Environmental factors in the development and progression of late-onset Alzheimer's disease

Moses N. Wainaina^{1,3}, Zhichun Chen¹, Chunjiu Zhong^{1,2}

1 *Department of Neurology, Zhongshan Hospital; State Key Laboratory of Medical Neurobiology, Fudan University, Shanghai 200032, China*

2 *Institutes of Brain Science; Fudan University, Shanghai 200032, China* 3 *Pwani University, Kilifi , Kenya* Corresponding author: Chunjiu Zhong*.* E-mail: Zhongcj@163.com

© Shanghai Institutes for Biological Sciences, CAS and Springer-Verlag Berlin Heidelberg 2014

Late-onset Alzheimer's disease (LOAD) is an age-related neurodegenerative disorder characterized by gradual loss of synapses and neurons, but its pathogenesis remains to be clarified. Neurons live in an environment constituted by neurons themselves and glial cells. In this review, we propose that the neuronal degeneration in the AD brain is partially caused by diverse environmental factors. We first discuss various environmental stresses and the corresponding responses at different levels. Then we propose some mechanisms underlying the specific pathological changes, in particular, hypothalamic-pituitary adrenal axis dysfunction at the systemic level; cerebrovascular dysfunction, metal toxicity, glial activation, and Aβ toxicity at the intercellular level; and kinase–phosphatase imbalance and epigenetic modification at the intracellular level. Finally, we discuss the possibility of developing new strategies for the prevention and treatment of LOAD from the perspective of environmental stress. We conclude that environmental factors play a significant role in the development of LOAD through multiple pathological mechanisms.

Keywords: Alzheimer's disease; environmental factors; corticotrophin-releasing factor; cerebrovascular; metal toxicity; glia; astrocyte; microglia; Aβ; kinase; phosphatase; tau; hyperphosphorylation; epigenetic modification; DNA methylation; histone acetylation

Introduction

The most important feature of organisms is their adaptability to external environmental changes by modulating their internal metabolism or external behaviors. But when the environmental variations exceed a certain level, environmental stress, a term originating from ecology, is induced^[1]. For example, life-forms under disadvantageous environmental conditions exhibit poor growth and responses different from those under fertile conditions^[2]. Excessive environmental stress or abnormal responses to environmental factors can induce morbidity and even $death^{[3, 4]}$. Human beings are also influenced by numerous natural, psychological, and social factors from birth.

The cerebrovascular system, especially the bloodbrain barrier (BBB), plays an essential role in the transport and excretion of environmental factors. Besides, the environment of neurons, constituted by neurons themselves and glial cells^[5, 6], markedly impacts their functional status and fate^[7, 8]. Thus, environmental factors may participate in aging or aging-related diseases $[9, 10]$, including late-onset Alzheimer's disease (LOAD)^[11].

To better understand the relationship between environmental factors and LOAD, we decided to explore LOAD pathophysiology from a new direction in this review. To date, many environmental factors have been proposed as risks of LOAD, including metals $[12, 13]$, air pollution $[11, 14]$, pesticides^[15-17], chronic psychological stress^[18-20],

starvation^[21, 22], hyperthermia/ hypothermia^[23-25], and brain trauma^[26-28]. However, their intracellular and/or extracellular mechanisms of inducing LOAD are still controversial. Although previous studies have reported many mechanisms to interpret the pathological alterations in LOAD patients $[29-31]$, some intermediate components that link the systemic level with intracellular level are still lacking. Here we propose that dysfunction of the hypothalamic-pituitary adrenal (HPA) axis at the systemic level; cerebrovascular dysfunction, glial activation, metals malmetabolism, and extracellular Aβ toxicity at the intercellular level; and intracellular pathways, including kinase/phosphatase imbalance, epigenetic modifications, and oxidative stress, are the main mechanisms involved in the effects of environmental factors on the pathogenesis of LOAD.

Effects of Environmental Factors on LOAD Pathogenesis at the Systemic Level

Mammals generally respond to environmental stimuli at the whole-individual level through complicated mechanisms, such as the activation of HPA axis and autonomic system $[32, 33]$, to direct and control multiple reflex activities to deal with environmental pressures. At the systemic level, behavioral stress induced by external stimulation is the leading process associated with LOAD.

Kirby and co-workers demonstrated that acute stress increases the proliferation of hippocampal cells, neurogenesis, and the expression of astrocytic fibroblast growth factor 2 (FGF2)^[34]. A behavioral study focusing on the memory functions of the hippocampus has also shown that moderate acute stress enhances memory performance^[35]. In contrast to acute stress, chronic behavioral stress suppresses cell proliferation and differentiation of new neurons in the adult dentate gyrus^[36]. Unpredictable chronic mild stress preferentially reduces cell proliferation and neurogenesis in the ventral hippocampus, and the reductions can be reversed by fluoxetine^[37]. Cumulative evidence demonstrates that the release of stress hormones is responsible for the brain changes caused by chronic behavioral stress. Chronically stressed rats show enhanced corticosterone feedback on acute stress-induced HPA activity^[38] and the attenuated stress response of adrenocorticotropic hormone (ACTH) after chronic morphine treatment is associated with enhanced sensitivity to glucocorticoids^[39]. Kim *et al.* (2008) reported that corticotrophin-releasing factor (CRF) mRNA expression in the hypothalamus and ACTH levels in serum are significantly increased by chronic administration of corticosterone^[40]. Some investigators reported similarly increased CRF gene transcription in the paraventricular nucleus, consistent with other reports that chronic stress upregulates CRF mRNA expression $[41-43]$.

Effects of Environmental Factors on LOAD at the Intercellular Level

Abnormality of the cerebrovascular system, especially disruption of the BBB, is an important factor associated with the transport of environmental factors^[44]. Recent studies have shown co-morbidity between AD and cerebrovascular disease $^{[45]}$, and that AD is associated with atherosclerosis $^{[46, 47]}$, amyloid angiopathy^[48, 49], and cerebral microvascular pathology^[50, 51]. Extracelluar A β causes inflammation^[52, 53], apoptosis^[54, 55], oxidative stress^[56, 57], and excitotoxicity^[56, 58]. Metals are also associated with neuronal malfunction in AD, though the effects of iron, aluminum, zinc, and copper remain inconclusive $[12, 59]$. Most of these metals induce oxidative stress and increase $\text{A}\beta$ accumulation^[12, 13]. Surrounding glia may influence neuronal function by secreting inflammatory factors^[60-62], participating in the clearance of amyloid and exacerbating Aβ accumulation^[63-65].

External factors also influence neuronal microenvironment through multiple pathogenic mechanisms. Air pollution is one of the most common sources of environmental toxins. Since various air pollutants lead to ROS production, and as the human population exposed to air pollution increases, the risk of LOAD increases^[66]. In a related study, exposed urbanites displayed differential regulation of apolipoprotein E (APOE) genes and tau hyperphosphorylation with pre-tangle material, while 51% had diffuse Aβ plaques^[14]. Correspondently, air pollution accelerates Aβ42 accumulation^[67]. Traumatic brain injury (TBI) is recognized as one of the most detrimental environmental risk factors for the later development of cognitive impairment^[68, 69]. Repetitive neurotrauma can lead to the development of a progressive form of dementia reminiscent of early onset $LOAD^{[70, 71]}$. Aß plaques have also been identified in patients following a single TB $I^{[72, 73]}$, even in children[74]. Similarly, Uryu *et al*. (2002) reported

increased Aβ levels and urine isoprostanes in Tg2576 mice, together with induced cognitive impairments after mild TBI^[75]. Anesthesia is another risk factor associated with LOAD. It is linked to cognitive dysfunction and acceleration of senile dementia^[76, 77]. The commonly-used inhaled anesthetic sevoflurane induces caspase activation and apoptosis, and also increases the levels of β-site APP-cleaving enzyme (BACE), leading to elevated Aβ levels both in H4-APP cells and mice^[78]. Other anesthetics, such as isoflurane^[79], isoflurane with hypoxia^[80], desflurane with hypoxia^[81], and isoflurane with nitrous oxide $[82]$, also alter APP processing and subsequently increase Aβ production.

Effects of Environmental Factors on LOAD at the Intracellular Level

Nutrient deficiency plays a significant role in glucose metabolism dysfunction in LOAD. In a recent review^[58], we illustrated thiamine deficiency as an important contributor to the induction of dysfunctional glucose metabolism. Deficiency of folate and vitamin B_{12} also influences neuronal metabolism by inducing hyperhomocysteinemia and decreasing S-adenosylmethionine (SAM) levels. Under conditions of energy deficiency, the balance between kinases and phosphatases, protein aggregation and clearance, as well as oxidative stress and reduction capacity, also tends to be disturbed, which further leads to Aβ accumulation, tau hyperphosphorylation, and neuronal $death^{[58]}$.

Overall, exposure to environmental insults or stressors such as psychological stress^[83, 84], environmental toxins^[11], hypothermia^[24, 25], anesthesia^[84, 85], brain trauma and injury^[75, 86], heat^[87], starvation and glucose hypometabolism^[22, 58], can induce tau hyperphosphorylation, and Aβ aggregation and oligomerization, which are tightly linked to LOAD pathogenesis. Here we propose that multiple mechanisms at different levels mediate the pathological roles of environmental factors in LOAD. At the systemic level, HPA axis dysfunction is the main mechanism to induce a LOAD phenotype by behavioral stress^[88, 89]. At the intercellular level, cerebrovascular dysfunction, Aβ accumulation^[57, 90], glial activation^[91-93], and metal toxicity^[11-13] are generally responsible for an abnormal microenvironment surrounding neurons, and contribute to their dysfunction. Imbalance between the activities of tau-related kinases

and phosphatases induced by environmental stimuli or insults leads to the highly-phosphorylated form of tau protein in LOAD^[94, 95]. Furthermore, intracellular epigenetic modification through experience, especially DNA methylation and histone acetylation, also plays a vital role in LOAD^[96-98], as discussed in detail below. In all, complicated pathological networks at different levels are correlated with LOAD (Fig. 1).

Environmental Factors in the Development of LOAD

HPA Axis Dysfunction at the Systemic Level

Some studies have shown that behavioral stress promotes the pathological changes in animal models of LOAD. Lee *et al*. (2009) demonstrated that behavioral stress aggravates LOAD pathology through the generation of metabolic oxidative stress and down-regulation of MMP-2, which may be mediated by corticotropin-releasing hormone receptor (CRFR)[100]. Rissman *et al*. (2012) showed that CRFR1 and CRFR double-knockout mice do not show repeated stressinduced alterations in tau-P or solubility, indicating that CRF-induced tau phosphorylation is CRFR1-dependent^[19]. Consistently, CRF antagonism inhibits stress-induced Aβ deposition within the cortex of transgenic mice. Kang *et al*. (2007) demonstrated that behavioral stressors rapidly increase interstitial fluid Aβ through neuronal activity, which is mimicked by exogenous CRF administration but not corticosterone^[101]. In addition, stress-level glucocorticoid administration exacerbates Aβ accumulation by increasing the levels of APP and $β$ -APP-cleaving enzyme^[102]. Glucocorticoids also augment tau accumulation, indicating that this hormone accelerates the development of neurofibrillary tangles (NFTs)^[89]. Catecholamines may be also involved in AD phenotype, which is demonstrated by the fact that blockade of β-adrenergic receptors (β-ARs) prevents the release of IL-1β during acute stress, and stimulation of β-ARs induces the release of IL-1β and IL- 6 ^[103-105]. By injecting the β2-AR-selective agonist clenbuterol hydrochloride, acute stress-induced Aβ production is enhanced, while injecting the β2-AR-selective antagonist ICI 118,551 reduces Aβ production^[105]. All of this evidence suggests that stress hormones are directly related to pathological changes.

Tran *et al*. (2011) found that chronic psychological

Fig. 1. Environmental factors in the development and progression of late-onset Alzheimer's disease (LOAD). Multiple mechanisms at different levels mediate the pathological roles of environmental factors in LOAD. At the systemic level, HPA axis dysfunction is the main mechanism to induce AD phenotype[42, 88, 89]. At the intercellular level, cerebrovascular dysfunction, Aβ accumulation[57, 90], glial activation[91-93, 99], and metal toxicity[11-13] are responsible for creating an abnormal microenvironment around neurons. At the intracellular level, the imbalance between the activities of tau-related kinases and phosphatases[29, 94, 95], and abnormal epigenetic modification, especially DNA methylation and histone acetylation^[96-98], are associated with neuronal dysfunction in LOAD. ACTH, **adrenocorticotropic hormone; CRF, corticotrophin-releasing factor; HPA axis, hypothalamic-pituitary adrenal axis; ROS, reactive oxygen species; TBI, traumatic brain injury.**

stress leads to lower scores in the radial arm water maze, impairs long-term potentiation (LTP), enhances long-term depression, and increases total CaMKII and calcineurin in a rat AD model^[18, 106]. Other investigators also found that chronic stressors, like social isolation, are associated with abnormalities in LTP and dendritic branching, and decreased cued and contextual memory^[107]. Hippocampal neurogenesis is also impaired by isolation stress in APPsw (Tg2576) mutant mice^[108]. Other chronic stressors, including exposure to predator odor^[109], psychosocial stress^[110, 111] and restraint $[112, 113]$, as well as direct corticosterone administration^[114], all lead to impaired adult neurogenesis in the hippocampus. Even in neurologically normal individuals, excessive adverse stress causes cognitive impairment^[115]. Hence, there is no doubt that HPA axis dysfunction induces LOAD pathology and cognitive impairment (Fig. 2).

Environmental Factors at the Intercellular Level

Cerebrovascular dysfunction Recent findings indicate that neurovascular dysfunction contributes to cognitive decline and neurodegeneration in AD. To begin with, the BBB is responsible for the clearance of Aβ through receptors for advanced glycation end products (RAGE) and low-density lipoprotein receptor-related protein (LRP)^[44, 116]. Acting as a cell-surface receptor for $AB^{[117]}$, RAGE binds monomeric, oligomeric, and aggregated Aβ^[118, 119]. RAGE also mediates Aβ-induced neurotoxicity directly by inducing oxidative stress and indirectly by activating microglia^[117]. RAGE leads to mitochondrial dysfunction *via* intraneuronal Aβ transport^[120]. LRP also transports Aβ *via* the BBB^[121, 122]. Reduced LRP levels in brain microvessels are correlated with endogenous Aβ deposition in a chronic hydrocephalus model in rats^[123]. In addition, the formation of capillaries by brain capillary endothelial cells is greatly reduced in the Tg2576 mouse model of AD^[124], and high concentrations of the β-sheet form of Aβ are anti-angiogenic^[125]. Decreased length of brain capillaries in the CA1 region correlates well with increasing dementia rating scores^[51]. Zlokovic (2005) proposed that senescence of the neurovascular unit severely reduces the normal responses of neurovascular cells to physiological and pathophysiological stimuli, which in turn may disrupt normal BBB functions^[116]. Furthermore, glucose transporters (GLUTs) in the BBB are responsible for the transport of sufficient glucose to astrocytes and then neurons^[58]. Reduced GLUTs such as GLUT1 and GLUT3 have been found in AD^[126, 127]. Thus BBB dysfunction

Fig. 2. Hypothalamic-pituitary adrenal axis dysfunction mediates the induction of LOAD-like pathology through behavioral stress. Behavioral stress causes HPA abnormality by producing excessive corticotrophin-releasing factor (CRF), corticotropin, and catecholamines and their receptors, which further leads to LOAD pathology, such as Aβ accumulation and neuronal and synaptic loss. ACTH, adrenocorticotropic hormone; ACTHR, adrenocorticotropic hormone receptor; CRF, corticotrophin-releasing factor; CRFR1, corticotropin-releasing hormone receptor 1; GCRs, glucocorticoid receptors; NFTs, neurofibrillary tangles.

leads to an abnormal brain energy supply, and hence induces the regional hypometabolism revealed by imaging techniques^[58].

Glial activation Reactive astrocytes have been suggested to participate in the clearance and degradation of Aβ in LOAD. Actually, activated astrocytes located close to senile plaques accumulate in the brains of transgenic APP mice, and astrogliosis parallels the increase of the expression of neprilysin, one of the enzymes responsible for Aβ clearance^[128]. Thus it seems that astrocytes are activated to protect neurons from Aβ toxicity. However, it is also possible that the excessive inflammatory factors or cytokines secreted by astrocytes stimulated by Aβ lead to a malignant microenvironment that further impairs neuronal functions. Besides, astrocytes are responsible for the metabolism of glucose/lactate, glutamate/glutamine, and glutathione precursors/glutathione; when these mechanisms are impaired, neuronal functions are compromised $[129]$.

Apart from astrocyte activation, microglial activation has received much attention and is recognized as an early event in LOAD. Imaging targeted at microglial activation has been explored for early diagnosis of AD. Microglia clusters are located in Aβ deposits in the brains of LOAD patients and APP transgenic mice^[53]. In addition, cultured microglia secrete Aβ and catalyze APP in a manner that promotes A β deposition^[130]. Furthermore, many laboratories have shown that microglia, both *in vivo* and *in vitro*, phagocytose exogenous fibrillar $AB^{[114, 115]}$, indicating that microglia participate in Aβ clearance.

CD45 (also known as leukocyte common antigen) is a transmembrane protein tyrosine phosphatase and plays an essential role in modulating immune responses. CD45 is expressed on microglia in the frontal cortex and hippocampus of normal aging subjects, and markedly increases in the regions close to Aβ plaques in LOAD patients and transgenic animal models of LOAD^[131, 132]. Wilcock *et al*. (2001) showed that increased CD45 expression is positively correlated with reduced Congo red staining of compact plaques^[133]. Zhu et al. (2011) found elevation of both cerebral intracellular and extracellular soluble oligomeric Aβ, insoluble Aβ, and the microglial neurotoxic cytokines tumor necrosis factor-α and IL-1β in PSAPP/CD45^{-/-}mice compared with CD45-sufficient PSAPP littermates^[131]. This further indicates that reduced CD45 activity leads to the accumulation of neurotoxic Aβ oligomers and validates the CD45-mediated microglial clearance of oligomeric Aβ as a novel therapeutic target for $LOAD^{[132]}$.

Aβ deposition Previous evidence has shown that Aβ leads to oxidative stress^[57], mitochondrial dysfunction^[134, 135], excitotoxicity^[136, 137], and inflammation^[138, 139]. However, the role of Aβ seems to be a mystery in the field of LOAD pathophysiology, though many mechanisms have been proposed. Studies have shown that Aβ accumulation in mitochondria precedes extracellular amyloid deposition and increases with age in LOAD patients and animal models^[140, 141]. Soluble Aβ oligomers also cause neuronal damage or mitochondrial dysfunction by disturbing the functions of the respiratory chain^[142] and other mitochondrial components, including cyclophilin D^[143], Aβbinding alcohol dehydrogenase^[144], and TOMM40^[145]. Most previous studies indicated that Aβ is a negative component in the pathogenesis of LOAD. Nevertheless, some studies found that picomolar or low nanomolar levels of Aβ are neurotrophic or neuroprotective^[146] and physiological concentrations of Aβ efficiently increase hippocampal $LTP^{[147]}$. From our perspective of environmental stress, Aβ tends to be recognized as an abnormal response to environmental stress and also acts as an environmental insult that results in the abnormal interstitial changes surrounding the neurons, which may be also associated with glial abnormalities, including astrocytic and microglial activation. Besides, based on the above discussion, the entire pathogenic pathway induced by environmental stress may contribute to Aβ production and accumulation. Thus, Aβ toxicity would also be the common link among different mechanisms.

Metal toxicityIron (Fe) accumulation has been demonstrated in cells associated with neuritic plaques in LOAD^[148, 149]. Fe regulates α -secretase activity to influence APP cleavage^[150]. Besides, congenital Fe overload (hemochromatosis or HFE) has been linked to $LOAD^{[71]}$. The occurrence of HFE mutations in LOAD indicates that HFE mutation is also a risk factor for LOAD^[151, 152]. More importantly, HFE mutations are associated with increased oxidative stress and the progression of disease $[153]$. Furthermore, the transferrin subtype C2 increases in LOAD patients^[154]. The presence of transferrin subtype C2 may have additive effects on the risk of LOAD with APOE ε4 and HFE mutations^[154]. The combined data on transferrin subtype C2 and HFE mutation indicate that an Fe metabolism-related genotype may increase the risk of LOAD.

The involvement of aluminum (Al) neurotoxicity in LOAD neurodegeneration is supported by considerable evidence. Compared with normal controls, the Al level is elevated in necropsy and biopsy samples from LOAD brains confirmed by histopathology^[155]. Besides, the risk of LOAD is correlated with increased Al levels in drinking water $[156, 157]$. Evidence also supports that AI participates in the formation of NFTs and neuritic plaques $^{[158-160]}$. By intracisternal injection of Al phosphate into rabbits, Forbes

et al.(2002) showed that Al causes pathological lesions similar to NFTs^[161]. With the same method, Vasudevaraju *et* al. (2008) showed that AI causes apoptosis in neurons^[162]. Other laboratory work further showed that Al-induced lesions share biochemical similarities with NFTs in LOAD, especially the presence of tau protein^{$[163, 164]$}. The role of Al in LOAD pathogenesis remains to be explored; further experiments need to be designed.

A relationship between zinc (Zn) and LOAD has been suggested, and Zn seems to be a key component of amyloid plaques and the cerebral amyloid angiography observed in LOAD^[165]. First, Zn is responsible for the aggregation of Aβ. At pH 7.4, Aβ is rapidly aggregated by Zn-induced resistance to cleavage^[166]; this differs from the pH range for Fe- and copper (Cu)-induced Aβ aggregation and toxicity. More importantly, Zn participates in the formation of toxic small oligomer intermediates, associated with the condensation of Aβ oligomers on the neuronal surface^[167, 168]. Previous evidence appears to indicate that Zn causes synaptic disruption induced by Aβ accumulation. Deshpande *et al*. (2009) have demonstrated that Zn promotes the binding of Aβ to NR2B, an N-methyl-*D*-aspartate receptor subunit that is responsible for the induction of excitotoxicity^[169]. By sequestering Zn, Aβ seems to affect the metabotropic ZnR (GPR39) and TkB receptors, resulting in LTP impairment^[170]. Moreover, several Zndependent metalloproteinases, such as neprilysin, insulindegrading enzyme, and matrix metalloproteinases, degrade Aβ in the extracellular milieu, which may offer a possible interpretation of the inverse relationship between cerebrospinal fluid (CSF) Zn and Cu levels and CSF Aβ levels found in normal healthy people^[171]. Furthermore, Zn induces tau hyperphosphorylation in neuronal cell lines, and intracellular Zn increases in NFT-bearing neurons^[172].

It is well known that acute exposure to arsenic impairs brain functions^[173, 174]. Exposure to the heavy metal arsenic results in a 4-fold increase in tau phosphorylation at many of the sites that are hyperphosphorylated in paired helical filament tau^[175]. Further, tau is a major substrate for the enzymatic activities affected by arsenic. The arseniccontaining compound phenylarsine oxide induces tau phosphorylation within its microtubule-binding domain *in situ* by a staurosporine-sensitive protein kinase in cultured cells $[176]$.

Apart from the above-mentioned metals, Cu is also thought to be involved in LOAD, but it is still unknown whether deficiency or overload occurs. Some studies have shown that Cu deficiency is associated with LOAD, and excessive dietary Cu on a high-cholesterol diet in rabbits and in a mouse model of LOAD induces LOADlike pathology^[177]. Another study also indicated that chronic Cu exposure contributes to LOAD in humans $[178]$. There is strong evidence that low-dose lead (Pb) exposureive is causally associated with deficits in cognition $[179]$. Coincidentally, the highest levels of Pb in the brain have been found in the hippocampus and cerebral cortex, areas associated with learning and memory $[180]$. Recently, Pbinduced impairment of learning and memory has been attributed to over-expression or over-activation of serine/ threonine protein phosphatases, suggesting a novel mechanism of Pb neurotoxicity^[181]. Further studies are needed to explore the role of metal malmetabolism in AD pathogenesis.

Kinase–Phosphatase Imbalance and Epigenetic **Modification at the Intracellular Level**

Phosphatase hypoactivity Accumulating studies suggest that an imbalance of kinase-phosphatase is intimately associated with the pathophysiology of LOAD. Previous studies have shown that the brain protein phosphatases (PPs) PP1, PP2A, PP2B, and PP5 dephosphorylate the hyperphosphorylated tau *in vitro*, but PP2C does not dephosphorylate at any of the sites studied^[182-184]. Hyperphosphorylated tau in LOAD brain is mainly dephosphorylated by PP2A and PP2B and at a lesser extent by PP1. PP2A is the main phosphatase that regulates tau phosphorylation, accounting for 71% of the total tau activity in the human brain, with PP1, PP5, and PP2B accounting for 11%, 10%, and 7%, respectively^[184]. The PP2A and PP5 activity and expression are downregulated in the LOAD brain and this results in aberrant phosphorylation of tau and neurofilament proteins^[185, 186]. The enzymatic activity levels of PP2A and PP5 are reduced by 50% and 20%, respectively, in the LOAD brain^{$[187, 188]$}. A minor reduction in the PP2A methylation rate is related to tau hyperphosphorylation and increased Aβ production^[189]. In the LOAD brain, the neocortical levels of the endogenous inhibitors of PP2A, I₁PP2A and I₂PP2A, are significantly increased^[190]. In addition, transgenic mice expressing the negative form of the PP2A catalytic subunit show increased tau phosphorylation^[191]. These findings imply that a downregulation of tau phosphatases in the LOAD brain

might underlie the abnormal hyperphosphorylation of tau and other neuronal proteins.

Kinase hyperactivity Recent *in vitro* studies have shown that several protein kinases are involved in tau phosphorylation at specific sites^[192]. These include prolinedirected kinases such as glycogen synthase kinase-3 $(GSK-3)^{[193]}$, cycle-independent kinase $5^{[192]}$, mitogenactivated protein kinase/extracellular signal regulatedkinases (MAPK/ERK)^[194], AMP-activated protein kinase^[195], stress-activated protein kinase/c-Jun N-terminal kinase $(SAPK/JNK)^{[196]}$, p-38 kinase $(p38)^{[197]}$, microtubule affinityregulating kinases (MARKs)^[198], cAMP-dependent protein kinase (PKA)^[182], Ca²⁺/calmodulin-dependent protein kinase II (CaMKII)^[199], tyrosine-specific kinases, and Src family kinases such as c-Abl kinase^[200] and lck ^[201]. In addition, JNK kinase 1, an upstream activator of JNK/SAPK^[202], and mitogen-activated kinase kinase 6, an upstream activator of p38^[203], are also activated in LOAD. Based on the literature, GSK-3β, also known as tau protein kinase 1, seems to be the most important enzyme in determining tau phosphorylation status^[184].

Tau hyperphosphorylation The hyperphosphorylation of tau is one of the key pathogenic features of LOAD. It has been hypothesized that it is induced by the overactivation of kinase or the inactivation of phosphatase. Thus kinases such as MARKs, PKA, CaMKII, and checkpoint kinase 2, which phosphorylate tau on Ser²⁶² in *vitro*[204], or GSK-3β[205, 206], CDK5[207, 208], and other kinases are all responsible for tau hyperphosphorylation. Abnormal interactions between kinase and phosphatase produce an imbalance of phosphorylation and dephosphorylation, which could be the fundamental mechanism for abnormal tau hyperphosphorylation (Fig. 3). Some studies have shown that the disruption of the balance between kinase and phosphatase induces tau hyperphosphorylation. In the study by Leclerc *et al*. (2001), it was proposed that GSK-3β and CDK5 are responsible for most of the abnormal hyperphosphorylation in LOAD, and indirubins from a traditional Chinese herbal formula (Danggui Longhui Wan) inhibit GSK-3β and CDK5/P25 and hence reduce abnormal tau phosphorylation^[205]. When the brain is exposed to acidosis, asparaginyl endopeptidase is released from lysosomes into the cytoplasm, which inhibits PP2A and promotes tau hyperphosphorylation, indicating that acidosis triggers tau hyperphosphorylation^[209]. Calcyclin-binding

Fig. 3. Imbalance between tau-related kinases and phosphatases causes tau hyperphosphorylation. Hyperphosphorylation of tau is induced by the over-activation of kinases such as GSK-3β and MARK, or the inactivation of phosphatases like PP2A, which further promotes the formation of neurofi brillary tangles.

protein and Siah-1-interacting protein (CacyBP/SIP) play a role in the organization of microtubule^[210]. Wasik *et al.* (2013) showed that in LOAD patients, CacyBP/SIP is almost exclusively present in neuronal somata, while it occurs in the somata and processes in control patients. Besides, they found a similar pathogenic distribution of CacyBP/SIP in tau transgenic mice, but not in APP/PS1 mice^[210]. Further research is needed to understand the role of abnormal kinases and phosphatases in the development of LOAD.

Epigenetic Modifications Induced by Environmental *Stress in LOAD*

DNA methylation Because of the diversity of individual genetic backgrounds and life experiences, factors late in life may influence the susceptibility and progression of LOAD through epigenetic modification by DNA methylation or histone acetylation^[96, 211, 212]. Based on previous data, it is quite possible that abnormal DNA methylation is induced by deficiency of vitamins B_6 , B_{12} , and folic acid, and this may be involved in gene transcription associated

with AD pathology. In the review by Marques *et al*. (2011) it was hypothesized that nutrient deficiency leads to hyperhomocysteinemia and further induces a consequent decrease in the levels of SAM, a key donor of methyl groups in DNA methylation^[211]. Thus, low SAM levels could demethylate DNA and result in the activation and overexpression of genes involved in LOAD pathology. A study has shown that the deficiency of folate and vitamin B_{12} in culture medium decreases the SAM levels, upregulates PSEN1 and BACE, and leads to more Aβ production^[213]. PSEN1, APOE, methylenetetrahydrofolate reductase, and DNA methyltransferases (DNMT1), which are all responsible for methylation homeostasis, show significant inter-individual epigenetic variability in the brain and lymphocytes of LOAD patients^[182, 214]. One finding suggested that APP promoter hypomethylation could be a risk factor for LOAD. Particularly, Aβ40 accumulation induces abnormal methylation patterns such as global hypomethylation and hypermethylation of specific loci, including the promoter of neprilysin^[215]. Infantile exposure to Pb causes increased BACE1 and Sp1 transcription, and their levels parallel the DNMT1 activity, indicating that the toxicity of Pb may be mediated by DNMT1 by increasing Aβ levels through hypomethylation of BACE1 or Sp1 216 . In the TGCRND8 model, Sp1 is also regulated by its methylation pattern in response to metabolic stimuli, such as vitamin B deficiency and SAM administration $[214]$. Oxidative stress may cause DNA damage and related deficiencies in Ogg1, which is thought to be a significant factor in the process of aging-related diseases such as LOAD^[96].

Histone modification Recent studies have shown the association of histone acetylation with the AD pathology, and this seems to be a new field to be explored. One of the most exciting studies to date was led by Li-Huei Tsai and André Fischer. They reported that a drug that promotes histone acetylation improves learning and memory in a mouse model of AD^[217]. Besides, specific overexpression of histone deacetylase 2 (HDAC2) in mouse neurons, reduces dendritic spine density, synapse number, synaptic plasticity, and memory formation, but not of HDAC1 $[217]$. On the contrary, HDAC2 knockout results in elevated synapse number and memory facilitation. Correspondently, oral administration of an HDAC inhibitor (HDACi)^[218] to the $3 \times Tg$ -LOAD mouse model clearly prevents cognitive deficits and reduces tau hyperphosphorylation^[219]. Consistent with this study, treatment with the HDACi sodium 4-phenylbutyrate for 5 weeks also reverses the memory impairment and improves spatial learning in a transgenic mouse model (Tg2576) of LOAD^[220]. Environmental enrichment increases histone acetylation and furthermore enables the rescue of impaired learning and memory in another mouse model of $\mathsf{LOAD}^{[217]}$. In a similar study, the HDACi sodium butyrate helps with the maintenance and recovery of long-term memory in the same model^[217, 221]. Trichostatin A, another HDACi, also rescues the memory defects and hippocampal synaptic dysfunction in APP/PS1 mice, as well as reducing tau hyperphosphorylation, but does not disrupt the interaction between HDAC6 and tau $^{[222]}$. By quantification with targeted proteomics, Zhang *et al*. (2012) showed that histone acetylation is significantly lower in the LOAD temporal lobe than in aged controls^[223]. Overall, the role of histone acetylation is still an area to be explored, and further studies are needed.

New Perspectives on the Management of LOAD

As described in our review of glucose metabolism in LOAD^[58], previous studies seem to have led to a frustrating impasse in LOAD research. Considering that most studies have long been focused on the two pathological hallmarks, their related mechanisms and consequences, we argue that it is time to understand the disease from an ecological perspective, using the concept of environmental stress. In this review, we admit that the typical pathological changes play a significant role in understanding LOAD; however, we tend to focus on the precedents of the pathology, and emphasize environmental risk factors and their pathological pathways in LOAD. This new orientation changes the concepts underlying LOAD management, such that early prevention of environmental risk factors and blocking the intermediate pathological pathways induced by environmental stress are the most attractive treatment. Based on this concept, we argue that avoidance or reduction of exposure to environmental factors is the first step in LOAD prevention and treatment. Metals, nutrients, air pollution, pesticides, and chronic psychological stress can all be controlled by individual or social measures. Though there is no confirmed evidence to support the sufficiency of AD prevetion from environmental factors, this seems to be a practical and effective mean of reducing

the occurrence or initiation of LOAD. Besides, blocking the intermediate pathological pathways based on the mechanisms proposed here is a potential approach to prevent the initiation and progression of the disease. By modulating the HPA axis, repairing the cerebrovascular system, balancing the activities of phosphatases and kinases, reducing metal exposure, regulating DNA methylation or histone acetylation, inhibiting inflammatory factors, and antagonizing Aβ toxicity, the initial pathological cascades may be blocked and LOAD would not occur. However, further studies are required to test the efficacy of these new mechanisms as LOAD research progresses.

Conclusions

Most previous studies have focused on the mechanisms and consequences of the internal pathological changes occurring in neurons in LOAD. In this review, we explore a new field based on the concept of environmental stress. We conclude that environmental stress may produce a microenvironment that induces neuronal dysfunction in LOAD *via* multiple pathological mechanisms, including HPA axis dysfunction, cerebrovascular dysfunction, imbalance of kinases and phosphatases, metal toxicity, epigenetic modification, glial activation, and Aβ toxicity. This perspective of environmental stress may shed new light on understanding the etiology, pathophysiology, prevention, and treatment of LOAD.

ACKNOWLEDGEMENTS

This review was supported by National Basic Research Development Program (973 Program) of China (2011CBA00400), the National Natural Science Foundation of China (91332201), the Natural Science Foundation of Shanghai Municipality, China (13JC1401500) and the Fund for Medical Emerging Cutting-edge Technology of Shanghai Municipality, China (SHDC12012114).

Received date: 2013-11-25; Accepted date: 2014-01-23

REFERENCES

- [1] Kagias K, Nehammer C, Pocock R. Neuronal responses to physiological stress. Front Genet 2012, 3: 222.
- [2] Stanton ML, Roy BA, Thiede DA. Evolution in stressful environments. I. Phenotypic variability, phenotypic selection, and response to selection in five distinct environmental

stresses. Evolution 2000, 54: 93–111.

- [3] Anderegg WR, Berry JA, Field CB. Linking definitions, mechanisms, and modeling of drought-induced tree death. Trends Plant Sci 2012, 17: 693–700.
- [4] Han HW, Ohn JH, Moon J, Kim JH. Yin and Yang of disease genes and death genes between reciprocally scale-free biological networks. Nucleic Acids Res 2013, 41: 9209–9217.
- [5] Fields RD, Araque A, Johansen-Berg H, Lim SS, Lynch G, Nave KA*, et al.* Glial biology in learning and cognition. Neuroscientist 2013. doi: 10.1177/1073858413504465.
- [6] Pirttimaki TM, Parri HR. Astrocyte plasticity: implications for synaptic and neuronal activity. Neuroscientist 2013, 19: 604–615.
- [7] Kagias K, Nehammer C, Pocock R. Neuronal responses to physiological stress. Front Genet 2012, 3: 222.
- [8] Caldji C, Hellstrom IC, Zhang TY, Diorio J, Meaney MJ. Environmental regulation of the neural epigenome. FEBS Lett 2011, 585: 2049–2058.
- [9] Migliore L, Coppede F. Genetics, environmental factors and the emerging role of epigenetics in neurodegenerative diseases. Mutat Res 2009, 667: 82–97.
- [10] Debacq-Chainiaux F, Leduc C, Verbeke A, Toussaint O. UV, stress and aging. Dermatoendocrinol 2012, 4: 236–240.
- [11] Moulton PV, Yang W. Air pollution, oxidative stress, and Alzheimer's disease. J Environ Public Health 2012, 2012: 472751.
- [12] Ayton S, Lei P, Bush AI. Metallostasis in Alzheimer's disease. Free Radic Biol Med 2013, 62: 76–89.
- [13] Bush AI. The metal theory of Alzheimer's disease. J Alzheimers Dis 2013, 33 Suppl 1: S277–281.
- [14] Calderon-Garciduenas L, Kavanaugh M, Block M, D'Angiulli A, Delgado-Chavez R, Torres-Jardon R*, et al.* Neuroinflammation, hyperphosphorylated tau, diffuse amyloid plaques, and down-regulation of the cellular prion protein in air pollution exposed children and young adults. J Alzheimers Dis 2012, 28: 93–107.
- [15] Cannas A, Costa B, Tacconi P, Pinna L, Fiaschi A. Dementia of Alzheimer type (DAT) in a man chronically exposed to pesticides. Acta Neurol (Napoli) 1992, 14: 220–223.
- [16] Baldi I, Lebailly P, Mohammed-Brahim B, Letenneur L, Dartigues JF, Brochard P. Neurodegenerative diseases and exposure to pesticides in the elderly. Am J Epidemiol 2003, 157: 409–414.
- [17] Thany SH, Reynier P, Lenaers G. Neurotoxicity of pesticides: its relationship with neurodegenerative diseases. Med Sci (Paris) 2013, 29: 273–278.
- [18] Alkadhi KA, Alzoubi KH, Srivareerat M, Tran TT. Chronic psychosocial stress exacerbates impairment of synaptic plasticity in beta-amyloid rat model of Alzheimer's disease: prevention by nicotine. Curr Alzheimer Res 2011, 8: 718–731.
- [19] Rissman RA, Staup MA, Lee AR, Justice NJ, Rice KC, Vale W*, et al.* Corticotropin-releasing factor receptor-dependent effects of repeated stress on tau phosphorylation, solubility, and aggregation. Proc Natl Acad Sci U S A 2012, 109: 6277– 6282.
- [20] Sierksma AS, Prickaerts J, Chouliaras L, Rostamian S, Delbroek L, Rutten BP*, et al.* Behavioral and neurobiological effects of prenatal stress exposure in male and female APPswe/PS1dE9 mice. Neurobiol Aging 2013, 34: 319–337.
- [21] Ma JF, Wang HM, Li QY, Zhang Y, Pan J, Qiang Q*, et al.* Starvation triggers Abeta42 generation from human umbilical vascular endothelial cells. FEBS Lett 2010, 584: 3101–3106.
- [22] Yanagisawa M, Planel E, Ishiguro K, Fujita SC. Starvation induces tau hyperphosphorylation in mouse brain: implications for Alzheimer's disease. FEBS Lett 1999, 461: 329–333.
- [23] Planel E, Miyasaka T, Launey T, Chui D-H, Tanemura K, Sato S*, et al.* Alterations in glucose metabolism induce hypothermia leading to tau hyperphosphorylation through differential inhibition of kinase and phosphatase activities: Implications for Alzheimer's disease. J Neurosci 2004, 24: 2401–2411.
- [24] Whittington RA, Papon MA, Chouinard F, Planel E. Hypothermia and Alzheimer's disease neuropathogenic pathways. Curr Alzheimer Res 2010, 7: 717–725.
- [25] Bretteville A, Marcouiller F, Julien C, El Khoury NB, Petry FR, Poitras I*, et al.* Hypothermia-induced hyperphosphorylation: a new model to study tau kinase inhibitors. Sci Rep 2012, 2: 480.
- [26] Sivanandam TM, Thakur MK. Traumatic brain injury: a risk factor for Alzheimer's disease. Neurosci Biobehav Rev 2012, 36: 1376–1381.
- [27] Fakhran S, Yaeger K, Alhilali L. Symptomatic white matter changes in mild traumatic brain injury resemble pathologic features of early Alzheimer dementia. Radiology 2013, 269: 249–257.
- [28] Washington PM, Morffy N, Parsadanian M, Zapple D, Burns MP. Experimental traumatic brain injury induces rapid aggregation and oligomerization of amyloid-beta in an Alzheimer's disease mouse model. J Neurotrauma 2013, 31(1):125–134.
- [29] Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wisniewski HM, Binder LI. Abnormal phosphorylation of the microtubuleassociated protein tau (tau) in Alzheimer cytoskeletal pathology. Proc Natl Acad Sci U S A 1986, 83: 4913–4917.
- [30] Barton AJ, Harrison PJ, Najlerahim A, Heffernan J, McDonald B, Robinson JR*, et al.* Increased tau messenger RNA in Alzheimer's disease hippocampus. Am J Pathol 1990, 137: 497–502.
- [31] Hardy JA, Higgins GA. Alzheimer's disease: the amyloid

cascade hypothesis. Science 1992, 256: 184–185.

- [32] Young EA, Abelson JL, Cameron OG. Interaction of brain noradrenergic system and the hypothalamic-pituitary-adrenal (HPA) axis in man. Psychoneuroendocrinology 2005, 30: 807–814.
- [33] Rabasa C, Munoz-Abellan C, Daviu N, Nadal R, Armario A. Repeated exposure to immobilization or two different footshock intensities reveals differential adaptation of the hypothalamic-pituitary-adrenal axis. Physiol Behav 2011, 103: 125–133.
- [34] Kirby ED, Muroy SE, Sun WG, Covarrubias D, Leong MJ, Barchas LA*, et al.* Acute stress enhances adult rat hippocampal neurogenesis and activation of newborn neurons via secreted astrocytic FGF2. Elife 2013, 2: e00362.
- [35] Conrad CD, Lupien SJ, McEwen BS. Support for a bimodal role for type II adrenal steroid receptors in spatial memory. Neurobiol Learn Mem 1999, 72: 39–46.
- [36] Wong EY, Herbert J. The corticoid environment: a determining factor for neural progenitors' survival in the adult hippocampus. Eur J Neurosci 2004, 20: 2491–2498.
- [37] Tanti A, Rainer Q, Minier F, Surget A, Belzung C. Differential environmental regulation of neurogenesis along the septotemporal axis of the hippocampus. Neuropharmacology 2012, 63: 374–384.
- [38] Mizoguchi K, Yuzurihara M, Ishige A, Sasaki H, Chui DH, Tabira T. Chronic stress differentially regulates glucocorticoid negative feedback response in rats. Psychoneuroendocrinology 2001, 26: 443–459.
- [39] Houshyar H, Galigniana MD, Pratt WB, Woods JH. Differential responsivity of the hypothalamic-pituitary-adrenal axis to glucocorticoid negative-feedback and corticotropin releasing hormone in rats undergoing morphine withdrawal: possible mechanisms involved in facilitated and attenuated stress responses. J Neuroendocrinol 2001, 13: 875–886.
- [40] Kim HG, Lim EY, Jung WR, Shin MK, Ann ES, Kim KL. Effects of treadmill exercise on hypoactivity of the hypothalamo-pituitary-adrenal axis induced by chronic administration of corticosterone in rats. Neurosci Lett 2008, 434: 46–49.
- [41] Imaki T, Nahan JL, Rivier C, Sawchenko PE, Vale W. Differential regulation of corticotropin-releasing factor mRNA in rat brain regions by glucocorticoids and stress. J Neurosci 1991, 11: 585–599.
- [42] Herman JP, Adams D, Prewitt C. Regulatory changes in neuroendocrine stress-integrative circuitry produced by a variable stress paradigm. Neuroendocrinology 1995, 61: 180–190.
- [43] Makino S, Smith MA, Gold PW. Increased expression of corticotropin-releasing hormone and vasopressin messenger ribonucleic acid (mRNA) in the hypothalamic paraventricular

nucleus during repeated stress: association with reduction in glucocorticoid receptor mRNA levels. Endocrinology 1995, 136: 3299–3309.

- [44] Sagare AP, Bell RD, Zlokovic BV. Neurovascular dysfunction and faulty amyloid beta-peptide clearance in Alzheimer disease. Cold Spring Harb Perspect Med 2012, 2.
- [45] Gorelick PB. Risk factors for vascular dementia and Alzheimer disease. Stroke 2004, 35: 2620–2622.
- [46] Casserly I, Topol E. Convergence of atherosclerosis and Alzheimer's disease: inflammation, cholesterol, and misfolded proteins. Lancet 2004, 363: 1139–1146.
- [47] Roher AE, Esh C, Rahman A, Kokjohn TA, Beach TG. Atherosclerosis of cerebral arteries in Alzheimer disease. Stroke 2004, 35: 2623–2627.
- [48] Greenberg SM, Gurol ME, Rosand J, Smith EE. Amyloid angiopathy-related vascular cognitive impairment. Stroke 2004, 35: 2616–2619.
- [49] Vinters HV, Farag ES. Amyloidosis of cerebral arteries. Adv Neurol 2003, 92: 105–112.
- [50] Farkas E, Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. Prog Neurobiol 2001, 64: 575–611.
- [51] Bailey TL, Rivara CB, Rocher AB, Hof PR. The nature and effects of cortical microvascular pathology in aging and Alzheimer's disease. Neurol Res 2004, 26: 573–578.
- [52] Capiralla H, Vingtdeux V, Zhao H, Sankowski R, Al-Abed Y, Davies P*, et al.* Resveratrol mitigates lipopolysaccharideand Abeta-mediated microglial inflammation by inhibiting the TLR4/NF-kappaB/STAT signaling cascade. J Neurochem 2012, 120: 461–472.
- [53] McGeer PL, McGeer EG. The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy. Acta Neuropathol 2013, 126: 479–497.
- [54] Sanphui P, Biswas SC. FoxO3a is activated and executes neuron death via Bim in response to [beta]-amyloid. Cell Death Dis 2013, 4: e625.
- [55] Woo JA, Jung AR, Lakshmana MK, Bedrossian A, Lim Y, Bu JH, et al. Pivotal role of the RanBP9-cofilin pathway in Abetainduced apoptosis and neurodegeneration. Cell Death Differ 2012, 19: 1413–1423.
- [56] Reddy PH. Amyloid precursor protein-mediated free radicals and oxidative damage: implications for the development and progression of Alzheimer's disease. J Neurochem 2006, 96: $1 - 13$.
- [57] De Felice FG, Velasco PT, Lambert MP, Viola K, Fernandez SJ, Ferreira ST*, et al.* Aβ Oligomers Induce Neuronal Oxidative Stress through an N-Methyl-D-aspartate Receptordependent Mechanism That Is Blocked by the Alzheimer Drug Memantine. J Biol Chem 2007, 282: 11590–11601.
- [58] Chen Z, Zhong C. Decoding Alzheimer's disease from

perturbed cerebral glucose metabolism: implications for diagnostic and therapeutic strategies. Prog Neurobiol 2013, 108: 21–43.

- [59] Eskici G, Axelsen PH. Copper and oxidative stress in the pathogenesis of Alzheimer's disease. Biochemistry 2012, 51: 6289–6311.
- [60] Lopategui Cabezas I, Herrera Batista A, Penton Rol G. The role of glial cells in Alzheimer's disease: Potential therapeutic implications. Neurologia 2012. doi: 10.1016/j.nrl.2012.10.006
- [61] Cunningham C. Microglia and neurodegeneration: the role of systemic inflammation. Glia 2013, 61: 71-90.
- [62] Skaper SD, Facci L, Giusti P. Mast cells, glia and neuroinflammation: partners in crime? Immunology 2013. doi: 10.1111/imm.12170
- [63] Paresce DM, Chung H, Maxfield FR. Slow degradation of aggregates of the Alzheimer's disease amyloid beta-protein by microglial cells. J Biol Chem 1997, 272: 29390–29397.
- [64] Frautschy SA, Yang F, Irrizarry M, Hyman B, Saido TC, Hsiao K*, et al.* Microglial response to amyloid plaques in APPsw transgenic mice. Am J Pathol 1998, 152: 307–317.
- [65] Wyss-Coray T, Lin C, Yan F, Yu G-Q, Rohde M, McConlogue L*, et al.* TGF-[beta]1 promotes microglial amyloid-[beta] clearance and reduces plaque burden in transgenic mice. Nat Med 2001, 7: 612–618.
- [66] Block ML, Calderon-Garciduenas L. Air pollution: mechanisms of neuroinflammation and CNS disease. Trends Neurosci 2009, 32: 506–516.
- [67] Campbell A. Inflammation, neurodegenerative diseases, and environmental exposures. Ann N Y Acad Sci 2004, 1035: 117–132.
- [68] Heyman A, Wilkinson WE, Stafford JA, Helms MJ, Sigmon AH, Weinberg T. Alzheimer's disease: a study of epidemiological aspects. Ann Neurol 1984, 15: 335–341.
- [69] Guo Z, Cupples LA, Kurz A, Auerbach SH, Volicer L, Chui H*, et al.* Head injury and the risk of AD in the MIRAGE study. Neurology 2000, 54: 1316–1323.
- [70] McKee AC, Cantu RC, Nowinski CJ, Hedley-Whyte ET, Gavett BE, Budson AE*, et al.* Chronic traumatic encephalopathy in athletes: progressive tauopathy after repetitive head injury. J Neuropathol Exp Neurol 2009, 68: 709–735.
- [71] Lehmann DJ, Worwood M, Ellis R, Wimhurst VL, Merryweather-Clarke AT, Warden DR*, et al.* Iron genes, iron load and risk of Alzheimer's disease. J Med Genet 2006, 43: e52.
- [72] Roberts GW, Gentleman SM, Lynch A, Murray L, Landon M, Graham DI. Beta amyloid protein deposition in the brain after severe head injury: implications for the pathogenesis of Alzheimer's disease. J Neurol Neurosurg Psychiatry 1994, 57: 419–425.
- [73] Adle-Biassette H, Duyckaerts C, Wasowicz M, He Y, Fornes P, Foncin JF*, et al.* Beta AP deposition and head trauma. Neurobiol Aging 1996, 17: 415–419.
- [74] Ikonomovic MD, Uryu K, Abrahamson EE, Ciallella JR, Trojanowski JQ, Lee VM*, et al.* Alzheimer's pathology in human temporal cortex surgically excised after severe brain injury. Exp Neurol 2004, 190: 192–203.
- [75] Uryu K, Laurer H, McIntosh T, Pratico D, Martinez D, Leight S*, et al.* Repetitive mild brain trauma accelerates Abeta deposition, lipid peroxidation, and cognitive impairment in a transgenic mouse model of Alzheimer amyloidosis. J Neurosci 2002, 22: 446–454.
- [76] Ritchie K, Polge C, de Roquefeuil G, Djakovic M, Ledesert B. Impact of anesthesia on the cognitive functioning of the elderly. Int Psychogeriatr 1997, 9: 309–326.
- [77] Ancelin ML, de Roquefeuil G, Scali J, Bonnel F, Adam JF, Cheminal JC*, et al.* Long-term post-operative cognitive decline in the elderly: the effects of anesthesia type, apolipoprotein E genotype, and clinical antecedents. J Alzheimers Dis 2010, 22 Suppl 3: 105–113.
- [78] Dong Y, Zhang G, Zhang B, Moir RD, Xia W, Marcantonio ER*, et al.* The common inhalational anesthetic sevoflurane induces apoptosis and increases beta-amyloid protein levels. Arch Neurol 2009, 66: 620–631.
- [79] Xie H, Guan J, Borrelli LA, Xu J, Serrano-Pozo A, Bacskai BJ. Mitochondrial alterations near amyloid plaques in an Alzheimer's disease mouse model. J Neurosci 2013, 33: 17042–17051.
- [80] Pan C, Xu Z, Dong Y, Zhang Y, Zhang J, McAuliffe S*, et al.* The potential dual effects of anesthetic isoflurane on hypoxiainduced caspase-3 activation and increases in beta-site amyloid precursor protein-cleaving enzyme levels. Anesth Analg 2011, 113: 145–152.
- [81] Zhang L, Zhang Y. Halothane increases neuronal cell death vulnerability by downregulating miR-214 and upregulating Bax. Int J Clin Exp Med 2013, 6: 452–460.
- [82] Zhen Y, Dong Y, Wu X, Xu Z, Lu Y, Zhang Y*, et al.* Nitrous oxide plus isoflurane induces apoptosis and increases betaamyloid protein levels. Anesthesiology 2009, 111: 741–752.
- [83] Alkadhi KA. Chronic psychosocial stress exposes Alzheimer's disease phenotype in a novel at-risk model. Front Biosci (Elite Ed) 2012, 4: 214–229.
- [84] Bohnen N, Warner MA, Kokmen E, Kurland LT. Early and midlife exposure to anesthesia and age of onset of Alzheimer's disease. Int J Neurosci 1994, 77: 181–185.
- [85] Andre D, Dartigues JF, Sztark F. [Alzheimer's disease and anaesthesia: potential relationships and clinical implications]. Ann Fr Anesth Reanim 2011, 30: 37–46.
- [86] Lye TC, Shores EA. Traumatic brain injury as a risk factor for Alzheimer's disease: a review. Neuropsychol Rev 2000, 10:

115–129.

- [87] Sinigaglia-Coimbra R, Cavalheiro EA, Coimbra CG. Postischemic hyperthermia induces Alzheimer-like pathology in the rat brain. Acta Neuropathol 2002, 103: 444–452.
- [88] Ostrander MM, Ulrich-Lai YM, Choi DC, Richtand NM, Herman JP. Hypoactivity of the hypothalamo-pituitaryadrenocortical axis during recovery from chronic variable stress. Endocrinology 2006, 147: 2008–2017.
- [89] Green KN, Billings LM, Roozendaal B, McGaugh JL, LaFerla FM. Glucocorticoids increase amyloid-beta and tau pathology in a mouse model of Alzheimer's disease. J Neurosci 2006, 26: 9047–9056.
- [90] Shinkai Y, Yoshimura M, Morishima-Kawashima M, Ito Y, Shimada H, Yanagisawa K*, et al.* Amyloid beta-protein deposition in the leptomeninges and cerebral cortex. Ann Neurol 1997, 42: 899–908.
- [91] Cornejo F, von Bernhardi R. Role of scavenger receptors in glia-mediated neuroinflammatory response associated with Alzheimer's disease. Mediators Inflamm 2013, 2013: 895651.
- [92] Morales I, Jimenez JM, Mancilla M, Maccioni RB. Tau oligomers and fibrils induce activation of microglial cells. J Alzheimers Dis 2013, 37: 849–856.
- [93] Ogundele OM, Omoaghe AO, Ajonijebu DC, Ojo AA, Fabiyi TD, Olajide OJ*, et al.* Glia activation and its role in oxidative stress. Metab Brain Dis 2013.
- [94] Alonso AC, Zaidi T, Grundke-Iqbal I, Iqbal K. Role of abnormally phosphorylated tau in the breakdown of microtubules in Alzheimer disease. Proc Natl Acad Sci U S A 1994, 91: 5562–5566.
- [95] Gendreau KL, Hall GF. Tangles, toxicity, and tau secretion in AD - new approaches to a vexing problem. Front Neurol 2013, 4: 160.
- [96] Chouliaras L, Rutten BP, Kenis G, Peerbooms O, Visser PJ, Verhey F*, et al.* Epigenetic regulation in the pathophysiology of Alzheimer's disease. Prog Neurobiol 2010, 90: 498–510.
- [97] Kwok JB. Role of epigenetics in Alzheimer's and Parkinson's disease. Epigenomics 2010, 2: 671–682.
- [98] Babenko O, Kovalchuk I, Metz GA. Epigenetic programming of neurodegenerative diseases by an adverse environment. Brain Res 2012, 1444: 96–111.
- [99] Lee DC, Rizer J, Hunt JB, Selenica ML, Gordon MN, Morgan D. Review: experimental manipulations of microglia in mouse models of Alzheimer's pathology: activation reduces amyloid but hastens tau pathology. Neuropathol Appl Neurobiol 2013, 39: 69–85.
- [100] Lee KW, Kim JB, Seo JS, Kim TK, Im JY, Baek IS*, et al.* Behavioral stress accelerates plaque pathogenesis in the brain of Tg2576 mice via generation of metabolic oxidative stress. J Neurochem 2009, 108: 165–175.

[101] Kang JE, Cirrito JR, Dong H, Csernansky JG, Holtzman

DM. Acute stress increases interstitial fluid amyloid-beta via corticotropin-releasing factor and neuronal activity. Proc Natl Acad Sci U S A 2007, 104: 10673–10678.

- [102] Green KN, Billings LM, Roozendaal B, McGaugh JL, LaFerla FM. Glucocorticoids increase amyloid-β and tau pathology in a mouse model of Alzheimer's disease. J Neurosci 2006, 26: 9047–9056.
- [103] Bierhaus A, Wolf J, Andrassy M, Rohleder N, Humpert PM, Petrov D*, et al.* A mechanism converting psychosocial stress into mononuclear cell activation. Proc Natl Acad Sci U S A 2003, 100: 1920–1925.
- [104] Johnson JD, Campisi J, Sharkey CM, Kennedy SL, Nickerson M, Greenwood BN*, et al.* Catecholamines mediate stressinduced increases in peripheral and central inflammatory cytokines. Neuroscience 2005, 135: 1295–1307.
- [105] Yu NN, Wang XX, Yu JT, Wang ND, Lu RC, Miao D*, et al.* Blocking beta2-adrenergic receptor attenuates acute stressinduced amyloid beta peptides production. Brain Res 2010, 1317: 305–310.
- [106] Tran TT, Srivareerat M, Alhaider IA, Alkadhi KA. Chronic psychosocial stress enhances long-term depression in a subthreshold amyloid-beta rat model of Alzheimer's disease. J Neurochem 2011, 119: 408–416.
- [107] Kamal A, Ramakers GM, Altinbilek B, Kas MJ. Social isolation stress reduces hippocampal long-term potentiation: Effect of animal strain and involvement of glucocorticoid receptors. Neuroscience 2013.
- [108] Dong H, Goico B, Martin M, Csernansky CA, Bertchume A, Csernansky JG. Modulation of hippocampal cell proliferation, memory, and amyloid plaque deposition in APPsw (Tg2576) mutant mice by isolation stress. Neuroscience 2004, 127: 601–609.
- [109] Tanapat P, Galea LA, Gould E. Stress inhibits the proliferation of granule cell precursors in the developing dentate gyrus. Int J Dev Neurosci 1998, 16: 235–239.
- [110] Czeh B, Welt T, Fischer AK, Erhardt A, Schmitt W, Muller MB*, et al.* Chronic psychosocial stress and concomitant repetitive transcranial magnetic stimulation: effects on stress hormone levels and adult hippocampal neurogenesis. Biol Psychiatry 2002, 52: 1057–1065.
- [111] Thomas RM, Hotsenpiller G, Peterson DA. Acute psychosocial stress reduces cell survival in adult hippocampal neurogenesis without altering proliferation. J Neurosci 2007, 27: 2734– 2743.
- [112] Wood GE, Young LT, Reagan LP, McEwen BS. Acute and chronic restraint stress alter the incidence of social conflict in male rats. Horm Behav 2003, 43: 205–213.
- [113] Barha CK, Brummelte S, Lieblich SE, Galea LA. Chronic restraint stress in adolescence differentially influences hypothalamic-pituitary-adrenal axis function and adult

hippocampal neurogenesis in male and female rats. Hippocampus 2011, 21: 1216–1227.

- [114] Diniz L, dos Santos TB, Britto LR, Cespedes IC, Garcia MC, Spadari-Bratfisch RC, et al. Effects of chronic treatment with corticosterone and imipramine on fos immunoreactivity and adult hippocampal neurogenesis. Behav Brain Res 2013, 238: 170–177.
- [115] Dubovicky M. Neurobehavioral manifestations of developmental impairment of the brain. Interdiscip Toxicol 2010, 3: 59–67.
- [116] Zlokovic BV. Neurovascular mechanisms of Alzheimer's neurodegeneration. Trends Neurosci 2005, 28: 202–208.
- [117] Yan SD, Chen X, Fu J, Chen M, Zhu H, Roher A*, et al.* RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. Nature 1996, 382: 685–691.
- [118] Sturchler E, Galichet A, Weibel M, Leclerc E, Heizmann CW. Site-specific blockade of RAGE-Vd prevents amyloid-beta oligomer neurotoxicity. J Neurosci 2008, 28: 5149–5158.
- [119] Yan SF, Ramasamy R, Schmidt AM. The RAGE axis: a fundamental mechanism signaling danger to the vulnerable vasculature. Circ Res 2010, 106: 842–853.
- [120] Takuma K, Fang F, Zhang W, Yan S, Fukuzaki E, Du H*, et al.* RAGE-mediated signaling contributes to intraneuronal transport of amyloid-beta and neuronal dysfunction. Proc Natl Acad Sci U S A 2009, 106: 20021–20026.
- [121] Shibata M, Yamada S, Kumar SR, Calero M, Bading J, Frangione B*, et al.* Clearance of Alzheimer's amyloid-β1-40 peptide from brain by LDL receptor–related protein-1 at the blood-brain barrier. J Clin Investig 2000, 106: 1489–1499.
- [122] Deane R, Wu Z, Sagare A, Davis J, Du Yan S, Hamm K*, et al.* LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. Neuron 2004, 43: 333-344.
- [123] Klinge PM, Samii A, Niescken S, Brinker T, Silverberg GD. Brain amyloid accumulates in aged rats with kaolin-induced hydrocephalus. Neuroreport 2006, 17: 657–660.
- [124] Paris D, Patel N, DelleDonne A, Quadros A, Smeed R, Mullan M. Impaired angiogenesis in a transgenic mouse model of cerebral amyloidosis. Neurosci Lett 2004, 366: 80–85.
- [125] Paris D, Townsend K, Quadros A, Humphrey J, Sun J, Brem S*, et al.* Inhibition of angiogenesis by Abeta peptides. Angiogenesis 2004, 7: 75–85.
- [126] Simpson IA, Carruthers A, Vannucci SJ. Supply and demand in cerebral energy metabolism: the role of nutrient transporters. J Cereb Blood Flow Metab 2007, 27: 1766– 1791.
- [127] Simpson IA, Chundu KR, Davies-Hill T, Honer WG, Davies P. Decreased concentrations of GLUT1 and GLUT3 glucose transporters in the brains of patients with Alzheimer's disease. Ann Neurol 1994, 35: 546–551.
- [128] Apelt J, Ach K, Schliebs R. Aging-related down-regulation

of neprilysin, a putative beta-amyloid-degrading enzyme, in transgenic Tg2576 Alzheimer-like mouse brain is accompanied by an astroglial upregulation in the vicinity of beta-amyloid plaques. Neurosci Lett 2003, 339: 183–186.

- [129] Fuller S, Munch G, Steele M. Activated astrocytes: a therapeutic target in Alzheimer's disease? Expert Rev Neurother 2009, 9: 1585–1594.
- [130] Barnum SJJ, Muller–Ladner U, Samimi A, Campbell IL. Chronic complement C3 gene expression in the CNS of transgenic mice with astrocyte-targeted IL-6 expression. Glia 1996, 18:107–117.
- [131] Zhu Y, Hou H, Rezai-Zadeh K, Giunta B, Ruscin A, Gemma C*,* et al. CD45 deficiency drives amyloid-beta peptide oligomers and neuronal loss in Alzheimer's disease mice. J Neurosci 2011, 31: 1355–1365.
- [132] Tan J, Town T, Mori T, Wu Y, Saxe M, Crawford F*, et al.* CD45 opposes beta-amyloid peptide-induced microglial activation via inhibition of p44/42 mitogen-activated protein kinase. J Neurosci 2000, 20: 7587–7594.
- [133] Wilcock DM, Gordon MN, Ugen KE, Gottschall PE, DiCarlo G, Dickey C*, et al.* Number of Abeta inoculations in APP+PS1 transgenic mice influences antibody titers, microglial activation, and congophilic plaque levels. DNA Cell Biol 2001, 20: 731–736.
- [134] Yan SD, Stern DM. Mitochondrial dysfunction and Alzheimer's disease: role of amyloid-beta peptide alcohol dehydrogenase (ABAD). Int J Exp Pathol 2005, 86: 161–171.
- [135] Du H, Guo L, Yan S, Sosunov AA, McKhann GM, Yan SS. Early deficits in synaptic mitochondria in an Alzheimer's disease mouse model. Proc Natl Acad Sci U S A 2010, 107: 18670–18675.
- [136] Lesort M, Terro F, Esclaire F, Hugon J. Neuronal APP accumulates in toxic membrane blebbings. J Neural Transm 1997, 104: 497–513.
- [137] Paula-Lima AC, Brito-Moreira J, Ferreira ST. Deregulation of excitatory neurotransmission underlying synapse failure in Alzheimer's disease. J Neurochem 2013, 126: 191–202.
- [138] Masilamoni JG, Jesudason EP, Jesudoss KS, Murali J, Paul SF, Jayakumar R. Role of fibrillar Abeta25-35 in the inflammation induced rat model with respect to oxidative vulnerability. Free Radic Res 2005, 39: 603–612.
- [139] Cai Z, Hussain MD, Yan LJ. Microglia, neuroinflammation, and beta-amyloid protein in Alzheimer's disease. Int J Neurosci 2013.
- [140] Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS*, et al.* Mitochondrial abnormalities in Alzheimer's disease. J Neurosci 2001, 21: 3017–3023.
- [141] Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, Reddy PH. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: implications

for free radical generation and oxidative damage in disease progression. Hum Mol Genet 2006, 15: 1437–1449.

- [142] Cardoso SM, Santos S, Swerdlow RH, Oliveira CR. Functional mitochondria are required for amyloid betamediated neurotoxicity. FASEB J 2001, 15: 1439–1441.
- [143] Devi L, Prabhu BM, Galati DF, Avadhani NG, Anandatheerthavarada HK. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. J Neurosci 2006, 26: 9057–9068.
- [144] Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N*, et al.* ABAD directly links Abeta to mitochondrial toxicity in Alzheimer's disease. Science 2004, 304: 448–452.
- [145] Devi L, Prabhu B, Galati D, Avadhani N, HK. A. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. J Neurosci 2006, 26: 9057– 9068.
- [146] Luo Y, Sunderland T, Roth GS, Wolozin B. Physiological levels of beta-amyloid peptide promote PC12 cell proliferation. Neurosci Lett 1996, 217: 125–128.
- [147] Puzzo D, Privitera L, Leznik E, Fa M, Staniszewski A, Palmeri A*, et al.* Picomolar amyloid-beta positively modulates synaptic plasticity and memory in hippocampus. J Neurosci 2008, 28: 14537–14545.
- [148] Falangola MF, Lee SP, Nixon RA, Duff K, Helpern JA. Histological co-localization of iron in Abeta plaques of PS/ APP transgenic mice. Neurochem Res 2005, 30: 201–205.
- [149] Ghribi O, Golovko MY, Larsen B, Schrag M, Murphy EJ. Deposition of iron and β-amyloid plaques is associated with cortical cellular damage in rabbits fed with long-term cholesterol-enriched diets. J Neurochem 2006, 99: 438–449.
- [150] Bodovitz S, Falduto MT, Frail DE, Klein WL. Iron levels modulate alpha-secretase cleavage of amyloid precursor protein. J Neurochem 1995, 64: 307–315.
- [151] Connor JR, Lee SY. HFE mutations and Alzheimer's disease. J Alzheimers Dis 2006, 10: 267–276.
- [152] Lin M, Zhao L, Fan J, Lian XG, Ye JX, Wu L*, et al.* Association between HFE polymorphisms and susceptibility to Alzheimer's disease: a meta-analysis of 22 studies including 4,365 cases and 8,652 controls. Mol Biol Rep 2012, 39: 3089–3095.
- [153] Pulliam JF, Jennings CD, Kryscio RJ, Davis DG, Wilson D, Montine TJ*, et al.* Association of HFE mutations with neurodegeneration and oxidative stress in Alzheimer's disease and correlation with APOE. Am J Med Genet B Neuropsychiatr Genet 2003, 119B: 48–53.
- [154] Namekata K, Imagawa M, Terashi A, Ohta S, Oyama F, Ihara Y. Association of transferrin C2 allele with late-onset Alzheimer's disease. Hum Genet 1997, 101: 126–129.
- [155] Crapper DR, Krishnan SS, Dalton AJ. Brain aluminum distribution in Alzheimer's disease and experimental neurofibrillary degeneration. Science 1973, 180: 511-513.
- [156] McLachlan DR, Bergeron C, Smith JE, Boomer D, Rifat SL. Risk for neuropathologically confirmed Alzheimer's disease and residual aluminum in municipal drinking water employing weighted residential histories. Neurology 1996, 46: 401–405.
- [157] Rondeau V, Commenges D, Jacqmin-Gadda H, Dartigues JF. Relation between aluminum concentrations in drinking water and Alzheimer's disease: An 8-year follow-up study. J Epidemiol 2000, 152: 59–66.
- [158] Langui D, Probst A, Anderton B, Brion JP, Ulrich J. Aluminium-induced tangles in cultured rat neurones. Enhanced effect of aluminium by addition of maltol. Acta Neuropathol 1990, 80: 649–655.
- [159] Mera SL. Aluminium, amyloid, and Alzheimer's disease. Med Lab Sci 1991, 48: 283–295.
- [160] Exley C. The aluminium-amyloid cascade hypothesis and Alzheimer's disease. Subcell Biochem 2005, 38: 225–234.
- [161] Forbes MS, Ghribi O, Herman MM, Savory J. Aluminuminduced dendritic pathology revisited: cytochemical and electron microscopic studies of rabbit cortical pyramidal neurons. Ann Clin Lab Sci 2002, 32: 75–86.
- [162] Vasudevaraju P, Govindaraju M, Palanisamy AP, Sambamurti K, Rao KS. Molecular toxicity of aluminium in relation to neurodegeneration. Indian J Med Res 2008, 128: 545–556.
- [163] Savory J, Herman MM, Ghribi O. Intracellular mechanisms underlying aluminum-induced apoptosis in rabbit brain. J Inorg Biochem 2003, 97: 151–154.
- [164] Kawahara M. Effects of aluminum on the nervous system and its possible link with neurodegenerative diseases. J Alzheimers Dis 2005, 8: 171–182; discussion 209-115.
- [165] Sensi SL, Paoletti P, Bush AI, Sekler I. Zinc in the physiology and pathology of the CNS. Nat Rev Neurosci 2009, 10: 780– 791.
- [166] Bush AI, Pettingell WH Jr, Paradis MD, Tanzi RE. Modulation of A beta adhesiveness and secretase site cleavage by zinc. J Biol Chem 1994, 269: 12152–12158.
- [167] Lesne S, Koh MT, Kotilinek L, Kayed R, Glabe CG, Yang A*, et al.* A specific amyloid-beta protein assembly in the brain impairs memory. Nature 2006, 440: 352–357.
- [168] Dukes KD, Rodenberg CF, Lammi RK. Monitoring the earliest amyloid-beta oligomers via quantized photobleaching of dyelabeled peptides. Anal Biochem 2008, 382: 29–34.
- [169] Deshpande A, Kawai H, Metherate R, Glabe CG, Busciglio J. A role for synaptic zinc in activity-dependent Aβ oligomer formation and accumulation at excitatory synapses. J Neurosci 2009, 29: 4004–4015.
- [170] Huang YZ, Pan E, Xiong ZQ, McNamara JO. Zinc-mediated transactivation of TrkB potentiates the hippocampal mossy

fiber-CA3 pyramid synapse. Neuron 2008, 57: 546-558.

- [171] Bush AI. Drug development based on the metals hypothesis of Alzheimer's disease. J Alzheimers Dis 2008, 15: 223–240.
- [172] Bjorkdahl C, Sjogren MJ, Winblad B, Pei JJ. Zinc induces neurofilament phosphorylation independent of p70 S6 kinase in N2a cells. Neuroreport 2005, 16: 591–595.
- [173] Freeman JW, Couch JR. Prolonged encephalopathy with arsenic poisoning. Neurology 1978, 28: 853–855.
- [174] Lee VM, Balin BJ, Otvos L Jr, Trojanowski JQ. A68: a major subunit of paired helical filaments and derivatized forms of normal Tau. Science 1991, 251: 675–678.
- [175] Giasson BI, Sampathu DM, Wilson CA, Vogelsberg-Ragaglia V, Mushynski WE, Lee VMY. The Environmental Toxin Arsenite Induces Tau Hyperphosphorylation. Biochemistry 2002, 41: 15376–15387.
- [176] Jenkins SM, Johnson GVW. Microtubule/MAP-Affinity Regulating Kinase (MARK) is activated by phenylarsine oxide in situ and phosphorylates tau within its microtubule-binding domain. J Neurochem 2000, 74: 1463–1468.
- [177] Sparks DL, Schreurs BG. Trace amounts of copper in water induce beta-amyloid plaques and learning deficits in a rabbit model of Alzheimer's disease. Proc Natl Acad Sci U S A 2003, 100: 11065–11069.
- [178] Brewer GJ. The risks of copper toxicity contributing to cognitive decline in the aging population and to Alzheimer's disease. J Am Coll Nutr 2009, 28: 238–242.
- [179] Needleman HL, Gatsonis CA. Low-level lead exposure and the IQ of children. A meta-analysis of modern studies. JAMA 1990, 263: 673–678.
- [180] Lefauconnier JM, Bernard G, Mellerio F, Sebille A, Cesarini E. Lead distribution in the nervous system of 8-month-old rats intoxicated since birth by lead. Experientia 1983, 39: 1030–1031.
- [181] Rahman A, Brew BJ, Guillemin GJ. Lead dysregulates serine/threonine protein phosphatases in human neurons. Neurochem Res 2011, 36: 195–204.
- [182] Wang SC, Oelze B, Schumacher A. Age-specific epigenetic drift in late-onset Alzheimer's disease. PLoS One 2008, 3: e2698.
- [183] Gong CX, Shaikh S, Wang JZ, Zaidi T, Grundke-Igbal I, Igbal K. Phosphatase activity toward abnormally phosphorylated tau: decrease in Alzheimer disease brain. J Neurochem 1995, 65: 732–738.
- [184] Liu F, Iqbal K, Grundke-Iqbal I, Rossie S, Gong CX. Dephosphorylation of tau by protein phosphatase 5: impairment in Alzheimer's disease. J Biol Chem 2005, 280: 1790–1796.
- [185] Rudrabhatla P, Pant HC. Role of protein phosphatase 2A in Alzheimer's disease. Curr Alzheimer Res 2011, 8: 623–632.
- [186] Sontag E, Luangpirom A, Hladik C, Mudrak I, Ogris E,

Speciale S*, et al.* Altered expression levels of the protein phosphatase 2A ABalphaC enzyme are associated with Alzheimer disease pathology. J Neuropathol Exp Neurol 2004, 63: 287–301.

- [187] Gong CX, Singh TJ, Grundke-Iqbal I, Iqbal K. Phosphoprotein phosphatase activities in Alzheimer disease brain. J Neurochem 1993, 61: 921–927.
- [188] Liu R, Zhou XW, Tanila H, Bjorkdahl C, Wang JZ, Guan ZZ*, et al.* Phosphorylated PP2A (tyrosine 307) is associated with Alzheimer neurofibrillary pathology. J Cell Mol Med 2008, 12: 241–257.
- [189] Zhou XW, Gustafsson JA, Tanila H, Bjorkdahl C, Liu R, Winblad B*, et al.* Tau hyperphosphorylation correlates with reduced methylation of protein phosphatase 2A. Neurobiol Dis 2008, 31: 386–394.
- [190] Tanimukai H, Grundke-Iqbal I, Iqbal K. Up-regulation of inhibitors of protein phosphatase-2A in Alzheimer's disease. Am J Pathol 2005, 166: 1761–1771.
- [191] Kins S, Crameri A, Evans DR, Hemmings BA, Nitsch RM, Gotz J. Reduced protein phosphatase 2A activity induces hyperphosphorylation and altered compartmentalization of tau in transgenic mice. J Biol Chem 2001, 276: 38193– 38200.
- [192] Avila J, Perry G, Martinez-Martin P. Prospects on the origin of Alzheimer's disease. J Alzheimers Dis 2010, 20: 669–672.
- [193] Hanger DP, Hughes K, Woodgett JR, Brion JP, Anderton BH. Glycogen synthase kinase-3 induces Alzheimer's disease-like phosphorylation of tau: generation of paired helical filament epitopes and neuronal localisation of the kinase. Neurosci Lett 1992, 147: 58–62.
- [194] Reynolds CH, Betts JC, Blackstock WP, Nebreda AR, Anderton BH. Phosphorylation sites on tau identified by nanoelectrospray mass spectrometry. J Neurochem 2000, 74: 1587–1595.
- [195] Vingtdeux V, Davies P, Dickson D, Marambaud P. AMPK is abnormally activated in tangle- and pre-tangle-bearing neurons in Alzheimer's disease and other tauopathies. Acta Neuropathologica 2011, 121: 337–349.
- [196] Reynolds CH, Nebreda AR, Gibb GM, Utton MA, Anderton BH. Reactivating kinase/p38 phosphorylates tau protein *in vitro*. J Neurochem 1997, 69: 191–198.
- [197] Reynolds CH, Utton MA, Gibb GM, Yates A, Anderton BH. Stress-activated protein kinase/c-jun N-terminal kinase phosphorylates tau protein. J Neurochem 1997, 68: 1736– 1744.
- [198] Drewes G, Trinczek B, Illenberger S, Biernat J, Schmitt-Ulms G, Meyer HE*, et al.* Microtubule-associated Protein/ Microtubule Affinity-regulating Kinase (p110mark): A novel protein kinase that regulates tau-microtubule interactions and dynamic instability by phosphorylation at the Alzheimer-

specific site Serine 262. J Biol Chem 1995, 270: 7679–7688.

- [199] Gupta RP, Abou-Donia MB. Tau phosphorylation by diisopropyl phosphorofluoridate (DFP)-treated hen brain supernatant inhibits its binding with microtubules: role of Ca2+/Calmodulin-dependent protein kinase II in tau phosphorylation. Arch Biochem Biophys 1999, 365: 268–278.
- [200] Derkinderen P, Scales TME, Hanger DP, Leung KY, Byers HL, Ward MA*, et al.* Tyrosine 394 is phosphorylated in Alzheimer's paired helical filament tau and in fetal tau with c-Abl as the candidate tyrosine kinase. J Neurosci 2005, 25: 6584–6593.
- [201] Williamson R, Scales T, Clark BR, Gibb G, Reynolds CH, Kellie S*, et al.* Rapid tyrosine phosphorylation of neuronal proteins including tau and focal adhesion kinase in response to amyloid-beta peptide exposure: involvement of Src family protein kinases. J Neurosci 2002, 22: 10–20.
- [202] Zhu X, Sun Z, Lee HG, Siedlak SL, Perry G, Smith MA. Distribution, levels, and activation of MEK1 in Alzheimer's disease. J Neurochem 2003, 86: 136–142.
- [203] Zhu X, Rottkamp CA, Hartzler A, Sun Z, Takeda A, Boux H*, et al.* Activation of MKK6, an upstream activator of p38, in Alzheimer's disease. J Neurochem 2001, 79: 311–318.
- [204] Iijima-Ando K, Zhao L, Gatt A, Shenton C, Iijima K. A DNA damage-activated checkpoint kinase phosphorylates tau and enhances tau-induced neurodegeneration. Hum Mol Genet 2010, 19: 1930–1938.
- [205] Leclerc S, Garnier M, Hoessel R, Marko D, Bibb JA, Snyder GL*, et al.* Indirubins inhibit glycogen synthase kinase-3β and CDK5/P25, two protein kinases involved in abnormal tau phosphorylation in Alzheimer's disease: A property common to most cyclin-dependent kinase inhibitors? J Biol Chem 2001, 276: 251–260.
- [206] Peng CX, Hu J, Liu D, Hong XP, Wu YY, Zhu LQ*, et al.* Disease-modified glycogen synthase kinase-3beta intervention by melatonin arrests the pathology and memory deficits in an Alzheimer's animal model. Neurobiol Aging 2013, 34: 1555–1563.
- [207] Cheung ZH, Ip NY. Cdk5: a multifaceted kinase in neurodegenerative diseases. Trends Cell Biol 2012, 22: 169–175.
- [208] Sundaram JR, Poore CP, Sulaimee NH, Pareek T, Asad AB, Rajkumar R*, et al.* Specific inhibition of p25/Cdk5 activity by the Cdk5 inhibitory peptide reduces neurodegeneration in vivo. J Neurosci 2013, 33: 334–343.
- [209] Basurto-Islas G, Grundke-Iqbal I, Tung YC, Liu F, Iqbal K. Activation of asparaginyl endopeptidase leads to Tau hyperphosphorylation in Alzheimer disease. J Biol Chem 2013, 288: 17495–17507.
- [210] Wasik U, Schneider G, Mietelska-Porowska A, Mazurkiewicz M, Fabczak H, Weis S*, et al.* Calcyclin binding protein and

Siah-1 interacting protein in Alzheimer's disease pathology: neuronal localization and possible function. Neurobiol Aging 2013, 34: 1380–1388.

- [211] Marques SC, Oliveira CR, Pereira CM, Outeiro TF. Epigenetics in neurodegeneration: a new layer of complexity. Prog Neuropsychopharmacol Biol Psychiatry 2011, 35: 348– 355.
- [212] Zawia NH, Lahiri DK, Cardozo-Pelaez F. Epigenetics, oxidative stress, and Alzheimer disease. Free Radic Biol Med 2009, 46: 1241–1249.
- [213] Fuso A, Seminara L, Cavallaro RA, D'Anselmi F, Scarpa S. S-adenosylmethionine/homocysteine cycle alterations modify DNA methylation status with consequent deregulation of PS1 and BACE and beta-amyloid production. Mol Cell Neurosci 2005, 28: 195–204.
- [214] Fuso A, Nicolia V, Pasqualato A, Fiorenza MT, Cavallaro RA, Scarpa S. Changes in Presenilin 1 gene methylation pattern in diet-induced B vitamin deficiency. Neurobiol Aging 2011, 32: 187–199.
- [215] Chen KL, Wang SS, Yang YY, Yuan RY, Chen RM, Hu CJ. The epigenetic effects of amyloid-beta(1-40) on global DNA and neprilysin genes in murine cerebral endothelial cells. Biochem Biophys Res Commun 2009, 378: 57–61.
- [216] Wu J, Basha MR, Brock B, Cox DP, Cardozo-Pelaez F, McPherson CA*, et al.* Alzheimer's disease (AD)-like pathology in aged monkeys after infantile exposure to environmental metal lead (Pb): evidence for a developmental origin and

environmental link for AD. J Neurosci 2008, 28: 3–9.

- [217] Fischer A, Sananbenesi F, Wang X, Dobbin M, Tsai L-H. Recovery of learning and memory is associated with chromatin remodelling. Nature 2007, 447: 178–182.
- [218] Perez M, Santa-Maria I, De Barreda EG, Zhu X, Cuadros R, Cabrero JR*, et al.* Tau – an inhibitor of deacetylase HDAC6 function. J Neurochem 2009, 109: 1756–1766.
- [219] Green KN, Steffan JS, Martinez-Coria H, Sun X, Schreiber SS, Thompson LM*, et al.* Nicotinamide restores cognition in Alzheimer's disease transgenic mice via a mechanism involving sirtuin inhibition and selective reduction of Thr231 phosphotau. J Neurosci 2008, 28: 11500–11510.
- [220] Ricobaraza A, Cuadrado-Tejedor M, Perez-Mediavilla A, Frechilla D, Del Rio J, Garcia-Osta A. Phenylbutyrate ameliorates cognitive deficit and reduces tau pathology in an Alzheimer's disease mouse model. Neuropsychopharmacology 2009, 34: 1721–1732.
- [221] Sweatt JD. Behavioural neuroscience: Down memory lane. Nature 2007, 447: 151–152.
- [222] Francis YI, Fa M, Ashraf H, Zhang H, Staniszewski A, Latchman DS*, et al.* Dysregulation of histone acetylation in the APP/PS1 mouse model of Alzheimer's disease. J Alzheimers Dis 2009, 18: 131–139.
- [223] Zhang K, Schrag M, Crofto n A, Trivedi R, Vinters H, Kirsch W. Targeted proteomics for quantification of histone acetylation in Alzheimer's disease. Proteomics 2012, 12: 1261–1268.