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Themis2: setting the threshold for B-cell selection

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The strength of signal downstream of the B-cell receptor (BCR) determines the positive and negative selection of B cells. Expression of a functional BCR is required for the selection of immature B cells into the peripheral repertoire in the absence of antigen; but selection is also modulated by avidity and affinity-dependent interactions with self and foreign antigens. These antigendependent events shape the B-cell repertoire during ontogeny, development and the response to pathogens, and generation of B-cell memory; however, the mechanisms underlying these processes are not very well understood.

During early ontogeny, interactions with intracellular or low abundance self-antigens allow positive selection of innate-like B1 B cells.¹ In contrast, the presence of abundant soluble or lowavidity self-antigen leads to B-cell anergy, whereas stronger signals from highly abundant or multivalent antigens trigger receptor editing and/or negative selection of immature B cells through central tolerance.² These processes must be determined by the state and environment of the B cells, defined in part by the presence of co-stimulation or other signals. Later in the periphery, antigens of similar affinity or avidity elicit B-cell activation, leading to rapid proliferation, plasma cell differentiation and antibody production. During an immune response to pathogens, the majority of B-cell activation involves interactions with lower avidity and/or low-affinity antigens in a soluble, potentially tolerogenic form or coupled via complement or Fc receptors, notably arrayed on the membranes of macrophages, B cells and follicular dendritic cells.^{3,4} However, despite the importance of antigen quality and quantity in modulating B-cell responses, very little is understood about the process that defines the signalling threshold between positive and negative selection, allowing B cells to discriminate between antigens of varying strength and form.

In a recent paper published in Nature Immunology, Cheng et al.5 show how Themis2, a member of the newly described Themis-family of proteins, modulates the positive selection of B cells in response to self and foreign antigen. Themis2 determines the threshold for activation of B cells by lowaffinity and low-avidity ligands via PLCy2 activation and ERK1/2-dependent pathways (Figure 1). These effects were detected using the immunoglobulin transgenic MD4 mouse model and the cognate antigen 'hen egg lysozyme' (HEL). Themis2 has no known catalytic domain and potentially functions as an adaptor protein by strengthening the interactions between PLCy2 and the upstream kinase Lyn. Loss of Themis2

raises the threshold for the antigeninduced upregulation of CD69, which is required for the retention and proliferation of activated B cells in the lymph node, and reduces the expression of CD86, which is required for receiving 'co-stimulatory' T-cell help. Therefore, Themis2 may play an important part in the initial immune response, by increasing the sensitivity of naive B cells to rare and low-affinity antigens.

The Themis-family of proteins is evolutionarily conserved in metazoans from mammals to cnidarians and is characterized by one or more copies of a novel cysteine containing globular CABIT domain.⁶ Themis2 has two tandem CABIT domains and has two close homologues: mammalian Themis1, expressed primarily in T cells, and Themis3, expressed in the intestine. In addition, mammals express two family members of unknown function, with single copies of the CABIT domain and a proline-rich region, Fam59A (Garem) and FAM59B. Like Themis2, Themis1 is proposed to be an analogue-to-digital convertor, which translates low-affinity signals into a selection event during the positive and negative selection of CD4⁺ CD8⁺ thymocytes. In the absence of Themis1, T-cell development is blocked at the CD4⁺CD8^{lo} positive selection point.^{6–9} Themis1 is constitutively associated with Grb2 via its proline-rich domain and is rapidly phosphorylated by Lck and ZAP70 and recruited to LAT upon T-cell activation.

Although research to elucidate the function of Themis1 demonstrates our

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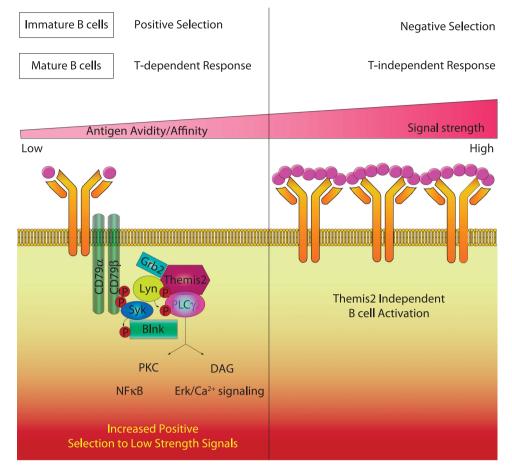


Figure 1 Themis2 increases the positive selection of B cells to low-avidity or low-affinity self and foreign antigens during formation of the repertoire and during the immune response.

relatively better understanding of the role of avidity and affinity during T-cell activation and development compared to B cells, there remain contrasting reports about the mode of action of Themis1. In 2013, it was suggested that Themis1 had a role in transmitting T-cell receptor (TCR) signals from low affinity, but not high affinity antigens by attenuating, rather than increasing, signal strength, thus modulating negative selection.¹⁰ The authors compared developmentally matched OVA-specific T cells and reported increased TCR-dependent signalling due to low-affinity ligands in the absence of Themis1. From these results, they proposed that in the absence of Themis1, signals generated from weak TCR-ligand interactions are enhanced, partly due to a failure to activate the inhibitory phosphatase SHP1 and/or recruit SHP1 to LAT. In contrast, Choi et al.11 more recently reported in a second Nature Immunology paper that Themis1-deficient cells have elevated SHP1 activity, and that Themis1 positively regulates T-cell development by a novel mechanism involving the suppression of SHP1 by the Themis2 CABIT domain. Deficiency in SHP1 rescues T-cell development in Themis1-deficient thymocytes. Themis2 is functionally interchangeable with Themis1 in T-cell development, implying that they may have a similar mechanism of action.¹² However, cell-specific dissimilarities, including redundancy and a significant difference in the level of expression of phosphatases and Themis1 and 2 in T and B cells may mean this interpretation is too simplistic. Competition between positive and negative regulators linked to Themis-family proteins as well as other binding partners, including inhibitory co-receptors might modulate

the differential signalling outcomes of B and T cells.

In reporting the characterization of a novel protein that modulates signal transduction downstream of the BCR, the paper by Cheng *et al.* raises questions about how B cells discriminate between qualitatively different antigenic signals received via the same BCR and how antigens of different quantity and quality fine-tune the immune response. It also emphasizes the need to adopt new approaches and develop tools to measure these effects experimentally.

Although polyvalent membranebound antigens are better able to induce antibody responses and activate B cells,¹³ it has long been known that soluble, low valency antigens are also able to activate B cells *in vitro* and *in vivo*.¹⁴ In addition to the maintenance of tolerance and survival by low-affinity interactions and tonic signalling, B-cell responses to antigens of relatively low avidity are likely to be important in most physiological settings. The decision to differentiate into an extrafollicular plasma cell versus becoming a germinal centre B cell during the initial phase of T-dependent antibody responses is likely governed by the nature of the interaction between antigen and BCR.¹⁵

However, in vitro and biochemical studies of B-cell activation have almost exclusively used high avidity analogues of T-independent antigens, typically anti-IgM $F(ab')_2$. These potent forms of antigenic stimulation are likely to miss subtle, but important, aspects of differential BCR signalling. Analogous to the use of antigens of different affinity T-cell responses, assess similar to approaches using cognate antigens of different affinity and avidity, and with a defined BCR specificity, may offer valuable insights into the selection of B cells.

It was reported in a separate study that Themis2-deficiency had no effect on B-cell development or antibody titres following immunization with NP-Ficoll and NP₂₁-CGG or B-cell activation by anti-IgM *in vitro*, all of which are higher avidity forms of antigen.¹⁶ Only when analysis of B-cell responses to lower valency forms of antigen such as NP₃-CGG or soluble HEL is carried out, does the role of Themis2 in establishing a signalling threshold and modulating responses to lower avidity antigens become evident.⁵

Future studies will want to investigate the role of other molecules that allow signal discrimination during situations where antigen is most limiting: for example, during the positive selection of B1 B cells and marginal zone B cells or during the germinal centre response, where B-cell selection must be exquisitely regulated to eliminate self-reactive clones, while allowing progressively enhanced affinity maturation via competition for limiting antigen. A layer of complexity may be added to these studies by employing antigens of variable avidity and affinity, thus better dissecting the processes that fine-tune BCR activation outcomes. Untangling the role of localized cellular signalling and how this is dependent on different environmental cues and developmental stages, as well as on the nature and dose of antigens, will give us a better understanding of how B cells integrate upstream signals and switch between different outcomes.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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