

APOE genotype and Alzheimer's immunotherapy

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Alzheimer's disease (AD) is a chronic neurodegenerative disease and the most common cause of dementia. AD risk is foremost modified by allelic composition of the *APOE* gene encoding apolipoprotein (apo) E-brain's main lipid carrying protein. Emerging evidence suggests that *APOE* genotype also may modulate efficacy and safety of AD immunotherapy, which is under development as a potential disease-modifying treatment.

There are three allelic forms of the *APOE* gene $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$. AD frequency reaches 91% and 47% in *APOE* $\epsilon 4$ homo- and heterozygous carriers compared to 20% in non-carriers, respectively; while the age of dementia onset averages 68 and 76 years in *APOE* $\epsilon 4$ homo- and heterozygous carriers compared to 84 years in non-carriers, respectively [1]. A protective effect of the *APOE* $\epsilon 2$ allele also can be seen among *APOE* $\epsilon 4$ non-carriers. *APOE* genotype effecting AD predisposition results from a combination of disease specific and constitutional biological effects differentially modulated by apo E isoforms encoded by various *APOE* alleles [2]. Apo E isoforms diversely affect rate of soluble β -amyloid ($A\beta$) clearance from the brain interstitial space and also promote formation of $A\beta$ plaques and vascular deposits, what effects a down-stream neurodegenerative cascade involving neurofibrillary pathology, inflammatory microglia response, and synaptic and neuronal loss. *APOE* $\epsilon 4$ carriers typically show much greater load of $A\beta$ deposits compared to non-carriers. Constitutional effects of apo E implicated in AD pathogenesis concern its involvement in microglia phagocytic function, synaptic plasticity and neuronal network repair. *APOE* $\epsilon 4$ carriers have an attenuated reparative response to the neurodegenerative cascade induced by $A\beta$ accumulation they are more prone to develop.

Development of anti- $A\beta$ immunotherapy as a disease-modifying treatment is being actively pursued. One tested approach concerns intravenous administration of monoclonal antibodies (mAbs) recognizing antigens exposed in brain deposited $A\beta$. A modest fraction of such mAbs permeates the blood-brain barrier and upon binding $A\beta$ plaques facilitates their clearance by macrophage transformed microglia. Bapinezumab was the first humanized mAb having this modus operandi, which came to clinical development. Its phase 2 trial evidenced potential effect of *APOE* genotype on efficacy and safety. Significant treatment effects on cognitive and functional endpoints were found in *APOE* $\epsilon 4$ non-carriers but not in *APOE* $\epsilon 4$ carriers [3], while amyloid related

imaging abnormalities (ARIA) (including vasogenic edema [ARIA-E] and microhemorrhages [ARIA-H]) occurred seven and three times more frequently in *APOE* $\epsilon 4$ homo- and heterozygous carriers compared to non-carriers, respectively [4]. Exact pathomechanism of ARIA remains elusive but it is linked to immune response against perivascular $A\beta$ causing transient increase in vascular wall permeability. Roughly 20% of ARIA-E affected patients reported clinical signs and symptoms. Enrolment to subsequent phase III trials was prospectively segregated based on *APOE* $\epsilon 4$ carrier status and the maximal bapinezumab dose was reduced from 2 mg/kg tested in phase 2 trial to 0.5 mg/kg and 1 mg/kg in *APOE* $\epsilon 4$ carrier and non-carrier groups, respectively [5]. Although dose reduction was a rationale measure to manage ARIA risk, in retrospect it can be viewed as one of several reasons these trials failed to meet efficacy endpoints.

Aducanumab is a newer anti- $A\beta$ mAb, which like bapinezumab binds deposited $A\beta$ exerting effector microglia response but can be tolerated in significantly

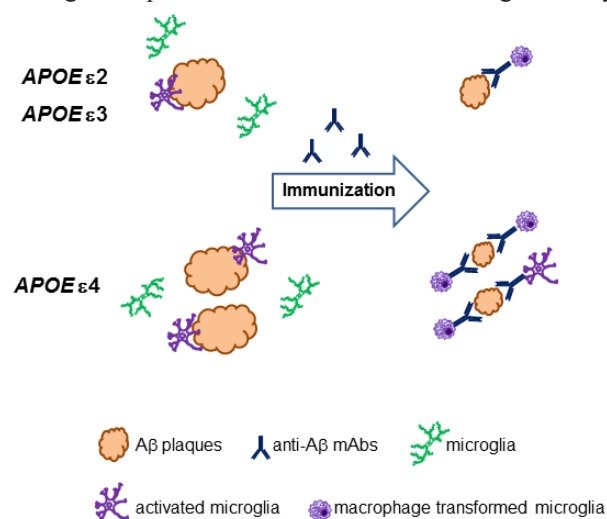


Figure 1: Interaction between *APOE* genotype, $A\beta$ deposition, and microglia. The *APOE* $\epsilon 4$ allele is associated with greater β -amyloid ($A\beta$) deposition than *APOE* $\epsilon 2$ and $\epsilon 3$ alleles. $A\beta$ parenchymal plaques attract microglia cells and cause their peri-plaque activation, which is proportional to $A\beta$ plaque load across all *APOE* genotypes. Therapeutic anti- $A\beta$ monoclonal antibodies (mAbs) bind deposited $A\beta$ and activate microglia cells to clear $A\beta$ through Fc receptor-mediated phagocytosis. The *APOE* $\epsilon 4$ allele is associated with greater microglia activation than *APOE* alleles $\epsilon 2$ and $\epsilon 3$ resulting in greater clearance of $A\beta$ plaque load during $A\beta$ -directed passive immunization.

higher doses [6]. Its phase 1b clinical trial showed dose dependent reduction of A β deposits. The maximal tested dose was 10 mg/kg administered every four weeks, while bapinezumab was infused quarterly. Unlike previous AD clinical trials, aducanumab development focuses on prodromal AD patients in attempt to contain A β pathology early and attenuate down-stream neurodegenerative cascade. In aforementioned phase 1b trial, 10 mg/kg aducanumab dose showed strong effect on cognitive endpoints. ARIA remains main adverse effect of aducanumab and its risk is mediated by the *APOE* ϵ 4 allele. Thus, for two enrolling phase 3 trials patients are prospectively segregated based on *APOE* ϵ 4 status. *APOE* ϵ 4 carriers are randomized to either placebo, 3 or 6 mg/kg aducanumab doses while non-carriers to either placebo, 6 or 10 mg/kg doses.

Given emerging evidence concerning effects of *APOE* genotype on efficacy and safety of AD immunotherapy we took “bedside-to-bench” approach re-testing outcomes of passive immunization in APP_{SWE}/PS1_{DE9} AD transgenic mice made homozygous for each of human *APOE* alleles. We used 10D5 anti-A β mAb with similar modus operandi to that of bapinezumab or aducanumab. The same 10D5 mAb dose (10 mg/kg/week) was used across all *APOE* genotypes. *APOE* ϵ 4 mice showed the greatest reduction in A β deposits and the most robust microglia activation adjusted for A β plaque load, which for the first time evidenced that the *APOE* ϵ 4 allele mediates stronger microglia response to anti-A β immunotherapy and enhances microglia phagocytic effect [7] (Figure 1). As aducanumab phase III clinical trials employ A β PET imaging, it would be interesting to see whether reduction in A β load with comparable dose is higher in *APOE* ϵ 4 carriers than non-carrier, counteracting typically higher A β plaque load of the former.

We also investigated effects of *APOE* genotype on vascular complications of anti-A β immunotherapy. Using μ MRI we showed occurrence of new microhemorrhages (ARIA-H) in *APOE* ϵ 4 mice undergoing 10D5 mAb treatment but we found no evidence of “vasogenic edema” (ARIA-E) [8]. Though ARIA-E remains the most troublesome and dose-limiting adverse effect of anti-A β immunotherapy its nature remains obscure largely

due to absence of animal models allowing to dissect its pathogenesis. Postmortem analysis of perivascular microhemorrhages across *APOE* genotypes showed their greatest incidence in *APOE* ϵ 2 mice evidencing for the first time the *APOE* ϵ 2 allele as a risk factor for A β immunotherapy related microhemorrhages [7]. When translating these observations from transgenic mice back to humans one needs to be mindful that ϵ 2/ ϵ 2 genotype is rare among AD patients, nevertheless careful monitoring of *APOE* ϵ 2 carriers during clinical trials of anti-A β immunotherapy may effect a reduction in ARIA-H events.

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