi1* with DP3, DP4. The reaction mixture was incubated with  $200 \text{ nM}$  of *EcChi1* and  $1 \text{ mM}$  of DP3/DP4 for different time periods from 0 to 720 min at  $40^\circ\text{C}$ . Products were analyzed by binary gradient HPLC. (A and B) HPLC profiles of reaction products from DP3 (A) and DP4 (B) substrates. The topmost profile shows a standard mixture of CHOS ranging from DP1 to DP6 (A and B). The other profiles show the reaction products from DP3 and DP4 substrates at the indicated incubation times. The inset shows a magnified view of the low-peak-area products. Control represents the substrate without *EcChi1*. Overview of concentrations of CHOS products generated during reaction time courses with DP3 (C) and DP4 (D) substrates. Products were quantified from respective peak areas using standard calibration curves of CHOS ranging from DP1 to DP6.

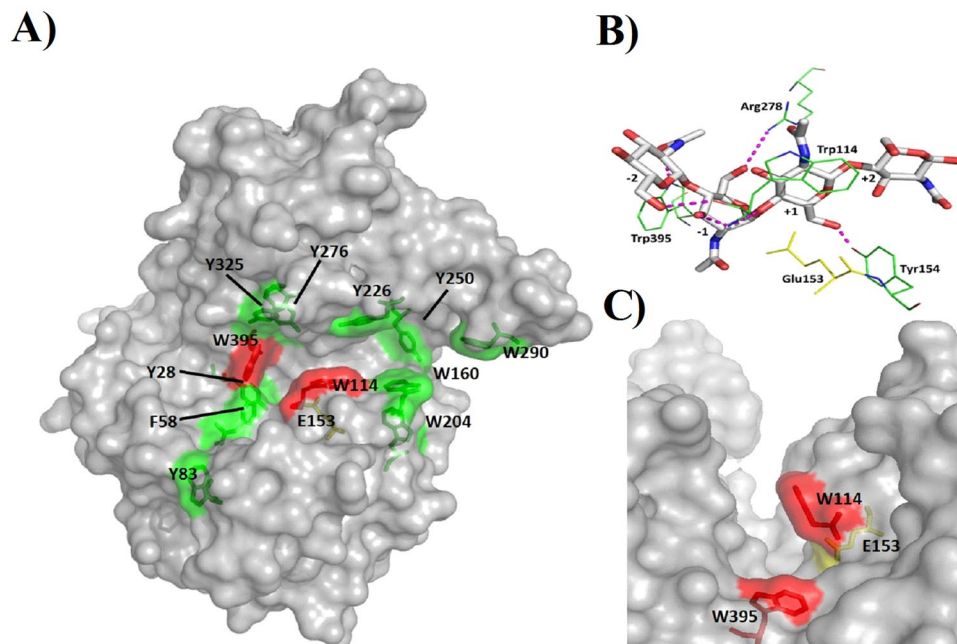


**Figure 5.** Time course hydrolysis and TG catalyzed by *EcChi1* with DP5 and DP6. The reaction mixture products were analyzed by binary gradient HPLC. (A and B) HPLC profiles of reaction products from DP5 (A) and DP6 (B) substrates. The topmost profile shows a standard mixture of CHOS ranging from DP1 to DP6 (A and B). The other profiles show the reaction products from DP5 and DP6 substrates at the indicated incubation times. The inset shows a magnified view of the low-peak-area products. Control represents the substrate without *EcChi1*. (C and D) Overview of concentrations of CHOS products generated during reaction time courses with DP5 (C) and DP6 (D) substrates. Products were quantified from respective peak areas using standard calibration curves of CHOS ranging from DP1 to DP6.



**Figure 6.** MALDI-TOF MS analysis of CHOS/TG products with DP5 and DP6 substrates catalyzed by *EcChi1*. The each 40  $\mu\text{L}$  of reaction sample was concentrated under reduced pressure at 25  $^{\circ}\text{C}$  and dissolved in 10  $\mu\text{L}$  of MilliQ  $\text{H}_2\text{O}$ . The 2  $\mu\text{L}$  of the reaction mixture was mixed with equal volume of 2,5-dihydroxybenzoic acid (2,5-DHB), and the resultant solution was subjected to mass measurements using an Ultraflex MALDI – TOF/TOF instrument. The masses shown here are with Na adducts.

and 5 min, respectively (Fig. 4B and D). With DP5 substrate, DP6-DP9 products formed due to TG. DP6 formed from 1 min that reached to a maximum (2.5%) by 45 min, that was hydrolyzed to shorter CHOS (Fig. 5A and C). The CHOS DP7-DP9 were detectable TG products with DP6 substrate (Fig. 5B and D). The TG products formed from 1 min onwards and remained until 240 min.



**Figure 7.** Substrate binding cleft of modelled *EcChi1*. (A) Aromatic residues lining along the substrate binding cleft (protein in surface representation and amino acids in green and red sticks) and amino acids interacting with chitin tetramer labeled in red colour. (B) Docked conformation of chitin tetramer (white sticks) and its possible interactions with residues (green lines) along with catalytic residue, Glu153 (yellow line) of *EcChi1*. (C) Deep active site cleft of *EcChi1* showing catalytic (yellow colour) and aromatic (red colour) amino acids.

The hydrolytic and TG activity of *EcChi1* were assayed at low substrate and enzyme concentrations with DP4 substrate. *EcChi1* exhibited remarkable TG activity even with the low substrate and enzyme concentrations. A 10-fold lower concentration of the substrate (100  $\mu\text{M}$ ) and enzyme (20 nM) resulted in the formation of DP5–DP7 as TG products until 30 min that was eventually degraded (Supplementary Fig. S4A). When the substrate concentration decreased by 20 fold (50  $\mu\text{M}$ ), the TG products, DP5 and DP6 formed between 0–8 min, which gradually decreased over time (Supplementary Fig. S4B). These results indicated that the TG activity of *EcChi1* was proportional to the substrate concentration.

**Docking of *EcChi1* with chitin tetramer.** Sequence analysis revealed that *EcChi1* is a single modular GH18 chitinase. The closest homologue of *EcChi1* with a known crystal structure was chitinase II from *Klebsiella pneumoniae* (PDB: 3QOK; 86% sequence identity) which was used as the template to generate a 3D model. The stereo chemical quality of the generated model was checked for its quality through Ramachandran plot generated from PROCHECK analysis. The model showed 92.1% amino acids of modelled protein fell in the most favored region. Verify-3D graph also showed that 95.71% of the residues had an averaged 3D-1D score  $\geq 0.2$ . Coordinates for chitin tetramer were extracted from the crystal structure of chitinase A from *Serratia marcescens* (PDB: 1NH6) and docked with modelled *EcChi1* protein. *EcChi1* complexed with DP4 substrate showed binding of DP4 from +2 to –2 subsites. Trp114 and Tyr154 and Arg278 and Trp395 are present at +2, +1 and –1, –2 positions, respectively. The residues forming interactions with ligand in the protein–ligand complex were Trp114, Tyr154, Arg278, and Trp395. Amino group in indole ring of Trp395 formed the hydrogen bond with a distance of 3.5 Å with the oxygen of acetamido group of non-reducing sugar moiety and a hydroxyl group on C6 of non-reducing sugar the formed hydrogen bond with 2.8 Å distance with the carboxylic group of Trp114, at –2 subsite (Fig. 7B). A terminal guanidium amino group of Arg278 and the amino group of Trp114 formed a hydrogen bond with 3.4 Å and 2.1 Å distances with the hydroxyl group of C6 and C4 of the penultimate non-reducing sugar moiety, respectively at –1 subsite. The hydroxyl group of Tyr154 formed a hydrogen bond with the hydroxyl group on C6 of sugar moiety at +1 subsite with 1.9 Å distance (Fig. 7B). *EcChi1* has deep substrate binding cleft lined with aromatic residues that may play a major role in TG activity. The two aromatic amino acids Trp114 and Trp395 are located near to the catalytic center Glu153 (Fig. 7A and C).

## Discussion

The recent advances in whole genome sequencing technologies have paved the way to mine bacterial genomes for the genes and proteins of industrial importance. Search for new chitinases by utilizing the available genome information from diverse bacteria may provide chitinases with high TG activity for generating longer-chain CHOS. Phylogenetic analysis of chitinases from diverse bacterial members showed clustering of chitinases belonging to Enterobacteriaceae members along with a well characterized hyper-TG chitinase *SpChiD* with a few exceptions<sup>17</sup>. This observation suggested that mining Enterobacteriaceae members could be an option for novel TG enzyme. In the present study, we have identified and characterized a hyper-TG chitinase (*EcChi1*) from *Enterobacter*

| Strain name                             | Chitinase                | Mol. Wt (kDa) | Optimum pH | Optimum Temp (°C) | TG activity  | Reference     |
|---|--------------------------|---------------|------------|-------------------|--------------|---------------|
| <i>Enterobacter agglomerans</i>         | ChiA                     | 61            | ND         | ND                | Not reported | 21            |
| <i>Enterobacter</i> sp.G-1              | N-acetyl glucosaminidase | 92            | 6.0        | 45                | Not reported | 18            |
|   | ChiA                     | 60            | ND         | ND                | Not reported | 61            |
| <i>Enterobacter</i> sp. NRG4            | ChiA                     | 61            | 5.5        | 45                | Not reported | 20            |
| <i>E. aerogenes</i>                     | Chitinase                | 42.5          | 6.0        | 55                | Not reported | 19            |
| <i>E. cloacae</i> subsp. <i>cloacae</i> | EcChi1                   | 42.5          | 5          | 40                | Yes          | Present study |

**Table 1.** Details of chitinases characterized from *Enterobacter* sp. ND – Not determined.

*cloacae*, a familiar representative species of family Enterobacteriaceae. Very few reports are available on chitinases and chitinolytic system of *Enterobacter* sp (Table 1). Among which, *Enterobacter* sp. G-1<sup>18</sup>, *E. aerogenes*<sup>19</sup>, and *Enterobacter* sp. NRG4<sup>20</sup> chitinases were characterized. ChiA was the first reported chitinase<sup>21</sup> from an *Enterobacter* sp and shared 26% sequence homology with EcChi1. EcChi1 exhibited 87% sequence similarity with *E. aerogenes* chitinase, which did not have TG activity. There is no correlation between overall sequence similarity and TG as it depends on active site architecture of the enzyme and the orientation of water molecules at the catalytic centre which favors binding of incoming carbohydrate molecules, through strong interactions in the aglycon subsites<sup>22</sup>. The amino acid sequence of EcChi1 showed 87% identity to that of GH18 chitinase II (PDB ID: 3QOK) from *Klebsiella pneumoniae* for which no characterization was available. EcChi1 shared 75% identity with a hyper-TG chitinase of *S. proteamaculans* (SpChiD) (PDB ID: 4NZC)<sup>11</sup>. Nevertheless, many chitinases showing TG such as ChiA1 from *B. circulans* WI-12<sup>23</sup>, ChiA from *Vibrio harveyi*<sup>24</sup>, and human chitotriosidase-1<sup>25</sup> showed only 31%, 26% and 29% sequence similarity to EcChi1, respectively.

Enzymes show a pH range at which they are most active. EcChi1 was optimally active at slightly acidic pH (pH 5.0–6.0) similar to the chitinases A, B, and D from *S. proteamaculans*<sup>11,26</sup>, chitinase B from *S. marcescens*<sup>27</sup>, *Paenibacillus* sp. D1<sup>28</sup> and *Enterobacter* sp. NRG4<sup>29</sup>. EcChi1 optimally active at 40 °C comparable to SpChiB, SpChiC<sup>26</sup>, SpChiD<sup>11</sup> and *Chitinibacter* sp. GC72<sup>30</sup>. The optimum temperature of EcChi1 was higher than many other chitinases such as *Vibrio* sp<sup>31</sup> and *S. antarcticus*<sup>32</sup>. However, some chitinases from *B. cereus* IO8<sup>33</sup> and *Paenibacillus barengoltzii*<sup>34</sup> exhibited high optimal temperature than EcChi1.

Unlike pH and temperature optima, the kinetic parameters of EcChi1 were different from SpChiD. The  $K_m$  and catalytic efficiency ( $k_{cat}/K_m$ ) of EcChi1 were 6 fold decrease and nearly 2 fold increase than SpChiD, respectively<sup>11</sup>. Among all the kinetic parameters, there is a significant correlation between  $K_m$  and TG, whereas no direct correlation between  $k_{cat}$  and TG activity was observed in chitinases<sup>35</sup>. The single mutants of ChiA-D313N and ChiB-D142N from *S. marcescens* showed 2 to 3 fold lower  $K_m$  and increased TG than the wild-type. The hydrophobicity of amino acids at substrate acceptor region and catalytic groove makes the enzyme efficiently bind to the substrates which resulted in less  $K_m$ . The overall catalytic efficiency ( $k_{cat}/K_m$ ) of EcChi1 was lower than ChiA, ChiB and, ChiC from *S. proteamaculans*<sup>26</sup> possible attributed to the lack of accessory domain (s) in EcChi1. The activity of EcChi1 on crystalline  $\alpha$ -chitin and  $\beta$ -chitin was also low in comparison to colloidal chitin due to lack of accessory domains. The lining of aromatic amino acids in a substrate binding groove of EcChi1 (Fig. 7A) might responsible towards chitinous polymeric substrates. Similar findings were observed for SpChiD and *StmChiA* but not for *StmChiB*<sup>11,12</sup>. EcChi1 had a very low activity on avicel (Fig. 1D). This could be due to the non-specific activity of EcChi1, as reported in a chitinase (ChiC) from *Pseudoalteromonas* sp. DL-6<sup>36</sup>. Various cations, particularly metal ions influence the activity of chitinases<sup>37</sup>. Certain ions such  $Cu^{+2}$  and  $Zn^{+2}$  act as either stimulators or inhibitors on chitinases<sup>38</sup>, not in the case of EcChi1.

The highly conserved catalytic motif (DxxDxDxE) and chitin-binding motif (SXGG) are positioned between 146–153 and 110–113 amino acids, respectively. Single amino acid residue present immediate to the chitin-binding motif and presence of  $\alpha/\beta$  fold insertion plays a pivotal role in processivity<sup>39</sup>. The two aromatic amino acids Trp97 and Trp220 were crucial for processivity in ChiB from *S. marcescens*<sup>40</sup>. Presumably, corresponding aromatic amino acids Trp114 and Tyr226 might tightly bind with the polymeric substrate and steer the processive action of EcChi1, implying that EcChi1 might be a processive enzyme (Fig. 7A and C). In addition, homology modelling studies of EcChi1 showed the presence of deep substrate binding cleft lined with twelve aromatic amino acids, and  $\alpha/\beta$  fold insertion further support the characteristic feature of a processive enzyme. Analysis of product profile for dominance of even number oligosaccharides and the ratio of DP2/DP1 have been employed to determine an apparent degree of processivity<sup>41,42</sup>. For example, the ratio of initial DP2/DP1 for two processive enzymes *SmChiA* and *SmChiB* had  $30.1 \pm 1.5$  and  $24.3 \pm 2.0$ , respectively<sup>42</sup>. Upon degradation of colloidal chitin by EcChi1, DP1 was not detectable up to 60 min indicating a possible processive mode (Fig. 3B). At 90 min the DP2/DP1 ratio was 1.45, which further decreased with increase in time possible because of chitinase activity of EcChi1.

HPLC analysis revealed that DP2 and DP1 are the major end products with colloidal chitin substrate similar to SpChiD and *StmChiA*<sup>11,12</sup>. Gradual formation of DP1 from DP2 indicated chitinase activity and DP2 was not a suitable substrate for TG. EcChi1 exhibited marginally lower  $k_{cat}$  when compared to SpChiD. The sequence analysis and modelling studies showed EcChi1 having a unique loop (Asn30 - Asp42) present at N-terminal region. The amino acids present at substrate binding cleft were occupied by the loop structure. The unique loop and aromatic residues highly conserved in single domain GH18 Enterobacteriaceae chitinases. The conserved amino acids Tyr28, Val35, and Thr36 might be responsible for chitinase activity in EcChi1 similar to SpChiD<sup>17</sup>.

TG is a far-flung phenomenon in GH18 family chitinases, little known about the structural basis underlying this integral activity. *EcChi1* showed significant TG activity on CHOS and produced long chain CHOS up to DP9. The product profile with DP3 substrate suggested that  $-1$  to  $+2$  or  $-2$  to  $+1$  subsites are possible for productive binding. DP3 was the minimum length of the substrate for *EcChi1* to show TG activity. DP1-DP8 products were detected with DP4 substrate in which DP5-DP8 were TG products, whereas DP5-DP9 were TG products in *SpChiD*<sup>11</sup>. DP9 was the longest TG product detected with both DP5 and DP6 substrates. Similarly, incubation of 5 mM chitopentaose (DP5) with recombinant human chitotriosidase (HCT) resulted in the formation of the DP1-DP9<sup>25</sup>. No TG activity was observed by *StmChiA* with DP5 substrate<sup>12</sup>. *EcChi1* synthesized long-chain CHOS even with low concentrations of substrate and enzyme. Amino acid residues positioned at acceptor binding site are assumed to be crucial for TG activity<sup>43</sup>. The known chitinases with TG activity were occupied with aromatic amino acids at  $+n$  subsites. Phe396&Trp275 and Trp220&Trp97 were occupied by the position at  $+2$  and  $+1$  subsites of ChiA and ChiB from *S. marcescens*, respectively<sup>44,45</sup>. Similarly, Trp114, Tyr154 at  $+1$ ,  $+2$  subsites might help further in delivering efficient TG activity in *EcChi1*. In addition, Trp285 and Trp164 are occupying in  $+2$  and  $+1$  subsites of ChiA from *B. circulans*<sup>46</sup>. It was further supported by a non-processive endo-chitinase from *S. marcescens*, *SmChiC2* which lacks TG activity and aromatic amino acids at  $+1$ ,  $+2$  subsites<sup>39</sup>. Thus bulk aromatic amino acids present at positive subsites and the lining of aromatic amino acids at substrate binding cleft might be increased the binding affinities for upcoming sugar acceptor molecule thereby promoted TG activity in *EcChi1*.

In addition to processivity, single amino acid residue, present next to this chitin-binding motif (SXGG), plays an important role in TG<sup>47,48</sup>. SXGG motif was extended with tryptophan in other well-known TG chitinases, viz., HCT<sup>25</sup>, *B. circulans* ChiA<sup>46</sup>, ChiA and ChiB from *S. marcescens*<sup>49</sup> and *SpChiD* in *S. proteamaculans*<sup>11</sup>. In non-TG and non-processive endo-chitinases, such as *SmChiC2* and *LlChi18A*, this tryptophan was substituted with small amino acid alanine indicated its probable role in TG<sup>47,50</sup>. Trp114 was the corresponding amino acid in *EcChi1* which may contribute to TG and processivity. In addition, the highly conserved novel loop was identified among the GH18 single catalytic domain chitinases from Enterobacteriaceae, showing their divergence from other groups<sup>17</sup>. This unique loop (Asn30 - Asp42) is present in *EcChi1* may favor the interaction with the incoming substrate molecule.

The degree of endo-activity is best assessed as a probability of endo-mode initiation where concentrations of both insoluble reducing groups (endo-initiation) and reducing end groups (exo-initiation) upon substrate degradation are determined<sup>51</sup>. Hence, the observation of CHOS with DP higher than DP2 is indicative of *EcChi1* having endo-activity as observed for *SmChiC*. Homology modelling based structural analysis also suggests that *EcChi1* has structural features of an endo-acting enzyme with an apparent processive mode of action. *EcChi1* was the first reported chitinase from *Enterobacter* sp with high TG. The capability to hydrolyze chitin with high TG activity suggests *EcChi1* could be a suitable enzyme for industrial production of long chain CHOS, while it would also be of interest to know the possible physiological role of *EcChi1*.

## Methods

**Chemicals and enzymes.** Genomic DNA of *E. cloacae* subsp. *cloacae* ATCC 13047 was used as a template to amplify the *EcChi1* gene. The plasmid pET-28a (+) and *E. coli* Rosetta-gami II (DE3) (Novagen, Madison, USA) were used for heterologous expression. *E. coli* was grown in Luria bertani (LB) broth (1% peptone, 0.5% yeast extract, 1% NaCl) at 37 °C. The antibiotics kanamycin (50 µg ml<sup>-1</sup>) and chloramphenicol (25 µg ml<sup>-1</sup>) were added to the medium as required. Oligonucleotide primers were designed based on the DNA sequence available in the database and procured from Integrated DNA Technologies (Bangalore, India). Restriction enzymes, T4 DNA ligase, and Q5 DNA polymerase were from New England Biolabs (NEB). Isopropyl- $\beta$ -D-thiogalactoside (IPTG), kanamycin and all other chemicals were purchased from Calbiochem, Merck (Darmstadt, Germany) or from Hi-Media Labs (Mumbai, India). The polymeric substrates  $\alpha$ -chitin (extracted from shrimp shells),  $\beta$ -chitin (extracted from squid pen), and high molecular weight chitosan substrates of a fraction of acetylation ( $F_A$ ) = 0.10% (Mahtani Chitosan Pvt. Ltd.) and  $F_A$  = 25% (Sigma-Aldrich) were used for enzyme assay. Colloidal chitin and glycol chitin were prepared as described earlier<sup>52</sup>. CHOS with different DP were purchased from Seikagaku Corporation (Tokyo, Japan). Avicel was procured from Sigma-Aldrich (St. Louis, MO).

**DNA manipulations and cloning of *EcChi1*.** *E. cloacae* genomic DNA (gDNA) was isolated as described by<sup>53</sup>. A gradient PCR, with appropriate gene-specific forward primer: 5'-CTC CCA TGG GAC TGA TGT CCG TG-3' and reverse primer: 5'-GGC CTC GAG CTT CGC TAA CTG GTT-3' was used to determine the suitable annealing temperature for maximum amplicon yield. The amplicon was purified using PCR cleanup kit (MACHEREY- NAGEL GmbH & Co., Germany) and cloned into the *Nco* I and *Xho* I sites of the pET-28a (+). The construct was used to transform *E. coli* Rosetta-gami-II (DE3) for expression of the cloned gene. Transformants were selected on kanamycin and chloramphenicol containing LB plates. The nucleotide sequence of the insert was confirmed by automated DNA sequencing (Scigenom, Kerala, India).

**Expression and purification of *EcChi1*.** Expression and purification of *EcChi1* were done as described by ref. 52. The expressed protein was purified using Ni-nitrilotriacetic acid (Ni-NTA) affinity chromatography. Fractions of eluate having *EcChi1* were pooled and the protein purity was assessed by performing 12% SDS-PAGE. The purified *EcChi1*, in 50 mM sodium citrate buffer (pH 5.0), was stored at 4 °C for further characterization. The protein content of the sample was measured using a bicinchoninic acid (BCA) protein assay kit (Thermo Scientific, USA) where bovine serum albumin (BSA) as a standard.



**Dot blot assay.** A dot blot assay was performed to detect the activity of purified *EcChi1* as described by ref. 11. A composite gel mixed with 0.1% glycol chitin was prepared. Five micrograms of *EcChi1* was spotted onto the gel and placed in the humid chamber at 37 °C overnight. After incubation, the gel was stained with 0.01% calcofluor white M2R in 0.5 M Tris-HCl (pH 8.9) for 10 min at 28 ± 2 °C. Finally, the brightener solution was removed. The gel was washed three times with distilled water for every 10 min at 28 ± 2 °C. The lytic zone was visualized on a UV transilluminator.

**Reducing end assay.** Chitinase activity was measured by a reducing-end assay as described by ref. 54 with slight modifications. Recombinant *EcChi1* was incubated with colloidal chitin in 50 mM sodium citrate pH 5.0 at 40 °C, 200 rpm for 1 h, followed by centrifugation at 13,600 × g, 4 °C for 30 min. A 40 μL of the clear supernatant containing reducing groups was mixed with 300 μL of the freshly prepared color reagent (0.5 M sodium carbonate, 0.05% potassium ferricyanide) and boiled for 15 min at 120 °C. Optical density was measured for 200 μL of each reaction mixture at 420 nm. One unit was defined as the amount of enzyme that liberated 1 μmol of reducing sugar per min. All the reactions were performed in triplicate.

**Optimum pH and temperature.** To determine the optimum pH, the purified *EcChi1*, and colloidal chitin was incubated at 40 °C for 1 h in 50 mM buffers of different pH (2.0 to 10.0). Glycine-HCl buffer (pH 2.0), sodium citrate buffer (pH 3.0–6.0), sodium acetate buffer (pH 4.0–5.6), sodium phosphate buffer (pH 6.0–8.0), and glycine-NaOH buffer (pH 9.0 and 10.0) were used to determine optimum pH.

The effect of temperature on the activity of *EcChi1* was assessed by incubating 3.5 μM enzyme with 30 mg ml<sup>-1</sup> colloidal chitin in 50 mM citrate buffer, (pH 5.0) at 10, 20, 30, 40, 50, 60, 70 and 80 °C and measured chitinase activity.

**Steady-state kinetics.** Kinetic parameters of *EcChi1* were determined using colloidal chitin and chitobiose as the substrate. The reaction mixture (200 μL) containing 0 to 60 mg ml<sup>-1</sup> of colloidal chitin and 0 to 1000 μM chitobiose substrate with *EcChi1* in 50 mM buffer was incubated at 40 °C for 1 h with constant shaking at 200 rpm for chitinase assay. The unit of chitinase was defined as the release of 1 μmol of GlcNAc per second under standard experimental conditions. Specific activity was calculated in nkat/mg of protein. Kinetic values were obtained from triplicates of data fitting to the Michaelis-Menten equation *via* Non-linear regression using GraphPad Prism version 5.0 (GraphPad Software Inc, San Diego, CA).

**Substrate specificity.** The substrate specificity of the purified *EcChi1* was determined by replacing the colloidal chitin in the reaction mixture with different insoluble chitinous and non-chitinous substrates *viz.*, α-chitin, β-chitin, colloidal chitin, glycol chitin (water-soluble chitin), chitosans ( $F_A = 0.1$  and 25%), or Avicel (microcrystalline cellulose). *EcChi1* (3.5 μM) was incubated with different substrates (2.5% wt/vol) at 40 °C for 1 h followed by reducing end assay. For each experiment, we individually calculated the reducing ends present in substrate alone and deducted the initial number of reducing groups from the value obtained from the enzyme-substrate reaction. One unit was defined as the amount of chitinase that liberated 1 μmol of reducing sugar per min.

**Soluble substrate binding.** Soluble substrate binding was assessed as described by ref. 52. Different soluble polymeric substrates, *viz.*, glycol chitin, laminarin, and CM-cellulose were incorporated in 8% polyacrylamide gels. BSA was loaded as a non-interacting protein. The electrophoresis was performed at 4 °C at a constant voltage of 80 V. The gels were stained with coomassie blue G-250 to detect the proteins. Binding was assessed visually or alternatively, the migration distances of the chitinases and protein marker were measured directly on the resolving gels.

**Effect of various additives.** To determine the effect of various additives *viz.*, metal ions (Ca<sup>2+</sup>, Cu<sup>2+</sup>, Fe<sup>2+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Hg<sup>2+</sup>, Zn<sup>2+</sup>), chelator (EDTA) and denaturants (Urea, SDS), of 1 mM each were added to *EcChi1* with colloidal chitin as a substrate. After a pre-incubation for 1 h at 40 °C at 200 rpm activity was measured by reducing end assay. For each additive, the residual activity was calculated, considering the activity without additive was 100%.

**Time-course of colloidal chitin and oligosaccharide degradation.** The hydrolytic activity of *EcChi1* was assessed in a reaction mixture containing 1 mM substrate and 200 nM purified *EcChi1* in 50 mM citrate buffer, (pH 5.0) at 40 °C over 720 min. The reaction was terminated by transferring 40 μL of the reaction mixture to an Eppendorf tube containing an equal amount of 70% acetonitrile and stored at -20 °C until analysis. Aliquots of 20 μL were injected into HPLC (Shimadzu, Tokyo, Japan) equipped with a ShodexAsahipack NH2P-504E column (4.6 mm [inner diameter] by 250 mm; Showa Denko K.K). Briefly, the mobile phase consisted of 70% acetonitrile and 30% MilliQ H<sub>2</sub>O, the flow rate was set to 0.7 ml min<sup>-1</sup>. The eluted CHOS were monitored at 210 nm. A CHOS mixture containing equal weights of oligomers ranging from DP1 to DP6 was used to generate standard chromatogram. Calibration curves were constructed separately for each CHOS in the mixture. The data points generated a linear curve for each CHOS with r<sup>2</sup> values of 0.997 to 1.0.

**MALDI-TOF MS analysis.** The products of the reaction mixture with DP5 and DP6 CHOS substrates were analyzed by matrix-assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF MS) after 45 min and 15 min, respectively. A 40 μL of the reaction mixture was concentrated and redissolved in 10 μL of HPLC-grade MilliQ H<sub>2</sub>O (Merck, Mumbai, India). Two microliters sample was mixed into the 2,5-dihydroxybenzoic acid (DHB) droplet and dried under a stream of air and analysis was done using Ultraflex MALDI-TOF/TOF (BrukerDaltonics GmbH, Bremen, Germany) with an autoflex 123 smart beam. The instrument was operated by the FlexControl 3.0 software package. All spectra were obtained using the reflectron mode

with an acceleration voltage of 25 kV, a reflector voltage of 26, and pulsed ion extraction of 40 ns in the positive ion mode. The acquisition range was from  $m/z$  800 to 2000. Peak lists were generated from the MS spectra using BrukerFlex Analysis software (version 3.0).

**Docking of EcChi1 with chitin tetramer.** The protein sequence of EcChi1 was retrieved from NCBI Database (<http://www.ncbi.nlm.nih.gov/>) with accession number ADF62010.1 for modeling. To select appropriate templates for constructing 3D structure models of the EcChi1 protein, The BLAST program was used for sequence search against known 3D structure available in the Protein Databank (PDB) (<http://www.rcsb.org/>), The homology model program MODELLER v9.12<sup>55</sup> was employed to generate 3D models of EcChi1 based on the crystal coordinates of chitinase II from *Klebsiella pneumoniae* (PDB: 3QOK). The stereochemical quality of the modeled protein structure was checked in Ramachandran plot<sup>56</sup> using PROCHECK<sup>57</sup>. The compatibility of the model with its sequence was measured by Verify-3D graph<sup>58,59</sup>. Chitin tetramer ligand was extracted from the 1NH6 crystal structure, using Discovery studio 4.0 and used for docking studies using Autodock 4.2<sup>60</sup> via Auto Dock Tools (ADT) graphical user interface. Interactions with ligand were viewed in PyMol Molecular Graphics System 1.7.

## References

- Das, S. N. *et al.* Biotechnological approaches for field applications of chitoooligosaccharides (COS) to induce innate immunity in plants. *Crit. Rev. Biotechnol.* **35**, 29–43 (2015).
- Hamed, I., Ozogul, F. & Regenstein, J. M. Industrial applications of crustacean by products (chitin, chitosan, and chitoooligosaccharides): A review. *Trends Food Sci. Technol.* **48**, 40–50 (2016).
- Gillard, L., Tran, A. T., Boyer, F. D. & Beau, J. M. Chitoooligosaccharide Synthesis Using an Ionic Tag. *European J. Org. Chem.* **2016**, 1103–1109 (2016).
- Dahiya, N., Tewari, R. & Hoondal, G. S. Biotechnological aspects of chitinolytic enzymes: A review. *Applied Microbiology and Biotechnology* **71**, 773–782 (2006).
- Divne, C. *et al.* The three-dimensional crystal structure of the catalytic core of cellobiohydrolase I from *Trichoderma reesei*. *Science*. **265**, 524–8 (1994).
- Rouvainen, J., Bergfors, T., Teeri, T., Knowles, J. K. & Jones, T. A. Three-dimensional structure of cellobiohydrolase II from *Trichoderma reesei*. *Science*. **249**, 380–6 (1990).
- Divne, C., Stahlberg, J., Teeri, T. T. & Jones, T. A. High-resolution crystal structures reveal how a cellulose chain is bound in the 50°A long tunnel of cellobiohydrolase I from *Trichoderma reesei*. *J. Mol. Biol.* **275**, 309–25 (1998).
- Varrot, A. *et al.* Structural basis for ligand binding and processivity in cellobiohydrolase Cel6A from *Humicola insolens*. *Structure*. **11**, 855–864 (2003).
- Umekawa, M. *et al.* Mutants of *Mucor hiemalis* endo beta N-acetyl glucosaminidase show enhanced transglycosylation and glycosynthase like activities. *J. Biol. Chem.* **283**, 4469–4479 (2008).
- Taira, T. *et al.* Transglycosylation reaction catalyzed by a class V chitinase from cycad, *Cycas revoluta*: A study involving site directed mutagenesis, HPLC, and real-time ESI-MS. *Biochim. Biophys. Acta - Proteins Proteomics* **1804**, 668–675 (2010).
- Purushotham, P. & Podile, A. R. Synthesis of long chain chitoooligosaccharides by a hyper transglycosylating processive endochitinase of *Serratia proteamaculans* 568. *J. Bacteriol.* **194**, 4260–4271 (2012).
- Suma, K. & Podile, A. R. Chitinase A from *Stenotrophomonas maltophilia* shows transglycosylation and antifungal activities. *Bioresour. Technol* **133**, 213–220 (2013).
- Huang, C. J. & Chen, C. Y. High level expression and characterization of two chitinases, ChiCH and ChiCW, of *Bacillus cereus* 28-9 in *Escherichia coli*. *Biochem. Biophys. Res. Commun.* **327**, 8–17 (2005).
- Frederiksen, R. F. *et al.* Bacterial chitinases and chitin binding proteins as virulence factors. *Microbiology*. **159**, 833–47 (2013).
- Prescott, J. F., Parreira, V. R., Mehdizadeh Gohari, I., Lepp, D. & Gong, J. The pathogenesis of necrotic enteritis in chickens: what we know and what we need to know: a review. *Avian Pathol.* **45**, 288–94 (2016).
- Hoffmann, H. *et al.* Reassignment of *Enterobacter dissolvens* to *Enterobacter cloacae* as *E. cloacae* sub species *dissolvens* comb. nov. and emended description of *Enterobacter asburiae* and *Enterobacter kobei*. *Syst. Appl. Microbiol.* **28**, 196–205 (2005).
- Madhuprakash, J. *et al.* Inverse relationship between chitobiase and transglycosylation activities of chitinase-D from *Serratia proteamaculans* revealed by mutational and biophysical analyses. *Sci. Rep* **5**, 15657 (2015).
- Matsuo, Y. *et al.* Purification, characterization and gene analysis of N-acetyl glucosaminidase from *Enterobacter* sp. G-1. *Biosci. Biotechnol. Biochem.* **63**, 1261–8 (1999).
- Tang, Y., Zhao, J., Ding, S., Liu, S. & Yang, Z. Purification and properties of chitinase from *Enterobacter aerogenes*. *Wei Sheng Wu Xue Bao* **41**, 82–6 (2001).
- Salam, M. *et al.* Cloning, characterization and expression of the chitinase gene of *Enterobacter* sp. NRG4. *Indian J. Microbiol.* **48**, 358–364 (2008).
- Chernin, L. S. *et al.* Molecular cloning, structural analysis, and expression in *Escherichia coli* of a chitinase gene from *Enterobacter agglomerans*. *Appl. Environ. Microbiol.* **63**, 834–9 (1997).
- Williams, S. J. & Withers, S. G. Glycosyl fluorides in enzymatic reactions. *Carbohydrate Research.* **327**, 27–46 (2000).
- Sasaki, C. *et al.* Comparative study of the reaction mechanism of family 18 chitinases from plants and microbes. *J. Biochem.* **131**, 557–564 (2002).
- Suginta, W., Vongsuwan, A., Songsiriritthigul, C., Svasti, J. & Prinz, H. Enzymatic properties of wild type and active site mutants of chitinase A from *Vibrio carchariae*, as revealed by HPLC-MS. *FEBS J* **272**, 3376–3386 (2005).
- Aguilera, B. *et al.* Transglycosidase Activity of Chitotriosidase: Improved enzymatic assay for the human macrophage chitinase. *J. Biol. Chem.* **278**, 40911–40916 (2003).
- Purushotham, P., Sarma, P. V. S. R. N. & Podile, A. R. Multiple chitinases of an endophytic *Serratia proteamaculans* 568 generate chitin oligomers. *Bioresour. Technol* **112**, 261–269 (2012).
- Brurberg, M. B., Nes, I. F. & Eijsink, V. G. H. Comparative studies of chitinases A and B from *Serratia marcescens*. *Microbiology*. **142**, 1581–1589 (1996).
- Singh, A. K. & Chhatpar, H. S. Purification and characterization of chitinase from *Paenibacillus* sp. D1. *Appl. Biochem. Biotechnol.* **164**, 77–88 (2011).
- Dahiya, N., Tewari, R., Tiwari, R. P. & Hoondal, G. S. Chitinase from *Enterobacter* sp. NRG4: Its purification, characterization and reaction pattern. *Electron. J. Biotechnol.* **8**, 134–145 (2005).
- Gao, C. *et al.* Characterization of extracellular chitinase from *Chitinibacter* sp. GC72 and its application in GlcNAc production from crayfish shell enzymatic degradation. *Biochem. Eng. J.* **97**, 59–64 (2015).
- Bendt, A., Huller, H., Kammel, U., Helmke, E. & Schweder, T. Cloning, expression, and characterization of a chitinase gene from the Antarctic psychrotolerant bacterium *Vibrio* sp. strain Fi:7. *Extremophiles*. **5**, 119–26 (2001).
- Lee, S. G. *et al.* Expression of recombinant endochitinase from the Antarctic bacterium, *Sanguibacter antarcticus* KOPRI 21702 in *Pichia pastoris* by codon optimization. *Protein Expr. Purif.* **71**, 108–114 (2010).

33. Hammami, I. *et al.* Partial purification and characterization of chiIO8, a novel antifungal chitinase produced by *Bacillus cereus* IO8. *J. Appl. Microbiol.* **115**, 358–366 (2013).
34. Yang, S. *et al.* Cloning, expression, purification and application of a novel chitinase from a thermophilic marine bacterium *Paenibacillus barengoltzii*. *Food Chem.* **192**, 1041–8 (2016).
35. Zakarlassen, H. *et al.* Aromatic residues in the catalytic center of chitinase A from *Serratia marcescens* affect processivity, enzyme activity, and biomass converting efficiency. *J. Biol. Chem.* **284**, 10610–10617 (2009).
36. Wang, X. *et al.* Characterization of a cold-adapted and salt-tolerant exo-chitinase (ChiC) from *Pseudoalteromonas* sp. DL-6. *Extremophiles.* **20**, 167–176 (2016).
37. Zhang, J., Koppurapu, N. K., Yan, Q., Yang, S. & Jiang, Z. Purification and characterisation of a novel chitinase from persimmon (*Diospyros kaki*) with antifungal activity. *Food Chem.* **138**, 1225–1232 (2013).
38. Yuli, P., Suhartono, M. T., Rukayadi, Y., Hwang, J. K. & Pyun, Y. R. Characteristics of thermostable chitinase enzymes from the Indonesian *Bacillus* sp.13.26. *Enzyme Microb. Technol.* **35**, 147–153 (2004).
39. Payne, C. M. *et al.* Hallmarks of processivity in glycoside hydrolases from crystallographic and computational studies of the *Serratia marcescens* chitinases. *J. Biol. Chem.* **287**, 36322–36330 (2012).
40. Jana, S. *et al.* Aromatic-Mediated Carbohydrate Recognition in Processive *Serratia marcescens* Chitinases. *J. Phys. Chem. B.* **120**, 1236–1249 (2016).
41. Sorbotten, A., Horn, S. J., Eijsink, V. G. H. & Varum, K. M. Degradation of chitosans with chitinase B from *Serratia marcescens*: Production of chito-oligosaccharides and insight into enzyme processivity. *FEBS J* **272**, 538–549 (2005).
42. Stockinger, L. W. *et al.* The effect of the carbohydrate binding module on substrate degradation by the human chitotriosidase. *Biochim. Biophys. Acta - Proteins Proteomics* **1854**, 1494–1501 (2015).
43. Eide, K. B., Lindbom, A. R., Eijsink, V. G. H., Norberg, A. L. & Sorlie, M. Analysis of productive binding modes in the human chitotriosidase. *FEBS Lett* **587**, 3508–3513 (2013).
44. Aronson, N. N. *et al.* Family 18 chitinase oligosaccharide substrate interaction: subsite preference and anomer selectivity of *Serratia marcescens* chitinase A. *Biochem. J.* **376**, 87–95 (2003).
45. Horn, S. J. *et al.* Costs and benefits of processivity in enzymatic degradation of recalcitrant polysaccharides. *Proc. Natl. Acad. Sci. USA* **103**, 18089–94 (2006).
46. Watanabe, T. *et al.* Aromatic residues within the substrate-binding cleft of *Bacillus circulans* chitinase A1 are essential for hydrolysis of crystalline chitin. *Biochem. J.* **376**, 237–244 (2003).
47. Lu, Y. *et al.* Mutation of Trp137 to glutamate completely removes transglycosyl activity associated with the *Aspergillus fumigatus* AfChiB1. *Glycoconj. J.* **26**, 525–534 (2009).
48. Madhuprakash, J., Tanneeru, K., Purushotham, P., Guruprasad, L. & Podile, A. R. Transglycosylation by Chitinase D from *Serratia proteamaculans* improved through altered substrate interactions. *J. Biol. Chem.* **287**, 44619–44627 (2012).
49. Horn, S. J. *et al.* Endo/exo mechanism and processivity of family 18 chitinases produced by *Serratia marcescens*. *FEBS J.* **273**, 491–503 (2006).
50. Vaaje Kolstad, G., Bunaes, A. C., Mathiesen, G. & Eijsink, V. G. H. The chitinolytic system of *Lactococcus lactis* ssp. *lactis* comprises a nonprocessive chitinase and a chitin-binding protein that promotes the degradation of  $\alpha$ - and  $\beta$ -chitin. *FEBS J* **276**, 2402–2415 (2009).
51. Kurasin, M., Kuusk, S., Kuusk, P., Sorlie, M. & Valjamae, P. Slow Off-rates and Strong Product Binding Are Required for Processivity and Efficient Degradation of Recalcitrant Chitin by Family 18 Chitinases. *J. Biol. Chem.* **290**, 29074–85 (2015).
52. Neeraja, C., Moerschbacher, B. & Podile, A. R. Fusion of cellulose binding domain to the catalytic domain improves the activity and conformational stability of chitinase in *Bacillus licheniformis* DSM13. *Bioresour. Technol.* **101**, 3635–41 (2010).
53. Sharma, A. D. & Singh, J. A nonenzymatic method to isolate genomic DNA from bacteria and actinomycete. *Anal. Biochem.* **337**, 354–356 (2005).
54. Imoto, T. & Yagishita, K. A Simple Activity Measurement of Lysozyme. *Agric. Biol. Chem.* **35**, 1154–1156 (1971).
55. Sali, A. & Blundell, T. L. Comparative protein modelling by satisfaction of spatial restraints. *J. Mol. Biol.* **234**, 779–815 (1993).
56. Ramachandran, G. N., Ramakrishnan, C. & Sasisekharan, V. Stereochemistry of polypeptide chain configurations. *J. Mol. Biol.* **7**, 95–99 (1963).
57. Laskowski, R. A., MacArthur, M. A., Moss, D. S. & Thornton, J. M. 20.19. PROCHECK - a program to check the stereochemical quality of protein structures. *J. App. Cryst* **26**, 283–291 (1993).
58. Bowie, J. U., Luthy, R. & Eisenberg, D. A method to identify protein sequences that fold into a known three dimensional structure. *Science.* **253**, 164–170 (1991).
59. Luthy, R., Bowie, J. U. & Eisenberg, D. Assessment of protein models with three dimensional profiles. *Nature.* **356**, 83–85 (1992).
60. Morris, G. & Huey, R. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* **30**, 2785–2791 (2009).
61. Park, J. *et al.* Purification and characterization of the chitinase (ChiA) from *Enterobacter* sp. G-1. *Biosci Biotechnol Biochem* **61**, 684–689 (1997).

## Acknowledgements

We thank the Department of Science and Technology, Government of India, Funds for Infrastructure in Science and Technology, Level II support, and Special Assistance Programme–University Grants Commission (SAP-UGC) to the Department of Plant Sciences. The authors also thank UGC-supported University with Potential for Excellence (UPE) (Phase II) and DBT sponsored Bioinformatics Infrastructure Facility (BIF), at University of Hyderabad. MKM and VPR thank Indian Council Medical Research (ICMR) and Department of Biotechnology (DBT), Government of India, respectively, for Senior Research Fellowship. ARP thanks DBT for Tata Innovation Fellowship.

## Author Contributions

M.K.M., P.R.V., S.N.D and A.R.P. designed the work. M.K.M. constructed the clone of *EcChi1*, purified the recombinant protein, performed enzyme characterization and H.P.L.C. studies. B.B.C. conducted docking studies. The data was analyzed by M.K.M., P.R.V., S.N.D and A.R.P. Manuscript was written by M.K.M. by taking critical inputs from S.N.D and A.R.P.

## Additional Information

**Supplementary information** accompanies this paper at doi:10.1038/s41598-017-05140-3

**Competing Interests:** The authors declare that they have no competing interests.

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2017