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Protecting Mitochondrial Health: A Unifying Mechanism in Adult Neurogenesis

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Synaptic Function Section, National Institute of Neurological Disorder and Stroke, National Institutes of Health, Bethesda, Maryland 20892 Review of Voloboueva et al.

Adult neurogenesis occurs in the hippocampal region of the brain of most mammals including humans. Adultborn neurons play a critical role in hippocampus-dependent memory formation. The importance of adult neurogenesis has been indicated by multiple studies in which dysfunctional adult neurogenesis is associated with aging and neurodegenerative disorders (Gonçalves et al., 2016). In fact, adult neurogenesis is critical for recovery from brain injury (Fallon et al., 2000; Magavi et al., 2000).

Adult neurogenesis involves a multistep process of lineage progression starting with neural stem cells (NSCs), which give rise to intermediate progenitor cells (IPCs), which in turn produce neuroblasts, which mature into neurons (Ming and Song, 2005; Fig. 1). Multiple reports have indicated that metabolic reprogramming events help to regulate the differentiation processes involved in adult neurogenesis. (Gonçalves et al., 2016). For example, NSCs are glycolytic, but mature neurons require mitochondrial respiration for ATP generation. Indeed, cellular mitochondria are essential for the differentiation

of NSCs into mature neurons. However, after brain injury, including stroke and head trauma, inflammatory responses from activated microglia cause mitochondrial damage in newly generated adult neurons, leading to significant loss in neurogenesis at the injury site (Ekdahl et al., 2003).

One way to overcome the problem of inflammation-induced mitochondrial dysfunction is to identify cellular and molecular factors that regulate mitochondrial health and dysfunction. One such factor is microRNA-210 (miR-210), which in vitro studies have shown have a role in downregulating mitochondrial function and mediating the switch to glycolysis (Chen et al., 2010; White et al., 2015). miRNAs are short, single-stranded, noncoding RNAs (~22 nucleotide long) that regulate gene expression by binding to target mRNAs (Huang and Zuo, 2014). In a recent issue of The Journal of Neuroscience, Voloboueva et al. (2017) investigated the effect of the loss of miR-210 on mitochondrial health and neurogenesis under conditions of inflammation in vitro.

Voloboueva et al. (2017) treated NSC cultures with media preconditioned by proinflammatory activated microglia to mimic inflammatory conditions and examined the effect of blocking miR-210 on the survival of NSC cultures under these conditions. They observed an increase in survival of differentiated NSCs and a decrease in apoptotic (TUNEL-positive)

mature neurons when miR-210 was inhibited. Moreover, a significant percentage of surviving NSCs differentiated into mature neurons under inflammatory conditions with miR-210 inhibition compared with NSC cultures exposed to inflammatory injury but transfected with negative control. These effects were accompanied by an increase in the activity of two major mitochondrial enzymes, cytochrome c oxidase (a component of the electron transport chain) and aconitase (a Krebs cycle enzyme). In contrast, an increase in miR-210 levels resulted in a decrease in total NSC survival and differentiated NSCs during inflammatory injury.

In the above experiments, NSC cultures were transfected with miR-210 inhibitor or negative control on days 4–5 of differentiation and exposed to preconditioned proinflammatory media 24 h later. Because the previous results indicated an increase in mitochondrial function and NSC survival rate during inflammatory injury, Voloboueva et al. (2017) predicted that transfecting NSC cultures on days 1-2 after the start of differentiation would further increase mitochondrial energetics. To their surprise, miR-210 inhibition at an earlier time point resulted in a significant decrease in total NSC density (low proliferation) and differentiated NSCs compared with controls. Their observation was consistent with earlier findings suggesting that dividing stem cells depend on glycolytic metabolism rather than mito-

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chondrial bioenergetics for ATP production (Gonçalves et al., 2016). Together, these results indicate that (1) inhibiting miR-210 promotes neurogenesis by protecting mitochondria in NSC cultures against inflammatory injury; and (2) for therapeutic purposes, the timing of miR-210 inhibition is critical because it induces metabolic changes that reduce NSC proliferation capacity.

Voloboueva et al. (2017) observed that blocking miR-210 at an earlier time point altered NSC metabolism with suppression of the glycolytic pathway and an increase in mitochondrial respiratory function. To address the mechanisms for the shift in metabolic activity associated with early miR-210 inhibition in NSCs, Voloboueva et al. (2017) focused on the AMP-activated protein kinase (AMPK) signaling pathway. The intracellular energy sensor AMPK is a master regulator of energy homeostasis that responds to changes in the cellular AMP/ ATP ratio (Carling, 2017). Earlier reports have indicated that AMPK also positively regulates NSC proliferation capacity. Voloboueva et al. (2017) observed that blocking miR-210 led to a decrease in AMPK activity, which likely explains the decreased rate in NSC proliferation.

To further delineate the molecular mechanism regulating NSC proliferation, the authors focused on retinoblastoma (Rb) protein, which is a downstream target of AMPK. Rb protein is one of the known negative regulators of cell-cycle progression in stem cell biology. It has been reported that AMPK directly phosphorylates Rb protein and that hyperphosphorylation of Rb acts as a positive regulator of NSC proliferation (Dasgupta and Milbrandt, 2009). Voloboueva et al. (2017) found that the inhibition of miR-210 led to hypophosphorylation of Rb protein in conjunction with low NSC proliferation. Since miR-210 does not interact directly with AMPK or Rb protein (source: miRBase.org database), this loss in AMPK and Rb protein activity may be a direct consequence of metabolic changes in NSCs resulting from the block in miR-210 function.

Loss of adult-born neurons resulting from defective mitochondrial function has been reported in multiple neurodegenerative disorders and in aging, as well as after brain injury. Indeed, several recent studies have examined factors that improve mitochondrial health and improve neurogenesis, often resulting in improved cognitive performance (Fig. 1). One such recent study (Richetin et al., 2017) highlighted the importance of manipulating

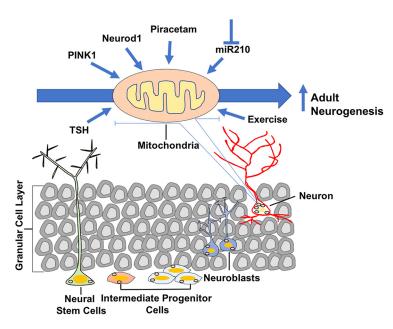


Figure 1. Schematic of the multistep lineage progression of neural stem cells to neurons during adult hippocampal neurogenesis and the factors that positively influence mitochondrial health and promote adult neurogenesis.

mitochondrial function to enhance adult neurogenesis in an Alzheimer's disease (AD) mice model. The authors observed defects in mitochondrial health that resulted in the loss of adult-born neurons in AD mice. They overexpressed a neurogenic bHLH transcription factor (Neurod1), which previously was reported to maintain mitochondrial mass and ATP levels under conditions of oxidative stress in neuroblastoma cells, in the brains of AD mice. Neurod1 overexpression rescued mitochondrial deficits of adult-born neurons and rescued spatial memory. Another study (Beckervordersandforth et al., 2017) investigated the effect of the memory-enhancing drug piracetam on adult neurogenesis in the brains of aged mice. Piracetam is known to enhance mitochondrial respiration, electron transport chain activity, and ATP production in cells with a loss in mitochondrial function (Keil et al., 2006). Interestingly, treatment with piracetam increased the population of IPCs and neuroblasts and ameliorated aging-related defects in hippocampal neurogenesis in aged mice. Exercise is another factor that has been linked to enhancing mitochondrial mass in adult-born neurons and promoting neurogenesis (Steib et al., 2014). A third study (Agnihotri et al., 2017) examined the effect of the loss of PTEN (phosphatase and tensin homolog)-induced kinase-1 (PINK1), a protein that helps to maintain mitochondrial health by regulating the mitophagy of dysfunctional mitochondria, on adult hippocampal neurogenesis in mice.

Loss of PINK1 was associated with mitochondrial deficits and loss in the maturation of adult born neurons in the hippocampus. Finally, multiple studies have indicated thyroid hormone therapy as a potential approach toward enhancing mitochondrial function and adult neurogenesis (Remaud et al., 2014). Defects in thyroid hormone signaling have been associated with alterations in both mitochondrial function and hippocampal neurogenesis in the aging brain (Remaud et al., 2014).

In conclusion, the findings of Voloboueva et al. (2017) provide a critical mechanism underlying inflammation-associated deficits in neurogenesis. This may aid in identifying novel therapeutic strategies for treatment in several neurodegenerative diseases. Recent findings, both *in vitro* and *in vivo*, indicate a direct role of mitochondria in the modulation of adult neurogenesis and provide a strong proof of concept that protecting mitochondrial health could enhance the rate of adult neurogenesis under different neuropathological conditions.

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