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## **Therapeutic Strategies Targeting Inherited Cardiomyopathies**

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## Abstract

**Purpose of review**—Cardiomyopathies due to genetic mutations are a heterogeneous group of disorders that comprise diseases of contractility, myocardial relaxation, and arrhythmias. Our goal here is to discuss a limited list of genetically inherited cardiomyopathies and the specific therapeutic strategies used to treat them.

**Recent findings**—Research into the molecular pathophysiology of the development of these cardiomyopathies is leading to the development of novel treatment approaches. Therapies targeting these specific mutations with gene therapy vectors are on the horizon, while other therapies which indirectly affect the physiologic derangements of the mutations are currently being studied and used clinically. Many of these therapies are older medications being given new roles such as mexiletine for Brugada syndrome and diflunisal for transthyretin amyloid cardiomyopathy. Newer targeted therapies such as the inhibitor of myosin ATPase MYK-461 has been shown to suppress the development of ventricular hypertrophy, fibrosis, and myocyte disarray and is being studied as a potential therapy in patients with hypertrophic cardiomyopathy.

**Summary**—While this field is too large to be completely contained in a single review, we present a large cross section of recent developments in the field of therapeutics for inherited cardiomyopathies. New therapies are on the horizon and their development will likely result in improved outcomes for patients inflicted by these conditions.

## Keywords

hypertrophic cardiomyopathy; arrhythmogenic cardiomyopathy; amyloidosis; Lamin mutations; hemochromatosis; Fabry's disease; SCN5A mutation; catecholaminergic polymorphic ventricular tachycardia

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**Conflicts of Interest** 

Kenneth Varian declares no conflict of interest.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

## Introduction

As we begin to understand the genetic basis for heart disease, more specific therapies are being developed. In the case of genetically inherited cardiomyopathies, therapies that target specific aspects of these diseases are being actively researched and developed. Specific targeted therapies are those that counteract the specific molecular derangement caused by the genetic mutation. The goal of this review is to look at a variety of genetically inherited syndromes that have cardiac involvement and treatments used in their specific instances. There are too many types and subtypes of inherited cardiomyopathies with specific treatments to discuss in one review. Many reviews are dedicated to the pathophysiology and specific treatments for one specific syndrome. Thus, this will be merely a sample of current therapy and therapies on the horizon.

## SCN5A mutations

The SCN5A gene encodes for a voltage gated sodium channel involved in myocardial electrical conduction and the rapid upstroke of the action potential. It is the predominant isoform in the heart. Mutations in the gene can play a causative role in malignant arrhythmia syndromes such as type III long QT syndrome (LQT3), Brugada syndrome<sup>1</sup> and even dilated cardiomyopathy.<sup>2</sup> A specific subgroup of idiopathic ventricular fibrillation later named Brugada syndrome was first described in 1992.<sup>3</sup> Several years later various types of mutations in the SCN5A gene were found to be present in families with a history of Brugada syndrome.<sup>4</sup> Around that same time a linkage was discovered between genetic polymorphisms within the SCN5A gene and LQT3.<sup>5</sup>

The mechanisms by which these mutations cause these syndromes were not elucidated until the early 2000s. In 2002, Baroudi et al reported that SCN5A mutations R123W and T1620M, responsible for the Brugada syndrome, resulted in the absence of channel function due to disruption in channel trafficking toward the plasma membrane<sup>6</sup>. That same year the SCN5A mutation M1766L was discovered in an infant with prolonged OT syndrome type III and torsades de pointes. This mutation caused a decrease in sodium current measured in HEK cells that improved with mexiletine.<sup>7</sup> In 2004 Valdivia et al reported that mexiletine rescued the membrane expression of the voltage gated sodium channel in HEK cells expressing the Brugada syndrome mutation G1743R by somehow acting as a molecular chaperone helping to transport the protein from the sarcoplasmic reticulum to the plasma membrane<sup>8</sup>. This rescue mechanism of mexiletine was supported by Tan et al who reported that the Brugada syndrome associated mutation, G1406R, also displayed a trafficking defect rescued by mexiletine.<sup>9</sup> Pfahnl et al showed that the Brugada syndrome causing the T353I mutation produced a failure to transport the protein to the sarcolemma resulting in decreased expression. However, while mexiletine rescued the trafficking defect and restored the membrane expression to near normal levels, the electrical current pattern included a significant late current consistent with a long QT syndrome phenotype.<sup>10</sup> This effect was also seen by Ruan et al who reported in 2010 that the SCN5A mutation F1473S (associated with long-QT syndrome type III in a child) resulted in reduced expression of the voltage gated channel. While expression was improved with mexiletine, treatment of the cells

resulted in prolongation of the action potential duration mimicking the clinically observed effect of worsening QT prolongation after mexiletine treatment in the patient.<sup>11</sup>

Most recently Mazzanti et al showed that mexiletine could shorten the QTc interval in patients with LQT3 mutations. In a non-randomized prospective study, 36 patients with LQT3 were given mexiletine. They found a reduction in the annual rate of arrhythmic events from 10.3% to 0.7%.<sup>12</sup> The use of sodium channel blockers in LQT3 syndrome patients is also supported by the most recent ESC clinical guidelines.<sup>13</sup>

#### Catecholaminergic polymorphic ventricular tachycardia

Catecholaminergic polymorphic ventricular tachycardia (CPVT) is a condition characterized by episodic polymorphic ventricular tachycardia brought on by any state that causes a heighted catecholaminergic state<sup>14</sup>. CPVT has been found to be associated with mutations in the ryanodine receptor<sup>15,16</sup> (RyR2), calsequestrin<sup>17</sup> (CASQ2), triadin<sup>18</sup> (TRDN), and calmodulin<sup>19</sup> (CALM1) and is considered generally to be a disorder of calcium handling in the individual myocytes with mutations. Specifically, these mutations cause an increase in delayed after-depolarization from spontaneous calcium release from the sarcoplasmic reticulum<sup>20</sup>. The gene KCNJ2 coding for an inwardly rectifying potassium channel may be associated with CPVT, though this remains controversial.<sup>21,22</sup> Of these mutations, the autosomal dominant inherited mutations in RyR2 are the most common (around 60%),<sup>22,23</sup> and may even explain a significant number of unexplained cases of sudden cardiac death in patients not previously diagnosed with CPVT.<sup>24</sup> Loss of function mutations in CASQ2 are autosomal recessive and account for far fewer cases. These mutations are thought to cause arrhythmia through a similar mechanism as the RyR2 mutations.<sup>25</sup>

While most patients who have recurrent ventricular tachycardia assessed to be CPVT get implantable cardioverter defibrillators (ICD), more specific therapies are being employed to prevent recurrence and minimize ICD shocks. Given its association with a heightened catecholaminergic state, the use of beta blockers is the mainstay of therapy, and has been shown to be associated with a lower rate of arrhythmia.<sup>26</sup> Nadolol is frequently chosen because it is long acting and has no intrinsic sympathomimetic activity. The use of nadolol over  $\beta$ 1 selective beta blockers such as metoprolol succinate has been shown to provide great efficacy at reducing the rate of exercise induced arrhythmias.<sup>27</sup>

In patients who have arrhythmias refractory to beta blocker and activity avoidance, the use of flecainide has been proposed as a viable therapeutic agent. While flecainide is best known as a class 1C sodium channel blocker, it also can inhibit cardiac RyR2 calcium release acting to help prevent the calcium flux associated polymorphic ventricular tachycardia.<sup>28,29</sup> The addition of flecainide to the regimen of 33 patients with CPVT suppressed 76% of exercise induced ventricular arrhythmias.<sup>30</sup> Even patients determined to have CPVT without an identifiable mutation culprit seem to benefit from flecainide by reducing exercise induced arrhythmias.<sup>31</sup> Specific cases of patients who have been refractory to calcium channel blocker and beta blocker therapy and with defibrillator induced storming of ventricular tachycardia rescued with flecainide have been reported.<sup>32,33</sup> The mechanism by which flecainide prevents arrhythmias in patients with CPVT is not full elucidated. It may be a

direct effect on the RyR2 receptor,<sup>34,35</sup> or it may act through its antagonistic activity on voltage gated sodium channels by increasing the threshold for triggered activity.<sup>20,36,37</sup> Finally, patients with calsequestrin associated CPVT (denoted CPVT2) may also benefit from flecainide by preventing exercise induced polymorphic VT.<sup>38</sup> While there is little controversy as to the beneficial effects of flecainide in patients with CPVT, the exact anti-arrhythmic mechanism has not been fully established.

In patients who have contraindications or are refractory to medical therapy, left cardiac sympathetic denervation is also an option. This was first described in 3 patients with medication refractory CPVT with excellent results.<sup>39</sup> Since then, only single center trials and low numbers of CPVT patients have been reported, some with good<sup>40–42</sup> and less certain results,<sup>43</sup> or with good results but short term follow up.<sup>44</sup>

Future therapeutics under study include JTV-519, an experimental drug that decreases intracellular calcium leak via the RyR2, which may be a viable option for patients with CPVT causing mutations in the future. This was initially shown from biochemical experiments<sup>45</sup> and later in dilated murine ventricular myocytes harboring a CPVT RyR2 mutation.<sup>46</sup> Most recently a related compound that decreases intracellular calcium leak, EL9, was shown to reduce ventricular tachyarrhythmias in mice carrying a CPVT causing RyR2 mutation.<sup>47</sup>

## Transthyretin Amyloid Cardiomyopathy

Transthyretin amyloid cardiomyopathy is a condition of amyloid involvement in the myocardium and can lead to heart failure (with preserved or reduced ejection fraction), a restrictive phenotype, and atrial arrhythmias.<sup>48,49</sup> Patients carrying certain mutations of the transthyretin (TTR) gene have a high risk of developing familial amyloid cardiomyopathy. This is secondary to decreased stability of the TTR tetramer which results in amyloid deposition. The V122I mutation is a common one found in 3–4% of African Americans.<sup>50</sup> In the Transthyretin Amyloidosis Cardiac Study, patients carrying the V122I mutation who were diagnosed with amyloid cardiomyopathy had a median survival from diagnosis of 25.6 months.<sup>51</sup> Definitive treatment for TTR cardiac amyloidosis in the past has been limited to heart transplant, liver transplant, or a combination of the two.<sup>48</sup>

Tafamidis meglumine (tafamidis; Pfizer Inc; New York, NY) is a novel compound being studied that targets the thyroxin binding site of the TTR tetramer and inhibits its dissociation<sup>52</sup> This compound has been used to treat familial amyloid polyneuropathy in Europe and can delay neurological impairment<sup>53</sup>. Recently, there have been published studies on the effects of tafamidis in wild type TTR cardiac amyloidosis. Maurer and colleagues published the effects of tafamidis on transthyretin stabilization and clinical outcomes in a phase 2 open-label trial. The trial included 31 patients NYHA class I-II with wild type TTR amyloid cardiomyopathy who received 20 mg daily of tafamidis for 12 months. At 6 weeks almost all patients achieved TTR stabilization as determined by an immunotrubidimetric assay. There were no clinically significant changes in the echocardiogram parameters at 12 months compared to the start of the study. Fifteen of 31 patients had progression of disease as determined by a rise in NT-proBNP > 1000 pg/mL, an

increase in serum creatinine of 0.5 mg/dL or a > 50 m decline in distance walked in the 6 minute walk test during the 12 months taking 20 mg of tafamidis. The study was designed to evaluate TTR stabilization which was achieved, but larger studies will need to be done to assess efficacy.<sup>54</sup> Damy and colleagues also performed a study on 21 patients with non-Val30Met and non-Val122IIe hereditary TTR cardiac amyloidosis administering tafamidis 20 mg per day for 12 months. In this small open label study, there were no clinically relevant changes in the mean echocardiographic or electrocardiographic variables. There were no safety concerns.<sup>55,56</sup> In patients with the Val30Met TTR mutation, tafamidis was found to lead to stabilization of cardiac biomarkers and echocardiographic parameters.<sup>57</sup>

Other potential targeted treatments include the diflunisal and doxycycline. Diflunisal is a non-steroidal anti-inflammatory drug which has been shown to stabilize the TTR tetramer and has been proposed as a treatment for the manifestations of TTR amyloidosis.<sup>58</sup> A preliminary study on the use of diflunisal in patients with TTR amyloidosis shows that it is well tolerated. In this 13 patient study that lasted about 1 year, there was no significant change in cardiac mass, ejection fraction, troponin I, or BNP.<sup>59</sup> Larger studies need to be performed for the statistical power to determine if diflunisal is a viable treatment modality. Doxycycline when combined with tauroursodeoxycholic acid was associated with no progression of cardiomyopathy during treatment in a small 20 patient phase II trial.<sup>60</sup>

Interfering RNA (RNAi) therapy is an emerging strategy that aims to reduce hepatic production of transthyretin by interfering with translation. In a recent phase I placebo controlled trial, a small RNAi delivered in lipid nanoparticles was studied in 32 patients with TTR amyloid and 17 healthy volunteers over the course of 28 days. Most of these patients harbored the V30M mutation, which causes polyneuropathy and less so cardiomyopathy. However, the concept would presumably be valid with any mutation or even wild type TTR amyloid cardiomyopathy. In this study, significant reduction in serum transthyretin levels was observed with only mild to moderate infusion related reactions. Clinical endpoints were not examined.<sup>61</sup>

## Hemochromatosis

Iron overload cardiomyopathy is frequently secondary to hereditary hemochromatosis. Most cases are caused by mutations in the HFE gene leading to elevated total body iron that subsequently gets deposited in the myocardium leading to myocardial damage. Clinically, patients may be initially characterized by diastolic dysfunction and restrictive hemodynamics and later (as the disease progresses) develop a dilated cardiomyopathy with reduced ejection fraction.<sup>62</sup> The mainstay of treatment is removal of excess iron from the tissues, preventing myocardial damage with early intervention being the goal. Regular phlebotomy disposes excess iron and has been shown to improve echocardiographic parameters of LV wall thickness and function.<sup>63,64</sup> However, this is not a treatment option in patients with anemia. Moreover, it is not an option for those with severe congestive heart failure.

Chelation therapy is an option for those with anemia or with more advanced cardiac disease. Three medications, deferoxamine, deferiprone, and deferasirox are all approved by the Food

and Drug Administration. Most of the data on iron chelation and improvement of myocardial iron overload is in transfusion dependent Thalassemia patients. Deferoxamine has been shown to reduce the risk of developing cardiac disease<sup>65</sup> and improve systolic function in patients with iron overload, secondary to Chronic transfusion.<sup>66,67</sup> Deferiprone and deferasirox are alternative chelating agents whose efficacy and long term safety data are not as well established as deferoxamine.<sup>68</sup>

## Anderson Fabry's cardiomyopathy

Anderson Fabry's disease results from a genetic deficiency in the enzyme alpha galactosidase A which results in globotriaosylceramide accumulation. It is the second most prevalent lysosomal storage disease second only to Gaucher's disease. It is an X linked condition that results in progressive problems in many systems including renal insufficiency, stroke, and it can result in hypertrophic cardiomyopathy<sup>69</sup>. In a review of 279 male and 168 female Fabry's patients, cardiac arrhythmia was seen in 42% of male patients and 27% of female patients, angina 13% and 14% of male and female patients, respectively, and heart failure was seen in 4% and 1% of the male and female patients, respectively<sup>70</sup>.

Prior to the development of enzyme replacement therapy, no specific treatments were available. The two current options include agalsidase alpha (Replagal) and agalsidase beta (Fabrazyme). Early studies showed that enzyme replacement therapy seemed to be safe, but also decreased the deposits of globotriaosylceramide in the liver and renal tubular cells<sup>71</sup> as well as deposits in the skin and myocardial tissue<sup>72,73</sup>. Soon after the initial safety study, it was found that echocardiographic parameters of left ventricular size and function (quantified by strain pattern) improved in patients with Fabry's disease treated with enzyme replacement therapy for 12 months<sup>74</sup>. In 2007, Banikazemi published a study on 82 Fabry's disease patients who received agalsidase beta or placebo over 35 months. The composite clinical outcome of renal, cardiac, and cerebrovascular complications and death were mitigated, though there was no significant difference in cardiac events between the groups<sup>75</sup>. Treatment of 181 adult Fabry patients with agalsidase alpha showed that, in patients with baseline cardiac hypertrophy, treatment resulted in a reduction in the left ventricular mass index after 5 years. Patients without baseline hypertrophy remained stable<sup>76</sup>. Furthermore, when patients without advanced cardiomyopathy were treated with enzyme replacement, reduced left ventricular mass, improved myocardial function and exercise capacity were observed after 3 years of treatment<sup>77</sup>. In a clinical trial of 58 patients who received 1 mg/kg of agalsidase beta every two weeks over a 10 year period, the left ventricular posterior wall thickness and interventricular septum thickness did not show a significant increase over the treatment period and remained normal<sup>78</sup>. Despite enzyme replacement therapy, cardiac complications are now the primary cause of death in both male and female Fabry's patients (surpassing renal disease) regardless of whether they receive enzyme replacement therapy or not<sup>79</sup>.

## Hypertrophic Cardiomyopathy

Mutations in sarcomeric proteins account for about 65% of all cases of hypertrophic cardiomyopathy<sup>80</sup>. Over 400 familial hypertrophic cardiomyopathy mutations have been

identified.<sup>81</sup> The mutations can occur in many sarcomeric proteins on both the thin and thick filament, and most of them occur in myosin and myosin binding protein C.<sup>81,82</sup> The discussion below pertains to targeted treatments for familial hypertrophic cardiomyopathy.

Losartan is an angiotensin II receptor blocker (ARB) used primarily to treat hypertension. In 2007, Yamazki et al performed a small study in 19 patients with hypertrophic nonobstructive cardiomyopathy where patients were treated with 50 mg of losartan or placebo and left ventricular mass was assessed by MRI. The study showed that left ventricular mass reduced by a ratio of 0.93 compared to 1.02 for placebo<sup>83</sup>. A similar study of 20 patients showed that 50 mg of losartan produced a non-significant (p=0.06) change in LV mass in patients with non-obstructive hypertrophic cardiomyopathy compared to placebo<sup>84</sup>. The ARB candesartan was also examined. In a randomized double blinded study of 24 patients with non-obstructive hypertrophic cardiomyopathy, after one year and using 32 mg as a target dose, there was a significant regression of left ventricular hypertrophy, improvement in left ventricular function (assessed by echocardiography), and improvement of exercise tolerance. The patients who had the largest regression of hypertrophy were those who carried beta myosin heavy chain mutations<sup>85</sup>. However, the findings of these smaller studies were not corroborated in a larger trial. The INHERIT trial studied losartan 100 mg in a randomized, double blinded, placebo controlled fashion in 133 patients with obstructive and non-obstructive hypertrophic cardiomyopathy and found no change in left ventricular mass assessed by cardiac MRI or CT86. The VANISH study (Valsartan for attenuating disease evolution in early sarcomeric HCM) is currently recruiting participates (NCT 01912534).<sup>87</sup> More studies will need to be done in the future, potentially using ARBs other than losartan to help answer the question of their role in hypertrophic cardiomyopathy.

Aldosterone has also been implicated in the pathogenesis of inherited hypertrophic cardiomyopathy. In mice carrying the hypertrophic cardiomyopathy associated troponin T Q92 mutation, antagonizing aldosterone with spironolactone reversed myocardial fibrosis, attenuated myocyte disarray by 50%, and improved diastolic function.<sup>88</sup>

Augmenting sarcoplasmic reticulum (SR) calcium uptake by augmenting gene expression of the SERCA2a calcium pump has also been speculated to be a mechanism by which the hypertrophic cardiomyopathy phenotype can be altered. Using a transgenic mouse model of familial hypertrophic cardiomyopathy with a mutation in tropomyosin, Pena et al showed that over-expression of SERCA2A improved overall whole heart morphology and augmented contractility and response to isoproterenol.<sup>89</sup> SR calcium reuptake can also be augmented by it phosphorylating or removing its inhibitor phospholamban. The augmentation of SR calcium uptake hypothesis was corroborated when the tropomyosin mutant mice were crossed with the phospholamban knockout mouse. These mice displayed improved ventricular function and less collagen deposition compared to the mice with the tropomyosin mutation only.<sup>90</sup>

While intracellular calcium handling has been a genetic target for hypertrophic cardiomyopathy, blocking the L-type calcium channel with diltiazem has also been studied. In 2002, Semsarian et al showed that in transgenic mice bearing the Arg403Gln missense mutation in the alpha cardiac myosin heavy chain, SR calcium, SR calsequestrin, and SR

ryanodine receptor levels were reduced prior to the development of pathologic changes seen in myocardial structure. Furthermore, early use of diltiazem restored the expression of calsequestrin and the ryanodine receptor and prevented the development of the myocardial hypertrophy and myocyte disarray.<sup>91</sup> Using a transgenic mouse with the hypertrophic cardiomyopathy associated troponin T I79N mutation. Westermann et al showed that pretreatment with diltiazem prevented the development of severe diastolic dysfunction in these mice.<sup>92</sup> Diltiazem was also shown to improve peak early diastolic velocity as assessed by echocardiography in a small group of patients who were carriers for a mutation in cardiac myosin binding protein C mutation, but did not show evidence of hypertrophy yet. It was speculated that diltiazem might help prevent development of later structural changes in hypertrophic cardiomyopathy.<sup>93</sup> Most recently, a randomized double blinded study of 38 patients who carried sarcomere mutations was performed where patients received either diltiazem or placebo for a median of 25 months. The results showed that left ventricular enddiastolic diameter and left ventricular wall thickness improved in the diltiazem group, but worsened in the placebo group.<sup>94</sup> Further studies will need to be completed to determine the long term efficacy in treating this patient population.

Increased myofilament calcium sensitivity is thought to be one of the mechanisms of hypertrophic cardiomyopathy<sup>81</sup>. Therefore treatments aimed at decreasing myofilament calcium sensitivity have been postulated as potential strategies. Small molecule inhibitors of cross bridge formation. One example is blebbistatin which specifically inhibits actin-myosin interaction.<sup>95</sup> The only studies looking at blebbistatin are in animal models. For example, Baudenbacher et al showed that blebbistatin could reduce arrhythmia susceptibility in a mouse model of hypertrophy with a troponin T mutation.<sup>96</sup> Coutu et al overexpressed parvalbumin (a calcium buffering compound) in mouse models of hypertrophic cardiomyopathy with a mutation in tropomyosin and found that the enhanced intracellular calcium buffering leads to improved diastolic function.<sup>97</sup> Another example, the company MyoKardia, is currently developing therapies for inherited hypertrophic cardiomyopathy. Their compound MYK-461 acts as an inhibitor of myosin ATPase by decreasing the steady state rate of the ATPase activity.<sup>98</sup> The molecule is being studied as a part of a phase I clinical trial (NCT02329184). Using a transgenic mouse model with a mutation in myosin heavy chain that develop hypertrophic cardiomyopathy, Green et al showed that MYK-461 suppressed the developed ventricular hypertrophy, fibrosis, and myocyte disarray, further supporting the notion that inhibiting hyperdynamic contraction may be of therapeutic benefit in patients with hypertrophic cardiomyopathy.99

Intracellular sodium fluxes have also been a focus of treatment for hypertrophic cardiomyopathy. Using human myocytes obtained from myectomy operations, Coppini et al showed that these myocytes exhibit a prolonged action potential related to increased late sodium current that led to prolonged plateau phase calcium current which resulted in augmented calcium calmodulin kinase II activity. Ranolazine is an antagonist to the late sodium current at therapeutic levels.<sup>100</sup> The drug was able to reduce acceleration of the contraction and relaxation cycle of HCM trabeculae.<sup>101</sup> A recent study conducted by Flenner et al showed that ranolazine improved tolerance to high work load in mice with Mybpc3-targeted knock out in mice with hypertrophic cardiomyopathy. However, the mechanism of this was antagonizing the beta adrenergic receptor. In addition, six months of

treatment with ranolazine did not reverse the cardiac hypertrophy or dysfunction in vivo.<sup>102</sup> The RHYME (Ranolazine for treatment of angina or dyspnea in hypertrophic cardiomyopathy patients) trial demonstrated some improved symptoms and quality of life in a small 14 patient study.<sup>103</sup> There is a multicenter, double blind, placebo-controlled study underway to assess the effects of ranolazine on exercise capacity, diastolic function and symptoms in patients with hypertrophic cardiomyopathy (The RESTYLE-HCM, EUDRA-CT 2011-004507-20) which should provide more clear answers to the question of ranolazine in this disease. Electazine (GS 6615) is another inhibitor of the late phase sodium current originally designed for patients with LQT3 syndrome that was studied as a means to improve exercise capacity in patients with symptomatic hypertrophic cardiomyopathy (LIBERTY-HCM NCT02291237).<sup>104</sup> This trial was recently terminated for reasons that are not entirely clear at the time of this writing.

N-acetylcysteine has been studied as a possible therapeutic agent in hypertrophic cardiomyopathy. In a study in 2004 by Marian et al, transgenic mice carrying the hypertrophic cardiomyopathy associated troponin T Q92 mutation were treated with N-acetyl cysteine. They found that N-acetylcysteine reduced myocardial fibrosis.<sup>105</sup> This finding was supported in 2009 by Lombardi et al who showed that in a rabbit model of hypertrophic cardiomyopathy, beta myosin heavy change mutation Q403, treatment with N-acetylcysteine reversed cardiac and myocyte hypertrophy, interstitial fibrosis, reduced the propensity for ventricular arrhythmias, and prevented cardiac dysfunction.<sup>106</sup> This hypothesis was further corroborated when Wilder et al showed that administration of N-acetylcysteine for 30 days to transgenic mice expressing the hypertrophic cardiomyopathy associated tropomyosin mutation Tm-E180G reversed the baseline diastolic dysfunction and hypertrophy.<sup>107</sup> A clinical trial to test its potential benefit on hypertrophic cardiomyopathy patients is at the recruiting stage (NCT01537926).

## Lamin Mutations Dilated Cardiomyopathy

Lamin A and C are filament proteins of the nuclear envelope, encoded by the same gene (LMNA), that have structural function in the nuclear membrane and are involved in transcriptional regulations.<sup>108,109</sup> Ever since Bonne et al published that mutations in the gene encoding lamin A/C caused Emery-Dreifuss muscular dystrophy which can result in a dilated cardiomyopathy with conduction system disease, it has been known that lamin A/C mutations have a link to inherited cardiomyopathies.<sup>110</sup> The prevalence of LMNA mutations in families with inherited dilated cardiomyopathy has been estimated at 8%.<sup>111</sup>

Currently, there are no approved treatments for LMNA mutation associated dilated cardiomyopathy other than standard evidence based therapies all other patients with dilated cardiomyopathy should receive. Animal studies have given us insight into what future treatments might target. Enhanced p38alpha signaling has been discovered in the hearts of mice with dilated cardiomyopathy harboring an LMNA mutation and implemented in the pathophysiology. Using the P38 alpha inhibitor AARY-371791, Muchir et al treated mice with LMNA mutation associated dilated cardiomyopathy and found that LV dimensions and fractional shortening improved.<sup>112</sup> Improvement in cardiac function was shown in other LMNA mutations, again in mouse models. Preventing apoptosis is a proposed mechanism of

action for how p38 inhibition improves heart function in mice with LMNA mutations. Currently, AARY-371791 is undergoing a phase II clinical trial in patients with LMNA mutation dilated cardiomyopathy (NCT 02057341).

## Conclusions

Inherited cardiomyopathies represent a rarer slice of cardiovascular disease when compared to conditions with a mixture of genetic and non-genetic risk factors such as coronary artery disease and atrial fibrillation. As such, targeted therapies for these conditions have been slower in development. The keys to treating some of these conditions lay in the rebranding of older medications for these new uses and in other cases the development of novel agents used to target the molecular derangements resulting from genetic mutations. New therapies and therapeutic strategies are on the horizon and their development will likely result in improved outcomes for patients inflicted by these conditions.

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#### References

- Amin AS, Asghari-Roodsari A, Tan HL. Cardiac sodium channelopathies. Pflugers Arch. 2010; 460:223–237. [PubMed: 20091048]
- Olson TM, Michels VV, Ballew JD, Reyna SP, Karst ML, Herron KJ, Horton SC, Rodeheffer RJ, Anderson JL. Sodium channel mutations and susceptibility to heart failure and atrial fibrillation. JAMA. 2005; 293:447–454. [PubMed: 15671429]
- Brugada P, Brugada J. Right bundle branch block, persistent st segment elevation and sudden cardiac death: A distinct clinical and electrocardiographic syndrome. A multicenter report. J Am Coll Cardiol. 1992; 20:1391–1396. [PubMed: 1309182]
- 4. Chen Q, Kirsch GE, Zhang D, Brugada R, Brugada J, Brugada P, Potenza D, Moya A, Borggrefe M, Breithardt G, Ortiz-Lopez R, Wang Z, Antzelevitch C, O'Brien RE, Schulze-Bahr E, Keating MT, Towbin JA, Wang Q. Genetic basis and molecular mechanism for idiopathic ventricular fibrillation. Nature. 1998; 392:293–296. [PubMed: 9521325]
- Wang Q, Shen J, Splawski I, Atkinson D, Li Z, Robinson JL, Moss AJ, Towbin JA, Keating MT. Scn5a mutations associated with an inherited cardiac arrhythmia, long qt syndrome. Cell. 1995; 80:805–811. [PubMed: 7889574]
- Baroudi G, Acharfi S, Larouche C, Chahine M. Expression and intracellular localization of an scn5a double mutant r1232w/t1620m implicated in brugada syndrome. Circ Res. 2002; 90:E11–16. [PubMed: 11786529]
- Valdivia CR, Ackerman MJ, Tester DJ, Wada T, McCormack J, Ye B, Makielski JC. A novel scn5a arrhythmia mutation, m1766l, with expression defect rescued by mexiletine. Cardiovasc Res. 2002; 55:279–289. [PubMed: 12123767]
- Valdivia CR, Tester DJ, Rok BA, Porter CB, Munger TM, Jahangir A, Makielski JC, Ackerman MJ. A trafficking defective, brugada syndrome-causing scn5a mutation rescued by drugs. Cardiovasc Res. 2004; 62:53–62. [PubMed: 15023552]
- Tan BH, Valdivia CR, Song C, Makielski JC. Partial expression defect for the scn5a missense mutation g1406r depends on splice variant background q1077 and rescue by mexiletine. Am J Physiol Heart Circ Physiol. 2006; 291:H1822–1828. [PubMed: 16632547]
- Pfahnl AE, Viswanathan PC, Weiss R, Shang LL, Sanyal S, Shusterman V, Kornblit C, London B, Dudley SC Jr. A sodium channel pore mutation causing brugada syndrome. Heart Rhythm. 2007; 4:46–53. [PubMed: 17198989]

- Ruan Y, Denegri M, Liu N, Bachetti T, Seregni M, Morotti S, Severi S, Napolitano C, Priori SG. Trafficking defects and gating abnormalities of a novel scn5a mutation question gene-specific therapy in long qt syndrome type 3. Circ Res. 2010; 106:1374–1383. [PubMed: 20339117]
- 12\*. Mazzanti A, Maragna R, Faragli A, Monteforte N, Bloise R, Memmi M, Novelli V, Baiardi P, Bagnardi V, Etheridge SP, Napolitano C, Priori SG. Gene-specific therapy with mexiletine reduces arrhythmic events in patients with long qt syndrome type 3. J Am Coll Cardiol. 2016; 67:1053–1058. In this retrospective cohort study, patients with LQT3 syndrome treated with mexiletine had shortening of their QTc interval and a reduction in arrhythmic events. [PubMed: 26940925]
- 13. Priori SG, Blomstrom-Lundqvist C, Mazzanti A, Blom N, Borggrefe M, Camm J, Elliott PM, Fitzsimons D, Hatala R, Hindricks G, Kirchhof P, Kjeldsen K, Kuck KH, Hernandez-Madrid A, Nikolaou N, Norekval TM, Spaulding C, Van Veldhuisen DJ. 2015 esc guidelines for the management of patients with ventricular arrhythmias and the prevention of sudden cardiac death: The task force for the management of patients with ventricular arrhythmias and the prevention of sudden cardiac death: The task force for the european society of cardiology (esc) Endorsed by: Association for european paediatric and congenital cardiology (aepc). Eur Heart J. 2015; 36:2793–2867. [PubMed: 26320108]
- Fisher JD, Krikler D, Hallidie-Smith KA. Familial polymorphic ventricular arrhythmias: A quarter century of successful medical treatment based on serial exercise-pharmacologic testing. J Am Coll Cardiol. 1999; 34:2015–2022. [PubMed: 10588218]
- Priori SG, Napolitano C, Tiso N, Memmi M, Vignati G, Bloise R, Sorrentino V, Danieli GA. Mutations in the cardiac ryanodine receptor gene (hryr2) underlie catecholaminergic polymorphic ventricular tachycardia. Circulation. 2001; 103:196–200. [PubMed: 11208676]
- Laitinen PJ, Brown KM, Piippo K, Swan H, Devaney JM, Brahmbhatt B, Donarum EA, Marino M, Tiso N, Viitasalo M, Toivonen L, Stephan DA, Kontula K. Mutations of the cardiac ryanodine receptor (ryr2) gene in familial polymorphic ventricular tachycardia. Circulation. 2001; 103:485– 490. [PubMed: 11157710]
- Postma AV, Denjoy I, Hoorntje TM, Lupoglazoff JM, Da Costa A, Sebillon P, Mannens MM, Wilde AA, Guicheney P. Absence of calsequestrin 2 causes severe forms of catecholaminergic polymorphic ventricular tachycardia. Circ Res. 2002; 91:e21–26. [PubMed: 12386154]
- 18. Roux-Buisson N, Cacheux M, Fourest-Lieuvin A, Fauconnier J, Brocard J, Denjoy I, Durand P, Guicheney P, Kyndt F, Leenhardt A, Le Marec H, Lucet V, Mabo P, Probst V, Monnier N, Ray PF, Santoni E, Tremeaux P, Lacampagne A, Faure J, Lunardi J, Marty I. Absence of triadin, a protein of the calcium release complex, is responsible for cardiac arrhythmia with sudden death in human. Hum Mol Genet. 2012; 21:2759–2767. [PubMed: 22422768]
- Nyegaard M, Overgaard MT, Sondergaard MT, Vranas M, Behr ER, Hildebrandt LL, Lund J, Hedley PL, Camm AJ, Wettrell G, Fosdal I, Christiansen M, Borglum AD. Mutations in calmodulin cause ventricular tachycardia and sudden cardiac death. Am J Hum Genet. 2012; 91:703–712. [PubMed: 23040497]
- 20. Venetucci L, Denegri M, Napolitano C, Priori SG. Inherited calcium channelopathies in the pathophysiology of arrhythmias. Nat Rev Cardiol. 2012; 9:561–575. [PubMed: 22733215]
- Tester DJ, Arya P, Will M, Haglund CM, Farley AL, Makielski JC, Ackerman MJ. Genotypic heterogeneity and phenotypic mimicry among unrelated patients referred for catecholaminergic polymorphic ventricular tachycardia genetic testing. Heart Rhythm. 2006; 3:800–805. [PubMed: 16818210]
- Watanabe H, Knollmann BC. Mechanism underlying catecholaminergic polymorphic ventricular tachycardia and approaches to therapy. J Electrocardiol. 2011; 44:650–655. [PubMed: 21872879]
- van der Werf C, Zwinderman AH, Wilde AA. Therapeutic approach for patients with catecholaminergic polymorphic ventricular tachycardia: State of the art and future developments. Europace. 2012; 14:175–183. [PubMed: 21893508]
- Tester DJ, Spoon DB, Valdivia HH, Makielski JC, Ackerman MJ. Targeted mutational analysis of the ryr2-encoded cardiac ryanodine receptor in sudden unexplained death: A molecular autopsy of 49 medical examiner/coroner's cases. Mayo Clin Proc. 2004; 79:1380–1384. [PubMed: 15544015]
- 25. Terentyev D, Nori A, Santoro M, Viatchenko-Karpinski S, Kubalova Z, Gyorke I, Terentyeva R, Vedamoorthyrao S, Blom NA, Valle G, Napolitano C, Williams SC, Volpe P, Priori SG, Gyorke S.

Abnormal interactions of calsequestrin with the ryanodine receptor calcium release channel complex linked to exercise-induced sudden cardiac death. Circ Res. 2006; 98:1151–1158. [PubMed: 16601229]

- 26. Hayashi M, Denjoy I, Extramiana F, Maltret A, Buisson NR, Lupoglazoff JM, Klug D, Takatsuki S, Villain E, Kamblock J, Messali A, Guicheney P, Lunardi J, Leenhardt A. Incidence and risk factors of arrhythmic events in catecholaminergic polymorphic ventricular tachycardia. Circulation. 2009; 119:2426–2434. [PubMed: 19398665]
- 27. Leren IS, Saberniak J, Majid E, Haland TF, Edvardsen T, Haugaa KH. Nadolol decreases the incidence and severity of ventricular arrhythmias during exercise stress testing compared with beta1-selective beta-blockers in patients with catecholaminergic polymorphic ventricular tachycardia. Heart Rhythm. 2016; 13:433–440. [PubMed: 26432584]
- Watanabe H, Chopra N, Laver D, Hwang HS, Davies SS, Roach DE, Duff HJ, Roden DM, Wilde AA, Knollmann BC. Flecainide prevents catecholaminergic polymorphic ventricular tachycardia in mice and humans. Nat Med. 2009; 15:380–383. [PubMed: 19330009]
- Hilliard FA, Steele DS, Laver D, Yang Z, Le Marchand SJ, Chopra N, Piston DW, Huke S, Knollmann BC. Flecainide inhibits arrhythmogenic ca2+ waves by open state block of ryanodine receptor ca2+ release channels and reduction of ca2+ spark mass. J Mol Cell Cardiol. 2010; 48:293–301. [PubMed: 19835880]
- 30\*. van der Werf C, Kannankeril PJ, Sacher F, Krahn AD, Viskin S, Leenhardt A, Shimizu W, Sumitomo N, Fish FA, Bhuiyan ZA, Willems AR, van der Veen MJ, Watanabe H, Laborderie J, Haissaguerre M, Knollmann BC, Wilde AA. Flecainide therapy reduces exercise-induced ventricular arrhythmias in patients with catecholaminergic polymorphic ventricular tachycardia. J Am Coll Cardiol. 2011; 57:2244–2254. In this retrospective analysis, patietns with gentoype positive CPVT previously uncontrolled by conventional drug therapy started on flexainide had a reduction in exercise induced ventricular arrhymias. [PubMed: 21616285]
- 31. Watanabe H, van der Werf C, Roses-Noguer F, Adler A, Sumitomo N, Veltmann C, Rosso R, Bhuiyan ZA, Bikker H, Kannankeril PJ, Horie M, Minamino T, Viskin S, Knollmann BC, Till J, Wilde AA. Effects of flecainide on exercise-induced ventricular arrhythmias and recurrences in genotype-negative patients with catecholaminergic polymorphic ventricular tachycardia. Heart Rhythm. 2013; 10:542–547. [PubMed: 23286974]
- 32. Pott C, Dechering DG, Reinke F, Muszynski A, Zellerhoff S, Bittner A, Kobe J, Wasmer K, Schulze-Bahr E, Monnig G, Kotthoff S, Eckardt L. Successful treatment of catecholaminergic polymorphic ventricular tachycardia with flecainide: A case report and review of the current literature. Europace. 2011; 13:897–901. [PubMed: 21292648]
- Hong RA, Rivera KK, Jittirat A, Choi JJ. Flecainide suppresses defibrillator-induced storming in catecholaminergic polymorphic ventricular tachycardia. Pacing Clin Electrophysiol. 2012; 35:794– 797. [PubMed: 22553997]
- 34. Hwang HS, Hasdemir C, Laver D, Mehra D, Turhan K, Faggioni M, Yin H, Knollmann BC. Inhibition of cardiac ca2+ release channels (ryr2) determines efficacy of class i antiarrhythmic drugs in catecholaminergic polymorphic ventricular tachycardia. Circ Arrhythm Electrophysiol. 2011; 4:128–135. [PubMed: 21270101]
- Smith GL, MacQuaide N. The direct actions of flecainide on the human cardiac ryanodine receptor: Keeping open the debate on the mechanism of action of local anesthetics in cpvt. Circ Res. 2015; 116:1284–1286. [PubMed: 25858058]
- 36. Liu N, Denegri M, Ruan Y, Avelino-Cruz JE, Perissi A, Negri S, Napolitano C, Coetzee WA, Boyden PA, Priori SG. Short communication: Flecainide exerts an antiarrhythmic effect in a mouse model of catecholaminergic polymorphic ventricular tachycardia by increasing the threshold for triggered activity. Circ Res. 2011; 109:291–295. [PubMed: 21680895]
- Bannister ML, Thomas NL, Sikkel MB, Mukherjee S, Maxwell C, MacLeod KT, George CH, Williams AJ. The mechanism of flecainide action in cpvt does not involve a direct effect on ryr2. Circ Res. 2015; 116:1324–1335. [PubMed: 25648700]
- Khoury A, Marai I, Suleiman M, Blich M, Lorber A, Gepstein L, Boulos M. Flecainide therapy suppresses exercise-induced ventricular arrhythmias in Patients with casq2-associated catecholaminergic polymorphic ventricular tachycardia. Heart Rhythm. 2013; 10:1671–1675. [PubMed: 23954267]

- Wilde AA, Bhuiyan ZA, Crotti L, Facchini M, De Ferrari GM, Paul T, Ferrandi C, Koolbergen DR, Odero A, Schwartz PJ. Left cardiac sympathetic denervation for catecholaminergic polymorphic ventricular tachycardia. N Engl J Med. 2008; 358:2024–2029. [PubMed: 18463378]
- 40. Schneider HE, Steinmetz M, Krause U, Kriebel T, Ruschewski W, Paul T. Left cardiac sympathetic denervation for the management of life-threatening ventricular tachyarrhythmias in young patients with catecholaminergic polymorphic ventricular tachycardia and long qt syndrome. Clin Res Cardiol. 2013; 102:33–42. [PubMed: 22821214]
- Hofferberth SC, Cecchin F, Loberman D, Fynn-Thompson F. Left thoracoscopic sympathectomy for cardiac denervation in patients with life-threatening ventricular arrhythmias. J Thorac Cardiovasc Surg. 2014; 147:404–409. [PubMed: 24268954]
- 42. McNamara C, Cullen P, Rackauskas M, Kelly R, O'Sullivan KE, Galvin J, Eaton D. Left cardiac sympathetic denervation: Case series and technical report. Ir J Med Sci. 2017
- 43. Costello JP, Wilson JK, Louis C, Peer SM, Zurakowski D, Nadler EP, Qureshi FG, Jonas RA, Greene EA, Berul CI, Moak JP, Nath DS. Surgical cardiac denervation therapy for treatment of congenital ion channelopathies in pediatric patients: A contemporary, single institutional experience. World J Pediatr Congenit Heart Surg. 2015; 6:33–38. [PubMed: 25548341]
- 44. Atallah J, Fynn-Thompson F, Cecchin F, DiBardino DJ, Walsh EP, Berul CI. Video-assisted thoracoscopic cardiac denervation: A potential novel therapeutic option for children with intractable ventricular arrhythmias. Ann Thorac Surg. 2008; 86:1620–1625. [PubMed: 19049760]
- 45. Lehnart SE, Wehrens XH, Laitinen PJ, Reiken SR, Deng SX, Cheng Z, Landry DW, Kontula K, Swan H, Marks AR. Sudden death in familial polymorphic ventricular tachycardia associated with calcium release channel (ryanodine receptor) leak. Circulation. 2004; 109:3208–3214. [PubMed: 15197150]
- 46. Sedej S, Heinzel FR, Walther S, Dybkova N, Wakula P, Groborz J, Gronau P, Maier LS, Vos MA, Lai FA, Napolitano C, Priori SG, Kockskamper J, Pieske B. Na+-dependent sr ca2+ overload induces arrhythmogenic events in mouse cardiomyocytes with a human cpvt mutation. Cardiovasc Res. 2010; 87:50–59. [PubMed: 20080988]
- 47. Li N, Wang Q, Sibrian-Vazquez M, Klipp RC, Reynolds JO, Word TA, Scott L Jr, Salama G, Strongin RM, Abramson JJ, Wehrens XH. Treatment of catecholaminergic polymorphic ventricular tachycardia in mice using novel ryr2-modifying drugs. Int J Cardiol. 2017; 227:668– 673. [PubMed: 27838126]
- Chakraborty R, Muchtar E, Gertz MA. Newer therapies for amyloid cardiomyopathy. Curr Heart Fail Rep. 2016; 13:237–246. [PubMed: 27497715]
- Rapezzi C, Quarta CC, Riva L, Longhi S, Gallelli I, Lorenzini M, Ciliberti P, Biagini E, Salvi F, Branzi A. Transthyretin-related amyloidoses and the heart: A clinical overview. Nat Rev Cardiol. 2010; 7:398–408. [PubMed: 20479782]
- 50. Connors LH, Prokaeva T, Lim A, Theberge R, Falk RH, Doros G, Berg A, Costello CE, O'Hara C, Seldin DC, Skinner M. Cardiac amyloidosis in African americans: Comparison of clinical and laboratory features of transthyretin v122i amyloidosis and immunoglobulin light chain amyloidosis. Am Heart J. 2009; 158:607–614. [PubMed: 19781421]
- 51. Ruberg FL, Maurer MS, Judge DP, Zeldenrust S, Skinner M, Kim AY, Falk RH, Cheung KN, Patel AR, Pano A, Packman J, Grogan DR. Prospective evaluation of the morbidity and mortality of wild-type and v122i mutant transthyretin amyloid cardiomyopathy: The transthyretin amyloidosis cardiac study (tracs). Am Heart J. 2012; 164:222–228. e221. [PubMed: 22877808]
- 52. Bulawa CE, Connelly S, Devit M, Wang L, Weigel C, Fleming JA, Packman J, Powers ET, Wiseman RL, Foss TR, Wilson IA, Kelly JW, Labaudiniere R. Tafamidis, a potent and selective transthyretin kinetic stabilizer that inhibits the amyloid cascade. Proc Natl Acad Sci U S A. 2012; 109:9629–9634. [PubMed: 22645360]
- 53. Coelho T, Maia LF, Martins da Silva A, Waddington Cruz M, Plante-Bordeneuve V, Lozeron P, Suhr OB, Campistol JM, Conceicao IM, Schmidt HH, Trigo P, Kelly JW, Labaudiniere R, Chan J, Packman J, Wilson A, Grogan DR. Tafamidis for transthyretin familial amyloid polyneuropathy: A randomized, controlled trial. Neurology. 2012; 79:785–792. [PubMed: 22843282]
- 54. Maurer MS, Grogan DR, Judge DP, Mundayat R, Packman J, Lombardo I, Quyyumi AA, Aarts J, Falk RH. Tafamidis in transthyretin amyloid cardiomyopathy: Effects on transthyretin stabilization and clinical outcomes. Circ Heart Fail. 2015; 8:519–526. [PubMed: 25872787]

- 55. Damy T, Judge DP, Kristen AV, Berthet K, Li H, Aarts J. Cardiac findings and events observed in an open-label clinical trial of tafamidis in patients with non-val30met and non-val122ile hereditary transthyretin amyloidosis. J Cardiovasc Transl Res. 2015; 8:117–127. [PubMed: 25743445]
- 56. Berk JL, Dyck PJ, Obici L, Zeldenrust SR, Sekijima Y, Yamashita T, Ando Y, Ikeda SI, Gorevic P, Merlini G, Kelly JW, Skinner M, Bisbee AB, Suhr OB. The diflunisal trial: Update on study drug tolerance and disease progression. Amyloid. 2011; (18 Suppl 1):196–197. [PubMed: 21838485]
- 57. Merlini G, Plante-Bordeneuve V, Judge DP, Schmidt H, Obici L, Perlini S, Packman J, Tripp T, Grogan DR. Effects of tafamidis on transthyretin stabilization and clinical outcomes in patients with non-val30met transthyretin amyloidosis. J Cardiovasc Transl Res. 2013; 6:1011–1020. [PubMed: 24101373]
- Adamski-Werner SL, Palaninathan SK, Sacchettini JC, Kelly JW. Diflunisal analogues stabilize the native state of transthyretin. Potent inhibition of amyloidogenesis. J Med Chem. 2004; 47:355– 374. [PubMed: 14711308]
- Castano A, Helmke S, Alvarez J, Delisle S, Maurer MS. Diflunisal for attr cardiac amyloidosis. Congest Heart Fail. 2012; 18:315–319. [PubMed: 22747647]
- Obici L, Cortese A, Lozza A, Lucchetti J, Gobbi M, Palladini G, Perlini S, Saraiva MJ, Merlini G. Doxycycline plus tauroursodeoxycholic acid for transthyretin amyloidosis: A phase ii study. Amyloid. 2012; 1(19 Suppl):34–36.
- 61\*. Coelho T, Adams D, Silva A, Lozeron P, Hawkins PN, Mant T, Perez J, Chiesa J, Warrington S, Tranter E, Munisamy M, Falzone R, Harrop J, Cehelsky J, Bettencourt BR, Geissler M, Butler JS, Sehgal A, Meyers RE, Chen Q, Borland T, Hutabarat RM, Clausen VA, Alvarez R, Fitzgerald K, Gamba-Vitalo C, Nochur SV, Vaishnaw AK, Sah DW, Gollob JA, Suhr OB. Safety and efficacy of rnai therapy for transthyretin amyloidosis. N Engl J Med. 2013; 369:819–829. In this non-randomised heathy volunteer controlled study, patients with transthyreting amyloidosis were treated with RNA interference which suppressed the production of transthyretin. This was prooved the concept that targeting messenger RNA was feasible in reducing the disease-causing mutant protein. [PubMed: 23984729]
- Gulati V, Harikrishnan P, Palaniswamy C, Aronow WS, Jain D, Frishman WH. Cardiac involvement in hemochromatosis. Cardiol Rev. 2014; 22:56–68. [PubMed: 24503941]
- Dabestani A, Child JS, Henze E, Perloff JK, Schon H, Figueroa WG, Schelbert HR, Thessomboon S. Primary hemochromatosis: Anatomic and physiologic characteristics of the cardiac ventricles and their response to phlebotomy. Am J Cardiol. 1984; 54:153–159. [PubMed: 6741807]
- Cecchetti G, Binda A, Piperno A, Nador F, Fargion S, Fiorelli G. Cardiac alterations in 36 consecutive patients with idiopathic haemochromatosis: Polygraphic and echocardiographic evaluation. Eur Heart J. 1991; 12:224–230. [PubMed: 2044557]
- 65. Brittenham GM, Griffith PM, Nienhuis AW, McLaren CE, Young NS, Tucker EE, Allen CJ, Farrell DE, Harris JW. Efficacy of deferoxamine in preventing complications of iron overload in patients with thalassemia major. N Engl J Med. 1994; 331:567–573. [PubMed: 8047080]
- 66. Davis BA, O'Sullivan C, Jarritt PH, Porter JB. Value of sequential monitoring of left ventricular ejection fraction in the management of thalassemia major. Blood. 2004; 104:263–269. [PubMed: 15001468]
- 67. Anderson LJ, Westwood MA, Holden S, Davis B, Prescott E, Wonke B, Porter JB, Walker JM, Pennell DJ. Myocardial iron clearance during reversal of siderotic cardiomyopathy with intravenous desferrioxamine: A prospective study using t2\* cardiovascular magnetic resonance. Br J Haematol. 2004; 127:348–355. [PubMed: 15491298]
- Gujja P, Rosing DR, Tripodi DJ, Shizukuda Y. Iron overload cardiomyopathy: Better understanding of an increasing disorder. J Am Coll Cardiol. 2010; 56:1001–1012. [PubMed: 20846597]
- 69. Germain DP. Fabry disease. Orphanet J Rare Dis. 2010; 5:30. [PubMed: 21092187]
- Schiffmann R, Warnock DG, Banikazemi M, Bultas J, Linthorst GE, Packman S, Sorensen SA, Wilcox WR, Desnick RJ. Fabry disease: Progression of nephropathy, and prevalence of cardiac and cerebrovascular events before enzyme replacement therapy. Nephrol Dial Transplant. 2009; 24:2102–2111. [PubMed: 19218538]
- 71. Schiffmann R, Murray GJ, Treco D, Daniel P, Sellos-Moura M, Myers M, Quirk JM, Zirzow GC, Borowski M, Loveday K, Anderson T, Gillespie F, Oliver KL, Jeffries NO, Doo E, Liang TJ,

Kreps C, Gunter K, Frei K, Crutchfield K, Selden RF, Brady RO. Infusion of alpha-galactosidase a reduces tissue globotriaosylceramide storage in patients with fabry disease. Proc Natl Acad Sci U S A. 2000; 97:365–370. [PubMed: 10618424]

- 72. Eng CM, Guffon N, Wilcox WR, Germain DP, Lee P, Waldek S, Caplan L, Linthorst GE, Desnick RJ. Safety and efficacy of recombinant human alpha-galactosidase a--replacement therapy in fabry's disease. N Engl J Med. 2001; 345:9–16. [PubMed: 11439963]
- 73. Eng CM, Banikazemi M, Gordon RE, Goldman M, Phelps R, Kim L, Gass A, Winston J, Dikman S, Fallon JT, Brodie S, Stacy CB, Mehta D, Parsons R, Norton K, O'Callaghan M, Desnick RJ. A phase 1/2 clinical trial of enzyme replacement in fabry disease: Pharmacokinetic, substrate clearance, and safety studies. Am J Hum Genet. 2001; 68:711–722. [PubMed: 11179018]
- 74. Weidemann F, Breunig F, Beer M, Sandstede J, Turschner O, Voelker W, Ertl G, Knoll A, Wanner C, Strotmann JM. Improvement of cardiac function during enzyme replacement therapy in patients with fabry disease: A prospective strain rate imaging study. Circulation. 2003; 108:1299–1301. [PubMed: 12952834]
- 75. Banikazemi M, Bultas J, Waldek S, Wilcox WR, Whitley CB, McDonald M, Finkel R, Packman S, Bichet DG, Warnock DG, Desnick RJ. Agalsidase-beta therapy for advanced fabry disease: A randomized trial. Ann Intern Med. 2007; 146:77–86. [PubMed: 17179052]
- 76\*. Mehta A, Beck M, Elliott P, Giugliani R, Linhart A, Sunder-Plassmann G, Schiffmann R, Barbey F, Ries M, Clarke JT. Enzyme replacement therapy with agalsidase alfa in patients with fabry's disease: An analysis of registry data. Lancet. 2009; 374:1986–1996. In this registry study of the 5 year outcomes of enzyme replacement therapy for patients with Fabry's disease, quality of life improved significantly. A reduction in left ventricular mass for those with baseline left ventricular hypertrophy was also observed. [PubMed: 19959221]
- 77. Weidemann F, Niemann M, Breunig F, Herrmann S, Beer M, Stork S, Voelker W, Ertl G, Wanner C, Strotmann J. Long-term effects of enzyme replacement Therapy on fabry cardiomyopathy: Evidence for a better outcome with early treatment. Circulation. 2009; 119:524–529. [PubMed: 19153271]
- Germain DP, Charrow J, Desnick RJ, Guffon N, Kempf J, Lachmann RH, Lemay R, Linthorst GE, Packman S, Scott CR, Waldek S, Warnock DG, Weinreb NJ, Wilcox WR. Ten-year outcome of enzyme replacement therapy with agalsidase beta in patients with fabry disease. J Med Genet. 2015; 52:353–358. [PubMed: 25795794]
- Mehta A, Clarke JT, Giugliani R, Elliott P, Linhart A, Beck M, Sunder-Plassmann G. Natural course of fabry disease: Changing pattern of causes of death in fos-fabry outcome survey. J Med Genet. 2009; 46:548–552. [PubMed: 19473999]
- Bos JM, Towbin JA, Ackerman MJ. Diagnostic, prognostic, and therapeutic implications of genetic testing for hypertrophic cardiomyopathy. J Am Coll Cardiol. 2009; 54:201–211. [PubMed: 19589432]
- Alves ML, Gaffin RD, Wolska BM. Rescue of familial cardiomyopathies by modifications at the level of sarcomere and ca2+ fluxes. J Mol Cell Cardiol. 2010; 48:834–842. [PubMed: 20079744]
- Hwang PM, Sykes BD. Targeting the sarcomere to correct muscle function. Nat Rev Drug Discov. 2015; 14:313–328. [PubMed: 25881969]
- 83. Yamazaki T, Suzuki J, Shimamoto R, Tsuji T, Ohmoto-Sekine Y, Ohtomo K, Nagai R. A new therapeutic strategy for hypertrophic nonobstructive cardiomyopathy in humans. A randomized and prospective study with an angiotensin ii receptor blocker. Int Heart J. 2007; 48:715–724. [PubMed: 18160763]
- 84. Shimada YJ, Passeri JJ, Baggish AL, O'Callaghan C, Lowry PA, Yannekis G, Abbara S, Ghoshhajra BB, Rothman RD, Ho CY, Januzzi JL, Seidman CE, Fifer MA. Effects of losartan on left ventricular hypertrophy and fibrosis in patients with nonobstructive hypertrophic cardiomyopathy. JACC Heart Fail. 2013; 1:480–487. [PubMed: 24621999]
- Penicka M, Gregor P, Kerekes R, Marek D, Curila K, Krupicka J. The effects of candesartan on left ventricular hypertrophy and function in nonobstructive hypertrophic cardiomyopathy: A pilot, randomized study. J Mol Diagn. 2009; 11:35–41. [PubMed: 19074594]
- 86. Axelsson A, Iversen K, Vejlstrup N, Ho C, Norsk J, Langhoff L, Ahtarovski K, Corell P, Havndrup O, Jensen M, Bundgaard H. Efficacy and safety of the angiotensin ii receptor blocker losartan for

hypertrophic cardiomyopathy: The inherit randomised, double-blind, placebo-controlled trial. Lancet Diabetes Endocrinol. 2015; 3:123–131. [PubMed: 25533774]

- Ammirati E, Contri R, Coppini R, Cecchi F, Frigerio M, Olivotto I. Pharmacological treatment of hypertrophic cardiomyopathy: Current practice and novel perspectives. Eur J Heart Fail. 2016; 18:1106–1118. [PubMed: 27109894]
- 88. Tsybouleva N, Zhang L, Chen S, Patel R, Lutucuta S, Nemoto S, DeFreitas G, Entman M, Carabello BA, Roberts R, Marian AJ. Aldosterone, through novel signaling proteins, is a fundamental molecular bridge between the genetic defect and the cardiac phenotype of hypertrophic cardiomyopathy. Circulation. 2004; 109:1284–1291. [PubMed: 14993121]
- Pena JR, Szkudlarek AC, Warren CM, Heinrich LS, Gaffin RD, Jagatheesan G, del Monte F, Hajjar RJ, Goldspink PH, Solaro RJ, Wieczorek DF, Wolska BM. Neonatal gene transfer of serca2a delays onset of hypertrophic remodeling and Improves function in familial hypertrophic cardiomyopathy. J Mol Cell Cardiol. 2010; 49:993–1002. [PubMed: 20854827]
- 90. Gaffin RD, Pena JR, Alves MS, Dias FA, Chowdhury SA, Heinrich LS, Goldspink PH, Kranias EG, Wieczorek DF, Wolska BM. Long-term rescue of a familial hypertrophic cardiomyopathy caused by a mutation in the thin filament protein, tropomyosin, via modulation of a calcium cycling protein. J Mol Cell Cardiol. 2011; 51:812–820. [PubMed: 21840315]
- 91. Semsarian C, Ahmad I, Giewat M, Georgakopoulos D, Schmitt JP, McConnell BK, Reiken S, Mende U, Marks AR, Kass DA, Seidman CE, Seidman JG. The l-type calcium channel inhibitor diltiazem prevents cardiomyopathy in a mouse model. J Clin Invest. 2002; 109:1013–1020. [PubMed: 11956238]
- 92. Westermann D, Knollmann BC, Steendijk P, Rutschow S, Riad A, Pauschinger M, Potter JD, Schultheiss HP, Tschope C. Diltiazem treatment prevents diastolic heart failure in mice with familial hypertrophic cardiomyopathy. Eur J Heart Fail. 2006; 8:115–121. [PubMed: 16214409]
- 93. McTaggart DR. Diltiazem reverses tissue doppler velocity abnormalities in pre-clinical hypertrophic cardiomyopathy. Heart Lung Circ. 2004; 13:39–40. [PubMed: 16352166]
- 94. Ho CY, Lakdawala NK, Cirino AL, Lipshultz SE, Sparks E, Abbasi SA, Kwong RY, Antman EM, Semsarian C, Gonzalez A, Lopez B, Diez J, Orav EJ, Colan SD, Seidman CE. Diltiazem treatment for pre-clinical hypertrophic cardiomyopathy sarcomere mutation carriers: A pilot randomized trial to modify disease expression. JACC Heart Fail. 2015; 3:180–188. [PubMed: 25543971]
- Dou Y, Arlock P, Arner A. Blebbistatin specifically inhibits actin-myosin interaction in mouse cardiac muscle. Am J Physiol Cell Physiol. 2007; 293:C1148–1153. [PubMed: 17615158]
- 96. Baudenbacher F, Schober T, Pinto JR, Sidorov VY, Hilliard F, Solaro RJ, Potter JD, Knollmann BC. Myofilament ca2+ sensitization causes susceptibility to cardiac arrhythmia in mice. J Clin Invest. 2008; 118:3893–3903. [PubMed: 19033660]
- Coutu P, Bennett CN, Favre EG, Day SM, Metzger JM. Parvalbumin corrects slowed relaxation in adult cardiac myocytes expressing hypertrophic cardiomyopathy-linked alpha-tropomyosin mutations. Circ Res. 2004; 94:1235–1241. [PubMed: 15059934]
- Hector M. Rodriguez SW-C, Raja Kawas, Yonghong Song, Arvinder Sran, Johan Oslob. Modulation of the cardiac sarcomere by a small molecule agent myk0000461: A potential therapeutic for the treatment of genetic hypertrophic cardiomyopathies. Biophys J. 2015:106. [PubMed: 26153707]
- 99\*. Green EM, Wakimoto H, Anderson RL, Evanchik MJ, Gorham JM, Harrison BC, Henze M, Kawas R, Oslob JD, Rodriguez HM, Song Y, Wan W, Leinwand LA, Spudich JA, McDowell RS, Seidman JG, Seidman CE. A small-molecule inhibitor of sarcomere contractility suppresses hypertrophic cardiomyopathy in mice. Science. 2016; 351:617–621. In this study, the small molecule MYK-461, which reduces contractility by inhibiting myosin ATPase suppressed the development of the typical phenotypic markers of hypertrophic cardiomyopathy in mice carrying human mutations in the myosin hevay chain. [PubMed: 26912705]
- 100. Antzelevitch C, Belardinelli L, Wu L, Fraser H, Zygmunt AC, Burashnikov A, Di Diego JM, Fish JM, Cordeiro JM, Goodrow RJ Jr, Scornik F, Perez G. Electrophysiologic properties and antiarrhythmic actions of a novel antianginal agent. J Cardiovasc Pharmacol Ther. 2004; (9 Suppl 1):S65–83. [PubMed: 15378132]
- 101. Coppini R, Ferrantini C, Yao L, Fan P, Del Lungo M, Stillitano F, Sartiani L, Tosi B, Suffredini S, Tesi C, Yacoub M, Olivotto I, Belardinelli L, Poggesi C, Cerbai E, Mugelli A. Late sodium

current inhibition reverses electromechanical dysfunction in human hypertrophic cardiomyopathy. Circulation. 2013; 127:575–584. [PubMed: 23271797]

- 102. Flenner F, Friedrich FW, Ungeheuer N, Christ T, Geertz B, Reischmann S, Wagner S, Stathopoulou K, Sohren KD, Weinberger F, Schwedhelm E, Cuello F, Maier LS, Eschenhagen T, Carrier L. Ranolazine antagonizes catecholamine-induced dysfunction in isolated cardiomyocytes, but lacks long-term therapeutic effects in vivo in a mouse model of hypertrophic cardiomyopathy. Cardiovasc Res. 2015
- 103. Gentry JL 3rd, Mentz RJ, Hurdle M, Wang A. Ranolazine for treatment of angina or dyspnea in hypertrophic cardiomyopathy patients (rhyme). J Am Coll Cardiol. 2016; 68:1815–1817. [PubMed: 27737749]
- 104. Olivotto I, Hellawell JL, Farzaneh-Far R, Blair C, Coppini R, Myers J, Belardinelli L, Maron MS. Novel approach targeting the complex pathophysiology of hypertrophic cardiomyopathy: The impact of late sodium current inhibition on exercise capacity in subjects with symptomatic hypertrophic cardiomyopathy (liberty-hcm) trial. Circ Heart Fail. 2016; 9:e002764. [PubMed: 26915375]
- 105. Marian AJ, Senthil V, Chen SN, Lombardi R. Antifibrotic effects of antioxidant n-acetylcysteine in a mouse model of human hypertrophic cardiomyopathy mutation. J Am Coll Cardiol. 2006; 47:827–834. [PubMed: 16487852]
- 106. Lombardi R, Rodriguez G, Chen SN, Ripplinger CM, Li W, Chen J, Willerson JT, Betocchi S, Wickline SA, Efimov IR, Marian AJ. Resolution of established cardiac hypertrophy and fibrosis and prevention of systolic dysfunction in a transgenic rabbit model of human cardiomyopathy through thiol-sensitive mechanisms. Circulation. 2009; 119:1398–1407. [PubMed: 19255346]
- 107. Wilder T, Ryba DM, Wieczorek DF, Wolska BM, Solaro RJ. N-acetylcysteine reverses diastolic dysfunction and hypertrophy in familial hypertrophic cardiomyopathy. Am J Physiol Heart Circ Physiol. 2015; 309:H1720–1730. [PubMed: 26432840]
- 108. Graham RM, Owens WA. Pathogenesis of inherited forms of dilated cardiomyopathy. N Engl J Med. 1999; 341:1759–1762. [PubMed: 10580077]
- 109. Fatkin D, MacRae C, Sasaki T, Wolff MR, Porcu M, Frenneaux M, Atherton J, Vidaillet HJ Jr, Spudich S, De Girolami U, Seidman JG, Seidman C, Muntoni F, Muehle G, Johnson W, McDonough B. Missense mutations in the rod domain of the lamin a/c gene as causes of dilated cardiomyopathy and conduction-system disease. N Engl J Med. 1999; 341:1715–1724. [PubMed: 10580070]
- 110. Bonne G, Di Barletta MR, Varnous S, Becane HM, Hammouda EH, Merlini L, Muntoni F, Greenberg CR, Gary F, Urtizberea JA, Duboc D, Fardeau M, Toniolo D, Schwartz K. Mutations in the gene encoding lamin a/c cause autosomal dominant emery-dreifuss muscular dystrophy. Nat Genet. 1999; 21:285–288. [PubMed: 10080180]
- 111. Taylor MR, Fain PR, Sinagra G, Robinson ML, Robertson AD, Carniel E, Di Lenarda A, Bohlmeyer TJ, Ferguson DA, Brodsky GL, Boucek MM, Lascor J, Moss AC, Li WL, Stetler GL, Muntoni F, Bristow MR, Mestroni L. Natural history of dilated cardiomyopathy due to lamin a/c gene mutations. J Am Coll Cardiol. 2003; 41:771–780. [PubMed: 12628721]
- 112. Muchir A, Wu W, Choi JC, Iwata S, Morrow J, Homma S, Worman HJ. Abnormal p38alpha mitogen-activated protein kinase signaling in dilated cardiomyopathy caused by lamin a/c gene mutation. Hum Mol Genet. 2012; 21:4325–4333. [PubMed: 22773734]

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