

Effects of *Plantago major* Extracts and Its Chemical Compounds on Proliferation of Cancer Cells and Cytokines Production of Lipopolysaccharide-activated THP-1 Macrophages

Kartini, Suratsawadee Piyaviriyakul¹, Suchitra Thongpraditchote², Pongpun Siripong¹, Omboon Vallisuta³

Pharmaceutical Biology Department, Faculty of Pharmacy, University of Surabaya, Surabaya 60293, Indonesia, ¹Research Division, Natural Products Research Section, National Cancer Institute, Departments of ²Physiology and ³Pharmacognosy, Faculty of Pharmacy, Mahidol University, Bangkok 10400, Thailand

Submitted: 16-09-2016

Revised: 23-11-2016

Published: 19-07-2017

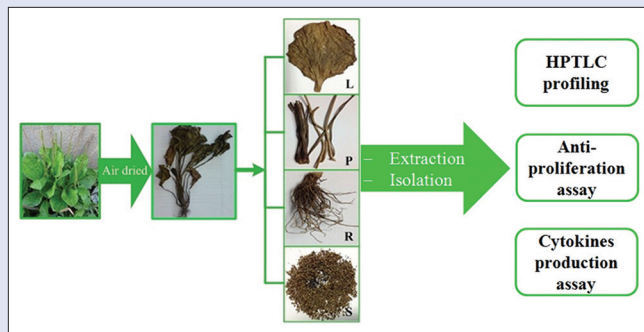
ABSTRACT

Background: *Plantago major* has been reported to have anticancer and anti-inflammatory properties. However, its antiproliferative and anti-inflammatory mechanisms have not been fully elucidated. Moreover, which plant parts are more suitable as starting materials has not been explored. **Objectives:** To investigate the antiproliferative activity of *P. major* extracts against MCF-7, MDA-MB-231, HeLaS3, A549, and KB cancer cell lines as well as their effects on inflammatory cytokines (tumor necrosis factor [TNF]- α , interleukin [IL]-1 β , IL-6, and interferon [IFN]- γ) production by lipopolysaccharide (LPS)-stimulated THP-1 macrophages. **Materials and Methods:** The methanol and aqueous extracts of *P. major* from different plant parts and its chemical compounds, i.e., ursolic acid (UA), oleanolic acid (OA), and aucubin were tested in this experiment. **Results:** Methanol and aqueous extracts of *P. major* seeds exhibited the greatest antiproliferative activity. The methanol extracts of seeds also demonstrated the highest inhibition of TNF- α , IL-1 β , IL-6, and IFN- γ production. Interestingly, the roots, which were commonly discarded, exhibited comparable activities to those of leaves and petioles. Furthermore, UA exhibited stronger activities than OA and aucubin. **Conclusions:** The seeds are being proposed as the main source for further development of anticancer and anti-inflammatory products, whereas the roots could be included in the preparation of *P. major* derived products with respect to anti-inflammatory. **Key words:** Anti-inflammatory, antiproliferative, cytokines, *Plantago major*, THP-1 macrophages

SUMMARY

- Amongst the parts of *Plantago major*, seeds exhibited the greatest antiproliferative activity against MCF-7, MDA-MB-231, HeLaS3, A549, and KB cell lines as well as the highest inhibition on TNF- α , IL-1 β , IL-6, and IFN- γ production
- The roots, which were commonly discarded, exhibited comparable antiproliferative and cytokines inhibition activities to those of leaves and petioles

- Ursolic acid, a chemical compound of *Plantago major*, exhibited stronger activities than oleanolic acid and aucubin
- The seeds are being proposed as the main source for further development of anticancer and anti-inflammatory products, whereas the roots could be included in the preparation of *Plantago major* derived products with respect to anti-inflammatory.



Abbreviations used: TNF: Tumor Necrosis Factor; IL: Interleukin; IFN: Interferon; HPTLC: High Performance Thin Layer Chromatography; UA: Ursolic Acid; OA: Oleanolic Acid; AUC: Aucubin.

Correspondence:

Dr. Kartini,
Faculty of Pharmacy, University of Surabaya,
Raya Kalirungkut Road, Surabaya 60293,
Indonesia.
E-mail: kartini@staff.ubaya.ac.id
DOI: 10.4103/pm.pm_406_16

Access this article online

Website: www.phcog.com

Quick Response Code:



INTRODUCTION

Plantago L. (Plantaginaceae) is a genus widely distributed all over the world, and more than 275 species have been reported.^[1] *Plantago major* [Figure 1] is an endemic species found in Indonesia. This plant has both perennial and annual growth habit, with ovoid or elliptic leaves. The leaves are normally green, sometimes with purple shading and are hairless or hairy on the surface, with 5–9 veins in parallel venation. Flowering of *P. major* is a spike which arises up to 30 cm in length. The fruit is a capsule (5 mm in length) which produced on a spike. Every centimeter of spike contains 23–26 capsules, and each capsule carries 4–15 seeds.^[2–4] It is used traditionally and commercially available.^[5] This herb can be found in extremely different geographical origins. Previous studies have indicated a variety of beneficial pharmacological effects of *P. major* such as anticancer,^[6–9] antioxidant,^[1,10,11] and immunomodulatory.^[6,12] Concerning its anti-inflammatory activity, several *in vivo* experiments

were carried out to prove this property.^[13,14] Inflammatory responses involve critical immune cells, macrophages, which are able to produce various inflammatory mediators including cytokines (tumor necrosis factor [TNF]- α , interleukin [IL]-1 α β , IL-6, IL-8, IL-10, and IL-12), eicosanoid derived from arachidonic acid, and reactive oxygen

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

Cite this article as: Kartini, Piyaviriyakul S, Thongpraditchote S, Siripong P, Vallisuta O. Effects of *Plantago major* extracts and its chemical compounds on proliferation of cancer cells and cytokines production of lipopolysaccharide-activated THP-1 macrophages. Phcog Mag 2017;13:393-9.



Figure 1: *Plantago major*

species.^[15] Various *in vitro* studies demonstrated the anti-inflammatory potency of *P. major* and its active compounds through inhibition of cyclooxygenases and lipoxygenases, enzymes involved in arachidonic acids pathways.^[16-18] Anti-inflammatory activity of *P. major* through modulation of inflammatory cytokines, however, has not been well characterized.

In terms of plant parts, leaves of *P. major* were most frequently used mainly as aqueous preparations, whereas roots were utilized in limited country, especially Iran.^[3] In many other countries, including Indonesia, roots are commonly discarded without any utilization. Nevertheless, our previous studies showed that root extracts exerted cytotoxicity on SiHa and HepG2 cell lines as well as possessed free radical scavenging activity which were comparable to those of the other plant parts.^[5] To more understand the anti-inflammatory actions of *P. major*, we investigated the effect on cytokines (TNF- α , IL-1 β , IL-6, and interferon [IFN]- γ) production in LPS-activated THP-1 macrophages cells by *P. major* extracts and its chemical compounds. Their antiproliferative effect against KB, MCF-7, MDA-MB-231, A-549, and HeLaS3 cell lines was determined as well.

MATERIALS AND METHODS

Chemicals

Silica gel, alumina, and high-performance thin-layer chromatography (HPTLC) plates precoated with silica gel 60 F₂₅₄ were purchased from Merck KGaA (Darmstadt, Germany). Ursolic acid (UA), oleanolic acid (OA), aucubin, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), sodium pyruvate, glucose, phorbol 12-myristate 13-acetate (PMA), and lipopolysaccharide (LPS, O127:B8) were obtained from Sigma (St. Louis, MO, USA). Dimethyl sulfoxide (DMSO) and cytokine kit (MILLIPLEX[®] MAP Kit, product code HCYTOMAG-60K-05) were obtained from Merck KGaA (Darmstadt, Germany). RPMI-1640 medium, Modified Eagle Medium (MEM), fetal bovine serum (FBS), 2-mercaptoethanol, phosphate buffer saline, and penicillin-streptomycin solution were purchased from Gibco Life Sciences (GibcoThai, Thailand). All solvents used were analytical grade and obtained from Labscan Asia (Bangkok, Thailand).

Plant materials

P. major samples were collected from two locations in Indonesia, i.e. Lumajang and Tawangmangu. Whole plant materials were harvested

and cleaned with tap water. Lumajang sample was separated into leaves (L1), petioles (P1), roots (R1), and seeds (S1) which were used for bioactivity assays. The aerial part of the sample from Tawangmangu (A1) was used for isolation of the chemical compounds. Authentication of the plants (No. 1101/D.T/XI/2013) was conducted by Prof. Sutarjadi (Center of Information and Development of Herbal Medicine, University of Surabaya, Indonesia). All samples were dried and ground as described in previous work.^[5]

Preparation of the extracts

P. major extracts were prepared according to the previous study.^[5] Briefly, 50 g of plant materials was extracted twice by boiling in 500 ml distilled water for 1 h each. The extracts were filtered and concentrated *in vacuo* by a rotary evaporator (Buchi, Switzerland) and then were lyophilized to dryness. For the methanol extracts, 10 g of plant materials was macerated with methanol (3 x 100 ml, 3 x 24 h) at room temperature. The extracts were combined, filtered, and evaporated under vacuum to dryness. The aqueous and methanol extracts of leaves, petioles, roots, and seeds were obtained.

All crude extracts were determined for the antiproliferative activity against several cancer cells in comparison with doxorubicin, an anticancer drug. Moreover, the effects of the methanol extracts on cytokines production of THP-1 macrophages were also investigated.

High-performance thin layer chromatography profiling and isolation of the chemical compounds from *Plantago major*

High-performance thin layer chromatography profiling

To check the TLC profile of *P. major* extracts, chromatography was performed with two different solvent systems, i.e., (A) toluene: acetone: formic acid (78:22:0.15) and (B) 1,4 dioxane: xylene: propane-2-ol: 12.5% NH₃ (1:2:5:2). Samples were spotted by Camag Linomat 5 sample applicator on the HPTLC plates precoated with silica gel 60 F₂₅₄. Before development under the first solvent system (A), spotted samples were prederivatized with iodine vapor as described earlier.^[5] Development was carried out with 10 ml of each solvent system in a Camag Twin Trough Chamber (10 cm x 10 cm) previously equilibrated with the mobile phase for 20 min at room temperature. After development, the plates were dried under warm air and subjected to derivatization by dipping in 5% sulfuric acid in methanol. TLC plates were further dried in a fume hood and then heated on a Camag TLC Plate Heater III (120°C, 3 min). The photographs were recorded by Camag Reprostar 3 under white light.

Isolation of the chemical compounds

Two kilograms of aerial part of the sample from Tawangmangu (A1) was extracted with methanol (6 x 10 L) by maceration at room temperature for 6 x 24 h. The combined extracts were then evaporated under vacuum to dryness (333 g). A portion (67 g) of this crude extract was subjected to a vacuum silica gel column chromatography eluted with n-hexane, dichloromethane, and methanol by increasing polarity. The like fractions were combined, evaporated *in vacuo* to dryness, and yielded five fractions (F1-F5). F4 and F5 were combined and further subfractionated in an alumina column and eluted by increasing polarity of ethyl acetate and methanol yielding 100 fractions. The combination of fractions 47-50 (325 mg) was then subjected to another silica gel column and development with ethyl acetate and methanol by increasing polarity afforded 177 subfractions. Subfraction 150 exhibited a single spot on a TLC plate at R_f value of 0.33 in the 1,4-dioxane: xylene: propane-2-ol:12.5%NH₃ (1:2:5:2) solvent system. After recrystallization in methanol: ethyl acetate, the white crystalline compound was obtained. The identification of this isolated compound

was confirmed by comparison of the spectral data of infrared (IR), proton nuclear magnetic resonance ($^1\text{H-NMR}$), Carbon-13 nuclear magnetic resonance ($^{13}\text{C-NMR}$), and electrospray ionization mass spectrometry (ESI-MS) with those in literature.

Antiproliferation assay on cancer cells

Cell culture

Human breast cancer (MCF-7, ATCC[®] HTB-22[™] and MDA-MB-231, ATCC[®] HTB-26[™]), human cervical cancer (HeLaS3, ATCC[®] CCL-2.2[™]), human lung cancer (A549, ATCC[®] HTB-22[™]), and human nasopharynx carcinoma (KB, ATCC[®] CCL-17[™]) cell lines were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). The cells were maintained in MEM or DMEM, supplemented with 10% heat-inactivated FBS, 100 U/ml penicillin and 100 $\mu\text{g/ml}$ streptomycin, and incubated at 37°C in a humidified atmosphere containing 5% CO_2 and 95% air.

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide cell proliferation assay

Cellular viability in the presence or absence of *P. major* extracts and three pure compounds, i.e. UA, OA, and aucubin, was examined using the MTT colorimetric assay as described previously.^[19] Briefly, cancer cells under log phase of growth at the density of 1×10^3 cells/ml in 100 μl of culture medium were seeded onto 96-well culture plate (Costar, Cambridge, MA, USA). After 24 h of preincubation, various concentrations of the tested extracts or pure compounds were added and then incubated for a further 72 h. At the end of incubation, 20 μl of MTT solution (5 mg/ml) was added to each well and further incubated at 37°C for 3 h. The MTT formazan product was dissolved in DMSO and evaluated by measuring the absorbance at 550 nm using a microplate reader (Benchmark 550, Bio-Rad, USA).

Quantification of pro-inflammatory cytokines production in lipopolysaccharide-activated THP-1 macrophages

Cell culture and differentiation

THP-1 (human monocytic leukemia, ATCC[®] TIB-202[™]) cell line was grown in suspension in RPMI-1640 medium, supplemented with 0.05 mM 2-mercaptoethanol, 2 mM glutamine, 1 mM sodium pyruvate, 990 mg/L glucose, 10% FBS, and 100 U/ml penicillin-100 $\mu\text{g/ml}$ streptomycin. The cells were maintained in a humidified incubator (5% CO_2 , at 37°C). Media were changed once every 3–4 days. Cells were disposed and changed by frozen cells every 20 passages.

The macrophage-like state was obtained by treating THP-1 monocytes for 48 h with 40 nM PMA in FBS-free medium at a density of 5×10^5 cells/ml in 96- and 24-well plates (100 μl and 1 ml, respectively). Furthermore, PMA-stimulated THP-1 cells (referred to as THP-1 macrophages) were washed with FBS-free medium and rested for another 24 h in the FBS-free medium and treatments were then initiated.

Cytotoxicity test

Cytotoxicity of *P. major* extracts and its pure compounds on THP-1 macrophages was assessed using the MTT method. THP-1 macrophages were incubated with a series concentration of extract (1–1000 $\mu\text{g/ml}$) in 96-well plate for 48 h. Cells were subsequently incubated with 100 μl of 1 mg/ml MTT in FBS-free medium for 3 h. After centrifugation (1200 rpm) at 4°C for 5 min, the medium was aspirated. The formazan product was dissolved in 100 μl DMSO, and the absorbance was further measured at 550 nm using a microplate reader. The absorbance of formazan formed in control cells (without extracts or pure compounds) was referred as 100% viability.

Multiplex quantification of pro-inflammatory cytokines production in lipopolysaccharide-activated THP-1 macrophages

THP-1 monocyte cells were plated at a density of 5×10^5 cells/ml in 24-well plate, differentiated with PMA, and subsequently incubated with the corresponding extracts or pure compounds in appropriate concentrations. One hour later, LPS was added to each well at the final concentration of 5 $\mu\text{g/ml}$ and incubated for another 47 h. At the end of incubation, the plates were centrifuged (1200 rpm) for 5 min at 4°C. The supernatant in each well was aspirated and kept in -80°C until cytokines determination. TNF- α , IL-1 β , IL-6, and IFN- γ concentrations in the supernatants were quantified simultaneously using MILLIPLEX[®] MAP kit according to the manufacturer's instruction (Merck KGaA, Darmstadt, Germany).

Statistical analysis

All values were presented as mean \pm standard error mean or mean \pm standard deviation, $n = 2-3$. For multiple variables comparison, data were analyzed by ANOVA followed by Dunnett's multiple comparison and Tukey's test when necessary using GraphPad Prism statistical software (GraphPad Software Inc., Sandiego, California, Windows Version 5.01). Differences were considered statistically significant at $P < 0.05$.

RESULTS

High-performance thin layer chromatography profiling and chemical structure of the isolated compound

To evaluate the chemical composition of *P. major*, HPTLC profiling of each extract was carried out using two different systems [Figure 2]. It was shown that all extracts contain UA (Rf value: 0.53), whereas OA (Rf value: 0.71) could be found in petioles and seeds only. Isolation of UA from the aerial part as well as determination of UA and OA concentration in different parts of *P. major* has been reported in our previous study.^[5] Contrastingly, aucubin (Rf value: 0.39) was detected in all extracts [Figure 2b].

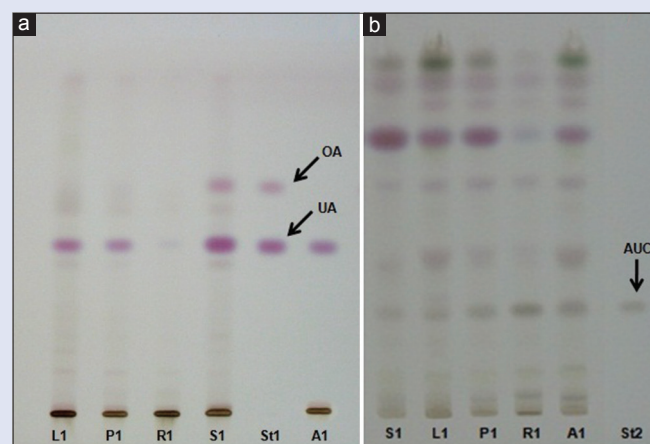


Figure 2: High-performance thin layer chromatography profile of *Plantago major* extracts. Solvent system (a) toluene:acetone:formic acid (78:22:0.15) and (b) 1,4 dioxane:xylene:propan-2-ol:12.5% NH_3 (1:2:5:2). L1, P1, R1, and S1 are methanol extracts of leaves, petioles, roots, and seeds of *Plantago major* from Lumajang, respectively. A1 is methanol extract of aerial part of *Plantago major* from Tawangmangu. St1 and St2 are ursolic acid-oleanolic acid and aucubin (AUC) standard, respectively

The isolated compound from this study was a white crystal with melting point 174–177°C, UV λ_{\max} (MeOH) 225.5 nm. IR ν_{\max} (KBr) 3289.83, 2914.38, 2882.50, 2359.80, 1652.67, and 1480.94 cm^{-1} ; ESI-MS m/z 369.1169 [M + Na]⁺; ¹H NMR (DMSO- d_6) δ 4.99 (1H, m, H-1), 6.27 (1H, dd, H-3), 4.81 (1H, d, H-4), 2.49 (1H, m, H-5), 4.27 (1H, br s, H-6), 5.62 (1H, br s, H-7), 2.71 (1H, br t, H-9), 3.93 (1H, d, H-10A), 4.13 (1H, d, H-10B), 4.48 (1H, d, H-1'), 2.94–3.65 (1H, H-2'-6'B); ¹³C NMR (DMSO- d_6) δ 95.4 (C-1), 140.1 (C-3), 105.1 (C-4), 44.6 (C-5), 80.5 (C-6), 129.2 (C-7), 146.2 (C-8), 46.5 (C-9), 59.6 (C-10), 98.2 (C-1'), 73.5 (C-2'), 76.2 (C-3'), 70.2 (C-4'), 77.2 (C-5'), 61.1 (C-6'). These results were in concordance with those previously reported for aucubin.^[20] Therefore, it can be concluded that the isolated compound was aucubin [Figure 3].

Effects of *Plantago major* extracts and its chemical compounds on cancer cells proliferation

Antiproliferative activity of *P. major* extracts and its chemical compounds against various cancer cells [Table 1] was evaluated *in vitro* by the MTT colorimetric assay. Among the extracts tested, methanol extracts of *P. major* seeds exhibited the most potent antiproliferative activity against five cancer cell lines used with IC_{50} values between 153.38 and 247.41 $\mu\text{g}/\text{ml}$. In terms of pure compounds, UA showed the strongest antiproliferative activity against all cancer cell lines used with IC_{50} values between 6.27 and 18.33 $\mu\text{g}/\text{ml}$, followed by OA (IC_{50}

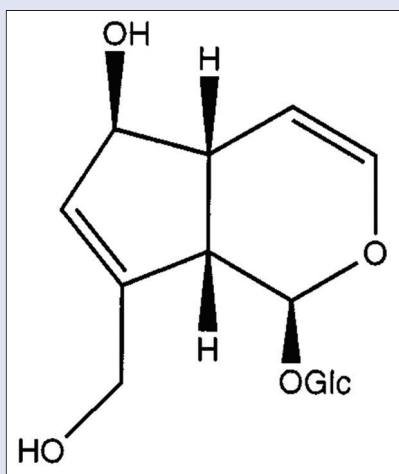


Figure 3: Chemical structure of aucubin

values between 17.63 and >100 $\mu\text{g}/\text{ml}$), whereas aucubin had the lowest effects.

Effects of *Plantago major* extracts and its chemical compounds on pro-inflammatory cytokines production in lipopolysaccharide-activated THP-1 macrophages

Cytotoxicity against THP-1 macrophage cells

In this study, methanol extracts of leaves, petioles, roots, and seeds of *P. major* were used for cytokines production assay. Before cytokines assay, potential cytotoxicity of the extracts and pure constituents on THP-1 macrophages was evaluated. Leaves, petioles, and seeds extracts reduced significantly cell viability at concentrations ≥ 300 , 1000, and 100 $\mu\text{g}/\text{ml}$, respectively. However, root extracts did not show any cytotoxic effect under the concentrations applied. Both UA and OA reduced significantly cells viability at concentrations ≥ 30 $\mu\text{g}/\text{ml}$, whereas cytotoxicity due to aucubin was observed at 1000 $\mu\text{g}/\text{ml}$ [Figure 4]. Considering this cytotoxic effect, the cytokines production assay was conducted at the concentrations having noncytotoxic effects.

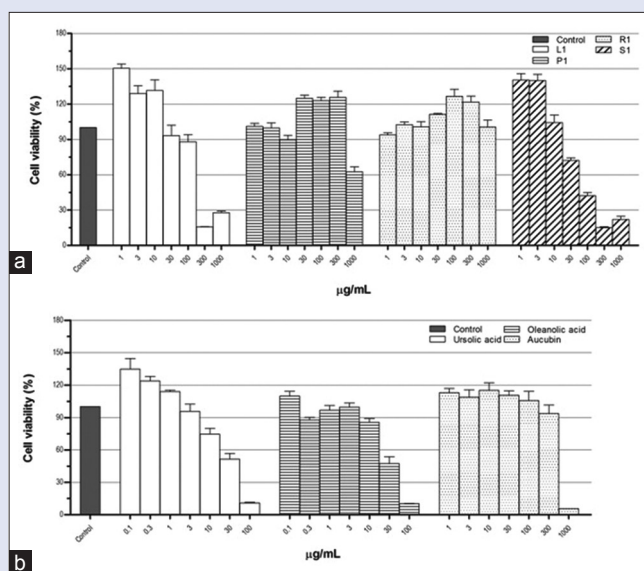


Figure 4: Cytotoxicity of *Plantago major* methanol extracts (a) and its chemical compounds (b) on THP-1 macrophages. Data represent the means \pm standard error mean of three replications

Table 1: Antiproliferative effect of *Plantago major* extracts and its chemical compounds

Sample	IC_{50} ($\mu\text{g}/\text{ml}$)				
	KB	MCF-7	MDA-MB-231	HeLaS3	A549
MeOH extract of L1	413.03 \pm 34.03	546.55 \pm 17.28	435.91 \pm 82.97	475.73 \pm 56.97	343.97 \pm 28.34
MeOH extract of P1	399.55 \pm 98.03	528.79 \pm 25.67	532.29 \pm 63.81	383.99 \pm 29.69	475.49 \pm 83.63
MeOH extract of R1	496.44 \pm 43.07	544.79 \pm 14.12	628.42 \pm 104.46	332.06 \pm 7.40	392.14 \pm 140.97
MeOH extract of S1	247.41 \pm 15.96	153.59 \pm 3.99	168.00 \pm 2.46	215.61 \pm 15.17	153.38 \pm 2.56
H ₂ O extract of L1	>1000	467.53 \pm 42.49	>1000	>1000	>1000
H ₂ O extract of P1	>1000	395.45 \pm 50.21	>1000	>1000	>1000
H ₂ O extract of R1	>1000	305.59 \pm 4.06	>1000	>1000	>1000
H ₂ O extract of S1	672.06 \pm 4.82	131.99 \pm 70.44	577.33 \pm 46.43	>1000	290.79 \pm 10.37
Ursolic acid	18.33 \pm 0.82	6.27 \pm 0.18	6.44 \pm 0.23	16.97 \pm 0.77	6.37 \pm 0.53
Oleanolic acid	17.63 \pm 1.43	40.14 \pm 5.77	>100	51.09 \pm 8.94	29.37 \pm 3.94
Aucubin	788.64 \pm 53.64	602.66 \pm 47.40	966.98 \pm 17.84	>100	>100
Doxorubicin	0.33 \pm 0.10	0.18 \pm 0	0.17 \pm 0.01	0.04 \pm 0.00	0.09 \pm 0.02

Results are means \pm SD of IC_{50} (inhibitory concentration 50, $n=3$). L1: Leaves; P1: Petioles; R1: Roots; S1: Seeds; SD: Standard deviation

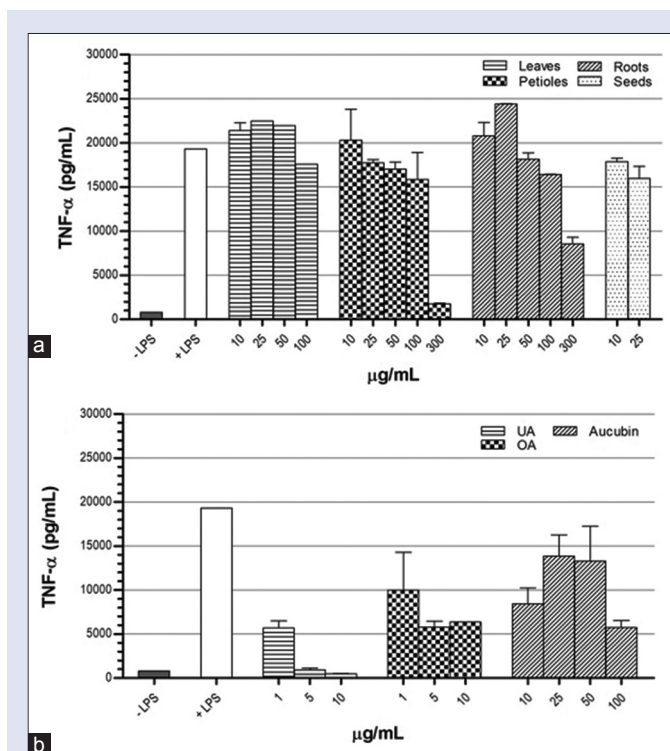


Figure 5: Effect of *Plantago major* methanol extracts (a) and its chemical compounds (b) on the production of tumor necrosis factor- α in THP-1 macrophages cells. Data represent the means \pm standard error mean of two replications

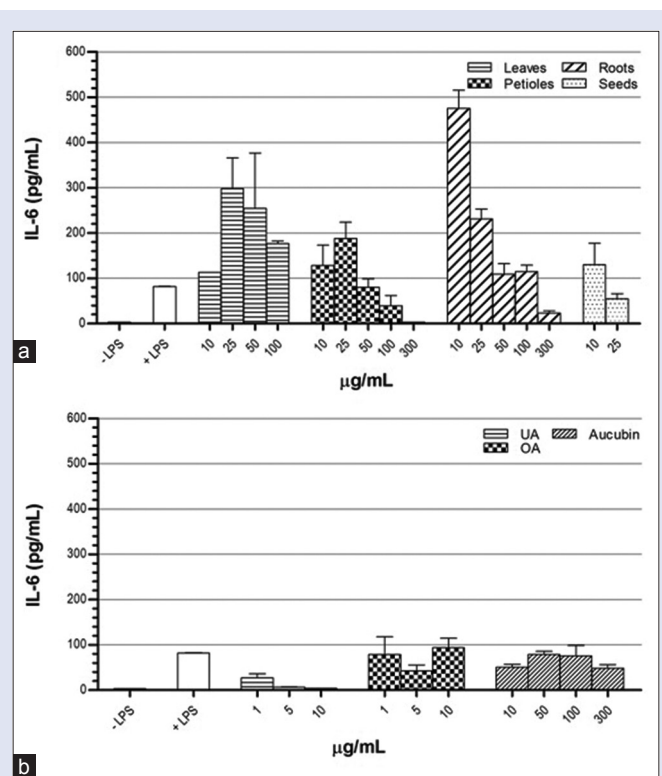


Figure 6: Effect of *Plantago major* methanol extracts (a) and its chemical compounds (b) on the production of interleukin-6 in THP-1 macrophages cells. Data represent the means \pm standard error mean of two replications

Pro-inflammatory cytokines production in lipopolysaccharide-activated THP-1 macrophages

THP-1 macrophage cells were incubated with various concentrations of either extracts or pure compounds for 48 h in the presence of LPS. In these exposed cells, secretion of cytokines was analyzed and compared to LPS-stimulated cells. Figure 5 shows that treatment of leaves, petioles, and roots extracts at low concentrations increased the level of TNF- α ; however, decreased it at high concentrations. Seeds extract slightly suppressed TNF- α production at both concentrations. UA, OA, and aucubin suppressed TNF- α production at all concentrations used, in which the effect of UA was in a dose-dependent manner.

Regarding IL-6 secretion [Figure 6], treatment of activated cells with any of *P. major* extracts increased this cytokine at low concentrations but inhibited its production at high concentrations. IL-6 secretion was reduced significantly by UA at all concentrations, and treatment with high concentration reached the basal production of nonactivated cells. In addition, the inhibition effect of UA was in a dose-dependent manner. OA and aucubin slightly decreased IL-6 secretion.

IL-1 β secretion [Figure 7] was stimulated when activated cells were incubated with leaves, petioles, and roots extracts. On the contrary, seeds extract slightly inhibited the production of IL-1 β . All pure compounds tested decreased significantly IL-1 β production.

In general, all of *P. major* extracts decreased the IFN- γ production [Figure 8]. The only one sample had a contrary effect, i.e., roots extract at the concentration of 100 μ g/ml. Inhibition effect was observed as well as in UA, OA, and aucubin.

DISCUSSION

P. major is one of the herbs widely used in cancer and pain relief management.^[3] The quality of herbs may fluctuate depending on plant

parts, growing environment, drying processes, solvents and methods of extraction, as well as many other postharvest factors.^[21] In this study, we evaluated antiproliferative activity of *P. major* extracts and its chemical compounds against various cancer cell lines. We investigated as well as their effect on the production of inflammatory cytokines. *P. major* extracts were prepared from different plant parts using methanol and water as extracting solvents.

HPTLC profiles, Figure 2, exhibited that UA, OA, and aucubin are clearly found in *P. major*. Anticancer activity of UA and OA has been actively investigated in recent years.^[22-25] Moreover, these two compounds along with aucubin have been known as anti-inflammatory agents.^[26-30] In this recent work, we isolated aucubin from the aerial part of *P. major*, whereas the isolation of UA was reported in our previous study.^[5]

In terms of antiproliferative activity, methanol extracts of all parts of *P. major* demonstrated cytotoxicities on five cancer cell lines in various potencies [Table 1], and the strongest activity was shown by the seeds extract (IC₅₀ value was between 153.38 and 247.41 μ g/ml). Water extract of *P. major* seed was active against KB, MCF-7, MDA-MB-231, and A549 cell lines, whereas the aqueous extracts of leaves, petioles, and root were active on MCF-7 only. These results were in concordance with the observations of Chiang *et al.* whose findings showed that aqueous extract of the whole plant of *P. major* showed cytotoxicities against various carcinoma cells with IC₅₀ values ranged from 283 to 1809 μ g/ml.^[6] We found also that *P. major* roots had comparable cytotoxic effect with leaves and petioles. This finding supported our previous report^[5] that roots could be included in the preparation of *P. major* derived herbal medicines. Further, toxicity and bioactivity tests are required to ensure its safety and efficacy.

Among the three pure chemicals utilized, UA showed the strongest cytotoxicity against five cancer cells tested, followed by OA. This finding confirmed our previous work which proposed UA as one of the active markers for quality assessment of *P. major*.^[5] Contrastingly,

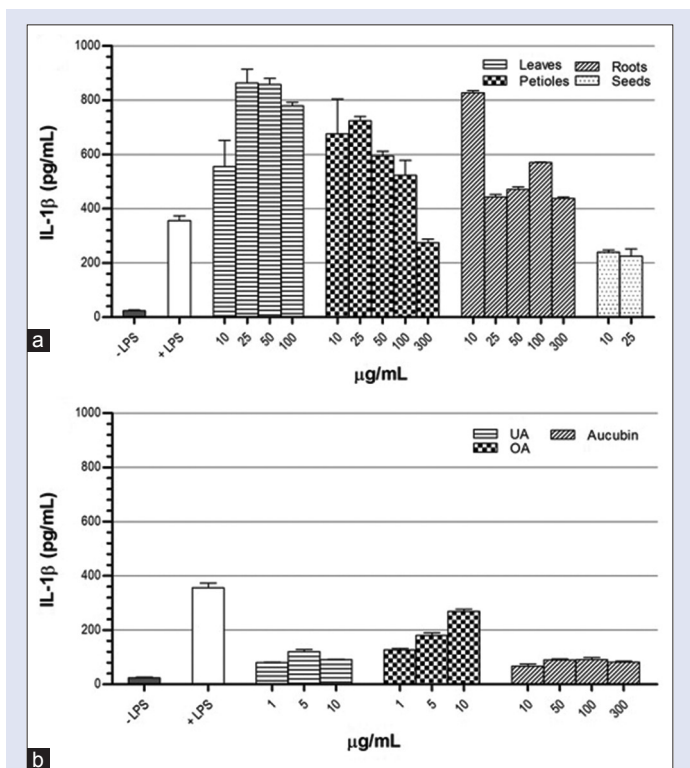


Figure 7: Effect of *Plantago major* methanol extracts (a) and its chemical compounds (b) on the production of interleukin-1β in THP-1 macrophage cells. Data represent the means ± standard error mean of two replications

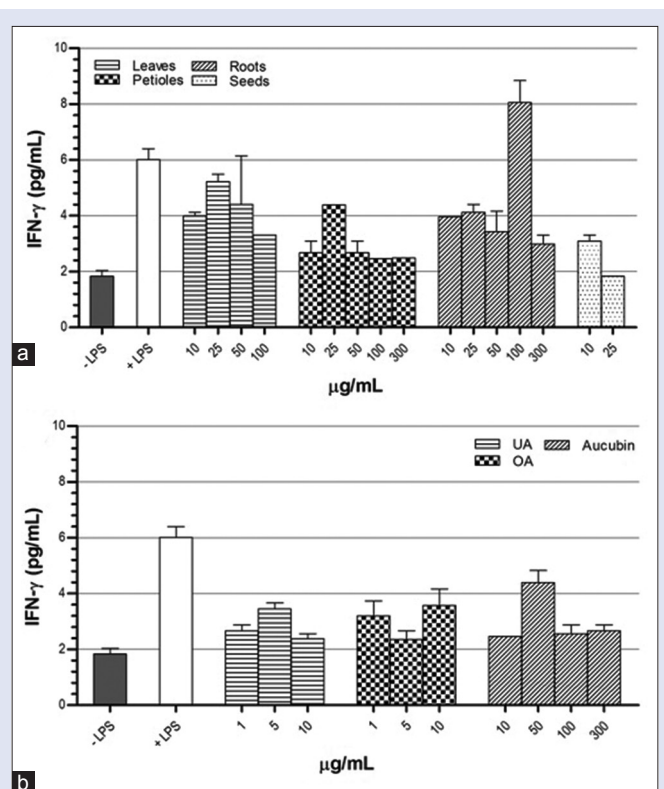


Figure 8: Effect of *Plantago major* methanol extracts (a) and its chemical compounds (b) on the production of interferon-γ in THP-1 macrophage cells. Data represent the means ± standard error mean of two replications

aucubin did not express cytotoxic effect ($IC_{50} > 100 \mu\text{g/ml}$). This finding was in agreement to Isiguro *et al.* whose investigation showed that aucubin did not have a cytotoxic effect toward leukemia P388, but its aglycone (i.e., aucubigenin) did.^[31]

Cytokines are soluble messengers having a fundamental role for cell–cell communication and regulation within the immune system. Macrophages release cytokines to activate and recruit other cells during inflammation or as direct killing agents.^[32] However, excessive production of this mediator in an inflammatory site may lead to chronic diseases such as cancer.^[26] The effects of *P. major* extracts on the production of pro-inflammatory cytokines (TNF- α , IL-1 β , IL-6, and IFN- γ) were determined.

Studies have shown that *P. major* extracts were able to alter the production of inflammatory cytokines. Levels and patterns of inflammatory cytokines production were found to be different for different parts of *P. major*, which may be due to the content of chemical compounds. Leaves, petioles, and roots extracts showed a similar effect on TNF- α and IFN- γ secretions [Figures 5 and 8]. These samples enhanced the production of TNF- α at low concentrations but decreased it at high concentrations and decreased the level of IFN- γ at any concentrations. Leaves extract increased IL-6 at all concentrations, whereas petioles and roots increased this cytokine at low concentrations and decreased it at high concentrations [Figure 6]. Leaves and roots extracts induced production of IL-1 β at all tested concentrations, whereas petioles extract increased its production at low concentrations and decreased it at high concentrations [Figure 7]. Our results seemed to be in concordance with the observations of Chiang *et al.* whose results revealed that *P. major* extracts enhanced the immune components at low concentrations, whereas inhibited these effect at high concentrations. Our findings support the concept of dual immunomodulatory activity owned by *P. major*.^[6]

Seeds extract exhibited different patterns. It inhibited secretion of TNF- α , IL-1 β , and IFN- γ in the range of concentrations used; however, it slightly increased IL-6 at low concentrations, whereas decreased it at high concentrations. If we compare among *P. major* parts at equivalent concentrations, seeds caused the lowest production of TNF- α , IL-1 β , IL-6, and IFN- γ . This effectiveness might be caused by its high content in UA^[5] which showed the greatest inhibition activity among the chemical compounds tested.

Various trends in the modulation of cytokines levels were exhibited by the pure compounds. UA was capable of inhibiting the production of TNF- α , IL-6, IL-1 β , and IFN- γ . UA is a unique triterpenoid, having contrary anti- and pro-inflammatory activities that are dependent on the biological status of cells and tissues.^[26] Our recent results, UA inhibited pro-inflammatory cytokines production in LPS-induced THP-1 macrophages, were in concordance with the previous study.^[26]

We observed that OA and aucubin inhibited TNF- α , IL-1 β , and IFN- γ ; however, they did not exhibit any effect on IL-6. These findings were in agreement with the previous work which showed that a series of OA analogs had weak inhibition toward IL-1 β production in LPS-induced RAW 264.7 and J774A.1 cells.^[33]

From this study, we observed that roots extracts possessed comparable antiproliferative and cytokines production activities to the other parts (leaves and petioles). This indicated that the roots could be included in the preparation of *P. major* derived herbal medicines. Seeds extract showed the highest antiproliferative effect as well as inhibition of inflammatory cytokines production. Among the pure constituents investigated, UA exhibited the strongest activities. Our previous study showed that UA was found abundantly in seeds, whereas very low in roots. Figure 2b showed that roots contain a high amount of aucubin. Therefore, in addition to UA,^[5] we proposed aucubin as another marker for standardization of *P. major*.

CONCLUSIONS

The results of this study support the existing data and the empirical uses of *P. major*. Five cancer cell lines were used in antiproliferative assay to strengthen the usage of *P. major* in cancer management. Anti-inflammatory assay of *P. major* through modulation of cytokines production was carried out to afford its comprehensive mechanism. We have demonstrated that methanol extract of seeds possessed the greatest activity both on antiproliferative and cytokines production assay. Roots extract exhibited comparable activities to leaves and petioles. The seeds are being proposed as the main source for further development of anticancer and anti-inflammatory products. The roots could be included in the preparation of *P. major* derived products with respect to anti-inflammatory. It could also be considered as the source of aucubin.

Acknowledgement

This research is part of Ph.D. thesis of Mahidol University, Bangkok, Thailand, which was funded by the Directorate of Higher Education of Indonesia, contract number 373.11/E4.4/K/2012. The authors acknowledged the Natural Products Research Section, Research Division, National Cancer Institute, Bangkok, Thailand, for providing the facilities and equipment for doing throughout the research work.

Financial support and sponsorship

This study was financially supported by Directorate of Higher Education of Indonesia, contract number 373.11/E4.4/K/2012.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Beara IN, Lesjak MM, Jovin ED, Balog KJ, Anackov GT, Orcic DZ, *et al.* Plantain (*Plantago* L.) species as novel sources of flavonoid antioxidants. *J Agric Food Chem* 2009;57:9268-73.
2. Sagar GR, Harper JL. *Plantago major* L., *P. Media* L. and *P. Lanceolata* L. *J Ecol* 1964;52:189-221.
3. Samuelsen AB. The traditional uses, chemical constituents and biological activities of *Plantago major* L. A review. *J Ethnopharmacol* 2000;71:1-21.
4. Warwick SI, Briggs D. The genecology of lawn weeds. V. The adaptive significance of different growth habit in lawn and roadside populations of *Plantago major* L. *New Phytol* 1980;85:289-300.
5. Kartini, Piyaviriyakul S, Siripong P, Vallisuta O. HPTLC simultaneous quantification of triterpene acids for quality control of *Plantago major* L. and evaluation of their cytotoxic and antioxidant activities. *Ind Crops Prod* 2014;60:239-46.
6. Chiang LC, Chiang W, Chang MY, Lin CC. *In vitro* cytotoxic, antiviral and immunomodulatory effects of *Plantago major* and *Plantago asiatica*. *Am J Chin Med* 2003;31:225-34.
7. Gálvez M, Martín-Cordero C, López-Lázaro M, Cortés F, Ayuso MJ. Cytotoxic effect of *Plantago* spp. on cancer cell lines. *J Ethnopharmacol* 2003;88:125-30.
8. Lithander A. Intracellular fluid of waybread (*Plantago major*) as a prophylactic for mammary cancer in mice. *Tumour Biol* 1992;13:138-41.
9. Ruffa MJ, Ferraro G, Wagner ML, Calcagno ML, Campos RH, Cavallaro L. Cytotoxic effect of Argentine medicinal plant extracts on human hepatocellular carcinoma cell line. *J Ethnopharmacol* 2002;79:335-9.
10. Kolak U, Boğa M, Uruşak EA, Ulubelen A. Constituents of *Plantago major* subsp. *intermedia* with antioxidant and anticholinesterase capacities. *Türk J Chem* 2011;35:637-45.
11. Stanisavljević IT, Stojičević SS, Veličković DT, Lazić ML, Veljković VB. Screening the antioxidant and antimicrobial properties of the extracts from plantain (*Plantago major* L.) leaves. *Sep Sci Technol* 2008;43:3652-62.
12. Gomez-Flores R, Calderon CL, Scheibel LW, Tamez-Guerra P, Rodriguez-Padilla C, Tamez-Guerra R, *et al.* Immunoenhancing properties of *Plantago major* leaf extract. *Phytother Res* 2000;14:617-22.
13. Núñez Guillén ME, da Silva Emim JA, Souccar C, Lapa AJ. Analgesic and anti-inflammatory activities of the aqueous extract of *Plantago major* L. *Pharm Biol* 1997;35:99-104.
14. Türel I, Ozbek H, Erten R, Oner AC, Cengiz N, Yılmaz O. Hepatoprotective and anti-inflammatory activities of *Plantago major* L. *Indian J Pharmacol* 2009;41:120-4.
15. Dudhgaonkar S, Thyagarajan A, Sliva D. Suppression of the inflammatory response by triterpenes isolated from the mushroom *Ganoderma lucidum*. *Int Immunopharmacol* 2009;9:1272-80.
16. Beara IN, Orcic DZ, Lesjak MM, Mimica-Dukic NM, Pekovic BA, Popovic MR. Liquid chromatography/tandem mass spectrometry study of anti-inflammatory activity of plantain (*Plantago* L.) species. *J Pharm Biomed Anal* 2010;52:701-6.
17. Ringbom T, Segura L, Noreen Y, Perera P, Bohlin L. Ursolic acid from *Plantago major*, a selective inhibitor of cyclooxygenase-2 catalyzed prostaglandin biosynthesis. *J Nat Prod* 1998;61:1212-5.
18. Stenholm A, Göransson U, Bohlin L. Bioassay-guided supercritical fluid extraction of cyclooxygenase-2 inhibiting substances in *Plantago major* L. *Phytochem Anal* 2013;24:176-83.
19. Buckberry LD. Cytotoxicity testing using cell lines. In: Jenkins N, editor. *Animal cell biotechnology: Methods and protocols*. Totowa, NJ: Humana Press; 1999. p. 239-52.
20. Akdemir ZŞ, Tatlı İİ, Bedir E, Khan IA. Acylated iridoid glycosides from *Verbascum lasianthum*. *Türk J Chem* 2009;28:101-10.
21. Gad HA, El-Ahmady SH, Abou-Shoer MI, Al-Azizi MM. Application of chemometrics in authentication of herbal medicines: A review. *Phytochem Anal* 2013;24:1-24.
22. Sun H, Fang WS, Wang WZ, Hu C. Structure-activity relationships of oleanane-and ursane-type triterpenoids. *Bot Stud* 2006;47:339-68.
23. Liu J. Pharmacology of oleanolic acid and ursolic acid. *J Ethnopharmacol* 1995;49:57-68.
24. Liu J. Oleanolic acid and ursolic acid: research perspectives. *J Ethnopharmacol* 2005;100:92-4.
25. Shanmugam MK, Dai X, Kumar AP, Tan BK, Sethi G, Bishayee A. Ursolic acid in cancer prevention and treatment: Molecular targets, pharmacokinetics and clinical studies. *Biochem Pharmacol* 2013;85:1579-87.
26. Ikeda Y, Murakami A, Ohigashi H. Ursolic acid: An anti- and pro-inflammatory triterpenoid. *Mol Nutr Food Res* 2008;52:26-42.
27. Jeong HJ, Koo HN, Na HJ, Kim MS, Hong SH, Eom JW, *et al.* Inhibition of TNF-alpha and IL-6 production by aucubin through blockade of NF-kappaB activation RBL2H3 mast cells. *Cytokine* 2002;18:252-9.
28. Park KS. Aucubin, a naturally occurring iridoid glycoside inhibits TNF-alpha-induced inflammatory responses through suppression of NF-kB activation in 3T3-L1 adipocytes. *Cytokine* 2013;62:407-12.
29. Park KS, Chang IM. Anti-inflammatory activity of aucubin by inhibition of tumor necrosis factor-alpha production in RAW 264.7 cells. *Planta Med* 2004;70:778-9.
30. Park KS, Kim BH, Chang IM. Inhibitory potencies of several iridoids on cyclooxygenase-1, cyclooxygenase-2 enzymes activities, tumor necrosis factor-alpha and nitric oxide production *in vitro*. *Evid Based Complement Alternat Med* 2010;7:41-5.
31. Isiguro K, Yamaki M, Takagi S, Ikeda Y, Kawakami K, Ito K, *et al.* Studies on iridoid-related compounds. IV. Antitumor activity of iridoid aglycones. *Chem Pharm Bull (Tokyo)* 1986;34:2375-9.
32. Stow JL, Low PC, Offenhäuser C, Sangermani D. Cytokine secretion in macrophages and other cells: Pathways and mediators. *Immunobiology* 2009;214:601-12.
33. Bhandari P, Patel NK, Gangwal RP, Sangamwar AT, Bhutani KK. Oleanolic acid analogs as NO, TNF-alpha and IL1b inhibitors: Synthesis, biological evaluation and docking studies. *Bioorg Med Chem Lett* 2014;24:4114-9.