

HHS Public Access

Author manuscript *J Invest Dermatol.* Author manuscript; available in PMC 2018 September 01.

Published in final edited form as:

J Invest Dermatol. 2017 September; 137(9): 2033–2035. doi:10.1016/j.jid.2017.04.022.

Activating CYSLTR2 and PLCB4 mutations in primary leptomeningeal melanocytic tumors

Johannes A.P. van de Nes^{1,3}, Christian Koelsche⁴, Marco Gessi^{5,6}, Inga Möller², Antje Sucker², Richard A. Scolyer^{7,9,10}, Michael E. Buckland^{8,9}, Torsten Pietsch⁵, Rajmohan Murali¹¹, Dirk Schadendorf², and Klaus G. Griewank^{2,12}

¹Institute of Pathology, Ruhr University Bochum, Bochum, Germany

²Department of Dermatology, University Hospital Essen, West German Cancer Center, University Duisburg-Essen and the German Cancer Consortium (DKTK), Germany

³Institute of Neuropathology, University Hospital Essen, West German Cancer Center, University Duisburg-Essen and the German Cancer Consortium (DKTK), Germany

⁴Department of Neuropathology, Ruprecht-Karls-University Heidelberg, and Clinical Cooperation Unit Neuropathology, and DKTK, DKFZ, Heidelberg, Germany

⁵Institute of Neuropathology, University of Bonn Medical Center, Bonn, Germany

⁶Division of Histopathology, Fondazione Policlinico Universitario "A.Gemelli", Università Cattolica del Sacro Cuore, Roma, Italy

⁷Tissue Pathology and Diagnostic Oncology, Camperdown, NSW, Australia

⁸Dept. of Neuropathology, Royal Prince Alfred Hospital, Camperdown, NSW, Australia

⁹The University of Sydney, Camperdown, NSW, Australia

¹⁰Melanoma Institute Australia, North Sydney, NSW, Australia

¹¹Department of Pathology, Memorial Sloan Kettering Cancer Center, New York NY, USA

¹²Dermatopathologie bei Mainz, Nieder-Olm, Germany

Melanocytic tumors arising in the central nervous system are also known as primary leptomeningeal melanocytic tumors (PLMTs). They frequently show a benign behavior and are designated as 'melanocytomas' in the World Health Organization classification of central nervous system tumors (Brat et al., 2016). Distinction of PLMTs from melanomas metastatic to the central nervous system can be challenging.

Recent studies have demonstrated that PLMTs have a very characteristic genetic profile, frequently harboring *GNAQ* and *GNA11* mutations (Gessi et al., 2014, Koelsche et al., 2015,

CORRESPONDENCE: Klaus G. Griewank^{1,2} (klaus.griewank@uk-essen.de), ¹Department of Dermatology, University Hospital Essen, University of Duisburg-Essen, Germany, ²Dermatopathologie bei Mainz, Nieder-Olm, Germany.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

van de Nes et al.

Kusters-Vandevelde et al., 2010a, Kusters-Vandevelde et al., 2015, Kusters-Vandevelde et al., 2010b, Murali et al., 2012). These mutations are very rare in cutaneous melanoma (Cancer Genome Atlas, 2015), but frequent in uveal melanoma (Van Raamsdonk et al., 2009, Van Raamsdonk et al., 2010). The close molecular relationship between PMLTs and uveal melanomas was further demonstrated by the finding that *EIF1AX*, *SF3B1* and *BAP1* mutations, previously identified in uveal melanomas, can also occur in PMLT (Kusters-Vandevelde et al., 2016, van de Nes et al., 2016a, van de Nes et al., 2016b). Similar to uveal melanomas, a proportion of PLMTs are not found to harbor activating mutations in *GNAQ* or *GNA11*.

Recent studies have identified activating *CYSLTR2* and *PLCB4* mutations in uveal melanomas, occurring at the L129 and D630 hotspots, respectively (Johansson et al., 2016, Moore et al., 2016). *CYSLTR2* mutations have also been detected in blue nevi (Moller et al., 2016). These mutations always occurred in tumors lacking *GNAQ* and *GNA11* mutations, and have not yet been reported in cutaneous melanomas (Cancer Genome Atlas, 2015). To determine whether *CYSLTR2* and *PLCB4* mutations also occur in PLMTs, we analyzed our previously published cohort of tumors (van de Nes et al., 2016a, van de Nes et al., 2016b), using a next-generation sequencing gene panel covering the mutational hot-spots in *CYSLTR2* and *PLCB4* as well as other gene mutations reported in uveal melanoma (described in Supplemental Material and previously reported (Moller et al., 2016)).

In 19 PLMTs, we found two tumors with activating L129Q (c.386T>A) mutations in *CYSLTR2* and one tumor harboring an activating D630Y (c.1888G>T) mutation in *PLCB4* (Figure 1, Table 1). These mutations were identified at the same hotspots previously described in uveal melanoma and predicted to be activating (Johansson et al., 2016, Moore et al., 2016). In our PLMT cohort, *CYSLTR2* and *PLCB4* mutations were found to be mutually exclusive of *GNAQ* and *GNA11* mutations. Together, mutations in *GNAQ*, *GNA11*, *CYSLTR2* or *PLCB4* were identified in 18/19 (94.7%) PLMTs.

The hotspot mutations in *GNAQ* and *GNA11* (R183 and Q209), *PLCB4* (D630) and *CYSLTR2* (L129) result in activation of the corresponding proteins and increased stimulation of a common downstream signaling pathway (Johansson et al., 2016, Moore et al., 2016, Van Raamsdonk et al., 2009, Van Raamsdonk et al., 2010). *CYSLTR2* codes for the CYSLTR2 receptor, a seven transmembrane receptor which signals through the products of the *GNAQ* and *GNA11* genes, the highly homologous heterotrimeric G proteins Gaq and Ga11. Gaq and Ga11 directly activate the *PLCB4* gene product, phospholipase-C β 4, which hydrolyses PIP2 (phosphatidylinositol 4,5-bisphosphate) releasing DAG (diacylglyerol) and IP3 (inositol-1,4,5-trisphosphate), subsequently releasing Ca+ (Suh et al., 2008, Waldo et al., 2010). The similar functional consequences of these gene mutations is demonstrated by the occurrence of these mutations in a mutually exclusive fashion.

The increased coverage and sensitivity of our sequencing assay identified multiple additional mutations in other genes, including *EIF1AX* (Figure 1), as was recently reported (Kusters-Vandevelde et al., 2016). One *EIF1AX* mutation (R13C, c.37C>T) was identified in a tumor that also harbored an activating *CYSLTR2*L129Q mutation (Supplemental Figure 1). In addition to a known *SF3B1* R625H (c.1874G>A) and inactivating *BAP1* R60* (c.178C>T)

J Invest Dermatol. Author manuscript; available in PMC 2018 September 01.

mutation (van de Nes et al., 2016a, van de Nes et al., 2016b), our targeted next-generation sequencing approach detected a *BAP1* E31del (c.91_93delGAG) mutation in the PLMT sample also harboring a *PLCB4* D630Y mutation (Supplemental Figure 1). The functional relevance of this *BAP1* in-frame one amino acid deletion is unclear. BAP1 immunohistochemistry showed retained nuclear protein expression (Supplemental Figure 2). However, copy number analysis (Supplemental Figure 3) demonstrated loss of chromosome 3 including the other wild-type *BAP1* allele which means the mutation would be highly relevant if it resulted in loss or impairment of protein function. The relevance of *EIF1AX*, *SF3B1* and *BAP1* mutations in terms of clinical behavior and prognosis needs to be fully elucidated in future larger studies.

In summary, our study demonstrates the occurrence of activating *CYSLTR2* or *PLCB4* mutations in PLMTs lacking *GNAQ* and *GNA11* mutations. Activating mutations in these genes are exceedingly rare in other melanomas, with the exception of uveal melanomas. The diagnosis of a PLMT requires exclusion of a more frequently occurring CNS metastasis of a non-CNS melanoma (most commonly of cutaneous origin). Presence of a *PLCB4, CYSLTR2, GNAQ* or *GNA11* mutation is strong evidence in favor of a PLMT, but a rare CNS uveal melanoma metastasis should also be considered.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

References

- Brat, D., Perry, A., Wesseling, P., Bastian, B. Melanocytic tumours. In: Louis, D.Ohgaki, H.Wiestler, O., Cavanee, W., editors. WHO Classification of Tumours of the Central Nervous System. Lyon: IARC Press; 2016. p. 266-70.
- Cancer Genome Atlas N. Genomic Classification of Cutaneous Melanoma. Cell. 2015; 161(7):1681– 96. [PubMed: 26091043]
- Gessi M, van de Nes J, Griewank K, Barresi V, Buckland ME, Kirfel J, et al. Absence of TERT promoter mutations in primary melanocytic tumours of the central nervous system. Neuropathology and applied neurobiology. 2014; 40(6):794–7. [PubMed: 24645797]
- Johansson P, Aoude LG, Wadt K, Glasson WJ, Warrier SK, Hewitt AW, et al. Deep sequencing of uveal melanoma identifies a recurrent mutation in PLCB4. Oncotarget. 2016; 7(4):4624–31. [PubMed: 26683228]
- Koelsche C, Hovestadt V, Jones DT, Capper D, Sturm D, Sahm F, et al. Melanotic Tumors of the Nervous System are Characterized by Distinct Mutational, Chromosomal and Epigenomic Profiles. Brain pathology. 2015; 25(2):202–8. [PubMed: 25399693]
- Kusters-Vandevelde HV, Creytens D, van Engen-van Grunsven AC, Jeunink M, Winnepenninckx V, Groenen PJ, et al. SF3B1 and EIF1AX mutations occur in primary leptomeningeal melanocytic neoplasms yet another similarity to uveal melanomas. Acta neuropathologica communications. 2016; 4:5. [PubMed: 26769193]
- Kusters-Vandevelde HV, Klaasen A, Kusters B, Groenen PJ, van Engen-van Grunsven IA, van Dijk MR, et al. Activating mutations of the GNAQ gene: a frequent event in primary melanocytic neoplasms of the central nervous system. Acta neuropathologica. 2010a; 119(3):317–23. [PubMed: 19936769]
- Kusters-Vandevelde HV, van Engen-van Grunsven IA, Coupland SE, Lake SL, Rijntjes J, Pfundt R, et al. Mutations in g protein encoding genes and chromosomal alterations in primary leptomeningeal melanocytic neoplasms. Pathology oncology research : POR. 2015; 21(2):439–47. [PubMed: 25315378]

J Invest Dermatol. Author manuscript; available in PMC 2018 September 01.

van de Nes et al.

- Kusters-Vandevelde HV, van Engen-van Grunsven IA, Kusters B, van Dijk MR, Groenen PJ, Wesseling P, et al. Improved discrimination of melanotic schwannoma from melanocytic lesions by combined morphological and GNAQ mutational analysis. Acta neuropathologica. 2010b; 120(6): 755–64. [PubMed: 20865267]
- Moller I, Murali R, Muller H, Wiesner T, Jackett LA, Scholz SL, et al. Activating cysteinyl leukotriene receptor 2 (CYSLTR2) mutations in blue nevi. Modern pathology : an official journal of the United States and Canadian Academy of Pathology, Inc. 2016
- Moore AR, Ceraudo E, Sher JJ, Guan Y, Shoushtari AN, Chang MT, et al. Recurrent activating mutations of G-protein-coupled receptor CYSLTR2 in uveal melanoma. Nature genetics. 2016; 48(6):675–80. [PubMed: 27089179]
- Murali R, Wiesner T, Rosenblum MK, Bastian BC. GNAQ and GNA11 mutations in melanocytomas of the central nervous system. Acta neuropathologica. 2012; 123(3):457–9. [PubMed: 22307269]
- Suh PG, Park JI, Manzoli L, Cocco L, Peak JC, Katan M, et al. Multiple roles of phosphoinositidespecific phospholipase C isozymes. BMB reports. 2008; 41(6):415–34. [PubMed: 18593525]
- van de Nes J, Gessi M, Sucker A, Moller I, Stiller M, Horn S, et al. Targeted next generation sequencing reveals unique mutation profile of primary melanocytic tumors of the central nervous system. Journal of neuro-oncology. 2016a; 127(3):435–44. [PubMed: 26744134]
- van de Nes J, Wrede K, Ringelstein A, Stiller M, Horn S, Sucker A, et al. Diagnosing a Primary Leptomeningeal Melanoma by Gene Mutation Signature. The Journal of investigative dermatology. 2016b; 136(7):1526–8. [PubMed: 27060446]
- Van Raamsdonk CD, Bezrookove V, Green G, Bauer J, Gaugler L, O'Brien JM, et al. Frequent somatic mutations of GNAQ in uveal melanoma and blue naevi. Nature. 2009; 457(7229):599–602. [PubMed: 19078957]
- Van Raamsdonk CD, Griewank KG, Crosby MB, Garrido MC, Vemula S, Wiesner T, et al. Mutations in GNA11 in uveal melanoma. The New England journal of medicine. 2010; 363(23):2191–9. [PubMed: 21083380]
- Waldo GL, Ricks TK, Hicks SN, Cheever ML, Kawano T, Tsuboi K, et al. Kinetic scaffolding mediated by a phospholipase C-beta and Gq signaling complex. Science. 2010; 330(6006):974–80. [PubMed: 20966218]

van de Nes et al.



Figure 1. Distribution of mutations in primary leptomeningeal melanocytic tumors

Distribution of mutations identified in primary leptomeningeal melanocytic tumors (PLMT). The histological diagnosis is demonstrated in the lowest row (M = melanocytoma, IGM = intermediate grade melanocytoma, Mel = primary leptomeningeal melanoma). $^{\circ}$ signifies previously pu blished mutations (van de Nes et al., 2016a, van de Nes et al., 2016b)

J Invest Dermatol. Author manuscript; available in PMC 2018 September 01.

Author
Manuscri

Author Manuscript

Author Manuscript

1
Ut
ho
۲ م
lar
ึมเ
ŝ
pt

BAPI	1	'	ı	ı	E31de	ı.	·	I	I	ı	ı	1	ı.	ı	T	1	'	R60* [*]	ı
SF3B1				ı	,	ı	ı		ï	·	·		·		ī				R625H°
EIFIAX	ı			ı	Ţ	R13C	ı	ı	ı	ı	G8V	G9D	·	ı	I	ı	R13P	·	G15D
CYSLTR2	ı	ı	·	ı		L129Q	ı	ı	ı	I	ı	·	ı	ı	ı	L129Q		ı	ı
PLCB4		·	ı	ı	D630Y		ı	ı	ı	ı	ı	ı	ı		ī			ı	ı
GNAII			·	ı	ı	ı	ı		ı	ı	ı		·	ı	ī	·	Q209L°	Q209L°	ı
10)L°	°do	9L			Чć	L°	Ľ	oP °	L°	L°)L°	9L	° d				°o
GN_{i}		Q209	Q209	Q20	'	1	Q20	Q209	Q209	Q209	Q209	Q209	Q209	Q20	Q209		'	·	R183
Sample GN/		P Q209	P Q209	P Q20	Ρ.	Ч.	P Q200	P Q209	R Q209	P Q209	P Q209	P Q209	P Q209	P Q20	P Q209	Ч.	R -	Ρ.	P R183
Diag. Sample GN	- P	M P Q209	M P Q209	M P Q20	М Р -	М Р-	IGM P Q20	IGM P Q209	IGM R Q209	IGM P Q209	IGM P Q209	IGM P Q209	IGM P Q209	IGM P Q20	IGM P Q209	IGM P -	IGM R -	IGM P -	Mel P R183
Location Diag. Sample GN	cavernous sinus M P -	spinal M P Q200	frontal M P Q209	spinal C2-3 M P Q20	temporal right M P -	spinal T7-8 M P -	spinal T10-12 IGM P Q20	spinal C1 IGM P Q209	spinal T4-5 IGM R Q209	spinal C5-6 IGM P Q209	spinal C3 IGM P Q209	spinal T8 IGM P Q205	spinal C5 IGM P Q200	NK IGM P Q20	frontal IGM P Q209	spinal T9-10 IGM P -	spinal T8-9 IGM R -	tentorium IGM P -	brain Mel P R183
Sex Location Diag. Sample GN	f cavernous sinus M P -	f spinal M P Q20	f frontal M P Q205	m spinal C2-3 M P Q20	f temporal right M P -	f spinal T7-8 M P -	m spinal T10-12 IGM P Q20	m spinal C1 IGM P Q205	m spinal T4-5 IGM R Q205	f spinal C5-6 IGM P Q205	m spinal C3 IGM P Q205	f spinal T8 IGM P Q205	m spinal C5 IGM P Q200	NK NK IGM P Q20	m frontal IGM P Q205	m spinal T9-10 IGM P -	f spinal T8-9 IGM R -	f tentorium IGM P -	m brain Mel P R183
Age Sex Location Diag. Sample GN	46 f cavernous sinus M P -	48 f spinal M P Q20	41 f frontal M P Q209	73 m spinal C2-3 M P Q20	73 f temporal right M P -	77 f spinal T7-8 M P -	34 m spinal T10-12 IGM P Q20	44 m spinal C1 IGM P Q205	79 m spinal T4-5 IGM R Q205	35 f spinal C5-6 IGM P Q200	65 m spinal C3 IGM P Q205	69 f spinal T8 IGM P Q205	64 m spinal C5 IGM P Q200	NK NK NK IGM P Q20	20 m frontal IGM P Q205	67 m spinal T9-10 IGM P -	48 f spinal T8-9 IGM R -	78 f tentorium IGM P -	66 m brain Mel P R183

NK

NK

A

A

8y 3y

A

NK NK NK

NK

NK

A

NK

NK

 $\begin{smallmatrix} 6y \\ 1y \\ 1 \end{smallmatrix}$

Þ

ŝ

Ω

5y

A, $R^{\#}$

NK

Clinicopathological and mutation information of primary leptomeningeal melanocytic tumors

J Invest Dermatol. Author manuscript; available in PMC 2018 September 01.

male; f = female; NK = not known; P = primary; R = recurrence; M = melanocytoma; IGM = intermediate grade melanocytoma; Mel = melanoma; del. = deletion; FU = follow-up period; NK = not known; unclear functional significances; Nr. = number; Diag. = diagnosis; m = $A = alive; R^{\#} = recurred after 3 years (mutation profile was identical to the primary); R^{\$} = recurred, then lost to follow-up; D = dead (disease related)$ 'II, BICY ncuon; reu expected to alter protein tu Dnin acuvaning m green

 $_o^\circ$ signifies mutations previously published (van de Nes et al., 2016a, van de Nes et al., 2016b)

FU NK NK NK NK NK NK NK Sy

Status

NK NK NK