

# **HHS Public Access**

Author manuscript *Am J Reprod Immunol.* Author manuscript; available in PMC 2017 July 26.

Published in final edited form as: *Am J Reprod Immunol.* 2017 May ; 77(5): . doi:10.1111/aji.12652.

# Mapping out p38MAPK

# Elizabeth A. Bonney, MD, MPH

<sup>1</sup>Department of Obstetrics, Gynecology and Reproductive Sciences, University of Vermont, Burlington, VT, United States

# Abstract

In order to generate new hypotheses, sometimes a "systems" approach is needed. In this review I focus on the mitogen activated kinase p38 because it has been recently shown to play an important role in the developmental programing and senescence of normal and stressed reproductive tissues. What follows is an overview of 1) pathways of p38 activation and their involvement in basic biological processes 2) evidence that p38 is involved in the homeostasis of reproductive tissues 3) how focus on p38 can be incorporated into investigation of normal and stressed pregnancies. Existence of excellent reviews will be mentioned as well as relevant animal models.

# Keywords

MAPK14 p38 pregnancy; reproduction; immunology

# 1. Why p38?

Efforts to delineate the molecular pathways critical to normal parturition and preterm birth have highlighted the importance of inflammation<sup>1–6</sup>. Further, analysis of the molecular events down-stream of senescence, oxidative stress, infection, and metabolic dysregulation in reproductive and other tissues have suggested that these events all involve the professional mediator p38 in the generation of inflammatory responses<sup>7–11</sup>. This and other data gives support to the development of a "p38-centric" (Fig. 1) view of cellular homeostasis that is relevant to reproduction, including normal and premature parturition.

# 2. Introduction to p38

Extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinases (JNK) and p38, are members of a serine/threonine kinases family with related molecules present in unicellular organisms, and thus have a long evolutionary history<sup>12, 13</sup>. These molecules are key in mediating cellular responses and are activated through the phosphorylation of a Thr-X-Tyr motif by upstream mitogen activated protein kinases (MAPK). While ERK pathways are typically associated with growth or proliferation signals, JNK and p38 are associated with the response to environmental and infectious/inflammatory stress. There are different

Corresponding author: Elizabeth A. Bonney, Division of Reproductive Science Research, Department of obstetrics, Gynecology, and Reproductive Sciences, University of Vermont College of Medicine, Given Building Rm C-246, 89 Beaumont Avenue, Burlington, Vermont 05405, USA. ebonney@uvm.edu.

pathways for activation of ERK, JNK, and p38, but it is likely that there is cross regulation between the pathways<sup>14</sup>, and frequently stimuli will activate more than one pathway. The p38s comprise a group of 4 proteins p38a, p38 $\beta$ , p38 $\gamma$ , and p38 $\delta$  that represent different branches on a phylogenetic tree and are each conserved among species <sup>15</sup>. The MAPK 14 family includes p38a in mice, rats, humans Xenopus laevis and zebra fish. The MAPK11 family similarly comprises p38β, while the MAPK 13 family corresponds to more loosely related p388, p38 $\gamma$ , also called ERK6 is part of the MAPK 12 family<sup>15</sup>. Of the p38 MAPK, p38a is most widely expressed and studied. While p38  $\beta$  and  $\delta^{16}$ , are expressed in immune cells,  $p38\gamma$  is not and seems to have a highly restricted expression that as yet is not completely elucidated. From here on discussion will focus on p38a and for convenience it will be referred to as p38. This molecule is activated by three upstream kinases MKK3, MKK4, and MKK6. While MKK3 and MKK6 are p38's main MAPKKs<sup>17</sup> specificity of the outcome of external stimuli as noted above, is determined by cell and tissue context and other associated molecules. For example, in mice, all three are involved in activation after ultra violet radiation, while only MKK3 and MKK6 are critical for TNF-a mediated activation.<sup>18</sup> Further upstream to the MKKs are the MEKKs 1 to 4 (also called the MAP3Ks for MAP kinase kinase kinases), MLK2/3, and ASK1 (Apoptosis signaling kinase 1), and TAK-1 (TGF- $\beta$  activated protein kinase 1). Other upstream molecules will likely be delineated with further study. Another point of specificity in p38 regulation comes from the site of phosphorylation. Sites for phosphorylation pf p38 include Thr180 and Tyr182 and another level of control is exemplified by cyclophilin-dependent isomerization of p38 that modifies the chance that upstream kinases can phosphorylate  $p38^{19}$ .

#### 3. UPSTREAM: how does p38 get activated?

There are several "stresses" that result in p38 activation. <sup>20</sup>

#### ER stress

Response to a stimulus includes generation of new proteins. The proper folding and transport of these proteins is monitored by the cell. The presence of excess unfolded protein generates the unfolded protein response that includes upregulation of PERK (Protein kinase (RNA)-like endoplasmic reticulum kinase), IRE1 (Inositol requiring protein 1) and ATF6 (Activating transcription factor 6), among others. This leads to an increase in XBP-1 (X-box binding protein 1) and free BiP (binding Ig protein) that can assist in chaperoning and folding the proteins. If however, such ER stress is not relived, this can lead to the interaction between ASK1 and TRAF2 and auto-phosphorylation of ASK1 which can feed into the pathway that causes MKK activation and leads to activation of p38<sup>21</sup>. Advanced glycation end products (AGEs), produced by the non-enzymatic glycation of macromolecules can accumulate in different tissues and mediate p38 activation through the generation of ER stress.<sup>22</sup> ER stress induced by infection can also elevate p38 activation.<sup>23</sup>

#### Oxidative stress

Oxidative stress leads to the activation of p38 in many tissues. For examples, direct application of cigarette smoke extract to fetal membranes leads to activation of p38.<sup>24</sup> In an endothelial cell line, lipid peroxidation can lead to activation of Src which in turn leads to

the activation of p38. A first trimester human trophoblast cell line responds to hypoxia followed by reoxygenation by increasing growth arrest and DNA damage-inducible 45 alpha (Gadd45alpha) expression which in turn activates p38. Data such as this supports the growing hypothesis of the close link between inflammation and oxidative stress and further places p38 as playing a role in the crosstalk between the two processes.

#### Inflammatory stress

Signaling through receptors for pathogen or "Danger" associated molecular patterns, such as the Toll like receptors leads to activation of the MAP3 Kinases, including TAK-1 (TGF-b activated protein kinase 1) which then activates one of the downstream MAP2 Kinases such as MKK3/6 or MKK4. This in turn activates p38. Cytokines involved in inflammatory processes, such as IL-1 and Tumor necrosis factor- $\alpha$  bind to their cognate receptors and via the small GTPases (e.g. RAC1 and CDC42) also lead to p38 activation (reviewed in<sup>25</sup>). Specific receptors on different types of inflammatory cells mediate their functions via activation of p38, and this includes the T cell receptor. Binding of this receptor includes activation of ZAP70 and LCK, and ultimately activation of p38.<sup>26</sup> Viral infection can also lead to p38 activation.<sup>27</sup> In neutrophils, TGF- $\beta$ , which with IL-6 helps to generate the TH-17 response<sup>28</sup>, activates p38 and MKK2 and promotes chemotaxis<sup>29</sup>.

#### **Metabolic stress**

This is a likely more broadly inclusive family of cellular stress that encompasses oxidative stress as well as other processes. Insulin, for example exerts an anti-proliferative and hypertrophic effect in an extra villous cell line, and this may be partially reversed by an inhibitor of p38,<sup>30</sup> suggesting a role for this molecule. Exposure of kidney cells to high salt conditions leads to p38 activation.<sup>31</sup> Embryonic stem cells starved of leukemia inhibitory factor die by apoptosis, and this is mediated by activated p38 and increased cleavage of caspase 3.<sup>32</sup>

Earlier studies have also delineated the potential role of p 38 in term trophoblast cell response to placenta derived growth factor and protection from serum withdrawal-induced apoptosis.<sup>20</sup> Other growth factors such as VEGF and erythropoietin all trigger p38 activation in tissue specific manner<sup>33</sup>.

#### DNA damage

Ionizing radiation and other inducers of DNA double strand breaks lead immediately to histone modifications and sensors of DNA damage, including the PCNA-like complex facilitate the activation of upstream kinases such as Ataxia telangiectasia mutated (ATM), and Ataxia-telangiectasia and rad3 (ATR) <sup>34, 35</sup>. This in turn can trigger phosphorylation p38 and cause its nuclear translocation.<sup>36</sup> ATM in addition can work with NEMO (NF- $\kappa$ B essential modifier) to release NF- $\kappa$ B from I $\kappa$ B kinase  $\alpha$  and  $\beta$ , Unprotected chromosomes induce DNA damage checkpoint cascades and has been associated with increased activation of p38<sup>37</sup> and may explain the association between telomere shortening and the presence of phosphorylated p38.<sup>38, 39</sup>

## 4. DOWNSTREAM: what happens when p38 gets activated?

#### p38 and inflammation

Several effector pathways are induced by the activation of p38. In the so-called 'classical' p38 pathway, phosphorylated p38 in the cytosol can mediate the activation and movement of several transcription factors including AP-1 and SP-1 to the nucleus which could be followed by transcription of several inflammatory cytokines and mediators<sup>33</sup>. A particular example is that p38 activates mitogen- and stress-activated protein kinases, MSK1 and MSK2, which can phosphorylate the trans-activating p65 subunit of the NF- $\kappa$ B complex at Ser276 and thus potentiate NF- $\kappa$ B signaling.<sup>40</sup> While in the cytoplasm, activated p38 can also mediate its effects through the activation of other specific proteins including MAP-kinase activated protein kinase 2 (MAPKAPK2, MK2) and MAPKAPK5 which in turn can phosphorylate heat shock (e.g. 25/27) and other proteins in and out of the nucleus <sup>41</sup> Downstream molecules of p38 activation can also stimulate the transcription factor STAT3 which can potentiate the actions of NK cells and IL-6<sup>42</sup>.

#### Autophagy<sup>43</sup>

This is an important mechanism for homeostasis in several tissues in response to stresses such as inflammation, and evidence of dysregulation of this process has been associated with preterm birth in humans<sup>44</sup>. p38 can up or down-regulate autophagy<sup>45, 46</sup>. p38 can block autophagy through the mechanistic target of rapamycin (mTOR) pathway and pathways independent of this molecule<sup>45</sup>

#### Senescence

Senescence occurs as the result of irreversible cell cycle arrest, and is associated with DNA fragments, and stereotypic histologic changes and inflammation<sup>40</sup>. Activated p38 can arrest cell division by multiple pathways, including activation of p53<sup>47</sup> and in the so-called 'non classical' pathway p38 can move to the nucleus itself and enhance the activity of molecules such as p21 and p16 to block cell cycling<sup>10</sup> at both the G2M<sup>48</sup> and G1/S checkpoints.<sup>47</sup> Genomic instability evoked by senescence triggers epigenetic changes, e.g. release of HMGB1 proteins which are also potent enhancers of inflammatory responses (see feedback loops, below). Another molecule upregulated by senescence is p19ARF that, along with are thought to also behave as tumor suppressors<sup>40</sup>. It is possible that senescent cells sit on a precipice between cancer and death and that p38 is critically involved in the balance.

#### Cell death

The death of a cell can proceed along several specific molecular pathways (reviewed in<sup>49, 50</sup>), and p38 may be directly or indirectly involved.<sup>51</sup> The two major apoptotic pathways that are well documented in mammalian cells are the death receptor and the mitochondrial pathway in the death receptor pathway. The death receptor pathway involves the cysteine proteases Caspase 8, 10, and 3, and is initiated by expression of molecules like TNF. The mitochondrial pathway is an intrinsic pathway is activated in response to ionizing radiation, growth factor withdrawal, chemotherapeutics drugs, reactive oxygen and nitrogen species, and DNA damage. Such insults lead to expression of BCL-2 family members,

mitochondrial membrane permeability, release of cytochrome c and activation of Caspases 9 and 3. Some of the pathways can be quite complex. For example, Glycogen synthase kinase  $3\beta$  (GSK3 $\beta$ ) is a constitutively active kinase that supports apoptosis through inhibition of the pro-survival transcription factors, and facilitating pro-apoptotic transcription factors such as  $p53^{52}$ . In developing T cells, GSK3 $\beta$  phosphorylates beta-catenin and causes its degradation<sup>53</sup> which prevents it from supporting the transcription of pro-survival genes. However, activated p38 can phosphorylate GSK3 $\beta$  on Ser 389 and inactivate it, thus preserving survival.<sup>54</sup> Recently, inability to inactivate GSK3 $\beta$  has been linked to promotion of cell death via a process now called necroptosis.<sup>49</sup>

#### Feedback loops and "rogue" pathways

The "upstream/downstream" construct for p38 may be a gross oversimplification. For example, molecules such as tumor progression locus 2 (TPL-2) TPL2 participate in a circuit that is dependent on p38 activation and itself activates the MKK3/6 which in turn activates p38<sup>55</sup>. For another example, stressors that generate p38 activation can also result in expression of High Mobility Group B1 protein (HMGB1), but this in turn can act via internal pathways to activate p38.<sup>56</sup> For yet a thrid example, senescent T cells emply neither the canonical nor alternative pathways and instead cause auto phosphorylation of p38 by engaging AMPK to cause p38 recruitment to the scaffold TAB1. This process leads to loss of telomerase<sup>57</sup> which may lead to shortened telomeres and may itself be associated with p38 activation.

# 5. Tissue specific regulation of p38 and implications for normal and abnormal function

#### The ovary

Activation of p38 might regulate proliferation. For example, in mouse granulosa cells, exposure to FSH leads to p38-mediated de-phosphorylation of the transcription factor STAT1. This leads to decreased transcription of the cytochrome P450 1 subfamily member Cyp1b1 and altered estrogen metabolism. The result is decreased proliferation, and it is hypothesized that this assists in the maintenance of estradiol levels in the dominant follicle in vivo.<sup>58</sup> In addition, p38 regulates the response to prolactin in a model premature ovarian failure<sup>59</sup>

#### The cervix

Cervical mucosal epithelium responds to infection with production of TNF- $\alpha$  and this is in part dependent on activation of p38<sup>60</sup>. In human cervix, p38 activation has been associated with parturition but this is likely due to inflammatory cell traffic macrophages<sup>61–63</sup> and or association with tissues such as the fetal membranes overlying the cervix at term<sup>64, 65</sup> as this tissue expresses an increased inflammatory signature.<sup>66</sup>

#### The myometrium

This kinase is expressed in myometrial cells where it has been found to mediate may mediate oxytocin receptor signaling and downstream inflammatory responses<sup>67</sup>. Histamine

also induces p38 activation in myometrial cells and this may in part explain the cross-talk between histamine and Toll like receptor triggered inflammation and further, that there is an increase in premature labor in patients with severe allergy<sup>68</sup> or asthma<sup>69</sup>. Stretch also activates the p38 pathway within myometrial cells<sup>2, 70</sup>, and this may contribute to the inflammatory signature occurring with multiple gestation, or with the end of a singleton pregnancy.

#### The decidua

Overall, activation of p38 increases with gestational age in mouse decidua <sup>38</sup> and may be increased by labor in humans.<sup>71</sup> The decidua is a complex tissue<sup>72, 73</sup> comprised of stromal, leukocyte, glandular, and vascular elements. Each is likely to have its own regulation with regard to the mitogen activated kinases. For examples, preimplantation factor, PIF supports implantation in a process that involves decreasing p38 activation and endo-cannabinoids can induce apoptosis in decidual cells in a process that includes activation of p38, mitochondrial stress, reactive oxygen species elaboration and apoptosis.<sup>74</sup> Decidual stromal cell dysfunction has been implicated in early and late pregnancy loss, related to a mouse model of progesterone receptor targeted insufficiency in p53, but the role of p38 in that model is unclear<sup>75</sup>. In humans, p38 may mediate the inflammation related to functional progesterone withdrawal<sup>7672</sup>.

The activation of leukocyte populations by Toll-like signalling largely involves activation of p38. For example, NK T cells are activated by Toll-like receptor, CD1, and inflammatory cytokine signaling that proceeds with activation of p38 and ERK pathways<sup>77</sup>. Decidual artery remodeling is an important aspect of normal pregnancy in the mouse<sup>7879, 80</sup> and deficient invasion of spiral arteries is thought to be an important initiator of preeclampsia. While many studies have reported the potential role of p38 activation or inhibition in systemic vascular remodelling<sup>81</sup>, few have examined decidual vessels. Human umbilical vein endothelial cells exposed to hypoxia/reperfusion upregulate growth arrest and DNA damage-inducible 45 alpha (Gadd45a) leading to p38 activation and ultimately an increase in fms-related tyrosine kinase-1 (sFlt-1) and soluble endoglin secretion, and both Gadd45a and activated p38 are found in placentas from preeclampsia pregnancies.<sup>82</sup>

#### Preimplantation embryo and trophoblast

Several studies have delineated the expression<sup>41</sup> and importance<sup>83</sup> of p38 signaling in the development and function of the preimplantation embryo. The p38 pathway is critical for glucose metabolism<sup>83</sup> and differentiation<sup>41</sup>, as well as in the generation of molecules important in implantation. Early in gestation, VEGF activates p38 and contributes to VEGF-induced EVT migration and capillary-like tube formation in human chorio-decidual trophoblast<sup>84</sup>.

#### **Fetal membranes**

The fetal membranes represent a unique tissue that balances proliferation, renewal, senescence and death. The fetal membranes are a multilayered and multicellular tissue that includes the amnion and the chorion.<sup>85</sup> Over time in normal pregnancy the fetal membranes undergo senescence associated with increased expression of p38<sup>346, 38</sup>, and express

increased p38 with oxidative stress,<sup>24</sup> and in the setting of premature rupture of membranes<sup>6, 71, 86, 87,72</sup>. Membranes express both classical and alternative p38 pathway molecules, but cell type specificity, and the exact up and downstream activators and effectors are not known.

## 6. The future of p38 in reproductive science

#### Animal models of reproductive outcome

There exist several models of reproductive outcome.<sup>88–90</sup> With regard to the role of p38, animal models, particularly mice offer the opportunity to examine both the kinetics and the specific molecular pathways by which activation of this molecule occurs and causes downstream effect. Studies in normal mice mice<sup>38</sup> suggest that progressive increase in activated p38 occurs in the decidua, placenta and fetal membranes. However, the relevant pathways and cells are unclear and will have to be examined in future studies and correlated with findings in humans. <sup>71, 91, 92,49</sup> Although some of the stressors leading to poor pregnancy outcome humans can be modeled in mice, the specific p38-involved pathways have yet to be fully explored.

#### **Biomarkers of p38 activation**

Peripheral blood cytokines and molecules such as sFlt have been used to predict abnormal pregnancy and may be down stream of p38 activation<sup>93</sup>. As activation of p38 is a pivotal event in many processes indicative of pregnancy stress or disregulation, it would be useful to have early indicators of p38 activation. In vitro, exosomes from aminon epithelial cells contain activated p38 and uptake into the cytoplasm of other cells activates inflammatory pathways.<sup>94</sup> Vesicles, including exosomes<sup>95</sup> eminate from several tissues of the reproductive tract and can be found in maternal blood. This raises the interesting idea that exosomes be used to diagnose early placental p38 activation.

#### Inhibitors and activators of p38 at the bedside

The clinical importance of modifiers of p38 activity and an approach to development of these drugs has been recently reviewed.<sup>96</sup> Focus has been on inhibition of the catalytic activity of p38 is through competitive or non-competitive binding of ATP<sup>97</sup>. Many inhibitors of p38 are being considered, including SB203580 (a competitive binder in the ATP pocket or p38), and BIRB 796<sup>97</sup> in diseases related to premature ageing (e.g. Werner Syndrome<sup>97</sup>) and autoimmunity. Focus on how to target drug activity and limit side effects continues. Further, there exists a pipeline to develop ways to modulate molecules down stream of p38 activation<sup>81, 96</sup> in specific cell types<sup>98</sup> order to increase specificity. Relevant clinical trials focusing on reproductive outcome need to be developed. Agents, such as statins<sup>99</sup> are being considered for improvement of reproductive outcome, and have an indirect action on the p38 pathway through decreasing upstream stressors such as oxidative stress<sup>100</sup>, but further understanding of these agents is also needed.

# 7. Summary

p38 is part of an evolutionarily conserved family of molecules that plays a pivotal role in homeostasis in the reproductive tract. Certain cell types, including immune cells, employ canonical, alternative or "rogue" p38 pathways. Consideration p38's roll in cell-cell interaction in the reproductive tract and beyond is crucial. Focus on the molecular stresses leading to increased expression of activated p38 as well as down-stream effector pathways will lead to greater understanding of normal and abnormal reproduction.

# Acknowledgments

I am supported in part by the Vermont Center for Immunology and Infectious Diseases (NIH P30 GM118228) and the Department of Obstetrics, Gynecology and Reproductive Sciences. I am grateful for discussions with my colleagues in PREBIC, the Preterm Birth International Collaborative and apologize to my colleagues for work not mentioned due to space or time considerations.

#### References

- 1. Murtha AP, Menon R. Regulation of fetal membrane inflammation: a critical step in reducing adverse pregnancy outcome. Am Journal Obstet Gynrcol. 2015; 213:447–448.
- Adams Waldorf KM, Singh N, Mohan AR, Young RC, Ngo L, Das A, Tsai J, Bansal A, Paolella L, Herbert BR, Sooranna SR, Gough GM, Astley C, Vogel K, Baldessari AE, Bammler TK, MacDonald J, Gravett MG, Rajagopal L, Johnson MR. Uterine overdistention induces preterm labor mediated by inflammation: observations in pregnant women and nonhuman primates. Am Journal Obstet Gynrcol. 2015; 213:830e831–830.e819.
- Kendal-Wright CE. Stretching, mechanotransduction, and proinflammatory cytokines in the fetal membranes. Reprod Sci. 2007; 14:35–41. [PubMed: 18089608]
- 4. Mendelson CR. Minireview: fetal-maternal hormonal signaling in pregnancy and labor. Molec Endocrinol. 2009; 23:947–954. [PubMed: 19282364]
- Tan H, Yi L, Rote NS, Hurd WW, Mesiano S. Progesterone receptor-A and -B have opposite effects on proinflammatory gene expression in human myometrial cells: implications for progesterone actions in human pregnancy and parturition. J Clin Endocrinol Metab. 2012; 97:E719–730. [PubMed: 22419721]
- Dutta EH, Behnia F, Boldogh I, Saade GR, Taylor BD, Kacerovsky M, Menon R. Oxidative stress damage-associated molecular signaling pathways differentiate spontaneous preterm birth and preterm premature rupture of the membranes. Molec Hum Reprod. 2016; 22:143–157. [PubMed: 26690900]
- Coppe JP, Patil CK, Rodier F, Sun Y, Munoz DP, Goldstein J, Nelson PS, Desprez PY, Campisi J. Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol. 2008; 6:2853–2868. [PubMed: 19053174]
- Gomez-Lopez N, Romero R, Plazyo O, Panaitescu B, Furcron AE, Miller D, Roumayah T, Flom E, Hassan SS. Intra-Amniotic Administration of HMGB1 Induces Spontaneous Preterm Labor and Birth. American J Reprod Immunol 1989. 2016; 75:3–7.
- 9. Rodier F, Campisi J. Four faces of cellular senescence. J Cell Biol. 2011; 192:547–556. [PubMed: 21321098]
- Krementsov DN, Thornton TM, Teuscher C, Rincon M. The emerging role of p38 mitogenactivated protein kinase in multiple sclerosis and its models. Molec Cell Biol. 2013; 33:3728– 3734. [PubMed: 23897428]
- Freund A, Patil CK, Campisi J. p38MAPK is a novel DNA damage response-independent regulator of the senescence-associated secretory phenotype. EMBO J. 2011; 30:1536–1548. [PubMed: 21399611]

- Garcia-Gomez C, Parages ML, Jimenez C, Palma A, Mata MT, Segovia M. Cell survival after UV radiation stress in the unicellular chlorophyte Dunaliella tertiolecta is mediated by DNA repair and MAPK phosphorylation. J Exp botany. 2012; 63:5259–5274. [PubMed: 22859678]
- 14. Busca R, Pouyssegur J, Lenormand P. ERK1 and ERK2 Map Kinases: Specific Roles or Functional Redundancy? Frontiers in Cell and Developmental Biol. 2016; 4:53.
- Krens SF, Spaink HP, Snaar-Jagalska BE. Functions of the MAPK family in vertebratedevelopment. FEBS letters. 2006; 580:4984–4990. [PubMed: 16949582]
- Hale KK, Trollinger D, Rihanek M, Manthey CL. Differential expression and activation of p38 mitogen-activated protein kinase alpha, beta, gamma, and delta in inflammatory cell lineages. J Immunol (Baltimore, Md : 1950). 1999; 162:4246–4252.
- Enslen H, Raingeaud J, Davis RJ. Selective activation of p38 mitogen-activated protein (MAP) kinase isoforms by the MAP kinase kinases MKK3 and MKK6. J Biol Chem. 1998; 273:1741– 1748. [PubMed: 9430721]
- Brancho D, Tanaka N, Jaeschke A, Ventura JJ, Kelkar N, Tanaka Y, Kyuuma M, Takeshita T, Flavell RA, Davis RJ. Mechanism of p38 MAP kinase activation in vivo. Genes Dev. 2003; 17:1969–1978. [PubMed: 12893778]
- Brichkina A, Nguyen NT, Baskar R, Wee S, Gunaratne J, Robinson RC, Bulavin DV. Proline isomerisation as a novel regulatory mechanism for p38MAPK activation and functions. Cell Death Differ. 2016; 23:1592–1601. [PubMed: 27233083]
- Desai J, Holt-Shore V, Torry RJ, Caudle MR, Torry DS. Signal transduction and biological function of placenta growth factor in primary human trophoblast. Biol Reprod. 1999; 60:887–892. [PubMed: 10084962]
- 21. Yan BC, Adachi T, Tsubata T. ER stress is involved in B cell antigen receptor ligation-induced apoptosis. Biochem Bioiophys Res Com. 2008; 365:143–148.
- 22. Rasheed Z, Haqqi TM. Endoplasmic reticulum stress induces the expression of COX-2 through activation of eIF2α, p38-MAPK and NF-κB in advanced glycation end products stimulated human chondrocytes. Biochimica Biophysica Acta. 2012; 1823:2179–2189.
- 23. Li YX, Ren YL, Fu HJ, Zou L, Yang Y, Chen Z. Hepatitis B Virus Middle Protein Enhances IL-6 Production via p38 MAPK/NF-kappaB Pathways in an ER Stress-Dependent Manner. PloS one. 2016; 11:e0159089. [PubMed: 27434097]
- Menon R, Fortunato SJ, Yu J, Milne GL, Sanchez S, Drobek CO, Lappas M, Taylor RN. Cigarette smoke induces oxidative stress and apoptosis in normal term fetal membranes. Placenta. 2011; 32:317–322. [PubMed: 21367451]
- Ashwell JD. The many paths to p38 mitogen-activated protein kinase activation in the immune system. Nature Rev Immunol. 2006; 6:532–540. [PubMed: 16799472]
- 26. Salvador JM, Mittelstadt PR, Guszczynski T, Copeland TD, Yamaguchi H, Appella E, Fornace AJ Jr, Ashwell JD. Alternative p38 activation pathway mediated by T cell receptor-proximal tyrosine kinases. Nature immunol. 2005; 6:390–395. [PubMed: 15735648]
- 27. Sreekanth GP, Chuncharunee A, Sirimontaporn A, Panaampon J, Noisakran S, Yenchitsomanus PT, Limjindaporn T. SB203580 Modulates p38 MAPK Signaling and Dengue Virus-Induced Liver Injury by Reducing MAPKAPK2, HSP27, and ATF2 Phosphorylation. PLoS One. 2016; 11:e0149486. [PubMed: 26901653]
- Brand S. Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease. Gut. 2009; 58:1152– 1167. [PubMed: 19592695]
- Hannigan M, Zhan L, Ai Y, Huang CK. The role of p38 MAP kinase in TGF-beta1-induced signal transduction in human neutrophils. Biochem Bioiophys Res Com communications. 1998; 246:55– 58.
- 30. Silva C, Nunes C, Correia-Branco A, Araújo JR, Martel F. Insulin Exhibits an Antiproliferative and Hypertrophic Effect in First Trimester Human Extravillous Trophoblasts. Reprod Sci. 2016 Sep 22.2016 pii: 1933719116667220 Epub ahead of print.

- Dmitrieva NI, Bulavin DV, Fornace AJ, Burg MB. Rapid activation of G(2)/M checkpoint after hypertonic stress in renal inner medullary epithelial (IME) cells is protective and requires p38 kinase. PNAS. 2002; 99:184–189. [PubMed: 11756692]
- 32. Duval D, Trouillas M, Thibault C, Dembele D, Diemunsch F, Reinhardt B, Mertz AL, Dierich A, Boeuf H. Apoptosis and differentiation commitment: novel insights revealed by gene profiling studies in mouse embryonic stem cells. Cell Death Differ. 2005; 13:564–575.
- Ono K, Han J. The p38 signal transduction pathway Activation and function. Cellular signalling. 2000; 12:1–13. [PubMed: 10676842]
- Tivey HS, Rokicki MJ, Barnacle JR, Rogers MJ, Bagley MC, Kipling D, Davis T. Small molecule inhibition of p38 MAP kinase extends the replicative life span of human ATR-Seckel syndrome fibroblasts. J Gerontol Series A. 2013; 68:1001–1009.
- 35. Misri S, Pandita S, Kumar R, Pandita TK. Telomeres, histone code, and DNA damage response. Cytogenetic Genome Res. 2008; 122:297–307.
- 36. Wood CD, Thornton TM, Sabio G, Davis RA, Rincon M. Nuclear localization of p38 MAPK in response to DNA damage. IJBS. 2009; 5:428–437. [PubMed: 23675168]
- 37. Derradji H, Bekaert S, De Meyer T, Jacquet P, Abou-El-Ardat K, Ghardi M, Arlette M, Baatout S. Ionizing radiation-induced gene modulations, cytokine content changes and telomere shortening in mouse fetuses exhibiting forelimb defects. Dev Biol. 2008; 322:302–313. [PubMed: 18722365]
- Bonney EA, Krebs K, Saade G, Kechichian T, Trivedi J, Huaizhi Y, Menon R. Differential senescence in feto-maternal tissues during mouse pregnancy. Placenta. 2016; 43:26–34. [PubMed: 27324096]
- Phillippe M. Cell-Free Fetal DNA, Telomeres, and the Spontaneous Onset of Parturition. Reprod Sci. 2015; 22:1186–1201. [PubMed: 26134037]
- 40. Salminen A, Kauppinen A, Kaarniranta K. Emerging role of NF-kappaB signaling in the induction of senescence-associated secretory phenotype (SASP). Cell Sig. 2012; 24:835–845.
- 41. Natale DR, Paliga AJ, Beier F, D'Souza SJ, Watson AJ. p38 MAPK signaling during murine preimplantation development. Dev Biol. 2004; 268:76–88. [PubMed: 15031106]
- 42. Braunschweig A, Poehlmann TG, Busch S, Schleussner E, Markert UR. Signal transducer and activator of transcription 3 (STAT3) and Suppressor of Cytokine Signaling (SOCS3) balance controls cytotoxicity and IL-10 expression in decidual-like natural killer cell line NK-92. Am J Reprod Immunol. 2011; 66:329–335. [PubMed: 21385272]
- Choi AMK, Ryter SW, Levine B. Autophagy in Human Health and Disease. New Eng J Med. 2013; 368:651–662. [PubMed: 23406030]
- 44. Avagliano L, Massa V, Zullino S, Doi P, Marconi AM, Ferrazzi E, Bulfamante GP. Inflammation modulates LC3 expression in human preterm delivery. J Maternal-fetal Neonat Med. 2016:1–7.
- 45. Henson SM, Lanna A, Riddell NE, Franzese O, Macaulay R, Griffiths SJ, Puleston DJ, Watson AS, Simon AK, Tooze SA, Akbar AN. p38 signaling inhibits mTORC1-independent autophagy in senescent human CD8(+) T cells. J Clin Invest. 2014; 124:4004–4016. [PubMed: 25083993]
- 46. Simone C. Signal-dependent control of autophagy and cell death in colorectal cancer cell: the role of the p38 pathway. Autophagy. 2007; 3:468–471. [PubMed: 17495519]
- Thornton TM, Rincon M. Non-classical p38 map kinase functions: cell cycle checkpoints and survival. IJBS. 2009; 5:44–51. [PubMed: 23675113]
- Bulavin DV, Higashimoto Y, Popoff IJ, Gaarde WA, Basrur V, Potapova O, Appella E, Fornace AJ. Initiation of a G2/M checkpoint after ultraviolet radiation requires p38 kinase. Nature. 2001; 411:102–107. [PubMed: 11333986]
- Tait SWG, Ichim G, Green DR. Die another way non-apoptotic mechanisms of cell death. J Cell Science. 2014; 127:2135–2144. [PubMed: 24833670]
- 50. Sanchez-Capelo A. Dual role for TGF-beta1 in apoptosis. Cytokine Growth Factor Rev. 2005; 16:15–34. [PubMed: 15733830]
- Cross TG, Scheel-Toellner D, Henriquez NV, Deacon E, Salmon M, Lord JM. Serine/Threonine Protein Kinases and Apoptosis. Exp Cell Res. 2000; 256:34–41. [PubMed: 10739649]
- 52. Watcharasit P, Bijur GN, Zmijewski JW, Song L, Zmijewska A, Chen X, Johnson GV, Jope RS. Direct, activating interaction between glycogen synthase kinase-3beta and p53 after DNA damage. PNAS. 2002; 99:7951–7955. [PubMed: 12048243]

- 53. Jacobs KM, Bhave SR, Ferraro DJ, Jaboin JJ, Hallahan DE, Thotala D. GSK-3β: A Bifunctional Role in Cell Death Pathways. Int J Cell Biol. 2012; 2012:930710. [PubMed: 22675363]
- 54. Thornton TM, Delgado P, Chen L, Salas B, Krementsov D, Fernandez M, Vernia S, Davis RJ, Heimann R, Teuscher C, Krangel MS, Ramiro AR, Rincon M. Inactivation of nuclear GSK3beta by Ser(389) phosphorylation promotes lymphocyte fitness during DNA double-strand break response. Nat Com. 2016; 7:10553.
- 55. Pattison MJ, Mitchell O, Flynn HR, Chen CS, Yang HT, Ben-Addi H, Boeing S, Snijders AP, Ley SC. TLR and TNF-R1 activation of the MKK3/MKK6-p38alpha axis in macrophages is mediated by TPL-2 kinase. Biochem J. 2016; 473:2845–2861. [PubMed: 27402796]
- 56. Bredeson S, Papaconstantinou J, Deford JH, Kechichian T, Syed TA, Saade GR, Menon R. HMGB1 promotes a p38MAPK associated non-infectious inflammatory response pathway in human fetal membranes. PLoS One. 2014; 9:e113799. [PubMed: 25469638]
- Lanna A, Henson SM, Escors D, Akbar AN. AMPK-TAB1 activated p38 drives human T cell senescence. Nature Immunol. 2014; 15:965–972. [PubMed: 25151490]
- Du XH, Zhou XL, Cao R, Xiao P, Teng Y, Ning CB, Liu HL. FSH-induced p38-MAPK-mediated dephosphorylation at serine 727 of the signal transducer and activator of transcription 1 decreases Cyp1b1 expression in mouse granulosa cells. Cell Sig. 2015; 27:6–14.
- 59. Devi YS, Seibold AM, Shehu A, Maizels E, Halperin J, Le J, Binart N, Bao L, Gibori G. Inhibition of MAPK by prolactin signaling through the short form of its receptor in the ovary and decidua: involvement of a novel phosphatase. J Biol Chem. 2011; 286:7609–7618. [PubMed: 21199871]
- 60. Yang JB, Quan JH, Kim YE, Rhee YE, Kang BH, Choi IW, Cha GH, Yuk JM, Lee YH. Involvement of PI3K/AKT and MAPK Pathways for TNF-alpha Production in SiHa Cervical Mucosal Epithelial Cells Infected with Trichomonas vaginalis. Korean J Parasitol. 2015; 53:371– 377. [PubMed: 26323834]
- Dubicke A, Ekman-Ordeberg G, Mazurek P, Miller L, Yellon SM. Density of Stromal Cells and Macrophages Associated With Collagen Remodeling in the Human Cervix in Preterm and Term Birth. Reprod Sci. 2015; 23(5):595–603. [PubMed: 26608218]
- Mackler AM, Iezza G, Akin MR, McMillan P, Yellon SM. Macrophage Trafficking in the Uterus and Cervix Precedes Parturition in the Mouse. Biol Reprod. 1999; 61:879–883. [PubMed: 10491619]
- 63. Dobyns AE, Goyal R, Carpenter LG, Freeman TC, Longo LD, Yellon SM. Macrophage gene expression associated with remodeling of the prepartum rat cervix: microarray and pathway analyses. PloS one. 2015; 10:e0119782. [PubMed: 25811906]
- 64. Larsen B, Hwang J. Progesterone interactions with the cervix: translational implications for term and preterm birth. Infect Dis Obstet Gynecol. 2011; 2011:353297. [PubMed: 22114461]
- 65. Lappas M, Riley C, Lim R, Barker G, Rice GE, Menon R, Permezel M. MAPK and AP-1 proteins are increased in term pre-labour fetal membranes overlying the cervix: regulation of enzymes involved in the degradation of fetal membranes. Placenta. 2011; 32:1016–1025. [PubMed: 21963187]
- 66. Lappas M, Odumetse TL, Riley C, Reti NG, Holdsworth-Carson SJ, Rice GE, Permezel M. Prelabour fetal membranes overlying the cervix display alterations in inflammation and NF-kappaB signalling pathways. Placenta. 2008; 29:995–1002. [PubMed: 18952281]
- Kim SH, MacIntyre DA, Firmino Da Silva M, Blanks AM, Lee YS, Thornton S, Bennett PR, Terzidou V. Oxytocin activates NF-kappaB-mediated inflammatory pathways in human gestational tissues. Mol Cell Endocrinol. 2015; 403:64–77. [PubMed: 25451977]
- Romero R, Kusanovic JP, Munoz H, Gomez R, Lamont RF, Yeo L. Allergy-induced preterm labor after the ingestion of shellfish. J Matern Fetal Neonatal Med. 2010; 23:351–359. [PubMed: 19900031]
- Bakhireva LN, Schatz M, Jones KL, Chambers CD. Asthma control during pregnancy and the risk of preterm delivery or impaired fetal growth. Ann Allergy Asthma Immunol. 2008; 101:137–143. [PubMed: 18727468]
- 70. Sooranna SR, Engineer N, Loudon JA, Terzidou V, Bennett PR, Johnson MR. The mitogenactivated protein kinase dependent expression of prostaglandin H synthase-2 and interleukin-8

messenger ribonucleic acid by myometrial cells: the differential effect of stretch and interleukin-1{beta}. J Clin Endocrinol Metab. 2005; 90:3517–3527. [PubMed: 15784717]

- 71. Menon R, Behnia F, Polettini J, Saade GR, Campisi J, Velarde M. Placental membrane aging and HMGB1 signaling associated with human parturition. Aging. 2016
- Menon R, Bonney EA, Condon J, Mesiano S, Taylor RN. Novel concepts on pregnancy clocks and alarms: redundancy and synergy in human parturition. Hum Reprod Update. 2016; 22:535–560. [PubMed: 27363410]
- 73. Norwitz ER, Bonney EA, Snegovskikh VV, Williams MA, Phillippe M, Park JS, Abrahams VM. Molecular Regulation of Parturition: The Role of the Decidual Clock. Cold Spring Harbor Perspect Med. 2015; 5:11.doi: 10.1101/cshperspect.a023143
- Fonseca BM, Correia-da-Silva G, Teixeira NA. The endocannabinoid anandamide induces apoptosis of rat decidual cells through a mechanism involving ceramide synthesis and p38 MAPK activation. Apoptosis. 2013; 18:1526–1535. [PubMed: 24048885]
- Hirota Y, Daikoku T, Tranguch S, Xie H, Bradshaw HB, Dey SK. Uterine-specific p53 deficiency confers premature uterine senescence and promotes preterm birth in mice. J Clin Invest. 2010; 120:803–815. [PubMed: 20124728]
- 76. Guzeloglu-Kayisli O, Kayisli UA, Semerci N, Basar M, Buchwalder LF, Buhimschi CS, Buhimschi IA, Arcuri F, Larsen K, Huang JS, Schatz F, Lockwood CJ. Mechanisms of chorioamnionitis-associated preterm birth: interleukin-1beta inhibits progesterone receptor expression in decidual cells. J Pathol. 2015; 237:423–434. [PubMed: 26175191]
- 77. Li L, Yang J, Jiang Y, Tu J, Schust DJ. Activation of decidual invariant natural killer T cells promotes lipopolysaccharide-induced preterm birth. Molec Hum Reprod. 2015; 21:369–381. [PubMed: 25589517]
- 78. Dixon ME, Chien EK, Osol G, Callas PW, Bonney EA. Failure of decidual arteriolar remodeling in the CBA/J x DBA/2 murine model of recurrent pregnancy loss is linked to increased expression of tissue inhibitor of metalloproteinase 2 (TIMP-2). Am J Obstet Gynecol. 2006; 194:113–119. [PubMed: 16389019]
- Felker AM, Chen Z, Foster WG, Croy BA. Receptors for non-MHC ligands contribute to uterine natural killer cell activation during pregnancy in mice. Placenta. 2013; 34:757–764. [PubMed: 23806179]
- Ashkar AA, Di Santo JP, Croy BA. Interferon gamma contributes to initiation of uterine vascular modification, decidual integrity, and uterine natural killer cell maturation during normal pregnancy. J Exp Med. 2000; 192:259–269. [PubMed: 10899912]
- Potthoff SA, Stamer S, Grave K, Konigshausen E, Sivritas SH, Thieme M, Mori Y, Woznowski M, Rump LC, Stegbauer J. Chronic p38 mitogen-activated protein kinase inhibition improves vascular function and remodeling in angiotensin II-dependent hypertension. JRAAS. 2016; 17(3) pii: 1470320316653284. doi: 10.1177/1470320316653284
- 82. Luo X, Yao Z-w, Qi H-b, Liu D-d, Chen G-q, Huang S, Li Q-s. Gadd45α as an upstream signaling molecule of p38 MAPK triggers oxidative stress-induced sFlt-1 and sEng upregulation in preeclampsia. Cell Tiss Res. 2011; 344:551.
- Sozen B, Ozturk S, Yaba A, Demir N. The p38 MAPK signalling pathway is required for glucose metabolism, lineage specification and embryo survival during mouse preimplantation development. Mech Dev. 2015; 138(Pt 3):375–398. [PubMed: 26025760]
- Lala N, Girish GV, Cloutier-Bosworth A, Lala PK. Mechanisms in decorin regulation of vascular endothelial growth factor-induced human trophoblast migration and acquisition of endothelial phenotype. Biol Reprod. 2012; 87:59. [PubMed: 22699486]
- 85. Bourne G. The Fœtal Membranes: A Review of the Anatomy of Normal Amnion and Chorion and Some Aspects of Their Function. Postgrad Med J. 1962; 38:193–201. [PubMed: 13871927]
- 86. Menon R, Boldogh I, Hawkins HK, Woodson M, Polettini J, Syed TA, Fortunato SJ, Saade GR, Papaconstantinou J, Taylor RN. Histological evidence of oxidative stress and premature senescence in preterm premature rupture of the human fetal membranes recapitulated in vitro. Am J Pathol. 2014; 184:1740–1751. [PubMed: 24832021]

- Menon R, Papaconstantinou J. p38 Mitogen activated protein kinase (MAPK): a new therapeutic target for reducing the risk of adverse pregnancy outcomes. Expert Opin Therapeut Targets. 2016:1–16.
- Nielsen BW, Bonney EA, Pearce BD, Donahue LR, Sarkar IN. A Cross-Species Analysis of Animal Models for the Investigation of Preterm Birth Mechanisms. Reprod Sci. 2016; 23:482– 491. [PubMed: 26377998]
- Bonney EA. Demystifying animal models of adverse pregnancy outcomes: touching bench and bedside. Am J Reprod Immunol. 2013; 69:567–584. [PubMed: 23448345]
- 90. Bonney EA, Brown SA. To drive or be driven: the path of a mouse model of recurrent pregnancy loss. Reproduction. 2014; 147:R153–167. [PubMed: 24472815]
- Behnia F, Taylor BD, Woodson M, Kacerovsky M, Hawkins H, Fortunato SJ, Saade GR, Menon R. Chorioamniotic membrane senescence: a signal for parturition? Am J Obstet Gynecol. 2015; 213:359e351–316. [PubMed: 26025293]
- Polettini J, Behnia F, Taylor BD, Saade GR, Taylor RN, Menon R. Telomere Fragment Induced Amnion Cell Senescence: A Contributor to Parturition? PLoS One. 2015; 10:e0137188. [PubMed: 26397719]
- Xiong Y, Liebermann DA, Tront JS, Holtzman EJ, Huang Y, Hoffman B, Geifman-Holtzman O. Gadd45a stress signaling regulates sFlt-1 expression in preeclampsia. J Cell Physiol. 2009; 220:632–639. [PubMed: 19452502]
- 94. Sheller S, Papaconstantinou J, Urrabaz-Garza R, Richardson L, Saade G, Salomon C, Menon R. Amnion-Epithelial-Cell-Derived Exosomes Demonstrate Physiologic State of Cell under Oxidative Stress. PLoS ONE. 2016; 11:e0157614. [PubMed: 27333275]
- 95. Foster BP, Balassa T, Benen TD, Dominovic M, Elmadjian GK, Florova V, Fransolet MD, Kestlerova A, Kmiecik G, Kostadinova IA, Kyvelidou C, Meggyes M, Mincheva MN, Moro L, Pastuschek J, Spoldi V, Wandernoth P, Weber M, Toth B, Markert UR. Extracellular vesicles in blood, milk and body fluids of the female and male urogenital tract and with special regard to reproduction. Crit Reviews Clinical Lab Sci. 2016; 53:379–395.
- 96. Menon R, Papaconstantinou J. p38 Mitogen activated protein kinase (MAPK): a new therapeutic target for reducing the risk of adverse pregnancy outcomes. Expert Opin Therapeut Targets. 2016; 20:1397–1412.
- Bagley MC, Davis T, Murziani PGS, Widdowson CS, Kipling D. Use of p38 MAPK Inhibitors for the Treatment of Werner Syndrome. Pharmaceuticals. 2010; 3:1842–1872. [PubMed: 27713332]
- Kragholm K, Newby LK, Melloni C. Emerging treatment options to improve cardiovascular outcomes in patients with acute coronary syndrome: focus on losmapimod. Drug Design, Dev Therapy. 2015; 9:4279–4286.
- 99. Ramma W, Ahmed A. Therapeutic potential of statins and the induction of heme oxygenase-1 in preeclampsia. J Reprod Immunol. 2014; 101–102:153–160.
- 100. Bao, X-m, Wu, C-f, Lu, G-p. Atorvastatin attenuates homocysteine-induced apoptosis in human umbilical vein endothelial cells via inhibiting NADPH oxidase-related oxidative stress-triggered p38MAPK signaling. Acta Pharmacologica Sinica. 2009; 30:1392–1398. [PubMed: 19767766]



#### Fig. 1. A p38-centric view of cellular homeostasis

A stylized cell with dark grey endoplasmic reticulum, light grey mitochondria and a while nucleus is shown. A sampling of factors leading to increased presence of activated (phosphorylated, P-p38) p38 are shown with blue arrows/black writing, while events downstream of p38 activation (green) are shown with red arrows and writing. ASK-1 (Apoptosis signaling kinase 1), TRAF2 (TNF receptor-associated factor 2), GADD45α (Growth arrest and DNA-damage-inducible protein GADD45 alpha), TNF (Tumor necrosis factor), IL-1 (Interleukin 1), NF-κb (nuclear factor kappa B) are all shown as representative molecules in the p38 'universe'.