

HHS Public Access

Author manuscript *Brain Lang.* Author manuscript; available in PMC 2018 September 01.

Published in final edited form as:

Brain Lang. 2017 September ; 172: 22-29. doi:10.1016/j.bandl.2017.04.008.

Genome-wide association study of language performance in Alzheimer's disease

Kacie D. Deters^{a,b,c}, Kwangsik Nho^{a,b}, Shannon L. Risacher^{a,b}, Sungeun Kim^{a,b}, Vijay K. Ramanan^{a,b}, Paul K. Crane^d, Liana G. Apostolova^{a,b,d,e,f}, Andrew J. Saykin^{a,b,d,e,f,*}, and the Alzheimer's Disease Neuroimaging Initiative[§]

^aCenter for Neuroimaging, Department of Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis, IN, USA

^bIndiana Alzheimer Disease Center, Indiana University School of Medicine, Indianapolis, IN, USA

^cProgram in Medical Neuroscience, Paul and Carole Stark Neurosciences Research Institute, Indiana University School of Medicine, Indianapolis, IN, USA

^dDepartment of Medicine, University of Washington, Seattle, WA, USA

^eDepartment of Neurology, Indiana University School of Medicine, Indianapolis, IN, USA

^fDepartment of Medical and Molecular Genetics, Indiana University School of Medicine, Indianapolis, IN, USA

Abstract

Language impairment is common in prodromal stages of Alzheimer's disease (AD) and progresses over time. However, the genetic architecture underlying language performance is poorly understood. To identify novel genetic variants associated with language performance, we analyzed brain MRI and performed a genome-wide association study (GWAS) using a composite measure of language performance from the Alzheimer's Disease Neuroimaging Initiative (ADNI; n=1,560). The language composite score was associated with brain atrophy on MRI in language and semantic areas. GWAS identified *GLI3* (GLI family zinc finger 3) as significantly associated with language performance ($p < 5 \times 10^{-8}$). Enrichment of GWAS association was identified in pathways related to nervous system development and glutamate receptor function and trafficking. Our results, which warrant further investigation in independent and larger cohorts, implicate *GLI3*, a developmental transcription factor involved in patterning brain structures, as a putative gene associated with language dysfunction in AD.

Conflicts of interest: The authors declare no conflict of interest.

^{*}Corresponding author: Dr. Saykin, 355 W. 16th St., Suite 4100, Center for Neuroimaging, Department of Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis, Indiana 46202, USA. asaykin@iupui.edu. *Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Alzheimer's disease; language performance; GWAS; GLI3; neuroimaging

1. Introduction

Alzheimer's disease (AD), the most common neurodegenerative disorder, is clinically characterized by progressive cognitive impairment primarily in memory in the earliest stages. However, as the disease progresses, deficits can also be observed in other cognitive domains such as judgment, orientation, executive function, visuospatial ability, and language. Language impairment is a prevalent clinical feature that often occurs early on – sometimes even before AD is diagnosed, and progresses during the course of the disease (Ahmed, Haigh, de Jager, & Garrard, 2013). Several language tests have been shown to discriminate between cognitively normal (CN) and those with AD (Bertola et al., 2014; Clark et al., 2014; Gomez & White, 2006; Henry, Crawford, & Phillips, 2004). Hence, language impairment is one of several areas of cognitive decline that can aid in the clinical diagnosis of AD dementia according to the most recent criteria (McKhann et al., 2011). Language deficits during prodromal AD include impairments in word finding (Bayles, Tomoeda, & Trosset, 1990), naming (Hodges, Salmon, & Butters, 1991), and verbal fluency (Monsch et al., 1992; Nutter-Upham et al., 2008).

A large portion of the neuropsychological tests used to assess language impairment, such as the Boston Naming Test (BNT) and Animal Fluency, are dependent on semantic memory (Verma & Howard, 2012). It has been suggested that the language impairment observed in AD is primarily the result of a decline in semantic processing due to structural and organizational deterioration of semantic memory (Chertkow & Bub, 1990). Other memory systems affected include episodic memory, one of the earliest and most severely affected systems in AD (Hodges & Graham, 2001). Language impairments in prodromal and clinical AD have been previously associated with cortical atrophy predominantly in left temporal and parietal lobe regions, in addition to several other brain regions (Ahn et al., 2011; Apostolova et al., 2008: Domoto-Reilly, Sapolsky, Brickhouse, Dickerson, & Alzheimer's Disease Neuroimaging, 2012; Dos Santos et al., 2011). Similarly, there are functional changes on semantic fMRI tasks manifest early stage AD (Saykin et al., 1999). The genetic factors that contribute to different modalities of language function have also been investigated in adolescents and young adults with and without language disorders. For example, genome-wide association studies (GWAS) have identified candidate genes related to reading and language in adolescents to young adults (Gialluisi et al., 2014; Luciano et al., 2013) and the morphology of Heschl's gyrus in young adults (Cai et al., 2014). However, the genetic architecture underlying language performance in older adults with and at risk for AD has not been previously studied. To identify novel genetic variants specifically associated with language performance, we conducted a GWAS as well as a pathway-based analysis using the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort.

2. Material and methods

2.1 Alzheimer's disease neuroimaging initiative (ADNI)

The ADNI was launched in 2003 to help researchers and clinicians develop new treatments for mild cognitive impairment (MCI) and early AD dementia, monitor their effectiveness, and decrease the time and cost of clinical trials. One of ADNI's major goals is to test whether serial magnetic resonance imaging (MRI) (Grundman et al.), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and AD. This multi-year multi-site longitudinal study was started by the National Institute on Aging (NIA), the National Institute of Biomedical Imaging and Bioengineering (NIBIB), the Food and Drug Administration (FDA), private pharmaceutical companies, and non-profit organizations as a \$60 million, 5-year public-private partnership. It was then extended for an additional 7 years through two additional phases of funding. The ADNI participants consist of AD dementia, MCI, and healthy elderly individuals with and without significant memory concern (i.e., SMC and CN, respectively). They were aged 55–90 years and recruited from 59 sites across the U.S. and Canada including medical and academic institutions. Further information can be found at http://www.adni-info.org/ and see (Aisen et al., 2010). The current study involved 1,575 subjects with genotyping: 370 CN, 94 SMC, 283 early MCI (EMCI), 515 late MCI (LMCI), and 313 AD.

2.2 Phenotypes

A composite measure of language was generated as the target phenotype for this study using the following language measures from the ADNI neuropsychological test battery - the BNT, the Animal Fluency test, and the naming portion of AD Assessment Schedule-Cog (ADAS-COG). Given the invariable presence of episodic memory deficits in the cognitively impaired ADNI population, we sought to capture language performance independent of episodic memory performance (Supplementary Table 1). Therefore, each language test (BNT, Animal Fluency, and ADAS-COG naming) was individually adjusted with a linear regression model for concomitant episodic memory deficits using a composite score for episodic memory (Crane et al., 2012) in IBM SPSS (version 22, Armonk, NY). The residuals were then used to generate a composite score using principal component analysis taking the resulting first component, which accounted for 66% of the variance, with the following loadings: animal fluency = 0.62, BNT = 0.84, and ADAS-COG naming = 0.76 (Supplementary Fig. 1). For each diagnostic group, a positive linear relationship was observed between each language measure with respect to episodic memory (Supplementary Table 2). Thus, diagnostic status was not considered in the derivation of the language composite score. This summary measure of language performance was used as the primary phenotype for GWAS. A total of 1,560 subjects (365 CN, 94 SMC, 280 EMCI, 511 LMCI, 310 AD) had all three language test scores available for analyses. Age and sex were significantly associated (p<0.001; age, r = 0.15; sex, r = 0.11) with the generated language composite score pre-adjusted for episodic memory and were used as covariates in the following analyses.

2.3 Structural MRI scans

Baseline 1.5T (ADNI-1) and 3T (ADNI-GO/2) magnetization-prepared rapid gradient-echo (MPRAGE) images were downloaded from the ADNI LONI site for all participants (http:// adni.loni.usc.edu/). Scan processing with voxel-based morphometry (VBM) in Statistical Parametric Mapping 8 (SPM8; Wellcome Trust Centre for Neuroimaging, http:// www.fil.ion.ucl.ac.uk/spm/software/spm8/) and quality control were done as previously described (Risacher et al., 2013). Overall, 71 subjects (17 CN, 5 SMC, 12 EMCI, 21 LMCI, 16 AD) were excluded due to missing MRI scan data or failed processing, leaving 1,489 subjects available for analyses. Analyses were performed separately for each magnetic field strength.

2.4 Genotyping and imputation

The Illumina610-Quad BeadChip was used for genotyping all ADNI-1 participants and the Illumina HumanOmni Express BeadChip or the Illumina Omni 2.5M was used for participants enrolled in ADNI-GO or ADNI-2 (Saykin et al., 2015). Un-genotyped single nucleotide polymorphisms (SNPs) were imputed as previously described (Nho et al., 2015). Since population stratification is known to cause spurious association in disease studies, from the ADNI participants, we restricted our analyses to only non-Hispanic Caucasian subjects that clustered with CEU (Utah residents with Northern and Western European ancestry from the CEPH collection) + TSI (Toscani in Italia) populations using HapMap 3 genotype data and the multidimensional scaling (MDS) analysis (www.hapmap.org). Standard sample and SNP quality control procedures were then implemented; SNPs were excluded if the genotyping call rate was < 95%, Hardy-Weinberg equilibrium test $p < 1 \times$ 10^{-6} , sample call rate < 95%, or the minor allele frequency < 1%. In addition, sample sex marker mismatch check and DNA sample SNP fingerprint to microarray identity checks were also performed. MACH and 1000 Genomes Project data (build 37, hg19) as a reference panel were used for imputation. After quality control steps including filtering for MAF 5% on the imputed SNPs were applied, 1,575 of 1,716 ADNI participants and 5,574,300 SNPs remained for subsequent analyses.

2.5 Statistical analysis

A linear regression model in SPM8 using MRI scans was performed across voxels to evaluate the relationship of the language composite scores (unadjusted and pre-adjusted for episodic memory performance) with grey matter (GM) density. This enabled delineation of the regional associations between the language composite score and structural brain changes. Voxel-wise analysis was also used to determine the anatomical distribution of the association of the most significant SNP identified in the GWAS (*see section below*) with GM density. Voxel-wise analyses included age, sex, years of education, intracranial volume, and apolipoprotein E (*APOE*) e4 status as covariates. Language abilities are typically lateralized to the left hemisphere, however some left handed individuals show right hemisphere or mixed dominance and thus handedness was also included as a covariate (Knecht et al., 2000). Significant results were displayed with a minimum cluster size (k;(Friston, Holmes, Worsley, Poline, & Frackowiak, 1995) of 100 contiguous voxels and a voxel-wise threshold of p < 0.001 (both uncorrected and corrected for multiple comparisons). Anatomical regions

were defined using the x-y-z coordinates for the most significant voxel within each cluster. These coordinates were entered into the Talairach daemon (http://www.talairach.org/daemon.html) to receive the anatomical names for the GM regions closest to that coordinate (Lancaster et al., 2000).

A GWAS across the whole sample (n=1560) was performed using a linear regression under an additive genetic model in PLINK (Purcell et al., 2007) with the language composite preadjusted for episodic memory performance as the endophenotype to specifically target language-specific cognitive function in a population with known memory impairment (Rabin et al., 2009). Age, sex, handedness, years of education, APOE e4 status, and genotyping platform were used as covariates. Bonferroni method was applied for correcting for multiple comparisons and SNPs with $p < 5 \times 10^{-8}$ were considered genome-wide significant (Pe'er, Yelensky, Altshuler, & Daly, 2008). Permutation testing (10⁹ permutations) of the most significant SNP was implemented in PLINK to ensure that the language composite measure had did not deviate from normal distribution. Manhattan and Quantile-Quantile plots were generated in R using the 'qqman' package and regional association plots were generated with LocusZoom (Pruim et al., 2010). Linkage disequilibrium (LD) was estimated and visualized with D' using haploview (http://www.broad.mit.edu/mpg/haploview). The pvalues obtained from the GWAS results and the GSA-SNP software (Nam, Kim, Kim, & Kim, 2010) were then utilized to identify pathways enriched with SNPs showing association to language performance (Ramanan, Kim, et al., 2012). We restricted downstream analysis to pathways containing 5-100 genes and used false discovery rate (FDR) to correct for pathway-level multiple comparisons. Pathway definitions were downloaded from the Molecular Signatures Database (http://www.broadinstitute.org/gsea/msigdb/index.jsp; canonical pathways set, version 4.0).

3. Results

Sample characteristics of the participants used in this study are presented in Table 1. To assess brain regions in which grey matter (GM) density is associated with the generated language composite score pre-adjusted for episodic memory, we used VBM. A significant association between poorer language performance, both adjusted and unadjusted for episodic memory performance, and reduced GM density throughout the brain was observed (Fig. 1 and supplemental Fig. 2–3). Specifically, worse performance on the unadjusted score was associated with reduced GM density in nearly the entire cortex (Supplemental Fig. 2). However, performance on the language composite pre-adjusted for episodic memory performance showed a more localized pattern of association, with poorer performance associated with reduced GM density in the middle and superior temporal gyrus (Brodmann area (BA) 21 and BA22), predominantly lateralized to the left hemisphere (Fig. 1 and supplemental Fig. 3). We observed a larger region significantly associated with language performance in ADNI-GO/2, likely due to the increased sensitivity of the 3T scans to degeneration.

A GWAS was performed to identify genetic variants that might influence language performance using the language composite score pre-adjusted for episodic memory as the phenotype. GWAS identified four SNPs (rs3801203, $p = 3.21 \times 10^{-9}$; rs3779159, $p = 7.12 \times 10^{-9}$)

 10^{-9} ; rs3801206, p = 6.13 × 10⁻⁹; and rs74745318, p = 7.12 × 10⁻⁹; supplemental Table 1) in the region of *GLI3* (GLI family zinc finger 3) that displayed genome-wide significant association with language function (Fig. 2a–b). The genomic inflation factor (λ) was 1.026 indicating no evidence for bias or inflation of our test statistics due to population stratification (Fig. 2c). These SNPs are intronic and demonstrated strong LD (Fig. 3a). To ensure our composite score had a normal distribution, permutation testing of the most significant SNP, rs3801203, resulted in the same p-value of 3.21×10^{-9} suggesting our phenotype had a normal distribution. Each SNP was also tested with BNT, ADAS-COG naming, and animal fluency, individually. All four *GLI3* SNPs were significantly associated with BNT and ADAS-COG naming (p< 1 × 10⁻⁵). The minor allele of each SNP (data not shown) including rs3801203, was associated with worse language performance across all subjects (Fig. 3b) and diagnostic groups with the exception of the smallest group, SMC (Fig. 3c). An additional 31 SNPs in five additional genes displayed suggestive association (p < 5 × 10⁻⁶) for language performance (Supplemental Table 3).

The four SNPs identified in the GWAS are in high LD thus we chose the most significant SNP, rs3801203, to determine if the genetic variants associated with language performance were also associated with GM density. *GLI3* SNP rs3801203 was associated with lower GM density in both the ADNI-1 and ADNI-GO/2 cohorts (Fig. 4). Specifically, in the ADNI-1 cohort, the minor allele of rs3801203 was associated with atrophy in the middle and superior temporal gyrus (BA 21 and BA22, respectively; Fig. 4a), amongst other regions such as the fusiform, cingulate gyrus, and precuneus (Supplemental Table 4). However, in the ADNI-GO/2 cohort, the minor allele of rs3801203 was only found to be associated with atrophy in the inferior frontal gyrus (orbitofrontal cortex; Fig. 4b).

Pathway- enrichment analysis identified 24 pathways associated (FDR p < 0.01) with language performance. The top 20 pathways are presented in Table 2 and include pathways related to nervous system development, such as neuronal migration, axon guidance, and cell differentiation. Additional pathways included glutamate receptor trafficking and function, immune activation, apoptosis, and others.

4. Discussion

We generated a language composite score using measures from animal fluency, BNT, and ADAS-COG naming. In the ADNI sample, MRI analysis identified significant GM atrophy across the whole-brain in relation to the language composite score unadjusted for episodic memory. However, GM atrophy in the left temporal, parietal, and frontal lobes were significantly associated with language performance pre-adjusted for episodic memory. These brain regions have previously been implicated in language and semantic memory processes (Apostolova et al., 2008; Domoto-Reilly et al., 2012; Joubert et al., 2010; McDonald, Bean, & Saykin, 2011; Saykin et al., 1999). We then investigated the effect of genetic variations on this language composite score using GWAS and pathway-enrichment analysis techniques in individuals at risk for and with AD from the ADNI cohort. GWAS identified variants in *GLI3* significantly associated with language impairment. Genetic variation in *GLI3* was also associated with lower GM density predominantly in the temporal lobes. Finally, neuronal development, glutamate receptor trafficking, immune function, and apoptotic pathways

showed enriched associated with language performance in this sample. To our knowledge, this study is the first to identify *GLI3* variation as associated with language performance.

GLI3 encodes one of three GLI zinc finger transcription factors expressed early in development that is normally involved in patterning brain structures (Blaess, Corrales, & Joyner, 2006; Ruppert, Vogelstein, Arheden, & Kinzler, 1990). The GLI3 protein is an important downstream mediator of the Sonic Hedgehog pathway and can act as an activator or repressor in the presence or absence of Sonic Hedgehog, respectively (Wang, Fallon, & Beachy, 2000). Mutations to this pathway are causative for developmental disorders which affect the limbs, head, and face (Kalff-Suske et al., 1999; Kang, Graham, Olney, & Biesecker, 1997).

GLI3 may also have a more direct role in development of the corpus callosum (Amaniti et al., 2013; Magnani et al., 2014) and hippocampus (Hasenpusch-Theil et al., 2012; Palma & Ruiz i Altaba, 2004). One study showed GLI3 expression was downregulated in the presence of Presenilin 1 (*PSEN1*), a protein which forms part of the γ -secretase complex involved in amyloid-beta production, ultimately leading to decreased neuronal differentiation (Paganelli et al., 2001). Notably, mutations in the *PSEN1* gene are causative for some forms of earlyonset autosomal dominant AD, including mutations with an aphasic phenotype (Denvir et al., 2015). Another study showed GLI3 expression is capable of repressing Pitrm1 (pitrilysin metallopeptidase) using limb tissue from GLI3 mutant mice (Town et al., 2009). The Pitrm1 protein is a metalloendopeptidase which is able to degrade amyloid-beta when it accumulates in mitochondria (Falkevall et al., 2006). Further Pitrm1 has been shown to have decreased expression in the temporal lobe of AD subjects (Alikhani et al., 2011), as well as decreased antisense expression in AD subjects compared to controls (Sekar et al., 2015). In light of these studies, our observed findings may suggest that this system could be dysregulated in older adults at risk for AD. Future studies exploring the relationships between GLI3, PSEN1, and Pitrm1 within the brain and in the context of AD would help to elucidate any potential role of this system in AD etiology.

To date, the four *GLI3* SNPs identified in this study have not been linked to any other biological pathway or pathology to our knowledge. We found that these genetic variants were independently associated with BNT and ADAS-COG naming, but not with animal fluency, suggesting the language composite score was more reflective of naming rather than fluency. Furthermore, the minor allele of rs3801203 in the ADNI-1 cohort was associated with auditory processing. However, this finding did not replicate in the ADNI-GO/2 cohort which could be due to differences in scanner strength. This could also be due to the sample characteristics of these two cohorts. The ADNI-1 cohort includes mainly subjects who are later in the disease course and ADNI-GO/2 includes predominantly subjects that are earlier in the disease course. This could account for the larger variation in atrophy in relation to the language composite score seen in ADNI-GO/2. Moreover, the effect of rs3801203 may occur later in the disease course which could account for why more atrophy is observed in the ADNI-1 cohort.

Pathway-based analysis has provided useful insights into the underlying biological processes of complex genetic diseases by integrating and assessing the contribution of multiple genes

and proteins across an intricate network of pathways and subnetworks (Ramanan, Shen, Moore, & Saykin, 2012). This pathway analysis identified several associations between nervous system development pathways and language performance. Specifically, one domain identified in the pathway analysis was glutamate receptor function and trafficking pathways. Glutamate is the primary excitatory neurotransmitter in the brain and is implicated in AD pathophysiology and neuronal cell death. Glutamate receptors have been under investigation as a therapeutic target for AD and other neurological disorders (Caraci et al., 2011; Morin et al., 2013; Olivares et al., 2012; Rosini, Simoni, Minarini, & Melchiorre, 2014). Memantine, a weak N-Methyl-D-aspartate channel blocker, has shown efficacy in reducing clinical decline in moderate AD and is widely used clinically (Reisberg et al., 2003; Tariot et al., 2004). Our results suggest that alterations in glutamate neurotransmission or receptor trafficking may play a critical role in language performance and functional semantic memory in AD.

One notable limitation of this study is the lack of a similar cohort to replicate our findings. Future studies investigating *GL13* in independent cohorts with similar neuroimaging and cognitive assessments should be done to confirm our results. In addition, further investigations about the functional impact of the observed significant *GL13* SNPs on a cellular or molecular level are warranted. Causal directions are yet to be determined but future studies may be able to capture the complicated relationship between the genetic, neuroimaging, and language domains using advanced statistical modeling such as mediation analysis or structural equation modeling. In view of the previously identified association of *GL13* with an amyloid-related protein (e.g.,*PSEN1*) as described above, future studies should also focus on evaluating the impact of *GL13* variation on amyloid phenotypes in older adults at risk for AD.

To our knowledge, this is the first reported GWAS of language performance in older adults at risk for or with AD. Our results identified novel associations of *GLI3*, a developmental transcription factor involved in patterning brain structures, with language dysfunction. Further, pathway analysis identified neuronal development and glutamate receptor pathways as enriched. Future studies will help to fully elucidate the underlying biology and importance of *GLI3* and identified pathways in AD etiology and/or their potential as therapeutic targets.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Data collection and sharing was funded by the ADNI (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: Alzheimer's Association; Alzheimer's Drug Discovery Foundation; BioClinica, Inc.; Biogen Idec Inc.; Bristol- Myers Squibb Company; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; GE Healthcare; Innogenetics, N.V.; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Medpace, Inc.; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Synarc Inc.; and

Takeda Pharmaceutical Company. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (http://www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Disease Cooperative Study at the University of California, San Diego. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

The specific analyses reported here were supported in part by grants from the NIH including P30 AG10133, R01 AG19771, R01 LM011360, R01 AG040770, R01 AG042437, and R00 LM011384, K02 AG048240, as well the Alzheimer's Association, Indiana CTSI (NIH grants U54 RR025761, RR027710-01, and RR020128), NIA grants R36 AG053445 and KO1 AG049050, and NSF grant IIS-1117335.

References

- Ahmed S, Haigh AM, de Jager CA, Garrard P. Connected speech as a marker of disease progression in autopsy-proven Alzheimer's disease. Brain. 2013; 136(Pt 12):3727–3737. DOI: 10.1093/brain/awt269 [PubMed: 24142144]
- Ahn HJ, Seo SW, Chin J, Suh MK, Lee BH, Kim ST, Na DL. The cortical neuroanatomy of neuropsychological deficits in mild cognitive impairment and Alzheimer's disease: a surface-based morphometric analysis. Neuropsychologia. 2011; 49(14):3931–3945. DOI: 10.1016/ j.neuropsychologia.2011.10.010 [PubMed: 22019776]
- Aisen PS, Petersen RC, Donohue MC, Gamst A, Raman R, Thomas RG. Alzheimer's Disease Neuroimaging, I. Clinical Core of the Alzheimer's Disease Neuroimaging Initiative: progress and plans. Alzheimers Dement. 2010; 6(3):239–246. DOI: 10.1016/j.jalz.2010.03.006 [PubMed: 20451872]
- Alikhani N, Guo L, Yan S, Du H, Pinho CM, Chen JX, Yan SS. Decreased proteolytic activity of the mitochondrial amyloid-beta degrading enzyme, PreP peptidasome, in Alzheimer's disease brain mitochondria. J Alzheimers Dis. 2011; 27(1):75–87. DOI: 10.3233/JAD-2011-101716 [PubMed: 21750375]
- Amaniti EM, Hasenpusch-Theil K, Li Z, Magnani D, Kessaris N, Mason JO, Theil T. Gli3 is required in Emx1+ progenitors for the development of the corpus callosum. Dev Biol. 2013; 376(2):113–124. DOI: 10.1016/j.ydbio.2013.02.001 [PubMed: 23396189]
- Apostolova LG, Lu P, Rogers S, Dutton RA, Hayashi KM, Toga AW, Thompson PM. 3D mapping of language networks in clinical and pre-clinical Alzheimer's disease. Brain Lang. 2008; 104(1):33–41. DOI: 10.1016/j.bandl.2007.03.008 [PubMed: 17485107]
- Bayles KA, Tomoeda CK, Trosset MW. Naming and categorical knowledge in Alzheimer's disease: the process of semantic memory deterioration. Brain Lang. 1990; 39(4):498–510. [PubMed: 2076493]
- Bertola L, Mota NB, Copelli M, Rivero T, Diniz BS, Romano-Silva MA, Malloy-Diniz LF. Graph analysis of verbal fluency test discriminate between patients with Alzheimer's disease, mild cognitive impairment and normal elderly controls. Front Aging Neurosci. 2014; 6:185.doi: 10.3389/ fnagi.2014.00185 [PubMed: 25120480]
- Blaess S, Corrales JD, Joyner AL. Sonic hedgehog regulates Gli activator and repressor functions with spatial and temporal precision in the mid/hindbrain region. Development. 2006; 133(9):1799–1809. DOI: 10.1242/dev.02339 [PubMed: 16571630]
- Cai DC, Fonteijn H, Guadalupe T, Zwiers M, Wittfeld K, Teumer A, Hagoort P. A genome-wide search for quantitative trait loci affecting the cortical surface area and thickness of Heschl's gyrus. Genes Brain Behav. 2014; 13(7):675–685. DOI: 10.1111/gbb.12157 [PubMed: 25130324]
- Caraci F, Molinaro G, Battaglia G, Giuffrida ML, Riozzi B, Traficante A, Nicoletti F. Targeting group II metabotropic glutamate (mGlu) receptors for the treatment of psychosis associated with Alzheimer's disease: selective activation of mGlu2 receptors amplifies beta-amyloid toxicity in cultured neurons, whereas dual activation of mGlu2 and mGlu3 receptors is neuroprotective. Mol Pharmacol. 2011; 79(3):618–626. DOI: 10.1124/mol.110.067488 [PubMed: 21159998]
- Chertkow H, Bub D. Semantic memory loss in dementia of Alzheimer's type. What do various measures measure? Brain. 1990; 113(Pt 2):397–417. [PubMed: 2328410]

- Clark DG, Kapur P, Geldmacher DS, Brockington JC, Harrell L, DeRamus TP, Marson DC. Latent information in fluency lists predicts functional decline in persons at risk for Alzheimer disease. Cortex. 2014; 55:202–218. DOI: 10.1016/j.cortex.2013.12.013 [PubMed: 24556551]
- Crane PK, Carle A, Gibbons LE, Insel P, Mackin RS, Gross A. Alzheimer's Disease Neuroimaging, I. Development and assessment of a composite score for memory in the Alzheimer's Disease Neuroimaging Initiative (ADNI). Brain Imaging Behav. 2012; 6(4):502–516. DOI: 10.1007/s11682-012-9186-z [PubMed: 22782295]
- Denvir J, Neitch S, Fan J, Niles RM, Boskovic G, Schreurs BG, Alkon DL. Identification of the PS1 Thr147Ile Variant in a Family with Very Early Onset Dementia and Expressive Aphasia. J Alzheimers Dis. 2015; doi: 10.3233/JAD-150051
- Domoto-Reilly K, Sapolsky D, Brickhouse M, Dickerson BC. Alzheimer's Disease Neuroimaging, I. Naming impairment in Alzheimer's disease is associated with left anterior temporal lobe atrophy. Neuroimage. 2012; 63(1):348–355. DOI: 10.1016/j.neuroimage.2012.06.018 [PubMed: 22728617]
- Dos Santos V, Thomann PA, Wustenberg T, Seidl U, Essig M, Schroder J. Morphological cerebral correlates of CERAD test performance in mild cognitive impairment and Alzheimer's disease. J Alzheimers Dis. 2011; 23(3):411–420. DOI: 10.3233/JAD-2010-100156 [PubMed: 21116054]
- Falkevall A, Alikhani N, Bhushan S, Pavlov PF, Busch K, Johnson KA, Glaser E. Degradation of the amyloid beta-protein by the novel mitochondrial peptidasome, PreP. J Biol Chem. 2006; 281(39): 29096–29104. DOI: 10.1074/jbc.M602532200 [PubMed: 16849325]
- Friston K, Holmes A, Worsley K, Poline J-B, Frackowiak R. Statistical parametric maps in functional imaging: a general linear approach. Hum Brain Mapp. 1995; 2:189–210.
- Gialluisi A, Newbury DF, Wilcutt EG, Olson RK, DeFries JC, Brandler WM, Fisher SE. Genome-wide screening for DNA variants associated with reading and language traits. Genes Brain Behav. 2014; 13(7):686–701. DOI: 10.1111/gbb.12158 [PubMed: 25065397]
- Gomez RG, White DA. Using verbal fluency to detect very mild dementia of the Alzheimer type. Arch Clin Neuropsychol. 2006; 21(8):771–775. DOI: 10.1016/j.acn.2006.06.012 [PubMed: 17011743]
- Grundman M, Petersen RC, Ferris SH, Thomas RG, Aisen PS, Bennett DA. Alzheimer's Disease Cooperative, S. Mild cognitive impairment can be distinguished from Alzheimer disease and normal aging for clinical trials. Arch Neurol. 2004; 61(1):59–66. DOI: 10.1001/archneur.61.1.59 [PubMed: 14732621]
- Hasenpusch-Theil K, Magnani D, Amaniti EM, Han L, Armstrong D, Theil T. Transcriptional analysis of Gli3 mutants identifies Wnt target genes in the developing hippocampus. Cereb Cortex. 2012; 22(12):2878–2893. DOI: 10.1093/cercor/bhr365 [PubMed: 22235033]
- Henry JD, Crawford JR, Phillips LH. Verbal fluency performance in dementia of the Alzheimer's type: a meta-analysis. Neuropsychologia. 2004; 42(9):1212–1222. DOI: 10.1016/j.neuropsychologia. 2004.02.001 [PubMed: 15178173]
- Hodges JR, Graham KS. Episodic memory: insights from semantic dementia. Philos Trans R Soc Lond B Biol Sci. 2001; 356(1413):1423–1434. DOI: 10.1098/rstb.2001.0943 [PubMed: 11571033]
- Hodges JR, Salmon DP, Butters N. The nature of the naming deficit in Alzheimer's and Huntington's disease. Brain. 1991; 114(Pt 4):1547–1558. [PubMed: 1832072]
- Joubert S, Brambati SM, Ansado J, Barbeau EJ, Felician O, Didic M, Kergoat MJ. The cognitive and neural expression of semantic memory impairment in mild cognitive impairment and early Alzheimer's disease. Neuropsychologia. 2010; 48(4):978–988. DOI: 10.1016/j.neuropsychologia. 2009.11.019 [PubMed: 19954747]
- Kalff-Suske M, Wild A, Topp J, Wessling M, Jacobsen EM, Bornholdt D, Grzeschik KH. Point mutations throughout the GLI3 gene cause Greig cephalopolysyndactyly syndrome. Hum Mol Genet. 1999; 8(9):1769–1777. [PubMed: 10441342]
- Kang S, Graham JM Jr, Olney AH, Biesecker LG. GLI3 frameshift mutations cause autosomal dominant Pallister-Hall syndrome. Nat Genet. 1997; 15(3):266–268. DOI: 10.1038/ng0397-266 [PubMed: 9054938]
- Knecht S, Drager B, Deppe M, Bobe L, Lohmann H, Floel A, Henningsen H. Handedness and hemispheric language dominance in healthy humans. Brain. 2000; 123(Pt 12):2512–2518. [PubMed: 11099452]

- Lancaster JL, Woldorff MG, Parsons LM, Liotti M, Freitas CS, Rainey L, Fox PT. Automated Talairach atlas labels for functional brain mapping. Hum Brain Mapp. 2000; 10(3):120–131. [PubMed: 10912591]
- Luciano M, Evans DM, Hansell NK, Medland SE, Montgomery GW, Martin NG, Bates TC. A genome-wide association study for reading and language abilities in two population cohorts. Genes Brain Behav. 2013; 12(6):645–652. DOI: 10.1111/gbb.12053 [PubMed: 23738518]
- Magnani D, Hasenpusch-Theil K, Benadiba C, Yu T, Basson MA, Price DJ, Theil T. Gli3 controls corpus callosum formation by positioning midline guideposts during telencephalic patterning. Cereb Cortex. 2014; 24(1):186–198. DOI: 10.1093/cercor/bhs303 [PubMed: 23042737]
- McDonald, BC., Bean, J., Saykin, AJ. fMRI Wada test: Prospects for presurgical mapping of language and memory. In: Faro, SH., Mohamed, FB., editors. Functional Neuroradiology: Principles and Clinical Applications. Springer; 2011.
- McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, Phelps CH. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011; 7(3):263–269. DOI: 10.1016/j.jalz.2011.03.005 [PubMed: 21514250]
- Monsch AU, Bondi MW, Butters N, Salmon DP, Katzman R, Thal LJ. Comparisons of verbal fluency tasks in the detection of dementia of the Alzheimer type. Arch Neurol. 1992; 49(12):1253–1258. [PubMed: 1449404]
- Morin N, Gregoire L, Morissette M, Desrayaud S, Gomez-Mancilla B, Gasparini F, Di Paolo T. MPEP, an mGlu5 receptor antagonist, reduces the development of L-DOPA-induced motor complications in de novo parkinsonian monkeys: biochemical correlates. Neuropharmacology. 2013; 66:355– 364. DOI: 10.1016/j.neuropharm.2012.07.036 [PubMed: 22884464]
- Nam D, Kim J, Kim SY, Kim S. GSA-SNP: a general approach for gene set analysis of polymorphisms. Nucleic Acids Res. 2010; 38(Web Server issue):W749–754. DOI: 10.1093/nar/ gkq428 [PubMed: 20501604]
- Nho K, Ramanan VK, Horgusluoglu E, Kim S, Inlow MH, Risacher SL, Saykin AJ. Comprehensive Gene- and Pathway-Based Analysis of Depressive Symptoms in Older Adults. J Alzheimers Dis. 2015; doi: 10.3233/JAD-148009
- Nutter-Upham KE, Saykin AJ, Rabin LA, Roth RM, Wishart HA, Pare N, Flashman LA. Verbal fluency performance in amnestic MCI and older adults with cognitive complaints. Arch Clin Neuropsychol. 2008; 23(3):229–241. DOI: 10.1016/j.acn.2008.01.005 [PubMed: 18339515]
- Olivares D, Deshpande VK, Shi Y, Lahiri DK, Greig NH, Rogers JT, Huang X. N-methyl D-aspartate (NMDA) receptor antagonists and memantine treatment for Alzheimer's disease, vascular dementia and Parkinson's disease. Curr Alzheimer Res. 2012; 9(6):746–758. [PubMed: 21875407]
- Paganelli AR, Ocana OH, Prat MI, Franco PG, Lopez SL, Morelli L, Carrasco AE. The Alzheimerrelated gene presenilin-1 facilitates sonic hedgehog expression in Xenopus primary neurogenesis. Mech Dev. 2001; 107(1-2):119–131. [PubMed: 11520668]
- Palma V, Ruiz i Altaba A. Hedgehog-GLI signaling regulates the behavior of cells with stem cell properties in the developing neocortex. Development. 2004; 131(2):337–345. DOI: 10.1242/dev. 00930 [PubMed: 14681189]
- Pe'er I, Yelensky R, Altshuler D, Daly MJ. Estimation of the multiple testing burden for genomewide association studies of nearly all common variants. Genet Epidemiol. 2008; 32(4):381–385. DOI: 10.1002/gepi.20303 [PubMed: 18348202]
- Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, Willer CJ. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics. 2010; 26(18):2336–2337. DOI: 10.1093/bioinformatics/btq419 [PubMed: 20634204]
- Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Sham PC. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet. 2007; 81(3):559–575. DOI: 10.1086/519795 [PubMed: 17701901]
- Rabin LA, Saykin AJ, West JD, Borgos MJ, Wishart HA, Nutter-Upham KE, Santulli RB. Judgment in Older Adults with Normal Cognition, Cognitive Complaints, MCI, and Mild AD: Relation to Regional Frontal Gray Matter. Brain Imaging Behav. 2009; 3(2):212–219. DOI: 10.1007/ s11682-009-9063-6 [PubMed: 24741381]

- Ramanan VK, Kim S, Holohan K, Shen L, Nho K, Risacher SL. Alzheimer's Disease Neuroimaging, I. Genome-wide pathway analysis of memory impairment in the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort implicates gene candidates, canonical pathways, and networks. Brain Imaging Behav. 2012; 6(4):634–648. DOI: 10.1007/s11682-012-9196-x [PubMed: 22865056]
- Ramanan VK, Shen L, Moore JH, Saykin AJ. Pathway analysis of genomic data: concepts, methods, and prospects for future development. Trends Genet. 2012; 28(7):323–332. DOI: 10.1016/j.tig. 2012.03.004 [PubMed: 22480918]
- Reisberg B, Doody R, Stoffler A, Schmitt F, Ferris S, Mobius HJ, Memantine Study G. Memantine in moderate-to-severe Alzheimer's disease. N Engl J Med. 2003; 348(14):1333–1341. DOI: 10.1056/ NEJMoa013128 [PubMed: 12672860]
- Risacher SL, Kim S, Shen L, Nho K, Foroud T, Green RC. Alzheimer's Disease Neuroimaging Initiative, d. The role of apolipoprotein E (APOE) genotype in early mild cognitive impairment (E-MCI). Front Aging Neurosci. 2013; 5:11.doi: 10.3389/fnagi.2013.00011 [PubMed: 23554593]
- Rosini M, Simoni E, Minarini A, Melchiorre C. Multi-target design strategies in the context of Alzheimer's disease: acetylcholinesterase inhibition and NMDA receptor antagonism as the driving forces. Neurochem Res. 2014; 39(10):1914–1923. DOI: 10.1007/s11064-014-1250-1 [PubMed: 24493627]
- Ruppert JM, Vogelstein B, Arheden K, Kinzler KW. GLI3 encodes a 190-kilodalton protein with multiple regions of GLI similarity. Mol Cell Biol. 1990; 10(10):5408–5415. [PubMed: 2118997]
- Saykin AJ, Flashman LA, Frutiger SA, Johnson SC, Mamourian AC, Moritz CH, Weaver JB. Neuroanatomic substrates of semantic memory impairment in Alzheimer's disease: patterns of functional MRI activation. J Int Neuropsychol Soc. 1999; 5(5):377–392. [PubMed: 10439584]
- Saykin AJ, Shen L, Yao X, Kim S, Nho K, Risacher SL. Alzheimer's Disease Neuroimaging, I. Genetic studies of quantitative MCI and AD phenotypes in ADNI: Progress, opportunities, and plans. Alzheimers Dement. 2015; 11(7):792–814. DOI: 10.1016/j.jalz.2015.05.009 [PubMed: 26194313]
- Sekar S, McDonald J, Cuyugan L, Aldrich J, Kurdoglu A, Adkins J, Liang WS. Alzheimer's disease is associated with altered expression of genes involved in immune response and mitochondrial processes in astrocytes. Neurobiol Aging. 2015; 36(2):583–591. DOI: 10.1016/j.neurobiolaging. 2014.09.027 [PubMed: 25448601]
- Tariot PN, Farlow MR, Grossberg GT, Graham SM, McDonald S, Gergel I, Memantine Study G. Memantine treatment in patients with moderate to severe Alzheimer disease already receiving donepezil: a randomized controlled trial. JAMA. 2004; 291(3):317–324. DOI: 10.1001/jama. 291.3.317 [PubMed: 14734594]
- Town L, McGlinn E, Fiorenza S, Metzis V, Butterfield NC, Richman JM, Wicking C. The metalloendopeptidase gene Pitrm1 is regulated by hedgehog signaling in the developing mouse limb and is expressed in muscle progenitors. Dev Dyn. 2009; 238(12):3175–3184. DOI: 10.1002/ dvdy.22126 [PubMed: 19877269]
- Verma M, Howard RJ. Semantic memory and language dysfunction in early Alzheimer's disease: a review. Int J Geriatr Psychiatry. 2012; 27(12):1209–1217. DOI: 10.1002/gps.3766 [PubMed: 22298328]
- Wang B, Fallon JF, Beachy PA. Hedgehog-regulated processing of Gli3 produces an anterior/posterior repressor gradient in the developing vertebrate limb. Cell. 2000; 100(4):423–434. [PubMed: 10693759]

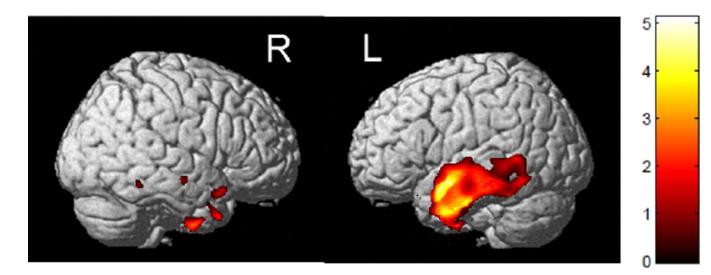
Author Manuscript

Highlights

• We identified novel genetic variants associated with language performance.

- Minor allele variants in *GLI3* are associated with worse language performance.
- Anatomical changes in language regions associated with language composite score.
- Developmental and glutamate pathways were related to lower language performance.

(a) ADNI-1



(b) ADNI-GO/2

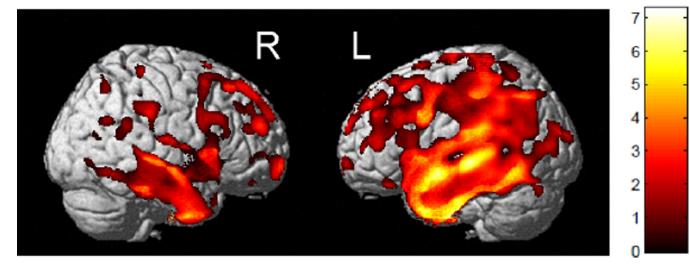


Figure 1.

Global regions associated with the language composite score pre-adjusted for episodic memory performance in ADNI participants. Grey matter density was positively correlated with the language composite score in the temporal lobe and language areas for both ADNI-1 (a) and ADNI-GO/2 (b). Covariates included age, sex, intracranial volume, *APOE* e4 status, and handedness. Results are displayed at p<0.001 (uncorrected) and at a threshold (k) of 100 voxels.

Deters et al.

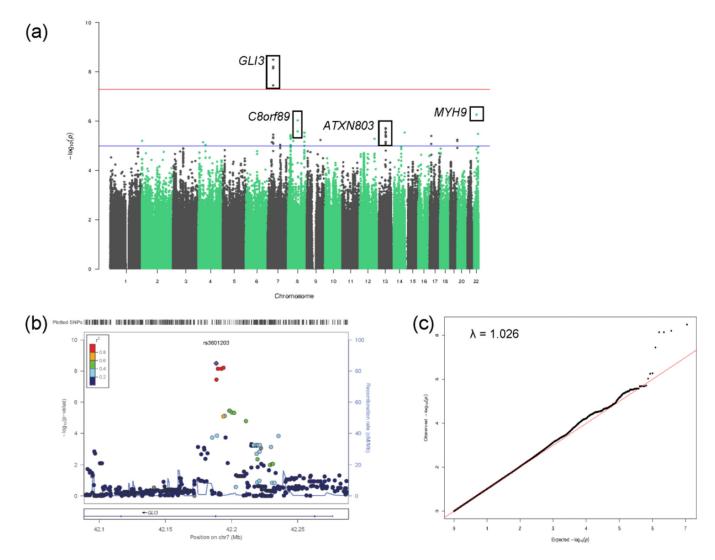


Figure 2.

Manhattan, regional association, and quantile-quantile plots for GWAS of language performance. (a) Observed $-\log^{10}$ p-values (y-axis) are displayed for all tested SNPs on each chromosome (x-axis). A SNP was considered genome-wide significant if $p < 5 \times 10^{-8}$ (above red line). Suggestive SNP associations were identified as those reaching genome-wide significance of $p < 1 \times 10^{-5}$ (above blue line). (b) Regional association plot showing the region around the most significant SNP in the GWAS. The SNPs within 500 kb of rs3801203 are plotted as their GWAS $-\log^{10} P$ -values against their NCBI 37 genomic position. The blue line indicates recombination rates estimated from the 1000 Genomes Project reference data. The color scale of r^2 values is used to label SNPs based on their correlation with rs3801203. (c) The genomic inflation factor (λ) was 1.026 suggesting no population stratification effect.

Deters et al.

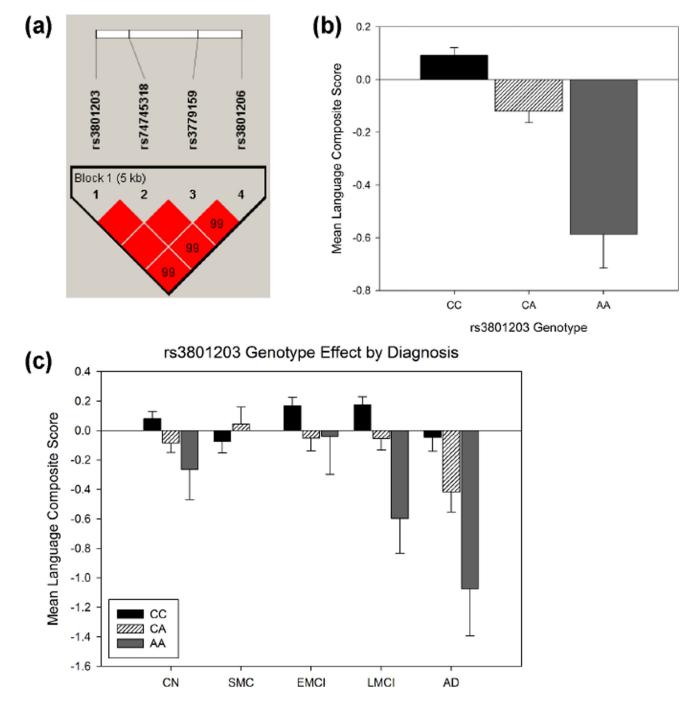
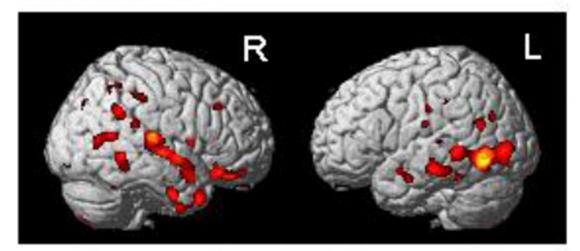


Figure 3.

GLI3 SNPs are associated with lower language performance. (a) Linkage disequilibrium map of the four *GLI3* SNPs significantly associated with language performance. The minor allele of rs3801203 (b-c, $p = 3.21 \times 10^{-9}$) was associated with lower language composite score in older adults at risk for or with AD, with the exception of the SMC group. Means adjusted for age, sex, years of education, genotyping platform, handedness, and *APOE* ϵ 4 status. Standard errors are displayed.

CN = cognitively normal; SMC = significant memory concern; EMCI = early mild cognitive impairment; LMCI = late mild cognitive impairment; AD = Alzheimer's disease



(b) ADNI-GO/2

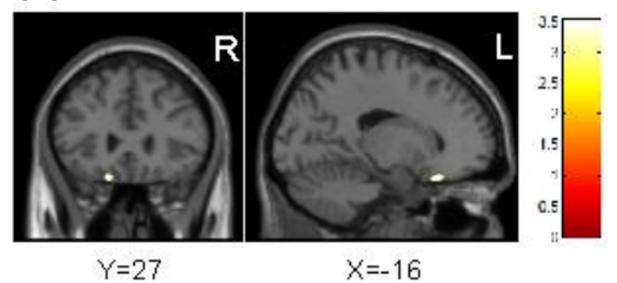


Figure 4.

The minor allele of *GLI3* SNP rs3801203 is associated with lower grey matter density in the temporal cortex of ADNI-1 participants (a) and the orbitofrontal cortex for ADNI-GO/2 participants (b). Covariates included age, sex, intracranial volume, *APOE* ϵ 4 status, and handedness. Results are displayed at p<0.001 (uncorrected) and at a threshold (k) of 100 voxels.

Baseline demographic characteristics.

Baseline Diagnosis	CN	SMC	EMCI	LMCI	AD
No. of Subjects	365	94	280	511	310
Male/Female	192/173	40/54	160/120	317/194	175/135
Baseline Years of Age (SD)	74.6 (5.6)	71.8 (5.7)	71.1 (7.4)	74.6 (5.6) 71.8 (5.7) 71.1 (7.4) 73.5 (7.7)	74.7 (7.8)
Baseline Years of Education (SD) 16.3 (2.6) 16.8 (2.6) 16.1 (2.7) 16 (2.9)	16.3 (2.6)	16.8 (2.6)	16.1 (2.7)	16 (2.9)	15.2 (3)
Right/Left Handed	338/27	82/12	251/29	457/54	286/24
APOE (e4-/e4+)	266/99	62/32	160/120	231/280	105/205
Lang. Comp. Mean $(SD)^{*}$	0.6 (0.6)	0.56 (0.5)	0.32 (0.7)	0.6(0.6) 0.56(0.5) 0.32(0.7) -0.14(0.9) -0.94(1.2)	-0.94 (1.2)

CN = cognitively normal; SMC = significant memory concern; EMCI = early mild cognitive impairment; LMCI = late mild cognitive impairment; AD = Alzheimer's disease; SD = standard deviation

* Not adjusted for memory composite score.

Table 2

Top 20 pathways associated with language performance.

Pathway description (Source database)	Size ^a	Uncorrected p ^b
Netrin-1 signaling (Reactome)	36	2.38E-07
L1CAM interactions (Reactome)	76	9.39E-07
Arrythmogenic right ventricular cardiomyopathy (KEGG)	71	9.78E-07
Ion channel transport (Reactome)	48	5.33E-06
SHH pathway (Biocarta)	15	6.81E-06
Phosphatidylinositol signaling system (KEGG)	75	9.84E-06
Trafficking of AMPA receptors (Reactome)	26	1.17E-05
NRAGE signals death through JNK (Reactome)	37	1.39E-05
Adherens junction (KEGG)	71	1.39E-05
Synthesis of PIPs at the plasma membrane (Reactome)	28	1.59E-05
Integrin cell surface reactions (Reactome)	74	1.72E-05
FC gamma R-mediated phagocytosis (KEGG)	90	4.91E-05
ECM-receptor interaction (KEGG)	81	5.64E-05
Cell death signaling via NGRAGE, NRIF, and NADE (Reactome)	51	7.69E-05
Semaphorin interactions (Reactome)	61	9.89E-05
Hypertrophic cardiomyopathy-HCM (KEGG)	79	1.15E-04
Unblocking of NMDA receptor glutamate binding and activation (Reactome)	14	1.53E-04
Interaction between L1 and Ankyrins (Reactome)	20	1.57E-04
PI metabolism (Reactome)	44	1.59E-04
Glioma (KEGG)	63	1.73E-04

^aNumber of genes in the pathway.

 $^b\mathrm{All}$ pathway displayed are significant at false discovery rate (FDR)-corrected p < 0.01.