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Genome-wide study links *PNPLA3* variant with elevated hepatic transaminase after acute lymphoblastic leukemia therapy

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Abstract

Remission induction therapy for acute lymphoblastic leukemia (ALL) includes medications that may cause hepatotoxicity, including asparaginase. We used a genome-wide association study (GWAS) to identify loci associated with elevated alanine transaminase (ALT) levels after induction

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Conflict of Interest

The authors have no conflicts of interest to disclose.

Authorship Contribution

M.V.R., Y.L., C.A.F., C.S., W.Y., C.C., C.P., N.K., C.L., L.R., S.E.K., L.J.J., E.C.L., N.W., W.L.C., M.L.L., E.A.R., S.P.H., M.D., J.J.Y., C.G.M., J.Z., W.E.E., S.J., and C-H.P. wrote the manuscript; M.V.R., Y.L., and C.A.F. designed the research; Y.L. and C.A.F. performed the research; M.V.R., Y.L., C.A.F., C.S., W.Y., and C.C. analyzed the data.

therapy in children with ALL enrolled on St. Jude Children's Research Hospital (SJCRH) protocols. Germline DNA was genotyped using arrays and exome sequencing. Adjusting for age, body mass index, ancestry, asparaginase preparation and dosage, the *PNPLA3* rs738409 (C>G) I148M variant, previously associated with fatty liver disease risk, had the strongest genetic association with ALT ($P = 2.5 \times 10^{-8}$). The *PNPLA3* rs738409 variant explained 3.8% of the variability in ALT, and partly explained race-related differences in ALT. The *PNPLA3* rs738409 association was replicated in an independent cohort of 2,285 patients treated on Children's Oncology Group protocol AALL0232 ($P = 0.024$). This is an example of a pharmacogenetic variant overlapping with a disease risk variant.

Keywords

Acute Lymphoblastic Leukemia; Asparaginase; Alanine Aminotransferase; Hepatotoxicity; Pharmacogenomics

Introduction

Although cure rates for childhood acute lymphoblastic leukemia (ALL) are approaching 90%,(1) therapy is complicated by drug toxicity. Patients who tolerate less treatment due to adverse drug effects may be at a higher risk of relapse.(2, 3) Hepatotoxicity is a frequent complication leading to treatment interruptions,(4–6) and several drugs used in ALL therapy are associated with the development of hepatotoxicity, including asparaginase, mercaptopurine, thioguanine, methotrexate, etoposide, and glucocorticoid.(7)

Some treatment regimens for adults with ALL avoid using and/or dose modify asparaginase and high-dose methotrexate in order to minimize treatment-related hepatotoxicity,(8, 9) which may contribute to the lower cure rates for adults with ALL. Therefore, studies that can elucidate the mechanism of drug-induced hepatotoxicity during the treatment of ALL may lead to therapeutic strategies that can reduce the frequency of hepatotoxicity, identify patients who are most likely to benefit from treatment, and possibly improve cure rates by reducing treatment interruptions due to hepatotoxicity.

We investigated coding and non-coding genetic variants associated with the elevation of alanine aminotransferase (ALT) after remission induction therapy. A total of 715 St. Jude Children's Research Hospital patients were evaluable with genotyping and post-induction chemotherapy ALT levels. We tested for replication in 2,285 patients treated on the Children's Oncology (COG) AALL0232 trial.

Results

Patients enrolled on St. Jude Children's Research Hospital (SJCRH) Total XV or Total XVI received similar induction therapy with a few notable differences (Table 1). The Total XV protocol included native *E. coli* asparaginase (Elspar), whereas the Total XVI protocol used PEGylated *E. coli* asparaginase (Oncaspar). Patients enrolled on Total XV received mercaptopurine, whereas most on Total XVI received thioguanine; those on Total XV received pre-induction window therapy with methotrexate, whereas those on Total XVI did

not (Table 1). In both protocols, patients with higher minimal residual disease (MRD) at day 15–19 of induction received additional asparaginase compared to those without.

Overall, 715 patients were evaluable for post-induction ALT data in the discovery cohort (Table 2, Figure S1). By univariate analysis (Table S1), treatment protocol (Figure 1A), additional asparaginase during induction (Figure 1B), race group (Figure 1C), age (Figure 1D), body mass index (BMI) percentile (Figure 1E), and gender were associated with post-induction ALT levels. In multivariate analysis (Table 3), higher ALT levels were associated with Total XVI treatment protocol ($P=0.041$), receiving additional induction asparaginase ($P=2.7\times 10^{-4}$), non-black race ($P=8.7\times 10^{-13}$ for whites and $P=2.6\times 10^{-7}$ for Hispanics), older age ($P=1.4\times 10^{-6}$), and higher BMI ($P=2.9\times 10^{-3}$). Race group accounted for 7.2% of the variability in ALT levels (Table 3). Focusing on the National Cancer Institute's common toxicity criteria (CTCAE) categories of ALT toxicity, Total XVI patients had more grade 2–3 ALT elevations compared to Total XV patients ($P=5.8\times 10^{-3}$, Figure S2A).

To identify genetic variants associated with post-induction ALT levels in the SJCRH discovery cohort, we used a multivariate linear regression model including treatment protocol, use of additional asparaginase doses, percentage of Northern European ancestry, West African ancestry and Native American ancestry, age, and BMI percentile as covariates along with genotypes. The strongest association ($P=2.5\times 10^{-8}$) with ALT levels was with the *PNPLA3* rs738409 (C>G) I148M variant (Figure 2A, Table 4), a missense polymorphism known as a quantitative trait locus (QTL) for ALT in the general population. (10) The median ALT level for patients of CC, GC, and GG diplotypes was 35, 45, and 58 IU/L, respectively. When ALT was translated into CTCAE categories of ALT toxicity, the frequency of higher ALT grade (grade > 2) was 2.6%, 3.9% and 5.3% respectively among patients of CC, GC, and GG diplotypes (Figure S2B). The association of *PNPLA3* rs738409 with ALT treated as a continuous variable was observed in both Total XV ($P=5.5\times 10^{-6}$) and Total XVI ($P=2.7\times 10^{-4}$) (Figure S3A), and differed within race groups (Figure 3A, Figure S3B). Interestingly, the minor allele frequency (MAF) of *PNPLA3* rs738409 was highest among Hispanics (44.2%), followed by whites (23.1%), and lowest in blacks (13.1%) in our cohort, as observed in previous studies,(11) and followed the same trend as their post-induction ALT levels (Figure 3B). Principal component analysis (PCA) within each race group did not uncover any stratification among the patients carrying the different *PNPLA3* rs738409 genotypes (Figure S4A–C).

Because the impact of *PNPLA3* rs738409 on ALT elevation differed by race (Figure 3A, Figure S3B), and because ALT levels differed among races in the same direction that the *PNPLA3* rs738409 MAF did (Figure 3B), we questioned whether the association between race and ALT elevation was accounted for mostly by this variant. After adding *PNPLA3* rs738409 genotype to a multivariate analysis for determinants of ALT levels, the *PNPLA3* SNP accounted for 3.8% of ALT variability, and race group still remained an important risk factor ($P=3.7\times 10^{-11}$, 6.6×10^{-5} and 2.0×10^{-3} for whites, Hispanics and others vs. blacks, respectively), and explained 5.3% of the variability in ALT elevation after incorporating the *PNPLA3* SNP (Table 3). Therefore, we performed three separate ancestry-specific GWAS among whites, blacks and Hispanics (Tables S2–S4) to identify additional possible race-specific genetic determinants of ALT, including the percentage of West African and Native

American ancestries as covariates in the GWASs within blacks and Hispanics, respectively. Two variants, *PIGV* rs12748152 and *GREB1* rs149940960, had P values approaching genome-wide significance and only in blacks ($P = 1.3 \times 10^{-8}$ and 2.0×10^{-8} respectively, Figure 2B, 3C&D). Coincidentally, the three subjects harboring the minor allele of *GREB1* rs149940960 also had the minor allele of *PIGV* rs12748152. Both variants were associated with lower ALT only in blacks ($P = 0.14$ in whites and $P = 0.59$ in Hispanics for *PIGV* rs12748152, Figure 3C; the *GREB1* rs149940960 was detected in blacks only). The *PIGV* rs12748152 had a higher MAF in whites (7.1%) than blacks (1.8%) and Hispanics (0.8%), whereas the *GREB1* rs149940960 had MAF = 1.7% in blacks, in accordance with the general population frequency (1000 Genomes). In addition, when race structure was calculated with PCA and the first five principal components were used as covariates in each race-specific GWAS, the top SNPs and P values for association with ALT did not change substantially (Tables S2–S4), and PCA within blacks did not uncover any stratification among the patients carrying the different *PIGV* rs12748152 genotypes (Figure S4D). *PNPLA3* rs6006460 (G>T) was previously reported to protect blacks from the development of nonalcoholic fatty liver disease (NAFLD).⁽¹²⁾ The *PNPLA3* rs6006460 minor allele was among the protective variants in our discovery cohort as well ($P = 3.2 \times 10^{-3}$, Figure S5A), and the effect was significant when restricted only to blacks in our cohort ($P = 0.041$, Figure S5B). The frequency of this protective variant was highest in the blacks of our cohort (MAF = 11%) and thus may have counteracted the adverse effects of carrying the risk allele of *PNPLA3* rs738409, therefore helping to explain why *PNPLA3* rs738409 was not significantly associated with increased ALT among blacks ($P = 0.20$, Figures 3A&S3B). In blacks, the *PNPLA3* rs738409 explained 1.3% of the variability in ALT level ($P = 0.065$) while *PNPLA3* rs6006460 explained another 5.8% ($P = 0.014$). In contrast, among non-blacks in the discovery cohort, *PNPLA3* rs738409 explained 4.2% of the variability whereas *PNPLA3* rs6006460 added only 1.2% ($P < 2.2 \times 10^{-16}$ for both).

A total of 109 variants within the *PNPLA3* gene were found in the SJCRH cohort, 65 of which had nominal P values of < 0.05 for their association with ALT, with 61/65 having the minor allele associated with higher ALT (Table S5). A gene-level GWAS including the same covariates as the single SNP GWAS (protocol, asparaginase dosage, percent ancestry, age, and BMI percentile) identified *PNPLA3* as an overall detrimental gene for ALT levels ($P = 0.0093$), reflecting the collective impact of *PNPLA3* variants.

In the replication cohort AALL0232, ALT elevation was classified into grade > 2 vs. grade 2. Accounting for the significant covariates of age, race, gender, and the induction steroid (Table S6), *PNPLA3* rs738409 was associated with the frequency of grade > 2 ALT elevations in the entire AALL0232 cohort ($P = 0.024$) (Figure S6) and when the analysis was limited to Hispanics ($P = 0.01$), thus independently replicating the polymorphism's association.

Because the *PNPLA3* variant rs738409 has been previously linked to elevated ALT levels and to the development of hepatic steatosis in the general population,⁽¹³⁾ its association with elevated ALT after remission induction therapy suggested that asparaginase may influence blood ALT levels through dysregulation of hepatic lipids. We confirmed that PEGylated *E. coli* asparaginase caused hepatic steatosis in mice, as evidenced by a doubling

in Oil-red O (ORO) staining in livers of mice treated with the drug compared to control (Figure S7).

Discussion

Patients with ALL are at increased risk for the development of toxicities, including hepatotoxicity, which is partly reflected by elevated serum aminotransferase levels.(3) Our objectives were to use a genome-wide approach to determine inherited variation associated with ALT elevation among a racially diverse set of pediatric patients with ALL at a uniform time point (the end of remission induction), adjusting for clinical and other covariates potentially associated with increased ALT levels. Although many ALL medications can contribute to elevations in serum ALT, our multivariate analysis suggested that elevated post-induction chemotherapy ALT levels were linked to asparaginase exposure (Figures 1A&B, Table 3). Patients on Total XVI (who all received PEGylated *E. coli* asparaginase) had higher ALT levels compared to the Total XV patients (who all received the less potent native *E. coli* asparaginase) ($P = 0.041$, Figure 1A, Table 3), despite the fact that patients on Total XV received pre-induction high-dose methotrexate whereas those on Total XVI did not (Table 1, Figure S9), suggesting an effect of asparaginase preparation on hepatotoxicity. Moreover, those who received additional asparaginase during induction had higher ALT levels compared to those that did not receive additional asparaginase ($P = 2.7 \times 10^{-4}$, Figure 1B, Table 3). Interestingly, serum bilirubin at end of induction did not differ by asparaginase exposure (data not shown). Adjusting for treatment, host-related covariates (e.g., age, BMI percentile) and ancestry, we found that the *PNPLA3* rs738409 I148M variant associated with higher ALT levels. This is the identical genetic variant that has previously been linked to ALT elevation in the general population,(10) non-alcoholic fatty liver disease (NAFLD), (12, 14, 15) NAFLD severity,(16) and hepatic fibrosis(15) in genome-wide studies. The effect on NAFLD susceptibility and severity was confirmed by meta-analyses.(17) This variant was also associated with poor chronic hepatitis C treatment response,(18) hepatocellular carcinoma,(19) and insulin resistance.(20) Overall in the discovery cohort of 715 patients, the median increase in ALT was 12 IU/L, or 33.3% of the median of the major allele homozygous (C/C) group, for each minor allele of *PNPLA3* ($P = 0.032$), compared to a 6.0% increase for each allele observed in the general population,(10) suggesting that the effect of the variant is amplified during ALL therapy, and consistent with larger effect sizes for pharmacogenetic traits.(21)

Because of hepatic infiltration of ALL at diagnosis and the prolonged use of potentially hepatotoxic chemotherapy, there is no optimal time to measure “baseline” ALT in ALL.(22) We did examine ALT levels at diagnosis within the SJCRH cohort and showed that serum ALT levels were lower at diagnosis than at end-of-induction ($P < 2.2 \times 10^{-16}$, paired t-test, Figure S8, Table 2), and that *PNPLA3* rs738409 remains a significant risk factor ($P = 4.4 \times 10^{-6}$) for post-induction ALT elevation even after including diagnostic ALT as a covariate.

Patatin-like Phospholipase Domain Containing Protein 3 (PNPLA3 or adiponutrin) is a member of the family of patatin-like lipolytic enzymes involved in triacylglycerol metabolism and signaling.(23) It has been shown that substituting methionine for isoleucine

at position 148 markedly compromised the catalytic velocity of PNPLA3.(24) This is reflected in knock-in mice studies suggesting that the *PNPLA3*I148M variant leads to an increased risk of hepatotoxicity due to accumulation of PNPLA3 on lipid droplets in the liver, a reduction in triglyceride hydrolase activity, and accumulation of hepatic triglycerides.(25) Our finding that the top ranked genetic variation associated with drug-induced liver dysfunction after ALL therapy is identical to one of the top genetic risk factors for constitutive liver dysfunction is remarkable, and has implications for the field of pharmacogenetics and its use to predict drug toxicities.

We confirmed that asparaginase exposure in mice indeed led to an accumulation of hepatic lipids and neutral triglycerides (Figure S7). The results suggest that the mechanism of asparaginase-induced hepatotoxicity may be similar to that of hepatic steatosis, and may be attenuated by therapies that can mitigate hepatic triglyceride accumulation. These results may have particular implications for adults with ALL, in whom hepatotoxicity is common. (26) Interestingly, an early postmortem study from 31 pediatric ALL patients that received native *E. coli* asparaginase found that 87% had developed steatosis after exposure to asparaginase.(27)

We observed that ALT was higher (Table 3, Table S6) in those with ancestry indicating Hispanic status or whites than in blacks, as others have reported.(11) Differences in the risk allele frequency of the *PNPLA3*rs738409 variant mirrored the differences in ALT among race groups (Figure 3B), although this variant did not reach statistical significance in blacks or Hispanics (Figures 3A&S3B), likely due to relatively low power because of small sample sizes for non-whites (with an effect size of 1.36 for the rs738409 variant, our power to detect a significant effect in separate GWASs of blacks and Hispanics was only 0.32 and 0.37, respectively). Although the *PNPLA3*rs738409 variant accounted for some variability in ALT across race groups, race remained a significant risk factor for ALT even after accounting for *PNPLA3*rs738409.

We found that some of the additional association of race with ALT might be due to other genetic variants. For example, black patients carrying the *PIGV*rs12748152 and *GREB1*rs149940960 variants had lower ALT levels compared to non-carriers (Figure 3C&D, Table S3). *PIGV*rs12748152 is an expression quantitative trait locus (eQTL) for *PIGV*, *ARID1A* and *ZDHHC18*,(28, 29) and is also a QTL for cholesterol and triglycerides levels,(30) whereas *GREB1*rs149940960 is in the 3' untranslated region (3'-UTR) of *GREB1*, which regulates circulating levels of chemoattractant protein-1 (CCL2) and insulin-like growth factor binding protein 3 (IGFBF-3).(31) High levels of the latter are associated with adiposity and insulin resistance(32) while the former is a cytokine that recruits immune cells into damaged areas of the liver.(33) Reports suggest that infiltrating immune cells directly kill distressed hepatocytes overloaded with triglycerides. Preventing this infiltration with NF- κ B inhibitors may be another possible avenue of intervention.(34) Although it has not been implicated in NAFLD, *PIGV* is involved in the biosynthesis of glycosylphosphatidylinositol. The *PIGV*rs12748152 variant explained 26.6% of the variability in ALT levels among black patients. However, there was no significant association in those with low West African ancestry ($P = 0.27$), and the variant was actually present at a lower frequency among blacks than other race groups (MAF = 2.0% in blacks,

13.3% in non-blacks). Limitations of these speculations lie in the small sample size of blacks in our cohort and low MAF of these genetic variants among them.

Although we think asparaginase was the most likely cause of post-induction ALT elevations in this study, we acknowledge that other elements of ALL induction therapy can also cause increases in ALT. Interestingly, high-dose methotrexate is well-known to cause acute elevations in ALT,(35) but this was unlikely to have contributed to the end-of-induction ALT (which was measured 42–46 days after a single dose of methotrexate for Total XV), and because high-dose methotrexate was used prior to induction only in Total XV, not Total XVI (Table 1, Figure S9), and yet ALT was higher in patients enrolled on Total XVI compared to Total XV (Figure 1A). Likewise, although thiopurines can cause elevations in ALT, they are more prominent with mercaptopurine (used in Total XV) than with thioguanine (used in Total XVI) (Table 1),(36, 37) and thus unlikely to explain the higher ALT on Total XVI compare to Total XV. The use of PEG-asparaginase at 3000 units/m² on Total XVI results in much higher and more prolonged serum asparaginase concentrations than the doses of native *E. coli* asparaginase that we used on Total XV.(38) Most tellingly, in both Total XV and Total XVI, ALT was higher in the 25% of patients who required extra doses of asparaginase due to high MRD during induction therapy than it was in those who did not receive extra doses (Figure 1B).

The association of *PNPLA3* rs738409 with ALT after induction therapy was replicated in the COG AALL0232 protocol. The replication is somewhat remarkable in that there were several important factors that differed between the discovery and replication cohorts. First, patients on AALL0232 were older than those in the SJCRH discovery cohort (Table 2), and, as expected, exhibited higher ALT grades. Second, discrete ALT values were not available for the AALL0232 cohort, only CTCAE grades; and very few cases of grade 1 and 2 ALT elevation were recorded. Thus, we used linear regression in the discovery cohort but used case (grade > 2)-control (grade = 2) logistic regression for the replication cohort (Figure S6). Finally, BMI was not available for patients in AALL0232. Despite these differences, the effect of *PNPLA3* rs738409 variant was still replicated.

In summary, we have identified that elevated post-induction chemotherapy ALT levels were likely due primarily to asparaginase, and we found that the same *PNPLA3* I148M variant that was previously shown to be the most important determinant of NAFLD in a general population of adults is associated with drug-induced increases in serum ALT. Together, our genetic data from patients with ALL and our preclinical data in mice suggest that hepatotoxicity caused by ALL remission-induction chemotherapy may be mediated by induction of fatty liver and that the toxicity can be modulated by inherited germline polymorphisms that affect hepatic triglyceride homeostasis.

Materials and Methods

Patients

The discovery cohort consisted of patients enrolled on SJCRH protocols Total XV(39) (n = 373, ClinicalTrials.gov identifier: NCT00137111) and Total XVI(40) (n = 342, ClinicalTrials.gov identifier: NCT00549848) for newly diagnosed ALL. The replication

cohort consisted of children with newly diagnosed ALL who were treated on the COG AALL0232 protocol (N = 2,285, ClinicalTrials.gov identifier: NCT00075725) for high-risk B-precursor ALL. Patients included in the genetic association analyses represented 85% (n = 715 of 840) of patients in the SJCRH discovery cohort, and 80% (n = 2285 of 2868) of participants in the COG replication cohort (Figure S1).

Informed consent from the parents or guardians or patients, and assent from the patients, where appropriate, were obtained according to guidelines used at the Institutional Review Boards of the participating institutions. The COG AALL0232 protocol was also approved by the National Cancer Institute.

For the SJCRH cohort, ALT values were graded using CTCAE version 3.0.(41) ALT values between 1.0 ~ 2.5 fold of normal upper limit were assigned as grade 1, between 2.5 ~5.0 fold as grade 2, between 5.0 ~20.0 fold as grade 3, and > 20.0 fold as grade 4. Patient BMI at the time of ALT sample collection was calculated and BMI percentile was determined as described previously with obesity defined as 95th percentile.(42) For the COG AALL0232 cohort, ALT values were graded using the CTCAE version 4.0. ALT values between 1.0 ~ 3.0 fold of normal upper limit were assigned as grade 1, between 3.0 ~5.0 fold as grade 2, between 5.0 ~20.0 fold as grade 3, and > 20.0 fold as grade 4.

Treatment

During induction therapy, patients enrolled on the SJCRH Total XV or Total XVI protocols received asparaginase, prednisone, vincristine, daunorubicin, cyclophosphamide, cytarabine, and thiopurine (Table 1, Figure S9).(39, 40) Total XV patients received native *E. coli* asparaginase (Elspar) at 10,000 units/m² IM, three times weekly for six doses and Total XVI patients received a single dose of PEGylated *E. coli* asparaginase (Oncaspar) at 3,000 units/m² IV. Patients on Total XV with day 19 MRD 5% received three additional doses of native *E. coli* asparaginase over a week's period, and patients on Total XVI with day 15 MRD 1% received an additional dose of PEGylated *E. coli* asparaginase (Table 1, Figure S9). Post-induction samples for ALT and total bilirubin measurement were drawn on day 46 (Total XV) and on day 42 (Total XVI) from the start of remission induction therapy, regardless of clinical suspicion of hepatotoxicity (Figure S9). ALT and total bilirubin were also measured at diagnosis (pre-induction therapy), before the patients were exposed to methotrexate or asparaginase (Figure S9).

During induction therapy, patients enrolled on COG AALL0232 received vincristine, daunorubicin, dexamethasone or prednisone, and PEGylated *E. coli* asparaginase.(43) Patients with day 29 MRD between 1% and 25% received extended induction consisting of additional doses of vincristine, daunorubicin, dexamethasone or prednisone, and PEGylated *E. coli* asparaginase. During consolidation, patients received cytarabine, vincristine, cyclophosphamide, mercaptopurine, and PEGylated *E. coli* asparaginase (Table S7). Measurement of ALT levels was a required observation on COG AALL0232 on day 1 of induction and at the start of consolidation and was performed at other times at the discretion of the treating physician. Sites were required to report all grade 3 and higher ALT elevation.

Genotyping

Germline DNA was genotyped using the Exome-24 BeadChip (Illumina, San Diego, CA) (689 patients in SJCRH cohort and 2,382 in AALL0232 trial) and either the Affymetrix Genome-Wide Human SNP Array 6.0 or the Affymetrix Human Mapping 500K Array Set (839 in SJCRH cohort and 2,666 for AALL0232 trial), as described previously.(44) MaCH-Admix (<http://www.unc.edu/~yunmli/MaCH-Admix>) was used to impute the remaining untyped SNPs using the 1000 Genomes Project (<http://www.1000genomes.org>) as described elsewhere.(45, 46) Germline DNA from 552 patients from the SJCRH cohort was also subjected to whole exome sequencing. For this, genomic libraries were prepared using the NimbleGen SeqCap EZ Exome Enrichment Kit v2.0 (Roche NimbleGen, Madison, WI) and sequencing was performed using the HiSeq 2000 Sequencing System (Illumina, San Diego, CA). The Hardy-Weinberg equilibrium (HWE) test was performed using PLINK on SNPs with a MAF \geq 1% and among patients of European ancestry. SNPs with genotyping call rates $<$ 95% and SNPs that were not in HWE (P values $<$ 0.001) were excluded from the association analysis (Figure S1).

Genetic Ancestry

The genetic ancestry of patients was determined using STRUCTURE (<http://pritchardlab.stanford.edu/structure.html>) as previously described,(47) and percent ancestry was treated as a continuous variable or used to assign patients to race groups: whites were defined as having $>$ 90% Northern European ancestry, blacks as having $>$ 70% West African ancestry, Hispanics as having Native American ancestry $>$ 10% and greater than their African ancestry percentage, Asians as having $>$ 90% East Asian ancestry, and others as those whose ancestry was outside the above boundaries. In the SJCRH cohort, patients of Asian ancestry were grouped with others due to their small sample size.

Preclinical *In vivo* Asparaginase-induced Hepatotoxicity Studies

Male Balb/cJ mice (4-week-old) (Jackson Laboratories, Bar Harbor, ME) received 1,500 IU/kg of PEGylated *E. coli* asparaginase by intraperitoneal injection twice weekly for 6 weeks. Livers were collected at the end of the experiment and snap frozen in liquid nitrogen. Frozen livers were immersed in OCT media and 4 μ m sections were stained with Oil-red O. The staining intensity was quantified using the Aperio Image analysis system (Aperio, Vista, CA). Three 1 mm² square regions for each sample were analyzed and the number of strongly positive pixels in the regions was used to compare experimental and control samples. All experiments involving the use of mice were approved by the Institutional Animal Care and Use Committee (IACUC) of St. Jude Children's Research Hospital. Mice were housed in an American Association of Laboratory Animal Care-accredited facility. The study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (NIH).

Data Analysis

Univariate and multivariate linear regression was performed using a general linear model with R statistical software (version 2.13.2) to identify clinical risk factors associated with post-induction ALT levels as a continuous variable. For the COG AALL0232 replication

cohort, associations between clinical risk factors and the frequency of grade > 2 hepatotoxicity were determined using Fisher's exact test in R. The genome-wide association analysis between SNP genotypes and ALT was performed in PLINK 1.9 (<https://www.cog-genomics.org/plink2>) using linear or logistic regression as appropriate, adjusting for significant covariates and assuming an additive genetic model.(48) Within each race group defined above (whites, blacks, and Hispanics), the first five principal components were calculated using GCTA version 1.26.0 (<http://cns.genomics.com/software/gcta/>) with genotype data acquired from the Affymetrix arrays. Gene-level association was done using the sequence kernel association test (SKAT) as implemented by PLINK/SEQ (<https://atgu.mgh.harvard.edu/plinkseq/>). Linkage disequilibrium based variant pruning was performed using PLINK. To assess the proportion of variability in ALT accounted for by specific genomic variants, a general linear model was used.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Study Highlights

What is the current knowledge on the topic?

Hepatotoxicity is a frequent complication leading to treatment interruptions, and several drugs used in ALL remission induction are associated with the development of hepatotoxicity, including asparaginase. There are no prior genome-wide association studies for hepatotoxicity in patients with ALL.

What question did this study address?

This study evaluated patient- and treatment- related variables associated with hepatotoxicity in two ancestrally diverse groups of pediatric ALL patients, using ALT levels measured at the end of remission induction as an indicator. The study identified genetic variants that contribute to the risk of hepatotoxicity on a genomic level.

What this study adds to our knowledge?

PNPLA3 rs738409 reached genome-wide significance as a risk factor of hepatotoxicity in patients that completed ALL remission induction. Asparaginase, especially in its PEGylated preparation, is likely a major contributor to higher ALT levels during this treatment phase.

How this might change clinical pharmacology or translational science?

The study accentuated asparaginase hepatotoxicity during ALL induction, and identified a pharmacogenetic risk factor in this setting, *PNPLA3* rs738409, which overlaps with that for non-alcoholic fatty liver disease.

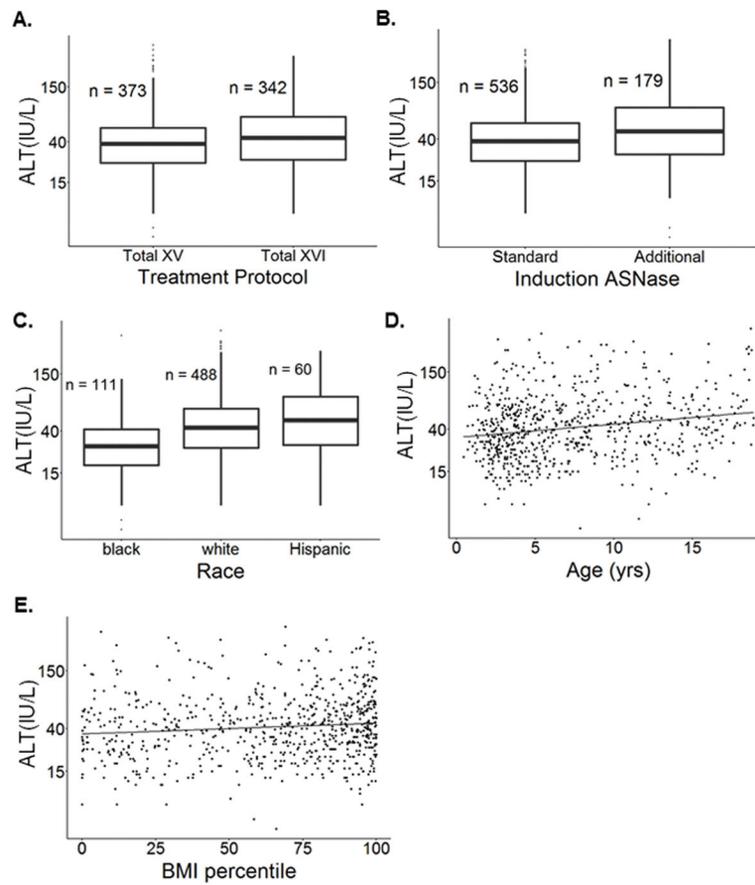


Figure 1. ALT level variation in pediatric ALL patients

(A) Total XVI patients receiving PEGylated *E. coli* asparaginase had higher ALT levels compared to Total XV patients receiving native *E. coli* asparaginase ($P = 3.0 \times 10^{-3}$). (B) Among Total XV and Total XVI patients, those receiving additional doses of asparaginase during induction due to a higher ALL burden at day 15–19 had higher ALT levels compared to patients receiving fewer doses of asparaginase ($P = 3.5 \times 10^{-6}$). (C) Among Total XV and Total XVI patients, whites ($P = 3.3 \times 10^{-10}$) and Hispanics ($P = 3.3 \times 10^{-7}$) had higher ALT levels compared to blacks. (D) ALT levels were correlated with age (yrs) in the SJCRH discovery cohort ($N = 715$, $r = 0.20$, $P = 9.4 \times 10^{-8}$). (E) ALT levels were positively correlated with BMI percentile in the SJCRH discovery cohort ($N = 715$, $r = 0.11$, $P = 5.2 \times 10^{-3}$). The P values indicated are from univariate analyses (Table S1).

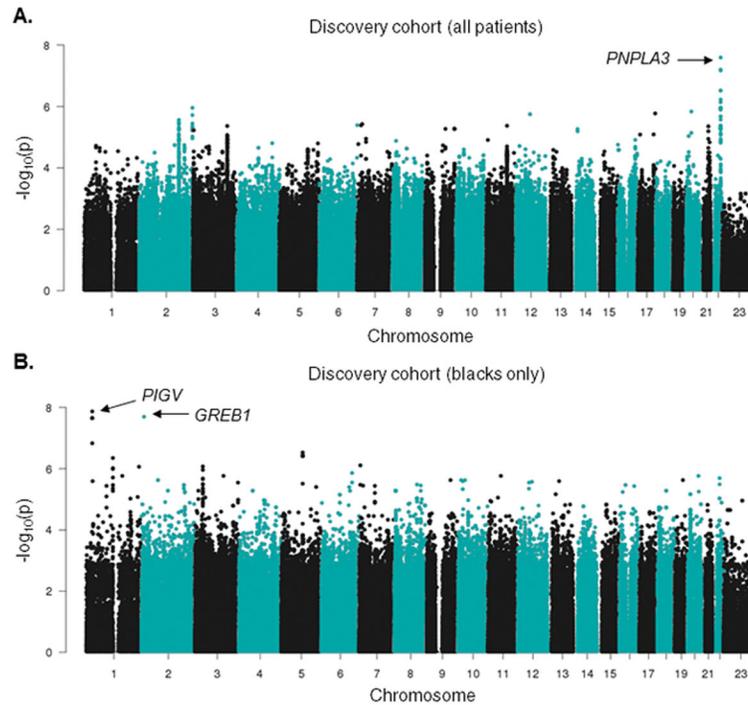


Figure 2. Manhattan plots of inverse P values for genome-wide SNP associations with ALT levels (A) Association between SNPs and natural logarithm ALT levels versus each chromosome for Total XV and XVI patients (N = 715). The analysis identified a variant in *PNPLA3* (rs738409) associated with ALT levels. (B) Association between SNPs and ALT levels versus each chromosome for black patients in Total XV and XVI patients (n = 111). A variant near *PIGV* and a variant in 3'-UTR of *GREB1* were associated with ALT levels with genome-wide significance.

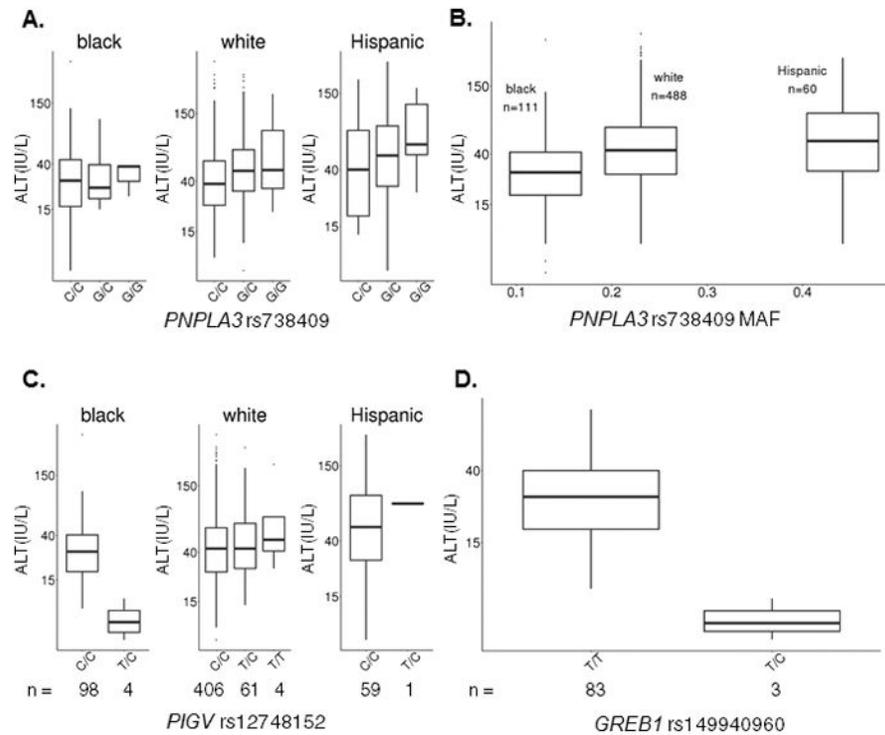


Figure 3. Post-induction ALT levels varied by genotype in 715 SJCRH Total XV and XVI patients

(A) Boxplots of ALT levels by *PNPLA3* rs738409 genotype in blacks ($n = 111$, $P = 0.20$), whites ($n = 488$, $P = 1.5 \times 10^{-6}$), and Hispanics ($n = 60$, $P = 0.058$) from the SJCRH discovery cohort. (B) Boxplots of ALT levels by *PNPLA3* rs738409 MAF in blacks, whites and Hispanics from the SJCRH discovery cohort. (P values of whites and Hispanics vs. blacks are 3.3×10^{-10} and 3.3×10^{-7} respectively in univariate analysis.) (C) Boxplots of ALT levels by *PIGV* rs12748152 genotype in blacks ($n = 102$, $P = 1.3 \times 10^{-8}$), whites ($n = 471$, $P = 0.14$), and Hispanics ($n = 60$, $P = 0.59$) from the SJCRH discovery cohort. (D) Boxplots of ALT levels by *GREB1* rs149940960 genotype in blacks ($n = 86$, $P = 2.0 \times 10^{-8}$) from the SJCRH discovery cohort. The boxes depict median with upper and lower quartiles.

Table 1

Chemotherapeutic agents used during Total XV and Total XVI remission induction therapy.

Agents	Dosage and Routes and days of induction therapy	Number of doses	Differences between Total XV and Total XVI
Prednisone ^d	40 mg/m ² /day, PO, T.I.D.; Total XV: days 5–32 Total XVI: days 1–28	84	-
Vincristine	1.5 mg/m ² /weekly, IV; Total XV: days 5, 12, 19, 26 Total XVI: days 1, 8, 15, 22	4	-
MTX ^b	1 g/m ² , IV, over a 4 or 24 hour infusion	1	Window MTX was not included on the Total XVI protocol
Daunorubicin	25 mg/m ² /weekly, IV; Total XV: days 5 & 12 Total XVI: days 1 & 8	2	-
ASNa ^c	Total XV: L-ASNa, 10,000 units/m ² , IM, thrice weekly; days 6, 8, 10, 12, 14, 16 Total XVI: PEG-ASNa, 3,000 units/m ² , IV; day 3	L-ASNa: 6 PEG-ASNa: 1	Preparation, dosage, and route
Cyclophosphamide	1,000 mg/m ² , IV; Total XV: day 26 Total XVI: day 22	1	Cyclophosphamide was intensified for standard/high risk patients with 5% of blast on day 15 of induction for Total XVI patients ^d
Cytarabine	75 mg/m ² /dose, IV; Total XV: days 27–30 & 34–37 Total XVI: days 23–26 & 30–33	8	-
Thiopurine	Total XV: mercaptopurine, 60 mg/m ² /day, PO; days 26–39 Total XVI: thioguanine ^e , 60 mg/m ² /day, PO; days 22–35	14	Total XVI received thioguanine; Total XV received mercaptopurine

^aOral prednisone was substituted with methylprednisolone at 20 mg/m²/day, IV for patients that could not tolerate oral administration.^bTotal XV patients were randomized to receive short (4 hour) or prolonged (24 hour) methotrexate infusion.^cAdditional 3 doses of L-ASNa were given if day 19 MRD 5% on days 19, 21, 23 to Total XV patients. Additional PEG-ASNa dose was given if day 15 MRD 1% on day 15 to Total XVI patients.^dTotal XVI patients with day 15 MRD 5% received cyclophosphamide at 300 mg/m², IV, q12 hours for 4 doses on days 22–23.^eTPMT heterozygote or TPMT deficient Total XVI patients received mercaptopurine at 60 mg/m²/dose, PO instead of thioguanine.

Abbreviations: PO, by mouth; IV, intravenous; IM, intramuscular; T.I.D., three times a day; MRD, minimal residual disease; MTX, methotrexate; ASNa, asparaginase

Table 2

Demographics of ALL patients with serum ALT measurement data.

Characteristics	Treatment protocol		
	SJCRH (Total XV)	SJRCH (Total XVI)	COG AALL0232 ^d
Sample Size (n)	373	342	2285
Age (n 10, mean in years)	102 (27.4%), ^a 7.1	92 (26.9%), ^a 7.2	1,468 (64.3%), ^a 10.6
Gender (n male)	206 (55.2%) ^a	198 (57.9%) ^a	1,244 (54.4%) ^a
Race group^b			
Whites	261 (70.0%) ^a	227 (66.4%) ^a	1,275 (55.8%) ^a
Blacks	64 (17.2%) ^a	47 (13.7%) ^a	139 (6.1%) ^a
Hispanics	24 (6.4%) ^a	36 (10.5%) ^a	601 (26.3%) ^a
Others	24 (6.4%) ^a	32 (9.4%) ^a	270 (11.8%) ^a
Diagnostic ALT (IU/L)^c			
Median (range)	21 (3 – 3866)	20 (4 – 2442)	NA
95% confidence interval	22.9 – 27.9	22.4 – 27.9	NA
Post-induction ALT (IU/L)			
Median (range)	38 (4 – 416)	44 (7 – 321)	NA
95% confidence interval	35.5 – 40.9	41.3 – 48.4	NA

^aThe percentage of patients with the characteristic described in the first column.^bWhites were defined as having > 90% Northern European ancestry (CEU), blacks as having > 70% West African ancestry (YRI), Hispanics as having Native American ancestry > 10% and greater than their African ancestry percentage, and others as those whose ancestry was outside the above boundaries.^cDiagnostic ALT (pre-induction therapy) was evaluable in 337 of 373 Total XV patients and 321 of 342 Total XVI patients, and ALT levels were lower at diagnosis than at end-of-induction ($P < 2.2 \times 10^{-16}$, paired t-test for 658 patients evaluable at both times, see also Figures S8&S9).^dNo discrete ALT levels were available for COG AALL0232 patients.

NA, not available

Table 3

Multivariate analyses of covariates associated with alanine transaminase levels during Total XV and Total XVI remission induction therapy.

Covariate	Without inclusion of <i>PNPLA3</i> rs738409			Including <i>PNPLA3</i> rs738409		
	Effect size ^a	P value	% of ALT level variability explained	Effect size ^a	P value	% of ALT level variability explained
Induction AS/Nase: Additional vs. standard schedule	1.26 (1.11–1.42)	2.7×10 ⁻⁴	1.7%	1.27 (1.12–1.43)	1.1×10 ⁻⁴	1.8%
Gender: Male vs. female	1.08 (0.97–1.20)	0.16	NA	NA	NA	NA
Age: Continuous variable	1.03 (1.02–1.04)	1.4×10 ⁻⁶	2.9%	1.03 (1.02–1.04)	8.6×10 ⁻⁸	3.4%
Race group^b:						
Whites vs. blacks	1.70 (1.48–1.96)	8.7×10 ⁻¹³		1.62 (1.41–1.87)	3.7×10 ⁻¹¹	
Hispanic vs. blacks	1.77 (1.43–2.20)	2.6×10 ⁻⁷	7.2%	1.57 (1.26–1.95)	6.6×10 ⁻⁵	5.3%
Others vs. blacks	1.52 (1.21–1.90)	3.2×10 ⁻⁴		1.41 (1.14–1.76)	2.0×10 ⁻³	
Protocol: Total XVI vs. Total XV	1.11 (1.00–1.24)	0.041	0.49%	1.12 (1.01–1.24)	0.025	0.58%
BMI percentile: Continuous variable	1.29 (1.09–1.52)	2.9×10 ⁻³	1.3%	1.28 (1.09–1.50)	2.7×10 ⁻³	1.0%
<i>PNPLA3</i> rs738409 genotype: Numeric variable ^c	NA	NA	NA	1.29 (1.18–1.40)	1.2×10 ⁻⁸	3.8%

^aThe effect size refers to the exponent of the beta coefficient estimated from the linear regression. Numbers in parentheses indicate 95% confidence interval.

^bWhites were defined as having > 90% Northern European ancestry, blacks as having > 70% West African ancestry, Hispanics as having Native American ancestry greater than 10% and Native American ancestry was greater than their percent African ancestry, and others were those whose ancestry was outside the above boundaries.

^cGenotype of SNP was treated as numeric variable, which equals to the number of minor allele in the genotype of each patient.
NA, not applicable

Table 4

Top 20 variants associated with post-induction ALT in the combined SJCRH Total XV and Total XVI cohort.

rsID	Hg19 position	Major allele ^a	Minor allele ^a	Platform	P value	Effect size ^b	Confidence interval	Gene	Function annotation
rs738409	22:44324727	C	G	exomechip	2.52E-08	1.29	1.04–1.58	PNPLA3	missense
rs144104656	2:119418867	A	G	imputed	1.60E-07	8.04	1.36–47.52	EN1	intergenic
rs2294915	22:44340904	C	T	imputed	5.72E-07	1.26	1.03–1.53	PNPLA3	intronic
rs147481775	22:32225363	C	A	imputed	5.77E-07	4.66	1.22–17.88	DEPDC5	intronic
rs74709575	13:38615256	A	C	imputed	1.23E-06	4.52	1.19–17.19	TRPC4	intergenic
rs17857135	17:78262161	T	C	exomeseq	1.68E-06	0.78	0.63–0.97	RNF213	missense
rs112769843	12:62308665	C	G	imputed	1.76E-06	0.54	0.32–0.94	FAM19A2	intronic
rs11696756	20:22982087	G	A	imputed	2.59E-06	1.39	1.03–1.86	SSTR4	intergenic
rs12530134	6:170919470	G	A	exomechip	3.37E-06	1.36	1.03–1.80	PDCD2	intergenic
rs117483095	9:132784775	C	T	imputed	3.43E-06	1.62	1.05–2.50	FNBP1	intronic
rs113941845	2:239501832	C	A	imputed	4.01E-06	1.24	1.02–1.51	ASB1	intergenic
rs145145722	7:17243557	T	C	imputed	4.03E-06	0.42	0.19–0.92	AHR	intergenic
rs11020478	11:93409448	A	G	imputed	4.30E-06	1.21	1.02–1.43	KIAA1731	intronic
rs1348850	2:178418575	G	A	exomechip	4.31E-06	1.22	1.02–1.46	TTC30B	intergenic
rs7024024	9:92210595	A	T	imputed	4.37E-06	0.82	0.68–0.98	GADD45G	intergenic
rs149940960	2:11781618	T	C	exomeseq	4.41E-06	0.16	0.03–0.85	GREB1	3' -UTR
rs143969105	12:58201465	A	G	exomeseq	4.48E-06	8.68	1.21–62.08	TSFM	missense
rs75870332	14:27753539	T	A	imputed	4.88E-06	1.51	1.04–2.19		intergenic
rs58800477	6:20759577	A	G	imputed	4.94E-06	4.10	1.13–14.91	CDKAL1	intronic
rs738409	22:44324727	C	G	exomechip	2.52E-08	1.29	1.04–1.58	PNPLA3	missense

^a Alleles were assigned as major or minor ones based on their frequency in the cohort of our study.

^b The effect size refers to the exponent of the beta coefficient of the minor allele estimated from the linear regression in PLINK. An effect size greater than 1 indicates the minor allele is the risk allele.