

# Biomedical Applications of Polyhydroxyalkanoates

Subhasree Ray<sup>1,2</sup> · Vipin Chandra Kalia<sup>1,2</sup> 

Received: 3 April 2017 / Accepted: 20 April 2017 / Published online: 22 April 2017  
© Association of Microbiologists of India 2017

**Abstract** Polyhydroxyalkanoates (PHA) are produced by a large number of microbes under stress conditions such as high carbon (C) availability and limitations of nutrients such as nitrogen, potassium, phosphorus, magnesium, and oxygen. Here, microbes store C as granules of PHAs—energy reservoir. PHAs have properties, which are quite similar to those of synthetic plastics. The unique properties, which make them desirable materials for biomedical applications is their biodegradability, biocompatibility, and non-toxicity. PHAs have been found suitable for various medical applications: biocontrol agents, drug carriers, biodegradable implants, tissue engineering, memory enhancers, and anticancer agents.

**Keywords** Antibacterials · Biocontrol agents · Biodegradable implants · Drug carriers · Memory enhancer · Tissue engineering

## Abbreviations

PHA	Polyhydroxyalkanoate
PHB	Polyhydroxybutyrate
3HB	3-Hydroxybutyric acid
3HV	3-Hydroxyvaleric acid
3HO	3-Hydroxyoctanoate
3HD	3-Hydroxydecanoic acid
4HB	4-Hydroxybutyric acid

P(3HB-3HV)	Poly-3hydroxybutyrate-co-3hydroxyvalerate
P(3HB-4HB-3HV)	Poly-3hydroxybutyrate-co-4hydroxybutyrate-co-3hydroxyvalerate
P(3HB-3HV-3HHx)	Poly-3hydroxybutyrate-co-3hydroxyvalerate-co-3hydroxyhexanoate
P(3HB-3HO)	Poly-3hydroxybutyrate-co-3hydroxyoctanoate
P(3HB-3HV-DHB)	Poly(3-hydroxybutyrate-co-3-hydroxyvalerate-co-2,3-dihydroxybutyrate)
3HA	Hydroxyalkanoic acid
OA	Octanoic acid
UA	Undecanoic acid

## Introduction

Plastics are synthetic polymers which find wide usage in our daily lives. The major limitation associated with plastics is their non-biodegradable nature. The production of plastics in large quantities makes their disposal a major issue to worry about among Environmentalists and Health Departments. The most well envisaged alternative is to produce biodegradable plastics. In Nature, microbes have been bestowed with ability to withstand environmental stress. Under stress caused by high carbon: nitrogen (or potassium, oxygen, magnesium, phosphorus) ratio, microbes manipulate their metabolic activities in such a manner that rather than following the tricarboxylic acid cycle, the acetyl-CoA is diverted towards the synthesis of

✉ Vipin Chandra Kalia  
vckalia@igib.res.in; vc\_kalia@yahoo.co.in

<sup>1</sup> Microbial Biotechnology and Genomics, CSIR - Institute of Genomics and Integrative Biology (IGIB), Delhi University Campus, Mall Road, Delhi 110007, India

<sup>2</sup> Academy of Scientific and Innovative Research (AcSIR), 2, Rafi Marg, Anusandhan Bhawan, New Delhi 110001, India

polyhydroxyalkanoates (PHAs), which can be categorized as completely degradable natural bioplastics [1–14].

The use of PHAs as biodegradable polymers has gained attention because of their biological (microbial) origin and non-toxic nature compared to synthetic plastics, which may be toxic [15]. Biocompatibility of PHA polymers—PHB and PHBV is an issue which has been the focus for their use in medical applications. In fact these polymers have caused prolonged and acute inflammatory responses. It was realized that we need PHAs which will be appropriate for engineering biological tissues and various applications in medical field [16, 17].

### Antibacterials

R-3HAs are chiral compounds, which have potential for being used as building blocks for compounds, for use in pharmaceutical industry [18]. 3HA can be transformed into different hydroxycarboxylic acids such as 2-alkylated 3HB and  $\beta$ -lactones. These can be employed as oral drugs [19]. R-3HAs can be formed by the degradation of PHAs [18]. The most important compounds are carbapenem or macrolide antibiotics [20, 21]. Depolymerase enzyme of *Pseudomonas fluorescens* GK13 encoded by gene *phaZGK13* can depolymerise PHAs to monomers [22, 23]. These monomeric units can reduce bacterial infection such as those by *Staphylococcus aureus* [24] and those conjugated with D-peptide prove anti-cancerous [25]. P3HB/P4HB helps in enhancing angiogenic properties of skin and wound healing (Table 1) [26, 27].

### Biocontrol Agents

Antibiotics are commonly used as feed supplement for animals. At low concentration, these antibiotics have been reported to influence the growth of the animals—livestock and aquaculture [28]. As the incorporation of antibiotic in a consistent manner can be a risky affair and the gastrointestinal microflora is likely to develop resistance [29, 30]. One has to ensure that there is complete elimination of antibiotic from the digestive system of the animal. It has been observed that Short-chain fatty acids (SCFAs) are effective in controlling agents against pathogens [31]. As PHAs are biopolymers of  $\beta$ -hydroxy SCFAs, these can be metabolized in the intestinal tract. The metabolites can be exploited as biocontrol agents for giant tiger prawn *Penaeus monodon* [32, 33].

### Drug Carriers

In order to improve the efficacy of the drugs, it has been envisaged that they should be delivered in a controlled and targeted manner. PHAs have been recognized as biomaterials which have desirable physical properties with high biocompatibility. Hence, their usage as raw material for producing tablets, nano-particles and as scaffolds for eluting drugs can be effective [34, 35].

3HB monomers have proved helpful in the synthesis of novel polymers such as Dendrimers. These polymers are biodegradable, have monodispersity, and surface-functional moieties. These features make them as potent drug carriers [19, 20]. Monomers—3HB and 4HB have been exploited for preparing novel  $\beta$ - and  $\gamma$ -peptides, which are resistant to the action of peptidases. This enables them to stay longer in mammalian serum i.e., improving its suitability for cargo-drug delivery. 3HB can inhibit glycolysis during haemorrhagic shock. These monomers are helpful in the synthesis of sex hormones and fragrance (S-citronellol). A few other properties of these monomers include antibacterial, anti-proliferative and haemolytic activities [36].

PHB microspheres carrying rifampicin were used as hemoembolizing agent. The microspheres proved effective in releasing drug. Implantable rods prepared with PHB, PHBV and their copolymers such as P(3HB-4HB) were used for delivery of antibiotics [37]. mcl-PHAs are more effective candidate for drug delivery as they have lower crystallinity and melting point. mcl-PHA have been employed for transdermal drug delivery. PHA copolymers containing 3HO and 3HHx shows adhesion to the python reticulatus skin and was used for dispersing three respective drugs such as tamsulosin, ketoprofen and clonidine. This drug shows good permeability through PHA matrix [38].

*P. fluorescens* produces mcl-PHAs, which helps in drug delivery, protein purification and immobilizing agent in clinical diagnostics (Table 1) [39]. These PHAs are bioengineered as biologically active beads for various medical applications. PHA beads are used in clinical diagnostics such as in recombinant protein production, vaccine delivery, bio-imaging, endotoxin removal, etc. (Table 1) [40–50]. PHA beads are also used as tuberculin skin test reagent for diagnosis of bovine tuberculosis (Table 1) [51, 52].

**Table 1** Potential applications of polyhydroxyalkanoates and their derivatives in medical industries

Bioproducts	Source	Applications	References
Polyhydroxybutyrate	Not known	Stomach wall patch	[77]
	<i>Staphylococcus aureus</i>	Peripheral nerve guide	[54]
Polyhydroxyalkanoate (mcl)	<i>P. fluorescens</i>	Drug delivery, protein microarray, protein purification, antibody immobilization in clinical diagnostics	[39]
Polyhydroxyalkanoate beads (antigen displaying polyester beads)	<i>Escherichia coli</i>	Tuberculin skin test reagent for diagnosis of bovine tuberculosis (TB)	[51, 52]
Polyhydroxyalkanoate beads	<i>E. coli</i>	Recombinant protein production	[47]
	<i>E. coli</i>	Protein purification	[49]
	<i>E. coli</i>	Vaccine delivery agent	[43]
	<i>Lactococcus lactis</i>	Hepatitis C vaccine delivery agent	[44]
	<i>Alcaligenes latus</i>	Microbeads	[40]
	<i>E. coli</i>	Nano/microdevices for bioimaging in medical approaches	[42]
	<i>E. coli</i>	Fused to specific antigen and applied as beads in Fluorescence activated cell sorting based diagnostics	[41]
	<i>E. coli</i>	Displays foreign antigens are immunogenic and presents a delivery system for vaccination against Hepatitis C virus	[50]
Polyhydroxybutyrate-valerate	<i>Ralstonia eutropha</i> B5785	Endotoxin removal and protein purification	[48]
	Sigma-Aldrich Company	Suture	[79]
	Not known	Myocardial patch	[78]
Poly-3-hydroxybutyrate, Poly-4-hydroxybutyrate	Not known	Bone regeneration	[57]
	<i>Cupriavidus eutrophus</i> B-10646	Elastic nonwoven membranes-helps in reducing inflammation, enhancing angiogenic properties of skin, facilitate wound healing capacity	[74]
Polyhydroxybutyrate-hydroxyhexanoate	Shantou Lianyi Biotech Company	Osteoblast attachment, proliferation and differentiation	[55, 56]
	Not known	Bone regeneration	[55]
	<i>Aeromonas hydrophila</i> 4AK4	Vessel stent, hemocompatibility and cytocompatibility	[58, 59]
	<i>A. hydrophila</i> 4AK4 (Recombinant) (20%)	Smooth muscle cells related graft scaffolds for tissue engineering	
	Not known	Cartilage tissue engineering	[56]
	–	Nerve conduit	[60]
	Not known	Scaffold for cartilage tissue engineering	[61]
	Not known	Nanoparticles	[34]
Polyhydroxybutyrate	<i>Biomer and Roth</i>	Scaffolds for tissue engineering	[67]
Polyhydroxyvalerate	<i>Goodfellow</i>		
Poly-3-hydroxyoctanoate-co-3-hydroxyundecanoate	<i>Procter and Gamble</i> <i>EMPA</i>		
Poly-3-hydroxy-acetylthioalkanoate-co-3-hydroxyalkanoate	<i>P. putida</i> KT2442	Antibacterial activity against methicillin resistant <i>S. aureus</i>	[24, 26]
	<i>P. putida</i> KT2442 fadB; <i>P. fluorescens</i> GK13		
3Hydroxybutyrate	<i>E. coli</i>	Immobilized cell factories for biocatalysis and bio-transformation, Chaperone protein levels	[46]
3Hydroxyalkanoic acid	<i>Pseudomonas</i> sp.	Helps in establishing PHA producers in soil and rhizosphere, and improves metabolism	[23]
3-hydroxydecanoic acids conjugated to D-peptide	<i>P. putida</i> CA-3	Anti-cancer activity	[25]
Polyhydroxybutyrate-valerate microspheres	Sigma-Aldrich Company	Neural tissue engineering	[66]
PHB-Hydroxyapatite composite	<i>R. eutropha</i> B5786	Bone regeneration	[74]
	Not known	Bone bonding between implants and biological tissue	[73]

**Table 1** continued

Bioproducts	Source	Applications	References
	Not known	Bone tissue engineering	[75]
Fibronectin and alginate coated PHB-fiber	Astra Tech Sweden	Spinal cord injury	[94]
PHBV-PLGA	Aldrich, UK	Nerve tissue engineering	[76]
Unsaturated m- and lcl-copolyesters: PHO-Sy series	<i>P. oleovorans</i> octanoic acid (OA) and soya oily acids (Sy)	Subcutaneous patches in rats	[92]
Gold embedded poly (3-OH octanoate-co-3-OH-10-undecenoate)	<i>P. oleovorans</i> (OA and 10-undecanoic acid, UA)	Subcutaneous implantation in rats	[93]
Polyhydroxyoctanoate +PGA conduit	PHO 3836; TEPHA Inc., Cambridge, MA PHO 3836; TEPHA Inc., Cambridge, MA PHO 3836; TEPHA Inc., Cambridge, MA PHO 3836; TEPHA Inc., Cambridge, MA PHO3836 TEPHA Inc, Cambridge, MA	Vascular tissue engineered structures	[72]

## Tissue Engineering

PHAs available in general were not targeted for use as medical implants and were thus lacking the quality which can get approval of the Drug Administrators. The need is to produce PHAs of high purity, check their biodegradation in vivo, fabrication of scaffolds, modify their surface [53]. PHAs with necessary modifications hold great potential to contribute to tissue engineering, developing tissue products for medical and therapeutic applications: (1) vascular grafts, (2) heart valves, (3) nerve tissue engineering, etc. (Table 1) [54–65].

PHAs can be used to produce scaffolds, which have higher mechanical strength. These scaffolds promote growth of the cells by supplying nutrition (Table 1) [66]. These products are available as screws, pins, sutures, films, etc. (Table 1) [67]. P(3HB-4HB-3HV) has been exploited for fiber meshes by providing support to stem cell growth for proliferation and cell adhesion [68]. P(3HB-3HV-3HHx) can be employed as scaffolds for engineering liver tissue [69]. On the other hand, 3-D scaffolds had been developed by using PHA nanofibers [70]. P(3HB-3HO) have found usage for scaffold formation for cartilage repair [71]. The new P(3-HB-3HV-2,3-diHB) produced by recombinant *Ralstonia eutropha* have also been exploited [53].

Further to enhance mechanical strength and flexibility of PHAs, inorganic bioceramics have been combined with PHAs, which produce novel composites for using them for engineering tissues. Composites of PHA and ceramic are employed to form different blends. Hydroxyapatite and PHA are also used in tissue engineering (Table 1) [72–76].

## Medical Devices

PHA have been envisaged to prove useful in making medical devices, as they are biocompatible, biodegradable and have strong mechanical characteristics. Some of the most potential devices are: adhesions barriers, articular cartilage repair, cardiovascular patch grafting, meniscus repair device, orthopedic pins, repair patch, rivets and tacks, staples and screws, stents, surgical mesh, sutured fastener, etc. (Table 1) [77–80]. Vessel stent produced from PHA copolymer (PHB-HHx) from *Aeromonas hydrophila* 4AK4 has high hemo- and bio-compatibility (Table 1) [58, 59]. The preparation of cardiovascular patches should have very high quality features, primary being resistance to infection and degradation. In addition, these bioproducts should be durable, lack immunogenicity and should not be toxic.

## Biodegradable Implants

Implants are used on a large scale in a very skilful manner. However, invariably the issue of their getting infected with pathogens comes as a major hurdle. Now-a-days biomaterials are also used as implants. Biomaterial associated infection is a serious health issue. In order to meet the functional demand, materials with desirable properties must be selected [81]. For example, their physical, chemical, biological, biochemical properties. Use of biopolymers as biodegradable implants has greatly influenced the modern medicine [82].

The use of PHA degradation product HAs for preparing biodegradable implants, and to fabricate systems to deliver antibiotics like Duocid and Sulperazone, in chronic osteomyelitis therapy have been gathering importance [19]. Rod of PHA biopolymer having antibiotics, such as combinations of sulbactam-ampicillin/cefoperazone, gentamicin, were prepared with the help of copolymer—P(3HB-3HV) as matrix [83–86]. Mixtures P3HB and P3HO were employed as a matrix system for coronary stents needed for eluting drugs. This can prove effective in reducing the recurrence of the blockages in the arteries [87].

To avoid biofilm formation and bacterial adhesion, there is a need to improve implant surfaces [88]. PHB and its copolymer P(3HB-3HHx) sheets loaded with lysozyme are being used for biofilm inhibition and in fabrication of wound dressing [89]. Implants coated with PHB copolymer fastens the degradation process for a stable drug release within a given time period as compared to its co-polymer [90, 91].

Gold-catalyzed oxidation of bacterial polyester produced by *Pseudomonas oleovorans* supplemented with octanoic acid and 10-undecanoic acid was employed for biodegradable subcutaneous implantation in rats (Table 1) [92, 93]. PHB fiber covered with fibronectin and alginate was used as implant in spinal cord injury (Table 1) [94].

## Anti-osteoporosis Effect

Ketoacidosis is induced in human beings by the accumulation of high concentration of 3HB [95]. Oligomers of 3HB have properties to act as good energy substrate for patients, where these compounds undergo rapid diffusion within peripheral tissues. It also prevents brain damage as it can enhance cardiac efficiency by regenerating mitochondrial energy. 3HB can potentially cure Parkinson's and Alzheimer's diseases, where they act by reducing the death rate of neuronal cells [96]. 3HB enhances osteoblasts growth and anti-osteoporosis activity, by rapidly depositing calcium and its strong serum alkaline phosphatase activity.

It helps in prevention lowering of bone mineral density and reducing the level of serum osteocalcin [97, 98].

## Memory Enhancer

Memory loss and related abilities are serious enough to disturb our daily routine. Among the different forms of dementia, Alzheimer's disease is the most common. As a consequence of memory loss, there are problems of thinking and behaviour. Derivatives of 3HB such as 3-hydroxybutyrate methyl ester (HBME) have the potential to act as drug against Alzheimer's disease. HBME acts by protecting mitochondrial damages [99]. During ketogenesis, D-β-HB prevents neuronal death, which is induced by glucose deprivation [100]. HA monomers derived from PHAs can stimulate the Ca<sup>2+</sup> channels, which can help in enhancing memory [98, 101–104].

## Challenges for Industrial Scale Production of PHAs

PHAs have unique characteristics, which make them suitable for commercial production. However, there are still a few challenges, which limit their upscaling [3]. The physical and chemical properties which demand attention include: lowering of melting point and glass transition temperature, elastic modulus, tensile strength and elongation. These characteristics depend up on the monomeric composition and the molecular weight of the polymer. Copolymers of PHAs with high molecular weights have the potential to overcome these limitations [3]. In addition, strategies to manipulate feed composition [14], culture conditions such as independence from nutritional imbalance [8, 107], increased microbial biomass, high expression of polymerase genes and genetic modifications to synchronize termination of PHA biosynthesis with cell lysis [1] will certainly help in economic production of PHAs on an industrial scale.

## Future Prospects

PHAs have been finding their applications in diverse fields [105–107]. The economic feasibility of using PHAs and their derivatives for medical purposes stands at the highest level. The usage of PHAs can be extended to other health related issues such as cancer therapy, fighting malnutrition, neurodegenerative and metabolic disorders, anti-diabetics agent, monitor environmental health, etc.



**Acknowledgements** The authors wish to thank the Director of CSIR-Institute of Genomics and Integrative Biology (CSIR-IGIB), CSIR-HRD project OLP1126 (ES Scheme No. 21(1022)/16/EMR-2), Delhi, India, for providing the necessary funds, facilities and moral support. Authors are also thankful to Academy of Scientific & Innovative Research (AcSIR), New Delhi.

## References

- Singh M, Patel SKS, Kalia VC (2009) *Bacillus subtilis* as potential producer for polyhydroxyalkanoates. *Microb Cell Fact* 8:38. doi:10.1186/1475-2859-8-38
- Singh M, Kumar P, Patel SKS, Kalia VC (2013) Production of polyhydroxyalkanoate co-polymer by *Bacillus thuringiensis*. *Indian J Microbiol* 53:77–83. doi:10.1007/s12088-012-0294-7
- Singh M, Kumar P, Ray S, Kalia VC (2015) Challenges and opportunities for the customizing polyhydroxyalkanoates. *Indian J Microbiol* 55:235–249. doi:10.1007/s12088-015-0528-6
- Kumar P, Patel SKS, Lee JK, Kalia VC (2013) Extending the limits of *Bacillus* for novel biotechnological applications. *Biotechnol Adv* 31:1543–1561. doi:10.1016/j.biotechadv.2013.08.007
- Kumar P, Singh M, Mehariya S, Patel SKS, Lee JK, Kalia VC (2014) Ecobiotechnological approach for exploiting the abilities of *Bacillus* to produce co-polymer of polyhydroxyalkanoate. *Indian J Microbiol* 54:1–7. doi:10.1007/s12088-014-0457-9
- Kumar P, Mehariya S, Ray S, Mishra A, Kalia VC (2015) Biodiesel industry waste: a potential source of bioenergy and biopolymers. *Indian J Microbiol* 55:1–7. doi:10.1007/s12088-014-0509-1
- Kumar P, Mehariya S, Ray S, Mishra A, Kalia VC (2015) Biotechnology in aid of biodiesel industry effluent (glycerol): biofuels and bioplastics. In: Kalia VC (ed) *Microbial factories*. Springer, New Delhi, pp 105–119. doi:10.1007/978-81-322-2598-0
- Kumar P, Ray S, Patel SKS, Lee JK, Kalia VC (2015) Bio-conversion of crude glycerol to polyhydroxyalkanoate by *Bacillus thuringiensis* under non-limiting nitrogen conditions. *Int J Biol Macromol* 78:9–16. doi:10.1016/j.ijbiomac.2015.03.046
- Raut S, Raut S, Sharma M, Srivastav C, Adhikari B, Sen SK (2015) Enhancing degradation of low density polyethylene films by *Curvularia lunata* SG1 using particle swarm optimization strategy. *Indian J Microbiol* 55:258–268. doi:10.1007/s12088-015-0522-z
- Patel SKS, Kumar P, Singh S, Lee JK, Kalia VC (2015) Integrative approach for hydrogen and polyhydroxybutyrate production. In: Kalia VC (ed) *Microbial factories waste treatment*. Springer, New Delhi, pp 73–85. doi:10.1007/978-81-322-2598-0\_5
- Patel SKS, Kumar P, Singh S, Lee JK, Kalia VC (2015) Integrative approach to produce hydrogen and polyhydroxybutyrate from biowaste using defined bacterial cultures. *Bioresour Technol* 176:136–141. doi:10.1016/j.biortech.2014.11.029
- Patel SKS, Lee JK, Kalia VC (2016) Integrative approach for producing hydrogen and polyhydroxyalkanoate from mixed wastes of biological origin. *Indian J Microbiol* 56:293–300. doi:10.1007/s12088-016-0595-3
- Kalia VC, Prakash J, Koul S (2016) Biorefinery for glycerol rich biodiesel industry waste. *Indian J Microbiol* 56:113–125. doi:10.1007/s12088-016-0583-7
- Ray S, Kalia VC (2016) Microbial cometabolism and polyhydroxyalkanoate co-polymers. *Indian J Microbiol* 57:39–47. doi:10.1007/s12088-016-0622-4
- Koller M, Marsalek L, de Sousa Dias MM, Braunegg G (2016) Producing microbial polyhydroxyalkanoate (PHA) biopolyesters in a sustainable manner. *New Biotechnol* 37:24–38. doi:10.1016/j.nbt.2016.05.001
- Williams SF, Martin DP (2005) Applications of polyhydroxyalkanoates (PHA) in medicine and pharmacy. *Biopolymers*. doi:10.1002/3527600035.bpol4004
- Hazer DB, Kılıçay E, Hazer B (2012) Poly(3-hydroxyalkanoate)s: diversification and biomedical applications: a state of the art review. *Mater Sci Eng C* 32:637–647. doi:10.1016/j.msec.2012.01.021
- Babel W, Ackermann JU, Breuer U (2001) Physiology, regulation, and limits of the synthesis of poly(3HB). *Adv Biochem Eng Biotechnol* 71:125–157. doi:10.1007/3-540-40021-4\_4
- Chen GQ, Wu Q (2005) The application of polyhydroxyalkanoates as tissue engineering materials. *Biomaterials* 26:6565–6578. doi:10.1016/j.biomaterials.2005.04.036
- Chen GQ, Wu Q (2005) Microbial production and applications of chiral hydroxyalkanoates. *Appl Microbiol Biotechnol* 67:592–599. doi:10.1007/s00253-005-1917-2
- Shivakumar S, Jagadish SJ, Zatakia H, Dutta J (2011) Purification, characterization and kinetic studies of a novel poly( $\beta$ )hydroxybutyrate (PHB) depolymerase *PhaZ* from *Penicillium citrinum* S2. *Appl Biochem Biotechnol* 164:1225–1236. doi:10.1007/s12010-011-9208-0
- Cai L, Yuan MQ, Liu F, Jian J, Chen GQ (2009) Enhanced production of medium-chain-length polyhydroxyalkanoates (PHA) by PHA depolymerase knockout mutant of *Pseudomonas putida* KT2442. *Bioresour Technol* 100:2265–2270. doi:10.1016/j.biortech.2008.11.020
- De Eugenio LI, Escapa IF, Morales V, Dinjaski N, Galan B, Garcia JL, Prieto MA (2010) The turnover of medium-chain-length polyhydroxyalkanoates in KT2442 and the fundamental role of *PhaZ* depolymerase for the metabolic balance. *Environ Microbiol* 12:207–221. doi:10.1111/j.1462-2920.2009.02061.x
- Martinez V, Dinjaski N, De Eugenio LI, De la Pena F, Prieto MA (2014) Cell system engineering to produce extracellular polyhydroxyalkanoate depolymerase with targeted applications. *Int J Biol Macromol* 71:28–33. doi:10.1016/j.ijbiomac.2014.04.013
- O'Connor S, Szejewicz E, Nikodinovic-Runic J, O'Connor A, Byrne AT, Devocelle M, O'Donovan N, Gallagher WM, Babu R, Kenny ST, Zinn M (2013) The anti-cancer activity of a cationic anti-microbial peptide derived from monomers of polyhydroxyalkanoate. *Biomaterials* 34:2710–2718. doi:10.1016/j.biomaterials.2012.12.032
- Dinjaski N, Fernandez-Gutierrez M, Selvam S, Parra-Ruiz FJ, Lehman SM, San Roman J, Garcia E, Garcia JL, Garcia AJ, Prieto MA (2014) PHACOS, a functionalized bacterial polyester with bactericidal activity against methicillin-resistant *Staphylococcus aureus*. *Biomaterials* 35:14–24. doi:10.1016/j.biomaterials.2013.09.059
- Shishatskaya EI, Nikolaeva ED, Vinogradova ON, Volova TG (2016) Experimental wound dressings of degradable PHA for skin defect repair. *J Mater Sci Mater Med* 27:165. doi:10.1007/s10856-016-5776-4
- Cabello FC (2006) Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environ Microbiol* 8:1137–1144. doi:10.1111/j.1462-2920.2006.01054.x
- Bangera R, Correa K, Lhorente JP, Figueroa R, Yáñez JM (2017) Genomic predictions can accelerate selection for resistance against *Piscirickettsia salmonis* in Atlantic salmon (*Salmo salar*). *BMC Genom* 18:121. doi:10.1186/s12864-017-3487-y

30. Martinez JL (2017) Effect of antibiotics on bacterial populations: a multi-hierarchical selection process. *F1000 Res* 6:51. doi:[10.12688/f1000research.9685.1](https://doi.org/10.12688/f1000research.9685.1)
31. Martínez V, de la Peña F, García-Hidalgo J, de la Mata I, García JL, Prieto MA (2012) Identification and biochemical evidence of a medium-chain-length polyhydroxyalkanoate depolymerase in the *Bdellovibrio bacteriovorus* predatory hydrolytic arsenal. *Appl Environ Microbiol* 78:6017–6026. doi:[10.1128/AEM.01099-12](https://doi.org/10.1128/AEM.01099-12)
32. Defoirdt T, Boon N, Sorgeloos P, Verstraete W, Bossier P (2009) Short-chain fatty acids and poly- $\beta$ -hydroxyalkanoates: (new) biocontrol agents for a sustainable animal production. *Biotechnol Adv* 27:680–685. doi:[10.1016/j.biotechadv.2009.04.026](https://doi.org/10.1016/j.biotechadv.2009.04.026)
33. Ludevese-Pascual G, Laranja JLQ, Amar EC, Sorgeloos P, Bossier P, De Schryver P (2016) Poly-beta-hydroxybutyrate-enriched *Artemia* sp. for giant tiger prawn *Penaeus monodon* larviculture. *Aquaculture* 23:422–429. doi:[10.1111/anu.12410](https://doi.org/10.1111/anu.12410)
34. Xiong YC, Yao YC, Zhan XY, Chen GQ (2010) Application of polyhydroxyalkanoates nanoparticles as intracellular sustained drug-release vectors. *J Biomater Sci* 21:127–140. doi:[10.1163/156856209X410283](https://doi.org/10.1163/156856209X410283)
35. Nigmatullin R, Thomas P, Lukaszewicz B, Puthussery H, Roy I (2015) Polyhydroxyalkanoates, a family of natural polymers, and their applications in drug delivery. *J Chem Technol Biotechnol* 90:1209–1221. doi:[10.1002/jctb.4685](https://doi.org/10.1002/jctb.4685)
36. Philip S, Keshavarz T, Roy I (2007) Polyhydroxyalkanoates: biodegradable polymers with a range of applications. *J Chem Technol Biotechnol* 82:233–247. doi:[10.1002/jctb.1667](https://doi.org/10.1002/jctb.1667)
37. Türesin F, Gursel I, Hasirci V (2001) Biodegradable polyhydroxyalkanoate implants for osteomyelitis therapy: in vitro antibiotic release. *J Biomater Sci Polym Ed* 12:195–207. doi:[10.1163/156856201750180924](https://doi.org/10.1163/156856201750180924)
38. Mokhtarzadeh A (2016) Recent advances on biocompatible and biodegradable nanoparticles as gene carriers. *Expert Opin Biol Ther* 16:771–785. doi:[10.1517/14712598.2016.1169269](https://doi.org/10.1517/14712598.2016.1169269)
39. Ihssen J, Magnani D, Thony-Meyer L, Ren Q (2009) Use of extracellular medium chain length polyhydroxyalkanoate depolymerase for targeted binding of proteins to artificial poly [(3-hydroxyoctanoate)-co-(3-hydroxyhexanoate)] granules. *Biomacromol* 10:1854–1864. doi:[10.1021/bm9002859](https://doi.org/10.1021/bm9002859)
40. Lee SJ, Park JP, Park TJ, Lee SY, Lee S, Park JK (2005) Selective immobilization of fusion proteins on poly(hydroxyalkanoate) microbeads. *Anal Chem* 77:5755–5759. doi:[10.1021/ac0505223](https://doi.org/10.1021/ac0505223)
41. Bäckström BT, Brockelbank JA, Rehm BH (2007) Recombinant *Escherichia coli* produces tailor-made biopolyester granules for applications in fluorescence activated cell sorting: functional display of the mouse interleukin-2 and myelin oligodendrocyte glycoprotein. *BMC Biotechnol* 7:3. doi:[10.1186/1472-6750-7-3](https://doi.org/10.1186/1472-6750-7-3)
42. Jahns AC, Haverkamp RG, Rehm BH (2008) Multifunctional inorganic-binding beads self-assembled inside engineered bacteria. *Bioconjug Chem* 19:2072–2080. doi:[10.1021/bc8001979](https://doi.org/10.1021/bc8001979)
43. Parlane NA, Wedlock DN, Buddle BM, Rehm BH (2009) Bacterial polyester inclusions engineered to display vaccine candidate antigens for use as a novel class of safe and efficient vaccine delivery agents. *App Environ Microbiol* 75:7739–7744. doi:[10.1128/AEM.01965-09](https://doi.org/10.1128/AEM.01965-09)
44. Parlane NA, Grage K, Lee JW, Buddle BM, Denis M, Rehm BH (2011) Production of a particulate hepatitis C vaccine candidate by an engineered *Lactococcus lactis* strain. *Appl Environ Microbiol* 77:8516–8522. doi:[10.1128/AEM.06420-11](https://doi.org/10.1128/AEM.06420-11)
45. Parlane NA, Gupta SK, Rubio-Reyes P, Chen S, Gonzalez-Miro M, Wedlock DN, Rehm BH (2016) Self-assembled protein-coated polyhydroxyalkanoate beads: properties and biomedical applications. *ACS Biomater Sci Eng*. doi:[10.1021/acsbiomaterials.6b00355](https://doi.org/10.1021/acsbiomaterials.6b00355)
46. Wang Q, Yu H, Xia Y, Kang Z, Qi Q (2009) Complete PHB mobilization in *Escherichia coli* enhances the stress tolerance: a potential biotechnological application. *Microb Cell Fact* 8:1. doi:[10.1186/1475-2859-8-47](https://doi.org/10.1186/1475-2859-8-47)
47. Geng Y, Wang S, Qi Q (2010) Expression of active recombinant human tissue-type plasminogen activator by using in vivo polyhydroxybutyrate granule display. *App Environ Microbiol* 76:7226–7230. doi:[10.1128/AEM.01543-10](https://doi.org/10.1128/AEM.01543-10)
48. Li J, Shang G, You M, Peng S, Wang Z, Wu H, Chen GQ (2011) Endotoxin removing method based on lipopolysaccharide binding protein and polyhydroxyalkanoate binding protein PhaP. *Biomacromol* 12:602–608. doi:[10.1021/bm101230n](https://doi.org/10.1021/bm101230n)
49. Hay ID, Du J, Reyes PR, Rehm BH (2015) In vivo polyester immobilized sortase for tagless protein purification. *Microb Cell Fact* 14:190. doi:[10.1186/s12934-015-0385-3](https://doi.org/10.1186/s12934-015-0385-3)
50. Martínez-Donato G, Piniella B, Aguilar D, Olivera S, Pérez A, Castañedo Y, Alvarez-Lajonchere L, Dueñas-Carrera S, Lee JW, Burr N, Gonzalez-Miro M (2016) Protective T cell and antibody immune responses against Hepatitis C virus achieved using a biopolyester-bead-based vaccine delivery system. *Clin Vaccine Immunol* 23:370–378. doi:[10.1128/CVI.00687-15](https://doi.org/10.1128/CVI.00687-15)
51. Chen S, Parlane NA, Lee J, Wedlock DN, Buddle BM, Rehm BH (2014) New skin test for detection of bovine tuberculosis on the basis of antigen-displaying polyester inclusions produced by recombinant *Escherichia coli*. *Appl Environ Microbiol* 80:2526–2535. doi:[10.1128/AEM.04168-13](https://doi.org/10.1128/AEM.04168-13)
52. Parlane NA, Chen S, Jones GJ, Vordermeier HM, Wedlock DN, Rehm BH, Buddle BM (2016) Display of antigens on polyester inclusions lowers the antigen concentration required for a bovine tuberculosis skin test. *Clin Vaccine Immunol* 23:19–26. doi:[10.1128/CVI.00462-15](https://doi.org/10.1128/CVI.00462-15)
53. Insomphun C, Chuah JA, Kobayashi S, Fujiki T, Numata K (2016) Influence of hydroxyl groups on the cell viability of polyhydroxyalkanoate (PHA) scaffolds for tissue engineering. *ACS Biomater Sci Eng*. doi:[10.1021/acsbiomaterials.6b00279](https://doi.org/10.1021/acsbiomaterials.6b00279)
54. Mosahebi A, Fuller P, Wiberg M, Terenghi G (2002) Effect of allogeneic Schwann cell transplantation on peripheral nerve regeneration. *Exp Neurol* 173:213–223. doi:[10.1006/exnr.2001.7846](https://doi.org/10.1006/exnr.2001.7846)
55. Wang YW, Wu Q, Chen J, Chen GQ (2005) Evaluation of three-dimensional scaffolds made of blends of hydroxyapatite and poly (3-hydroxybutyrate-co-3-hydroxyhexanoate) for bone reconstruction. *Biomaterials* 26:899–904. doi:[10.1016/j.biomaterials.2004.03.035](https://doi.org/10.1016/j.biomaterials.2004.03.035)
56. Wang Y, Bian YZ, Wu Q, Chen GQ (2008) Evaluation of three-dimensional scaffolds prepared from poly (3-hydroxybutyrate-co-3-hydroxyhexanoate) for growth of allogeneic chondrocytes for cartilage repair in rabbits. *Biomaterials* 29:2858–2868. doi:[10.1016/j.biomaterials.2008.03.021](https://doi.org/10.1016/j.biomaterials.2008.03.021)
57. Cool SM, Kenny B, Wu A, Nurcombe V, Trau M, Cassady AI, Grondahl L (2007) Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) composite biomaterials for bone tissue regeneration: in vitro performance assessed by osteoblast proliferation, osteoclast adhesion and resorption, and macrophage proinflammatory response. *J Biomed Mater Res* 3:599–610. doi:[10.1007/s00253-011-3099-4](https://doi.org/10.1007/s00253-011-3099-4)
58. Qu XH, Wu Q, Chen GQ (2006) In vitro study on hemocompatibility and cytocompatibility of poly (3-hydroxybutyrate-co-3-hydroxyhexanoate). *J Biomater Sci Polym Ed* 17:1107–1121. doi:[10.1163/156856206778530704](https://doi.org/10.1163/156856206778530704)
59. Qu XH, Wu Q, Liang J, Zou B, Chen GQ (2006) Effect of 3-hydroxyhexanoate content in poly (3-hydroxybutyrate-co-3-hydroxyhexanoate) on in vitro growth and differentiation of

- smooth muscle cells. *Biomaterials* 27:2944–2950. doi:[10.1016/j.biomaterials.2006.01.013](https://doi.org/10.1016/j.biomaterials.2006.01.013)
60. Bian YZ, Wang Y, Aibaidoula G, Chen GQ, Wu Q (2009) Evaluation of poly (3-hydroxybutyrate-co-3-hydroxyhexanoate) conduits for peripheral nerve regeneration. *Biomaterials* 30:217–225. doi:[10.1016/j.biomaterials.2008.09.036](https://doi.org/10.1016/j.biomaterials.2008.09.036)
  61. Ye C, Hu P, Ma MX, Xiang Y, Liu RG, Shang XW (2009) PHB/PHBHHx scaffolds and human adipose-derived stem cells for cartilage tissue engineering. *Biomaterials* 30:4401–4406. doi:[10.1016/j.biomaterials.2009.05.001](https://doi.org/10.1016/j.biomaterials.2009.05.001)
  62. Levine AC, Sparano A, Twigg FF, Numata K, Nomura CT (2015) Influence of cross-linking on the physical properties and cytotoxicity of polyhydroxyalkanoate (PHA) scaffolds for tissue engineering. *ACS Biomater Sci Eng* 1:567–576. doi:[10.1021/acsbiomaterials.6b00279](https://doi.org/10.1021/acsbiomaterials.6b00279)
  63. Goonoo N, Bhaw-Luximon A, Passanha P, Esteves SR, Jhurry D (2016) Third generation poly(hydroxyacid) composite scaffolds for tissue engineering. *J Biomed Mater Res B*. doi:[10.1002/jbm.b.33674](https://doi.org/10.1002/jbm.b.33674)
  64. Ke Y, Zhang XY, Ramakrishna S, He LM, Wu G (2017) Reactive blends based on polyhydroxyalkanoates: preparation and biomedical application. *Mater Sci Eng C Mater Biol Appl* 70:1107–1119. doi:[10.1016/j.msec.2016.03.114](https://doi.org/10.1016/j.msec.2016.03.114)
  65. Sangsanoh P, Israsena N, Suwanton O, Supaphol P (2017) Effect of the surface topography and chemistry of poly(3-hydroxybutyrate) substrates on cellular behavior of the murine neuroblastoma Neuro2a cell line. *Polym Bull*. doi:[10.1007/s00289-017-1947-9](https://doi.org/10.1007/s00289-017-1947-9)
  66. Chen W, Tong YW (2012) PHBV microspheres as neural tissue engineering scaffold support neuronal cell growth and axon-dendrite polarization. *Acta Biomater* 8:540–548. doi:[10.1016/j.actbio.2011.09.026](https://doi.org/10.1016/j.actbio.2011.09.026)
  67. Grande D, Ramier J, Versace DL, Renard E, Langlois V (2017) Design of functionalized biodegradable PHA-based electrospun scaffolds meant for tissue engineering applications. *New Biotechnol* 37:129–137. doi:[10.1016/j.nbt.2016.05.006](https://doi.org/10.1016/j.nbt.2016.05.006)
  68. Canadas RF, Cavalheiro JMBT, Guerreiro JD, de Almeida MCMD, Pollet E, da Silva CL, da Fonseca MMR, Ferreira FC (2014) Polyhydroxyalkanoates: waste glycerol upgrade into electrospun fibrous scaffolds for stem cells culture. *Int J Biol Macromol* 71:131–140. doi:[10.1177/0885328216639749](https://doi.org/10.1177/0885328216639749)
  69. Su Z, Li P, Wu B, Ma H, Wang Y, Liu G, Wei X (2014) PHBVHHx scaffolds loaded with umbilical cord-derived mesenchymal stem cells or hepatocyte-like cells differentiated from these cells for liver tissue engineering. *Mater Sci Eng C* 45:374–382. doi:[10.1016/j.msec.2014.09.022](https://doi.org/10.1016/j.msec.2014.09.022)
  70. Xu XY, Li XT, Peng SW, Xiao JF, Liu C, Fang G, Chen GQ (2010) The behaviour of neural stem cells on polyhydroxyalkanoate nanofiber scaffolds. *Biomaterials* 31:3967–3975. doi:[10.1016/j.biomaterials.2010.01.132](https://doi.org/10.1016/j.biomaterials.2010.01.132)
  71. Ching KY, Andriotis OG, Li S, Basnett P, Su B, Roy I, Stolz M (2016) Nanofibrous poly (3-hydroxybutyrate)/poly (3-hydroxyoctanoate) scaffolds provide a functional microenvironment for cartilage repair. *J Biomater Appl* 31:77–91. doi:[10.1177/0885328216639749](https://doi.org/10.1177/0885328216639749)
  72. Stock UA, Wiederschain D, Kilroy SM, Shum-Tim D, Khalil PN, Vacanti JP, Mayer JE, Moses MA (2001) Dynamics of extracellular matrix production and turnover in tissue engineered cardiovascular structures. *J Cell Biochem* 81:220–228. doi:[10.1002/1097-4644](https://doi.org/10.1002/1097-4644)
  73. Luklinska ZB, Schluckwerder H (2003) In vivo response to HA-polyhydroxybutyrate/polyhydroxyvalerate composite. *J Microsc* 211:121–129. doi:[10.1046/j.1365-2818.2003.01204.x](https://doi.org/10.1046/j.1365-2818.2003.01204.x)
  74. Shishatskaya EI, Khlusov IA, Volova TG (2006) A hybrid PHB–hydroxyapatite composite for biomedical application: production, in vitro and in vivo investigation. *J Biomater Sci* 17:481–498. doi:[10.1163/156856206776986242](https://doi.org/10.1163/156856206776986242)
  75. Xi J, Zhang L, Zheng ZA, Chen G, Gong Y, Zhao N, Zhang X (2008) Preparation and evaluation of porous poly (3-hydroxybutyrate-co-3-hydroxyhexanoate)—hydroxyapatite composite scaffolds. *J Biomater Appl* 22:293–307. doi:[10.1177/0885328207075425](https://doi.org/10.1177/0885328207075425)
  76. Yucel D, Kose GT, Hasirci V (2010) Polyester based nerve guidance conduit design. *Biomaterials* 31:1596–1603. doi:[10.1016/j.biomaterials.2009.11.013](https://doi.org/10.1016/j.biomaterials.2009.11.013)
  77. Lobler M, Sab M, Kunze C, Schmitz KP, Hopt UT (2002) Biomaterial implants induce the inflammation marker CRP at the site of implantation. *J Biomed Mater Res A* 61:165–167. doi:[10.1002/jbm.10155](https://doi.org/10.1002/jbm.10155)
  78. Kenar H, Kose GT, Hasirci V (2010) Design of a 3D aligned myocardial tissue construct from biodegradable polyesters. *J Mater Sci Mater Med* 21:989–997. doi:[10.1007/s10856-009-3917-8](https://doi.org/10.1007/s10856-009-3917-8)
  79. Volova T, Shishatskaya E, Sevastianov V, Efremov S, Mogilnaya O (2003) Results of biomedical investigations of PHB and PHB/PHV fibers. *Biochem Eng J* 16:125–133. doi:[10.1016/S1369-703X\(03\)00038-X](https://doi.org/10.1016/S1369-703X(03)00038-X)
  80. Valappil SP, Misra SK, Boccaccini AR, Roy I (2006) Biomedical applications of polyhydroxyalkanoates, an overview of animal testing and in vivo responses. *Expert Rev Med Dev* 3:853–868. doi:[10.1586/17434440.3.6.853](https://doi.org/10.1586/17434440.3.6.853)
  81. Romanò CL, Scarponi S, Gallazzi E, Romanò D, Drago L (2015) Antibacterial coating of implants in orthopaedics and trauma: a classification proposal in an evolving panorama. *J Orthop Surg Res* 10:157. doi:[10.1186/s13018-015-0294-5](https://doi.org/10.1186/s13018-015-0294-5)
  82. Ulery BD, Nair LS, Laurencin CT (2011) Biomedical applications of biodegradable polymers. *J Polym Sci Part B Polym Phys* 49:832–864. doi:[10.1002/polb.22259](https://doi.org/10.1002/polb.22259)
  83. Yagmurlu MF, Korkusuz F, Gursel I, Korkusuz P, Ors U, Hasirci V (1999) Sulbactam-cefoperazone polyhydroxybutyrate-co-hydroxyvalerate (PHBV) local antibiotic delivery system: In vivo effectiveness and biocompatibility in the treatment of implant-related experimental osteomyelitis. *J Biomed Mater Res A* 46:494–503
  84. Gursel I, Korkusuz F, Turesin F, Alaeddinoglu NG, Hasirci V (2000) In vivo application of biodegradable controlled antibiotic release systems for the treatment of implant-related osteomyelitis. *Biomaterials* 22:73–80. doi:[10.1016/S0142-9612\(00\)00170-8](https://doi.org/10.1016/S0142-9612(00)00170-8)
  85. Gursel I, Yagmurlu F, Korkusuz F, Hasirci V (2002) In vitro antibiotic release from poly (3-hydroxybutyrate-co-3-hydroxyvalerate) rods. *J Microencap* 19:153–164. doi:[10.1080/02652040110065413](https://doi.org/10.1080/02652040110065413)
  86. Korkusuz F, Korkusuz P, Eksioğlu F, Gursel İ, Hasirci V (2001) In vivo response to biodegradable controlled antibiotic release systems. *J Biomed Mater Res* 55:217–228. doi:[10.1002/\(SICI\)1097-4636](https://doi.org/10.1002/(SICI)1097-4636)
  87. Basnett P, Ching KY, Stolz M, Knowles JC, Boccaccini AR, Smith C, Locke IC, Keshavarz TK, Roy I (2013) Novel poly(3-hydroxyoctanoate)/poly(3-hydroxybutyrate) blends for medical applications. *React Funct Polym* 73:1340–1348. doi:[10.1016/j.reactfunctpolym.2013.03.019](https://doi.org/10.1016/j.reactfunctpolym.2013.03.019)
  88. Gallo J, Holinka M, Moucha CS (2014) Antibacterial surface treatment for orthopaedic implants. *Int J Mol Sci* 15:13849–13880. doi:[10.3390/ijms150813849](https://doi.org/10.3390/ijms150813849)
  89. Kehail AA, Brigham CJ (2017) Anti-biofilm activity of solvent-cast and electrospun polyhydroxyalkanoate membranes treated with lysozyme. *J Polym Environ*. doi:[10.1007/s10924-016-0921-1](https://doi.org/10.1007/s10924-016-0921-1)
  90. Raoga O, Sima L, Chirioiu M, Popescu-Pelin G, Fufă O, Grumezescu O, Socol M, Stănculescu A, Zgură I, Socol G (2017)



- Biocomposite coatings based on poly(3-hydroxybutyrate-co-3-hydroxyvalerate)/calcium phosphates obtained by MAPLE for bone tissue engineering. *Appl Surf Sci*. doi:[10.1016/j.apsusc.2017.01.205](https://doi.org/10.1016/j.apsusc.2017.01.205)
91. Rodríguez-Contreras A, García Y, Manero JM, Rupérez E (2017) Antibacterial PHAs coating for titanium implants. *Eur Polym J*. doi:[10.1016/j.eurpolymj.2017.03.004](https://doi.org/10.1016/j.eurpolymj.2017.03.004)
  92. Hazer DB, Hazer B, Kaymaz F (2009) Synthesis of microbial elastomers based on soybean oily acids. *Biocompatibility studies*. *Biomed Mater* 4:035011. doi:[10.1088/1748-6041/4/3/035011](https://doi.org/10.1088/1748-6041/4/3/035011)
  93. Hazer DB, Hazer B (2011) The effect of gold clusters on the autoxidation of poly(3-hydroxy 10-undecenoate-co-3-hydroxy octanoate) and tissue response evaluation. *J Polym Res* 18:251–262. doi:[10.1007/s10965-010-9413-5](https://doi.org/10.1007/s10965-010-9413-5)
  94. Novikov LN, Novikova LN, Mosahebi A, Wiberg M, Terenghi G, Kellerth JO (2002) A novel biodegradable implant for neuronal rescue and regeneration after spinal cord injury. *Biomaterials* 23:3369–3376. doi:[10.1016/S0142-9612\(02\)00037-6](https://doi.org/10.1016/S0142-9612(02)00037-6)
  95. Tokiwa Y, Calabia BP (2007) Biodegradability and biodegradation of polyesters. *J Polym Environ* 15:259–267. doi:[10.1007/s10924-007-0066-3](https://doi.org/10.1007/s10924-007-0066-3)
  96. Kashiwaya Y, Takeshima T, Mori N, Nakashima K, Clarke K, Veech RL (2000) d-β-Hydroxybutyrate protects neurons in models of Alzheimer's and Parkinson's disease. *Proc Natl Acad Sci USA* 97:5440–5444. doi:[10.1073/pnas.97.10.5440](https://doi.org/10.1073/pnas.97.10.5440)
  97. Zhao YH, Li HM, Qin LF, Wang HH, Chen GQ (2007) Disruption of the polyhydroxyalkanoate synthase gene in *Aeromonas hydrophila* reduces its survival ability under stress conditions. *FEMS Microbiol Lett* 276:34–41. doi:[10.1111/j.1574-6968.2007.00904.x](https://doi.org/10.1111/j.1574-6968.2007.00904.x)
  98. Chen GQ (2011) Biofunctionalization of polymers and their applications. In: *Biofunctionalization of polymers and their applications*. Springer, Berlin, pp 29–45. doi: [10.1007/10\\_2010\\_89](https://doi.org/10.1007/10_2010_89)
  99. Zhang J, Qian C, Shaowu L, Xiaoyun L, Yongxi Z, Ji-Song G, Jin-Chun C, Qiong W, Guo-Qiang C (2013) 3-Hydroxybutyrate methyl ester as a potential drug against Alzheimer's disease via mitochondria protection mechanism. *Biomater* 34:7552–7562. doi:[10.1016/j.biomaterials.2013.06.043](https://doi.org/10.1016/j.biomaterials.2013.06.043)
  100. Camberos-Luna L, Gerónimo-Olvera C, Montiel T, Rincon-Heredia R, Massieu L (2016) The ketone body, β-Hydroxybutyrate stimulates the autophagic flux and prevents neuronal death induced by glucose deprivation in cortical cultured neurons. *Neurochem Res* 41:600–609. doi:[10.1007/s11064-015-1700-4](https://doi.org/10.1007/s11064-015-1700-4)
  101. Cheng S, Chen GQ, Leski M, Zou B, Wang Y, Wu Q (2006) The effect of D,L-β-hydroxybutyric acid on cell death and proliferation in L929 cells. *Biomaterials* 27:3758–3765. doi:[10.1016/j.biomaterials.2006.02.046](https://doi.org/10.1016/j.biomaterials.2006.02.046)
  102. Xiao XQ, Zhao Y, Chen GQ (2007) The effect of 3-hydroxybutyrate and its derivatives on the growth of glial cells. *Biomaterials* 28:3608–3616. doi:[10.1016/j.biomaterials.2007.04.046](https://doi.org/10.1016/j.biomaterials.2007.04.046)
  103. Zou XH, Li HM, Wang S, Leski M, Yao YC, Yang XD, Huang QJ, Chen GQ (2009) The effect of 3-hydroxybutyrate methyl ester on learning and memory in mice. *Biomaterials* 30:1532–1541. doi:[10.1016/j.biomaterials.2008.12.012](https://doi.org/10.1016/j.biomaterials.2008.12.012)
  104. Magdouli S, Brar SK, Blais JF, Tyagi RD (2015) How to direct the fatty acid biosynthesis towards polyhydroxyalkanoates production? *Biomass Bioenerg* 74:268–279. doi:[10.1016/j.biombioe.2014.12.017](https://doi.org/10.1016/j.biombioe.2014.12.017)
  105. Gao X, Chen JC, Wu Q, Chen GQ (2011) Polyhydroxyalkanoates as a source of chemicals, polymers, and biofuels. *Curr Opin Biotechnol* 22:768–774. doi:[10.1128/AEM.01184-06](https://doi.org/10.1128/AEM.01184-06)
  106. Foster LJR, Saufi A, Holden PJ (2001) Environmental concentrations of polyhydroxyalkanoates and their potential as bioindicators of pollution. *Biotechnol Lett* 23:893–898. doi:[10.1023/A:1010528229685](https://doi.org/10.1023/A:1010528229685)
  107. Ray S, Kalia VC (2017) Co-metabolism of substrates by *Bacillus thuringiensis* regulates polyhydroxyalkanoate co-polymer composition. *Bioresour Technol* 224:743–747. doi:[10.1016/j.biortech.2016.11.089](https://doi.org/10.1016/j.biortech.2016.11.089)