



Published in final edited form as:

*Epilepsia*. 2016 October ; 57(10): 1651–1659. doi:10.1111/epi.13506.

## The spectrum of epilepsy and electroencephalographic abnormalities due to *SHANK3* loss of function mutations

J. Lloyd Holder Jr., MD, PhD<sup>1,2</sup> and Michael M. Quach, MD<sup>1</sup>

<sup>1</sup>Baylor College of Medicine and Texas Children's Hospital

### Summary

**Objective**—The coincidence of autism with epilepsy is 27% in those individuals with intellectual disability<sup>1</sup>. Individuals with loss of function mutations in *SHANK3* have intellectual disability, autism and variably, epilepsy<sup>2–5</sup>. The spectrum of seizure semiologies and electroencephalographic (EEG) abnormalities has never been investigated in detail. With the recent report that *SHANK3* mutations are present in approximately two percent of individuals with moderate to severe intellectual disabilities and one percent of individuals with autism, determining the spectrum of seizure semiologies and electrographic abnormalities will be critical for medical practitioners to appropriately counsel the families of patients with *SHANK3* mutations.

**Methods**—A retrospective chart review was performed of all individuals treated at the Blue Bird Circle Clinic for Child Neurology who have been identified as having either a chromosome 22q13 microdeletion encompassing *SHANK3* or a loss of function mutation in *SHANK3* identified through whole exome sequencing.

For each subject, the presence or absence of seizures, seizure semiology, frequency, age of onset and efficacy of therapy were determined. Electroencephalograms were reviewed by a board certified neurophysiologist. Neuroimaging was reviewed by both a board certified pediatric neuroradiologist and child neurologist.

**Results**—There is a wide spectrum of seizure semiologies, frequencies and severity in individuals with *SHANK3* mutations. There are no specific electroencephalographic abnormalities found in our cohort, and EEG abnormalities were present in individuals diagnosed with epilepsy and those without history of a clinical seizure.

**Significance**—All individuals with a mutation in *SHANK3* should be evaluated for epilepsy due to the high prevalence of seizures in this population. The most common semiology is atypical absence seizure which can be challenging to identify due to comorbid intellectual disability in individuals with *SHANK3* mutations; however, no consistent seizure semiology, neuroimaging

<sup>2</sup>Corresponding author. J. Lloyd Holder, Jr., MD, PhD, Assistant Professor of Pediatrics, Division of Neurology and Developmental Neuroscience, 1250 Moursund St., Suite 1350, Houston, Texas 77030, holder@bcm.edu, Telephone: 832-822-1750, Fax: 832-825-1717.

Dr. Holder has full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

None of the authors has any conflict of interest to disclose.

We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

findings or electroencephalogram findings were present in the majority of individuals with *SHANK3* mutations.

## Keywords

*SHANK3*; Phelan-McDermid Syndrome; epilepsy; electroencephalography; autism

---

## Introduction

Loss of function mutations in the *SHANK3* gene, either through chromosomal microdeletion in Phelan-McDermid Syndrome (PMS) or through missense/nonsense mutations, are a growing area of importance for neurologists and epileptologists<sup>3</sup>. Approximately two percent of individuals with moderate to severe intellectual disability and one percent of individuals with autism are estimated to harbor such mutations<sup>6</sup>.

The protein product of the *SHANK3* gene is a scaffolding protein that localizes primarily to the post-synaptic density of excitatory synapses<sup>7</sup>. At the post-synaptic density, it bridges transmembrane proteins critical for synapse formation such as Neuroligins and neurotransmitter receptors such as NMDA glutamate receptors with the underlying cytoskeleton, in particular F-actin elements<sup>8; 9</sup>. Loss of *SHANK3* in multiple model systems, cultured neurons to mice, has demonstrated synaptic dysfunction<sup>10-13</sup>. As such, *SHANK3* is a critical protein for biogenesis and maintenance of synapses as well as synapse function.

Epilepsy has been reported in individuals found to have 22q13 deletions including *SHANK3* with a prevalence ranging from 17–70% in multiple case series (Table I). Most of these studies, however, have provided limited information about the semiology, frequency or severity of seizures in this population. Nor have they provided information about the spectrum or frequency of electroencephalographic abnormalities of these patients. Here we present a detailed report of the spectrum of seizures and electroencephalographic findings in individuals with *SHANK3* loss of function mutations. We find significant heterogeneity in seizure types and frequency in our cohort. Furthermore, we identify a wide spectrum of electroencephalographic abnormalities in our subjects which are present in both individuals diagnosed with epilepsy and those without.

## Patients and Methods

### Ethics

This study was approved by the Institutional Review Board of Baylor College of Medicine. All participants or their guardians provided informed written consent to participate in this study.

### Patient Selection and Evaluation

We retrospectively collected data from a single center (Texas Children's Hospital) through the electronic medical record. Information about the seizure characteristics of 24 individuals (10 males and 14 females) with either a chromosomal deletion encompassing chromosome 22q13 including *SHANK3*, point mutation, indel or microduplication of *SHANK3* predicted

to result in a loss of function mutation were analyzed. Data regarding the onset of first clinical seizure, seizure frequency, duration, semiology, response to pharmaceutical and non-pharmaceutical therapies and neuroimaging abnormalities were collected and analyzed.

### Electroencephalogram interpretation

Electroencephalograms were obtained at a single site with review by a board certified neurophysiologist. The video-EEG recordings were obtained using the Nicolet video-EEG system (Natus Medical, Inc.) while patients were awake, drowsy and, when possible, asleep with a 21-channel EEG acquisition system using silver-chloride surface electrodes. The EEG data was reviewed in both bipolar and referential montages.

## Results

### Clinical findings

We retrospectively evaluated twenty-four subjects with *SHANK3* mutations including twenty individuals with chromosomal deletions of 22q13.33 including *SHANK3* and four individuals with point mutations identified by whole exome sequencing resulting in either frameshift or mis-splicing of the *SHANK3* coding frame (Table II). We found eleven out of 24 subjects had a history of at least one lifetime seizure (46%), including two of the four individuals with point mutations. The spectrum of seizure semiologies varied significantly from atypical absence seizures (90%) being the most common seizure type to tonic (54%), atonic (18%), tonic-clonic (9%) and myoclonic (9%) in those individuals with a history of clinical seizure. Six of the eleven subjects with a history of seizure had more than one seizure type (54%), and two patients from this cohort (8%) were diagnosed with Lennox-Gastaut Syndrome based upon the combination of multiple seizure types (including tonic seizures), intractability of their seizures and characteristic EEG finding of generalized bursts of 1.5–2 Hz spike and slow wave activity. Five of our 24 subjects (20%) had a history of status epilepticus requiring emergency intravenous medication for cessation of seizure activity. The onset of first seizure ranged from 14 months to 14 years with a mean age of onset of 5.2 years  $\pm$  3.9 (SD). The frequency of seizures similarly varied from a single lifetime seizure to hundreds of seizures per day (Table II).

### Electrographic findings

Electroencephalographic abnormalities were identified in fourteen out of twenty-one subjects (67%) for which EEG data was available. The spectrum of EEG abnormalities was wide from slowing or absence of the occipital dominant rhythm (42%) to focal spike and slow wave discharges (38%) to generalized spike and slow wave discharges (19%) (Figure 1). Several patients had combinations of the above abnormalities. Of the twenty-one patients with EEG data available, five had an abnormal EEG (either slow occipital dominant rhythm or focal spike and slow wave activity or both) but no history of clinical seizure (23%).

Most individuals with multiple seizure types had an abnormally slow occipital dominant rhythm (83%) while the majority of individuals with a single seizure type were found to have an age-appropriate occipital dominant rhythm (60%). Similarly, both individuals with high seizure burden of hundreds of seizure per day prior to anti-convulsant medication were

found to have a significantly slowed occipital dominant rhythm. However, it should be noted that three of the ten individuals (30%) for whom we have EEG data and without a history of seizure had abnormally slow occipital dominant rhythm.

### Neuroimaging

Of our 24 subjects, 21 had an anatomic brain magnetic resonance imaging (MRI) available for review (88%). The abnormalities identified varied greatly between individuals (Figure 2). The most common abnormalities were dysmorphisms of the corpus callosum (29%) and T2 hyperintensities of the deep white matter (24%). Anatomic brain abnormalities were detected in both individuals with history of seizures (33%) and without (38%).

### Treatments

The type and number of anti-convulsant medications prescribed varied greatly among subjects. The most commonly prescribed anti-convulsant was lamotrigine (17%) followed by levetiracetam and topiramate (12% each) and then several others in less than 10% of this population (rufinamide, zonisamide, perampanel, felbamate, valproate, carbamazepine, oxcarbazepine, lacosamide, phenytoin, methsuximide, chlorazepate, lorazepam, phenobarbital). While underpowered to evaluate for efficacy in this study, no medication was clearly superior for seizure prophylaxis. There was a trend for those with one seizure type to respond to fewer medications than those with multiple seizure semiologies. All individuals with only one semiology (six) were maintained on less than two anti-convulsant medications while three of five individuals with multiple seizure types required two or more maintenance anti-convulsant medications ( $p=0.06$ , Fisher's exact test). Two individuals were found to have pharmaco-resistant epilepsy and had implantation of vagus nerve stimulators with mild improvement in seizure frequency (<50% reduction in seizures).

**Longitudinal outcomes**—Of the eleven patients described here with *SHANK3* loss of function mutation and a history of seizure, five (subjects 1, 2, 4, 11 and 13) were evaluated at least twice in clinic.

Patient 2 developed epilepsy at eight years of age with the first seizure being generalized tonic-clonic and lasting more than five minutes thus requiring intravenous medication. Seizures evolved to include atonic and myoclonic seizures at 9 years of age and tonic seizures by 10 years of age at which time he was diagnosed with Lennox-Gastaut syndrome. Medication history included initiation of 14 different anti-convulsants in multiple combinations. Ketogenic diet was also attempted, initially with reduction of seizure burden by 90% but then return of multiple daily seizures leading to its discontinuation. A vagus nerve stimulator was also implanted at 10 years of age with initially >50% reduction in seizure burden following escalation of therapy, but subsequent increasing seizure burden. Following multiple injuries from atonic seizures despite use of a protective helmet, corpus callosotomy was performed at 15 years of age with significant reduction in frequency and severity of atonic seizures for more than two years. Initial neuroimaging was normal but developed into T2 hyperintensity of the left hippocampus suggestive of mesial temporal sclerosis.

Patient 4 developed epilepsy at two years of age. The initial seizure type was atypical absence seizure and this remains her only semiology. Initial treatment with lamotrigine was not efficacious, and she was transitioned to zonisamide therapy with less than one seizure per month.

Patient 1 had her first seizure at 14 months of age which was an episode of febrile status epilepticus. This initial event was characterized by a tonic seizure with decreased responsiveness. This was followed by multiple daily atypical absence seizures at which point she was initiated on levetiracetam therapy which reduced her seizure burden from daily to less than one per month.

Patient 11 had a history of two lifetime seizures both occurring with fever. The first was at one year of age. Both lasted less than one minute in duration. This patient was never placed on anti-convulsant medication.

Patient 13's only seizure occurred at 6 years of age and was associated with illness. The seizure lasted for approximately 30 minutes requiring intravenous medication. She has never required daily anti-convulsant medication.

## Discussion

Here we present extensive data of seizure characterization and electroencephalogram abnormalities in individuals with *SHANK3* mutations. *SHANK3* haploinsufficiency is believed to be the etiology of the cognitive abnormalities in individuals with Phelan-McDermid Syndrome due to a similar phenotype in an individual with a balanced translocation disrupting *SHANK3* and multiple studies which have identified individuals with point mutations, indels or small duplications involving the *SHANK3* gene who have similar autistic traits and intellectual disability as individuals with chromosomal deletions encompassing *SHANK3*<sup>11; 14; 15</sup>. Epilepsy is present in a substantial number of individuals with Phelan-McDermid Syndrome with a pooled prevalence of 32% in unique studies available from the literature including this one. Whether or not mutations in *SHANK3* underlie the increased propensity for epilepsy in Phelan-McDermid Syndrome is unclear. In our cohort, we identified four individuals by exome sequencing with variants in *SHANK3* predicted to be deleterious. Of these, two developed epilepsy. Furthermore, the largest study to date of 22q13 deletions found no significant differences in deletion size in those with epilepsy and those without<sup>2</sup>. These data provide preliminary evidence that loss of function mutations in *SHANK3* can increase the likelihood to develop epilepsy; however, confirmation will require larger cohorts of individuals with indels and nonsense mutations in *SHANK3*.

One previous study investigating the seizure types associated with Phelan-McDermid Syndrome suggested that epilepsy associated with this disorder is mild and easily pharmacologically controlled<sup>16</sup>. We find in our cohort some individuals with *SHANK3* deletions do have infrequent and easily managed epilepsy, but we also identified individuals with intractable seizures including Lennox-Gastaut Syndrome. Lennox-Gastaut Syndrome

has been previously reported in one adult with a chromosome 22q13 deletion including the *SHANK3* gene<sup>17</sup>.

It is unclear exactly what distinguishes those individuals who will have a single seizure semiology and infrequent seizures from those that have multiple seizure types. There was no significant difference in deletion size, neuroimaging finding or other clinical characteristic (Table II). The only parameter that trended with multiple seizure types was slow or absent occipital dominant rhythm (five of six) in individuals with multiple seizure types versus those with a single seizure type (two of five). Together, these data demonstrate that medical practitioners must educate families caring for individuals with *SHANK3* loss of function mutations that epilepsy can become pharmaco-resistant and intractable.

Why haploinsufficiency of *SHANK3* would predispose to epilepsy is unclear. Multiple lines of mice with loss of function mutations in *SHANK3* have been reported to date<sup>10; 12; 13</sup>. Only one (*Shank3* alpha/beta isoform knockout) has been reported to have spontaneous seizures although these seizures were not characterized in detail<sup>10</sup>. Excitatory neurons differentiated from patient iPSCs with *SHANK3* deletions have been reported<sup>18</sup>. These neurons display multiple molecular and neurophysiologic abnormalities. Importantly, they have increased input resistance compared to control neurons. This indicates a tendency to increased excitability potentially underlying the elevated prevalence of epilepsy in individuals with *SHANK3* mutations compared with the general population.

Recently, a mechanism for the increased input resistance of stem cell derived neurons has been proposed<sup>19</sup>. Utilizing neurons derived from human embryonic stem cells with a targeted mutation of *SHANK3*, impairment of  $I_h$  currents was identified due to reduced expression of subunits of hyperpolarization-activated cyclic nucleotide-gated (HCN) channels. Moreover mutations in HCN channels have been clinically associated with epilepsy<sup>20</sup>. Intriguingly while the seizures were polymorphic in individuals with *HCNI* mutations, all patients reported in this study were diagnosed with atypical absence seizures. Together, these data suggest a mechanism by which haploinsufficiency of *SHANK3* might predispose to epilepsy and why there is a wide spectrum of seizure semiologies with atypical absence being the most common type of seizure in patients with *SHANK3* haploinsufficiency.

There are no consistent neuroimaging abnormalities that are likely to predispose to seizures. In particular, there is no evidence in our population or from previous reports of migrational abnormalities that are likely causative of their seizures. In our cohort, we found neuroimaging abnormalities as frequently in subjects with a history of seizures as those without. What is less clear is whether individuals with *SHANK3* mutations have microscopic neuroanatomic abnormalities or abnormalities in connectivity that might be responsible for both the cognitive dysfunction and propensity for epilepsy. This will require more advanced neuroimaging techniques such as diffusion tensor imaging or functional magnetic resonance imaging to determine.

## Summary

Loss of function mutations in *SHANK3* either through chromosomal deletion or point mutation predispose to seizures and electroencephalographic abnormalities. Atypical absence seizures are the most common seizures in individuals with *SHANK3* mutations; however, there is a wide spectrum of seizure types. No specific electroencephalographic abnormality is present in our cohort, and abnormalities were seen in both individuals with a history of seizures and those without. Neuroimaging in our cohort did not identify a specific abnormality associated with seizures. No pharmacologic therapies were clearly superior for treating patients in our cohort. Individuals with loss of function mutations in *SHANK3* should be monitored for seizure activity.

## Acknowledgments

We are indebted to our patients and their guardians for participating in our study. We thank Sunita Misra and James Orengo for critical review of this manuscript. Dr. Holder is supported by a grant from the Thrasher Research Fund (Early Career Award) and a K08 Clinical Scientist Career Development Award (NS091381).

## References

1. Tuchman R, Hirtz D, Mamounas LA. NINDS epilepsy and autism spectrum disorders workshop report. *Neurology*. 2013; 81:1630–1636. [PubMed: 24089385]
2. Sarasua SM, Boccuto L, Sharp JL, Dwivedi A, Chen CF, Rollins JD, Rogers RC, Phelan K, DuPont BR. Clinical and genomic evaluation of 201 patients with Phelan-McDermid syndrome. *Human genetics*. 2014; 133:847–859. [PubMed: 24481935]
3. Soorya L, Kolevzon A, Zweifach J, Lim T, Dobry Y, Schwartz L, Frank Y, Wang AT, Cai G, Parkhomenko E, et al. Prospective investigation of autism and genotype-phenotype correlations in 22q13 deletion syndrome and *SHANK3* deficiency. *Molecular autism*. 2014; 4:18.
4. Dhar SU, del Gaudio D, German JR, Peters SU, Ou Z, Bader PI, Berg JS, Blazo M, Brown CW, Graham BH, et al. 22q13.3 deletion syndrome: clinical and molecular analysis using array CGH. *American journal of medical genetics Part A*. 2010; 152A:573–581. [PubMed: 20186804]
5. Cusmano-Ozog K, Manning MA, Hoyme HE. 22q13.3 deletion syndrome: a recognizable malformation syndrome associated with marked speech and language delay. *American journal of medical genetics Part C, Seminars in medical genetics*. 2007; 145C:393–398.
6. Betancur C, Buxbaum JD. *SHANK3* haploinsufficiency: a "common" but underdiagnosed highly penetrant monogenic cause of autism spectrum disorders. *Molecular autism*. 2013; 4:17. [PubMed: 23758743]
7. Boeckers TM, Liedtke T, Spilker C, Dresbach T, Bockmann J, Kreutz MR, Gundelfinger ED. C-terminal synaptic targeting elements for postsynaptic density proteins ProSAP1/Shank2 and ProSAP2/Shank3. *Journal of neurochemistry*. 2005; 92:519–524. [PubMed: 15659222]
8. Arons MH, Thynne CJ, Grabrucker AM, Li D, Schoen M, Cheyne JE, Boeckers TM, Montgomery JM, Garner CC. Autism-associated mutations in ProSAP2/Shank3 impair synaptic transmission and neurexin-neuroligin-mediated transsynaptic signaling. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2012; 32:14966–14978. [PubMed: 23100419]
9. Duffney LJ, Zhong P, Wei J, Matas E, Cheng J, Qin L, Ma K, Dietz DM, Kajiwara Y, Buxbaum JD, et al. Autism-like Deficits in Shank3-Deficient Mice Are Rescued by Targeting Actin Regulators. *Cell reports*. 2015; 11:1400–1413. [PubMed: 26027926]
10. Peca J, Feliciano C, Ting JT, Wang W, Wells MF, Venkatraman TN, Lascola CD, Fu Z, Feng G. Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. *Nature*. 2011; 472:437–442. [PubMed: 21423165]
11. Cochoy DM, Kolevzon A, Kajiwara Y, Schoen M, Pascual-Lucas M, Lurie S, Buxbaum JD, Boeckers TM, Schmeisser MJ. Phenotypic and functional analysis of *SHANK3* stop mutations identified in individuals with ASD and/or ID. *Molecular autism*. 2015; 6:23. [PubMed: 26045941]

12. Kouser M, Speed HE, Dewey CM, Reimers JM, Widman AJ, Gupta N, Liu S, Jaramillo TC, Bangash M, Xiao B, et al. Loss of predominant Shank3 isoforms results in hippocampus-dependent impairments in behavior and synaptic transmission. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2013; 33:18448–18468. [PubMed: 24259569]
13. Wang X, McCoy PA, Rodriguiz RM, Pan Y, Je HS, Roberts AC, Kim CJ, Berrios J, Colvin JS, Bousquet-Moore D, et al. Synaptic dysfunction and abnormal behaviors in mice lacking major isoforms of Shank3. *Human molecular genetics*. 2011; 20:3093–3108. [PubMed: 21558424]
14. Bonaglia MC, Giorda R, Borgatti R, Felisari G, Gagliardi C, Selicorni A, Zuffardi O. Disruption of the ProSAP2 gene in a t(12;22)(q24.1;q13.3) is associated with the 22q13.3 deletion syndrome. *American journal of human genetics*. 2001; 69:261–268. [PubMed: 11431708]
15. Hamdan FF, Gauthier J, Araki Y, Lin DT, Yoshizawa Y, Higashi K, Park AR, Spiegelman D, Dobrzyniecka S, Piton A, et al. Excess of de novo deleterious mutations in genes associated with glutamatergic systems in nonsyndromic intellectual disability. *American journal of human genetics*. 2011; 88:306–316. [PubMed: 21376300]
16. Figura MG, Coppola A, Bottitta M, Calabrese G, Grillo L, Luciano D, Del Gaudio L, Torniero C, Striano S, Elia M. Seizures and EEG pattern in the 22q13.3 deletion syndrome: clinical report of six Italian cases. *Seizure*. 2014; 23:774–779. [PubMed: 25027555]
17. Lund C, Brodtkorb E, Rosby O, Rodningen OK, Selmer KK. Copy number variants in adult patients with Lennox-Gastaut syndrome features. *Epilepsy research*. 2013; 105:110–117. [PubMed: 23415449]
18. Shcheglovitov A, Shcheglovitova O, Yazawa M, Portmann T, Shu R, Sebastiano V, Krawisz A, Froehlich W, Bernstein JA, Hallmayer JF, et al. SHANK3 and IGF1 restore synaptic deficits in neurons from 22q13 deletion syndrome patients. *Nature*. 2013; 503:267–271. [PubMed: 24132240]
19. Yi F, Danko T, Botelho SC, Patzke C, Pak C, Wernig M, Sudhof TC. Autism-associated SHANK3 haploinsufficiency causes Ih channelopathy in human neurons. *Science*. 2016; 352:aaf2669. [PubMed: 26966193]
20. Nava C, Dalle C, Rastetter A, Striano P, de Kovel CG, Nabbout R, Cances C, Ville D, Brilstra EH, Gobbi G, et al. De novo mutations in HCN1 cause early infantile epileptic encephalopathy. *Nature genetics*. 2014; 46:640–645. [PubMed: 24747641]
21. Phelan MC, Rogers RC, Saul RA, Stapleton GA, Sweet K, McDermid H, Shaw SR, Claytor J, Willis J, Kelly DP. 22q13 deletion syndrome. *American journal of medical genetics*. 2001; 101:91–99. [PubMed: 11391650]
22. Wilson HL, Wong AC, Shaw SR, Tse WY, Stapleton GA, Phelan MC, Hu S, Marshall J, McDermid HE. Molecular characterisation of the 22q13 deletion syndrome supports the role of haploinsufficiency of SHANK3/PROSAP2 in the major neurological symptoms. *Journal of medical genetics*. 2003; 40:575–584. [PubMed: 12920066]
23. Luciani JJ, de Mas P, Depetris D, Mignon-Ravix C, Bottani A, Prieur M, Jonveaux P, Philippe A, Bourrouillou G, de Martinville B, et al. Telomeric 22q13 deletions resulting from rings, simple deletions, and translocations: cytogenetic, molecular, and clinical analyses of 32 new observations. *Journal of medical genetics*. 2003; 40:690–696. [PubMed: 12960216]
24. Manning MA, Cassidy SB, Clericuzio C, Cherry AM, Schwartz S, Hudgins L, Enns GM, Hoyme HE. Terminal 22q deletion syndrome: a newly recognized cause of speech and language disability in the autism spectrum. *Pediatrics*. 2004; 114:451–457. [PubMed: 15286229]
25. Lindquist SG, Kirchoff M, Lundsteen C, Pedersen W, Erichsen G, Kristensen K, Lillquist K, Smedegaard HH, Skov L, Tommerup N, et al. Further delineation of the 22q13 deletion syndrome. *Clinical dysmorphology*. 2005; 14:55–60. [PubMed: 15770125]
26. Jeffries AR, Curran S, Elmslie F, Sharma A, Wenger S, Hummel M, Powell J. Molecular and phenotypic characterization of ring chromosome 22. *American journal of medical genetics Part A*. 2005; 137:139–147. [PubMed: 16059935]
27. Denayer A, Van Esch H, de Ravel T, Frijns JP, Van Buggenhout G, Vogels A, Devriendt K, Geutjens J, Thiry P, Swillen A. Neuropsychopathology in 7 Patients with the 22q13 Deletion Syndrome: Presence of Bipolar Disorder and Progressive Loss of Skills. *Molecular syndromology*. 2012; 3:14–20. [PubMed: 22855650]



28. Nesslinger NJ, Gorski JL, Kurczynski TW, Shapira SK, Siegel-Bartelt J, Dumanski JP, Cullen RF Jr, French BN, McDermid HE. Clinical, cytogenetic, and molecular characterization of seven patients with deletions of chromosome 22q13.3. *American journal of human genetics*. 1994; 54:464–472. [PubMed: 7906921]

Author Manuscript

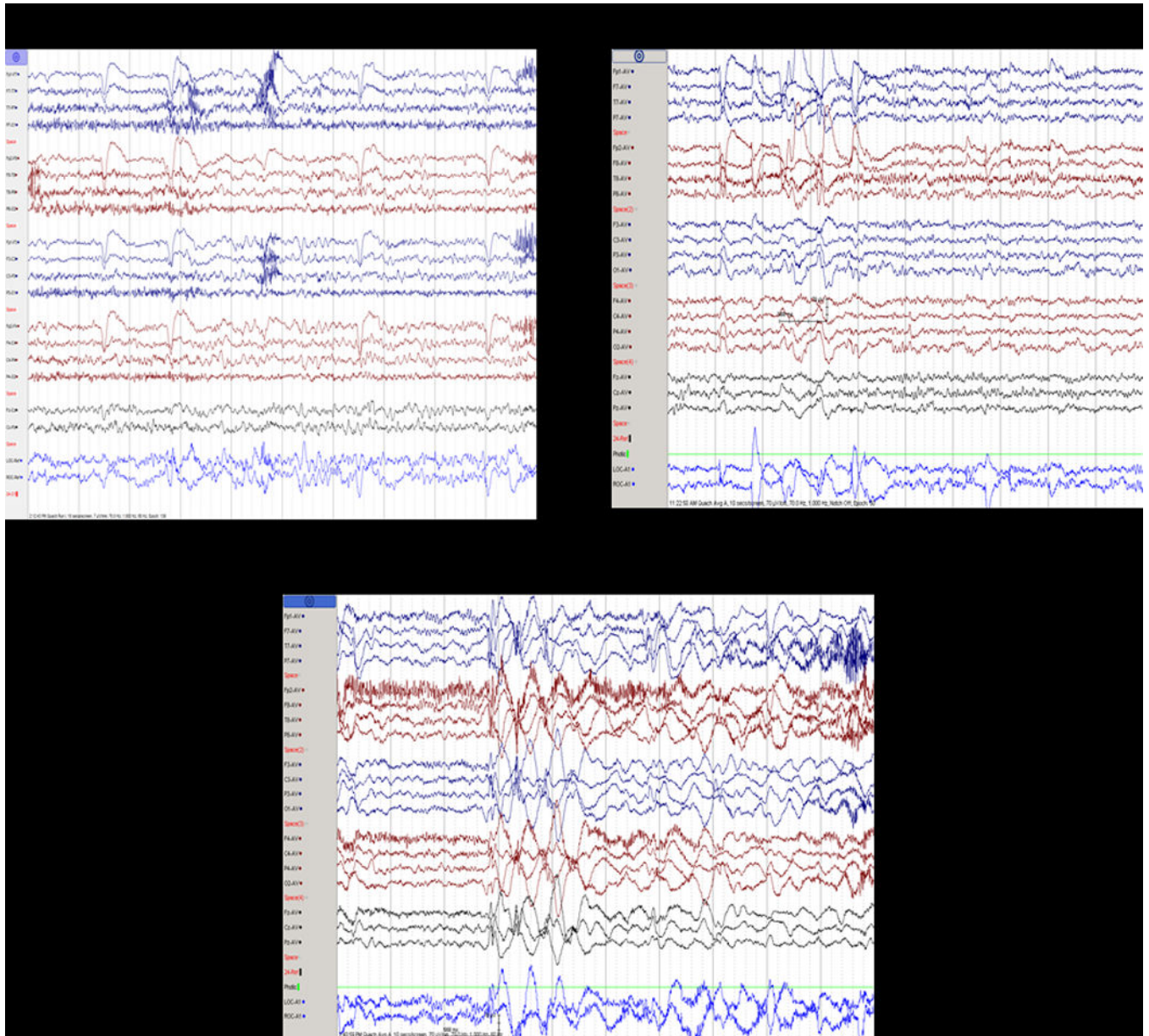
Author Manuscript

Author Manuscript

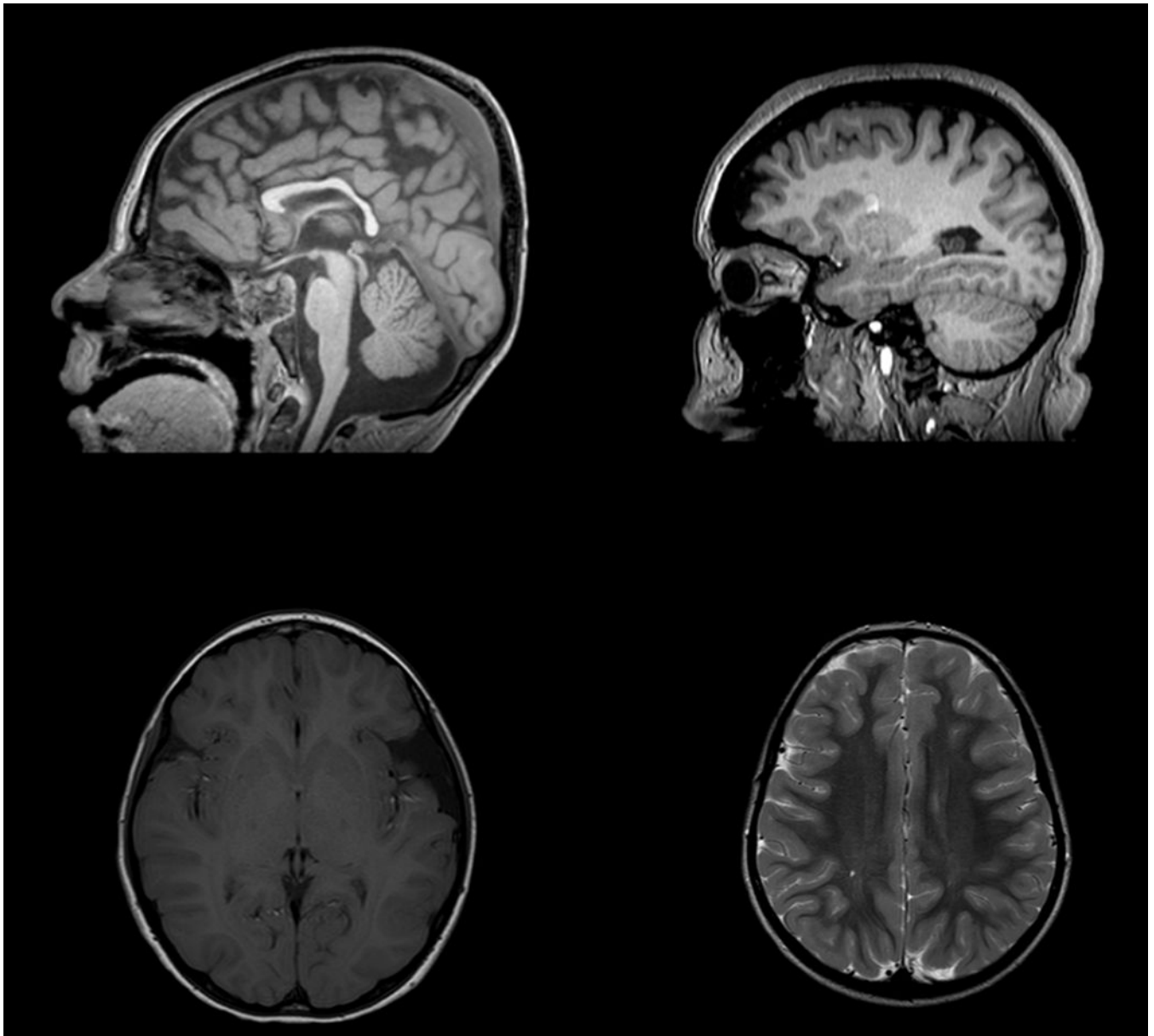
Author Manuscript

**Key Point Box**

- Seizures occur in greater than 30% of children with *SHANK3* mutations
- The most common seizure type is atypical absence
- A subset of patients with *SHANK3* mutations have medically intractable epilepsy
- There are no pathognomonic abnormalities on EEG in children with *SHANK3* mutations
- EEG abnormalities are seen in children harboring *SHANK3* mutations with and without history of seizures



**Figure 1.** Electroencephalograms in individuals with loss of function mutations in *SHANK3* (A) Subject 22 with generalized slowing of the occipital dominant rhythm (B) Subject 4 with frontal dominant generalized burst of 1.5 Hz spike and slow wave activity (C) Subject 2 with generalized burst of 1.5–2.5 Hz spike and slow wave activity.



**Figure 2.** Brain Magnetic Resonance Imaging (MRI) in individuals with loss of function mutations in *SHANK3* (A) Sagittal T1 weighted image of subject 9 demonstrating thinning of the corpus callosum (B) Sagittal T1 weighted image of subject 17 with mild cerebral volume loss (C) Axial T1 weighted image of subject 21 with left sylvian fissure arachnoid cyst (D) Axial T2 weighted image of subject 4 with mild T2 hyperintensity of the posterior centrum semiovale.

**Table I**

Seizure and EEG characterization from previous studies of Phelan-McDermid Syndrome

<b>Study</b>	<b>Prevalence of seizures</b>	<b>Types of seizures described</b>	<b>EEG findings</b>	<b>Number of patients</b>
<b>Phelan et al.<sup>21</sup></b>	27%	N/A	N/A	37
<b>Wilson et al.<sup>22</sup></b>	70%	N/A	N/A	51
<b>Luciani et al.<sup>23</sup></b>	24%	N/A	N/A	33
<b>Manning et al.<sup>24</sup></b>	27%	“Petit mal and focal”	N/A	11
<b>Lindquist et al.<sup>25</sup></b>	33%	N/A	N/A	6
<b>Jeffries et al.<sup>26</sup></b>	17%	N/A	N/A	30
<b>Cusmano et al.<sup>5</sup></b>	23%	N/A	N/A	107
<b>Dhar et al.<sup>4</sup></b>	30%	N/A	N/A	11
<b>Soorya et al.<sup>3</sup></b>	41%	“Generalized” (5/6), “partial-onset” (1/6)	34% with abnormality	32
<b>Sarasua et al.<sup>2</sup></b>	27%	N/A	N/A	151
<b>Figura et al.<sup>16</sup></b>	50%	“Myoclonic, tonic, generalized tonic-clonic”	Multifocal spike and wave	6
<b>Denayer et al.<sup>27</sup></b>	14%	N/A	N/A	7
<b>Nesslinger et al.<sup>28</sup></b>	14%	N/A	N/A	7

N/A = not available

**Table II**

Clinical Characteristics of Subjects

Sub-ject	Age	Genetic abnormality	Age first SZ	Frequency of SZ prior to medication	Frequency of SZ on medication	SZ semi-ology	Dura-tion of SZ	SE	EEG findings	Neuroimaging findings
1	2 years	22q deletion (44753814-51197838) hg19	14 month	daily	<1 per month	atypical absence, tonic	10-15 seconds	yes	excess beta activity, foci of SW activity - left central and parietal regions, intermittent slowing (delta) - bilateral occipital regions	thinning of corpus callosum
2	17 years		8 years	hundreds per day	daily	tonic, atypical absence, atonic, myoclonic, tonic-clonic	seconds to > 5 minute	yes	poorly organized ODR 6 Hz, frontocentral 8-10 Hz sharps evolving into 3-4 Hz spike and polyspike; bursts of generalized 1.5-2.5 Hz SW, intermittent generalized 3-5 Hz in 50% of recording	originally normal, evolved into left mesial temporal sclerosis
3	9 years	22q13.3 (GS-99K24 X1)	5 years	three times per week	2 times per month	atypical absence, tonic	15-20 seconds	no	ODR 7-8 Hz, frontal 1.5-2 Hz generalized SW/polyspike	normal
4	11 years		9 years	<1X/week	<1X/week	atypical absence	30 seconds	no	ODR 8 Hz with slower fused waveforms; multifocal SW/polyspike most frequent in the right frontal region, rare generalized SW	increased T2 signal in centrum semiovale, small corpus callosum
5	10 years	22q13.33 (49177451-49525470), 2p12 (76910620-77354922 x1) hg18	none	none	N/A	N/A	N/A	no	None	normal
6	6 years	22q13.33 (49484191-49525470 X1) hg18	2 years	10-20 x per day	N/A	atypical absence	20 sec - 1 min	no	ODR 8-8.5 Hz with some slower activity; active foci of SW in isolation and in runs in the left and right temporo-parietocentral regions	brachycephalic, otherwise normal
7	12 years	22q13.33 (48771374-51178264 X1); 7q31.1 (111056675-111191737 X1) hg19	none	none	N/A	N/A	N/A	no	ODR 6-7 Hz poorly developed and sustained; sharp waves and spikes over left and right temporal; anterior temporal and frontal regions independently	T2 prolongation of subcortical white matter bilateral temporal poles, slightly foreshortened corpus callosum

Sub- ject	Age	Genetic abnormality	Age first SZ	Frequency of SZ prior to medication	Frequency of SZ on medication	SZ semi- ology	Dura- tion of SZ	SE	EEG findings	Neuroimaging findings
8	15 years	22q13 deletion (no data)	5 years	hundreds per day	1–2 per day	atonic, atypical	seconds to >5 minute	yes	ODR 4–5 Hz; low amplitude spike discharges - left central; polyspike at right frontoparietal	arachnoid cyst
9	5 years	22q13.33 (189216-8185497 X3) hg19	none	none	N/A	N/A	N/A	no	ODR 8–9 Hz; normal	mild increased FLAIR signal in periventricular white matter of frontal, parietal and temporal lobes, moderate reduction in cerebral white matter volume including corpus callosum thinning, mild decreased size of the left hippocampus
10	12 years	22q13.2-3(43572964–51171678 X1)	3 years	daily	1/2–3 weeks	atypical absence, tonic	minute to hours	yes	no ODR, right centroparietooccipital sharps	mild thinning of the corpus callosum, generalized loss of white matter
11	4 years	22q13.2-3(43572964–51171678 X1)	1 year	2 lifetime with fever	N/A	tonic	1 minute	no	ODR 6 Hz with slower waveforms; during sleep - bursts of generalized anterior dominant SW/polyspike	mild hypoplasia of the posterior body of the corpus callosum, large cystic cavum septum pellucidum
12	17 years	22q13.2-3 (43014031–51171678 X1)	none	none	N/A	N/A	N/A	no	ODR 5–6 Hz; SW in right parietal and frontal	curvilinear cortical T2 hyperintensity right extreme capsule, infratentorial/ supratentorial volume loss, persistent cavum septum pellucidum et vergate, giant cisterna magna
13	13 years	22q13.31-3 (45680160–51178264 X1) 2q37.3 (241994956–242938241 X3) hg19	6 years	one	N/A	atypical absence	30 minute	yes	ODR 8 Hz, normal	none
14	28 years	22q13.31q13.33 deletion by FISH	none	none	N/A	N/A	N/A	no	normal per parental report	none
15	19 years	22q13.33 (49451411–49525130 X1) hg18	none	none	N/A	N/A	N/A	no	10–11 Hz, normal	cerebellar ectopia
16	7 years	22q13.33 (G248p86064C8– G248p86149G7 X1) BAC array	none	none	N/A	N/A	N/A	no	ODR 10–11 Hz, excessive beta activity	normal
17	16 years	22q13 deletion by FISH	none	none	N/A	N/A	N/A	no	ODR 9.5–10 Hz; occasional moderate voltage sharp waves right frontocentral regions near midline and right centrotemporal regions occasionally in serial	left sylvian fissure arachnoid cyst; mild cerebral volume loss, partial empty sella

Sub- ject	Age	Genetic abnormality	Age first SZ	Frequency of SZ prior to medication	Frequency of SZ on medication	SZ semi- ology	Dura- tion of SZ	SE	EEG findings	Neuroimaging findings
18	12 years	22q13.33 (49469317-49525130 X1) hg18	none	none	N/A	N/A	N/A	no	none	normal
19	9 years	22q13.33 (50103257-51178264 x1) hg19	none	none	N/A	N/A	N/A	no	none	none
20	3 years	22q13.33 (51137,176-51,197,838 X1)	none	none	N/A	N/A	N/A	no	poorly sustained ODR of 8 Hz	persistent cavum septum pellucidum et vergae
21	13 years	15q11.2 dup; SHANK3 c.2313+1G>A	none	none	N/A	N/A	N/A	no	ODR 8-9 Hz, normal	left sylvian fissure arachnoid cyst
22	14 years	SHANK3 indel (q51160326-51160327)	14 years	unknown	1/week	atypical absence	3 minute	no	no ODR	normal except mild cerebellar tonsillar ectopia
23	14 years	SHANK3 indel (q51160326-51160327)	7 years	multiple times per week	2-3 times per week	atypical absence, tonic	3-5 minutes	no	ODR 9-10 Hz, normal	normal
24	13 years	SHANK3 c.3727dupG(p.A1212fs)	none	none	none	none	none	no	Normal	bilateral T2 hyperintensities of posterior centrum semiovale

ODR = Occipital Dominant Rhythm; SW = spike and slow wave