

Changes to International Nonproprietary Names for antibody therapeutics 2017 and beyond: of mice, men and more

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ABSTRACT

Active pharmaceutical substances require an International Nonproprietary Name (INN) assigned by the World Health Organization (WHO) to obtain market authorization as a medicinal product. INNs are selected to represent a unique, generic name for a drug enabling unambiguous identification by stakeholders worldwide. INNs may be requested after initiating clinical development of an investigational drug. Pharmaceutical classes are indicated by a common stem or suffix. Currently, INNs for monoclonal antibody-based drugs are recognized by the suffix, -mab, preceded by a source infix such as -xi- (chimeric), -zu- (humanized) or -u- (human) designating the species from which the antibody was derived. However, many technological advances have made it increasingly difficult to accurately capture an antibody's source in its name. In 2014, the WHO and the United States Adopted Names (USAN) Council approached this challenge by implementing changes to antibody source infix definitions. Unfortunately, gaps and ambiguities in the definitions and procedures resulted in inconsistent source category assignments and widespread confusion. The Antibody Society, extensively supported by academic and industry scientists, voiced concerns leading to constructive dialog during scheduled consultations with WHO and USAN Council representatives. In June 2017, the WHO announced that use of the source infix will be discontinued for new antibody INNs effective immediately. We fully support this change as it better aligns antibody INNs with current and foreseeable future innovations in antibody therapeutics. Here we review the changes implemented. Additionally, we analyzed antibody INNs recently assigned under the previous 2014 definitions and provide recommendations for further alignment.

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Introduction

“The best laid schemes o’ mice an’ men / Gang aft a-gley.” This line of the well-known poem by Robert Burns¹ eloquently expresses the notion that things, even though carefully planned, can often go wrong. In fact, this is what happened with well-intended changes to the definitions used to assign the source infix (substem) for (generic) INN and USAN for antibody therapeutics (see appendix for details). Specific concerns with respect to the changes in the INN and USAN source designations have previously been discussed in detail elsewhere.²

Contemporary INNs lack transparency and consistency in source infix designations

The WHO and the USAN Council are not, to our knowledge, planning to change recommended INNs previously issued under the 2014 definitions. Therefore, we considered it important to reinvestigate nomenclature practices for contemporary INNs to identify and highlight shortcomings. We systematically analyzed all INN for antibody therapeutics as they occur in the most recent 2017 INNs Recommended List (RL77).³ The results summarized in Table 1 reinforce our previous

concerns,^{2,4} and show that multiple inconsistencies occur for antibodies with a chimeric or humanized source designation. The four antibodies at the top of Table 1 all received a chimeric (-xi-) or mixed (-xizu-) INN designation; the latter referring to antibodies containing both a chimeric and a humanized heavy or light chain. Dinutuximab beta is based on mouse variable (V) domains fused to human constant (C) domains and therefore represents a genuine chimeric antibody generated via classic domain exchange.⁵ The other three antibodies categorized as chimeras or mixed by their INNs, in contrast, were humanized using common methods.^{6–8} The chimeric designation of andecaliximab highlights the drawbacks of using linear sequence homology to categorize therapeutic antibodies by source. For this antibody, a humanization procedure was used that employs non-contiguous human framework regions (FRs) aimed at minimizing T cell epitope content and reducing immunogenicity risks.^{6,9} Next, rozanolixizumab represents the INN with the least transparent source designation in RL77. Despite its mixed source -xizu- designation and its annotation as a “humanized and chimeric antibody,” the INN description (i.e. additional information published by WHO),³ only shows alignments of rozanolixizumab’s variable region sequences

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Table 1. Antibody names in Recommended INN: List 77.

Antibody INN	Common name	Heavy chain VH ^a		Light chain VL ^a		Notes	Refs
		Top hit species	Homology (%)	Top hit species	Homology (%)		
dinutuximab beta	Ch14.18	<i>Mus musculus</i>	83.3	<i>Mus musculus</i>	87	Chimeric mouse Ab	5
andecaliximab	GS-5745	<i>Mus musculus</i>	82.5	<i>Mus musculus</i>	80	Humanized by Antitope's Composite Human Antibody technology	6
depatuximab mefadotin	ABT-806	<i>Homo sapiens</i>	84.5	<i>Mus musculus</i>	86.3	Humanized mouse Ab; <i>Antibody-drug conjugate</i>	7
rozanolixizumab	UCB-7665	<i>Homo sapiens</i>	86.5	<i>Homo sapiens</i>	76	Humanized rat Ab	8
camrelizumab	SHR-1210	<i>Homo sapiens</i>	90.8	<i>Homo sapiens</i>	87.4	Humanized mouse Ab	36
crizalimumab	SeIG1	<i>Homo sapiens</i>	81.6	<i>Homo sapiens</i>	86.9	Humanized mouse Ab	37
daclizumab beta	DAC HYP	<i>Homo sapiens</i>	82.7	<i>Homo sapiens</i>	84	Humanized mouse Ab	19
dezamizumab	GSK-2398852	<i>Homo sapiens</i>	85.7	<i>Homo sapiens</i>	85.3	Humanized mouse Ab	38
eptinezumab	ALD-403	<i>Homo sapiens</i>	81.4	<i>Homo sapiens</i>	86.2	Humanized rabbit Ab	39
fremanezumab	7E9	<i>Homo sapiens</i>	85.7	<i>Homo sapiens</i>	85.3	Humanized mouse Ab	40
gemtuzumab ozogamicin	CDP-771	<i>Homo sapiens</i>	72.9	<i>Homo sapiens</i>	81.8	Humanized mouse Ab; <i>Antibody-drug conjugate</i>	41
ifabotuzumab	KB004, IIIA4	<i>Homo sapiens</i>	91.8	<i>Homo sapiens</i>	91.6	Humanized mouse Ab (Kalobios Humanengineering)	42
lutikizumab	ABT-981, SK48-E26, X3	<i>Homo sapiens</i>	81.6 (Fab 1) 88.8 (Fab 2)	<i>Homo sapiens</i>	82.1 (Fab 1) 92.6 (Fab 2)	Humanized mouse Fab Human Fab, cloned from B cells from a patient with autoimmune disease; <i>DVD bispecific antibody</i>	15 16
rosmantuzumab	OMP-131R10	<i>Homo sapiens</i>	84.5	<i>Homo sapiens</i>	83.8	Humanized mouse Ab	43
sacituzumab	hRS7	<i>Homo sapiens</i>	85.7	<i>Homo sapiens</i>	82.2	Humanized mouse Ab	44
telisotuzumab vedotin	ABBY-399, ABT-700	<i>Homo sapiens</i>	92.9	<i>Homo sapiens</i>	85.1	Humanized mouse Ab; <i>Antibody-drug conjugate</i>	45, 46
trastuzumab duocarmazine	SYD985, 4D5-8	<i>Homo sapiens</i>	81.6	<i>Homo sapiens</i>	86.3	Humanized mouse Ab; <i>Antibody-drug conjugate</i>	47
vunakizumab	SHR-1314	<i>Homo sapiens</i>	82.7	<i>Homo sapiens</i>	80	Humanized mouse Ab	48
aprutumab ixadotin	BAY 1179470	<i>Homo sapiens</i>	98	<i>Homo sapiens</i>	90.7	Human Biolnvent n-CoDeR phage display library; <i>Antibody-drug conjugate</i>	49
burosomab	KRN-23, UX-023	<i>Homo sapiens</i>	94.9	<i>Homo sapiens</i>	97.9	Kirin-Medarex KM mouse platform	50
brazikumab	AMG-139, MEDI2070	<i>Homo sapiens</i>	99	<i>Homo sapiens</i>	96	Abgenix Xenomouse platform	51, 52
elezanumab	ABT-555, AE12-1Y	<i>Homo sapiens</i>	92.9	<i>Homo sapiens</i>	89.9	Phyllos Profusion mRNA display	53
lupartumab amadotin	BAY 1129980	<i>Homo sapiens</i>	92.9	<i>Homo sapiens</i>	87.9	Human Biolnvent n-CoDeR phage display; <i>Antibody-drug conjugate</i>	54
remtolumab	D2E7, A-1230717, ABT-122	<i>Homo sapiens</i>	93.9 (Fab1) 85.7 (Fab 2)	<i>Homo sapiens</i>	95.8 (Fab 1) 90.5 (Fab 2)	Guided selection phage display	55, 56
suptavumab	REGN 2222, SAR438584	<i>Homo sapiens</i>	87.9	<i>Homo sapiens</i>	94.7	Phyllos Profusion mRNA display; <i>DVD bispecific antibody</i>	57
utomilumab	PF-05082566	<i>Homo sapiens</i>	94.9	<i>Homo sapiens</i>	90	Regeneron VelocImmune mice	58
ranevetmab	INV-01	<i>Homo sapiens</i>	74.2	<i>Rattus norvegicus</i>	76.8	Morphosys HuCal phage library Veterinary use Rat Ab caninized by Nexvet PETization	59

^aAntibody INNs for which a different top hit relative to the species stated in RL77 was found are highlighted in red.

with *Homo sapiens* VH and VL reference genes with 86.5 and 76% homology, respectively (ref. 3 and Table 1). The IMGT/DomainGapAlign searches saved for rozanolixizumab in IMGT/mAb-DB^{10,11} also exclusively show the same human reference genes. So if the reference genes are human, then what is the mixed source designation based on? Notably, rerunning the IMGT reference gene database search as described in Methods reveals that the light chain aligns more closely to macaque VL genes with 7 *Macaca mulatta* reference alleles showing slightly greater homology compared with the first human VL hit (i.e., 77–79% versus 76%, respectively; Table 2). Differences are subtle, however, with rozanolixizumab VL showing 20 amino acid changes in FR1- FR3 compared with both the top macaque (i.e., IGKV1-13*01 with 9 changes in FR1-3 and 11 in complementary-determining region (CDR)1-2) as well as the top human (i.e., IGKV1-17*01 with 8 changes in FR1-3 and 12 in CDR1-2) reference allele hit in IMGT/DomainGapAlign. The lower score for the human references allele can therefore be attributed to a slightly greater dissimilarity of the CDR-L3 which, in fact, was grafted from the parental rat antibody during humanization. Rozanolixizumab's light chain, therefore, reasonably should also have obtained a humanized source designation. Unexpectedly, it was assigned a mixed chimeric stem instead which, in addition, is at odds with the documentation provided by WHO about this particular INN.³ Multiple inconsistencies were also found for 10 of 14 antibodies with a humanized INN designation upon redoing the IMGT/DomainGapAlign search (Table 2). In most cases, we obtained a macaque reference gene as a top hit instead of the human gene listed, although in 2 instances mouse reference genes were also found.

There may be multiple explanations for the observed inconsistencies. First, there is no precise definition of what constitutes “closer to human than to other species” in the reference database search. Presumably, an INN examiner may determine that a sequence aligns most closely to a human reference gene even if non-human genes score slightly higher but the observed homology is of similar magnitude (see appendix). This may be of particular relevance if the result is affected by differential

alignment in CDR sequences. Macaque reference genes, for example, may obtain a higher IMGT/DomainGapAlign score through a closer homology with rodent CDRs or, due to an artifact of the local alignment algorithm by having no identity or similarity in the V gene termini (CDR3 and FR3), even though alignment for the corresponding region in the human counterpart is better.² However, such subjectivity in assigning an appropriate source category is highly problematic as discussed above (Table 1). Second, results may vary over time as the composition of the IMGT gene reference database changes due to additional genome sequences becoming available in which specifically the addition of macaque germline reference genes with high allelic variation is of concern.² Finally, inconsistencies will occur when an INN for an antibody contained in a novel compound was issued before the 2014 change in source definitions. This is exemplified by gemtuzumab ozogamicin. The INN for the antibody portion of this antibody-drug conjugate (ADC) was issued in 2001 without its sequence being made available.¹² In fact, sequences have only been systematically disclosed in the INN description since RL57 released in 2007.¹³ The subjectivity and time-dependence of antibody INNs creates undesired uncertainties with respect to predicting and interpreting INN source categories.

As noted, the USAN definition differs from INN by using an 85% sequence cut-off definition for distinguishing chimeric from humanized antibodies (appendix). When using the USAN definition, only 8 of 19 antibody heavy chains and 9 of 19 light chains would have obtained a humanized designation. Interestingly, rozanolixizumab's nomenclature would be consistent with USAN's definitions for a mixed source antibody. This antibody however has not been assigned a USAN,¹⁴ so the pre-existence of a USAN cannot explain the discrepancies described.

Examining the 8 human antibodies in RL77, we observe no discrepancies, which is as expected due to the absence of a definition (Fig. 1; appendix). Ironically, the only antibody in RL77 derived from an immune response in a human individual (i.e., the second Fab in the bispecific DVD lutikizumab was cloned from a patient with auto-immune disease^{15,16}) did not earn the antibody a (mixed) human INN

Table 2. Inconsistencies in source infix designations analyzed for antibodies from RL77.

Antibody INN	Heavy chain			Light chain		
	Top hit species (searched May 2017) ^a	Homology to top hit species (%)	If different, homology to closest human (%)	Top hit species (searched May 2017) ^a	Homology to top hit species (%)	If different, homology to closest human (%)
andecaliximab	<i>Mus musculus</i>	82.5	71.1	<i>Macaca mulatta</i>	81.1	80
rozanolixizumab	<i>Homo sapiens</i>	86.5		<i>Macaca mulatta</i>	79.0 ^b	76
camrelizumab	<i>Homo sapiens</i>	90.8		<i>Macaca mulatta</i>	87.6	87.4
crizanlizumab	<i>Homo sapiens</i>	81.6		<i>Macaca mulatta</i>	87.1	86.9
daclizumab beta	<i>Macaca fascicularis</i>	83.7	82.7	<i>Homo sapiens</i>	84.0	
dezamizumab	<i>Homo sapiens</i>	85.7		<i>Macaca mulatta</i>	86.3	85.3
gemtuzumab ozogamicin	<i>Mus musculus</i>	77.1	72.9	<i>Homo sapiens</i>	81.9	
lutikizumab	Fab 1- <i>Homo sapiens</i>	81.6		<i>Macaca mulatta</i>	82.4	82.1
	Fab 2- <i>Homo sapiens</i>	88.8		<i>Macaca mulatta</i>	94.4	92.6
rosmantuzumab	<i>Homo sapiens</i>	84.5		<i>Macaca mulatta</i>	83.9	83.8
sacituzumab	<i>Homo sapiens</i>	85.7		<i>Mus musculus</i>	83.2	82.2
trastuzumab duocarmazine	<i>Homo sapiens</i>	81.6		<i>Macaca mulatta</i>	86.5	86.3
vunakizumab	<i>Macaca fascicularis</i>	84.7	82.7	<i>Mus musculus</i>	81.9	80

^aResults that differ from the closest reference gene or allele species shown in RL77 and as referenced in the IMGT/mAb-DB INN database are shown in red. New search was performed as stated in Methods. The homology to the closest human reference gene or allele is provided.

^bSearch provides 7 *Macaca mulatta* reference alleles that have greater homology than the *Homo sapiens* reference allele.

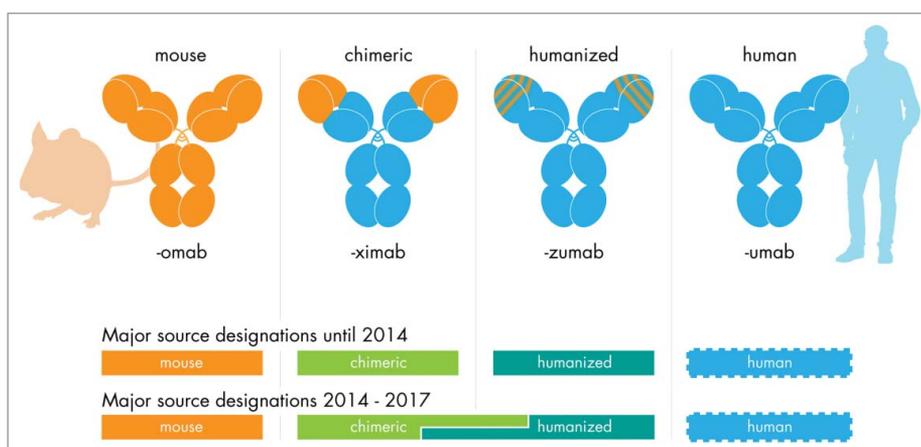


Figure 1. The INN source substem for therapeutic antibodies. Antibody INNs issued until June 2017 (with the exception of the first antibody INN muromonab-CD3) contain a source infix designating the species. The antibody's origin determined the source infix until 2014. For antibody INN issued between 2014 and early 2017, the source infix was determined using a sequence alignment procedure, which led to inconsistent source infix designations for chimeric and humanized antibodies. No definitions to determine a human source existed (see appendix).

designation. The wide range of technologies used to access human sequences for generating therapeutic antibodies is noteworthy. These technologies include mRNA-display, multiple distinct phage-display and several distinct transgenic mice platforms (Table 1), and exemplify that therapeutic antibodies can have many origins (Fig. 2). Additional similar technologies, often used in conjunction, are being used to fill early pharmaceutical development pipelines. INNs requests for such antibodies can be expected for submission in the near future.

In summary, therapeutic antibody INNs as well as the accompanying description published in the INN Recommended List RL77 lack consistency and transparency in source infix designations for chimeric and humanized antibodies.

Fixing the source infix

On behalf of its members and scientists who signed an online petition, The Antibody Society engaged in discussions with the WHO INN expert group and representatives from the USAN Council and Food and Drug Administration (FDA) during the 62nd open consultation on INN for Pharmaceutical Substances in April 2016 and an *ad hoc* meeting on Biologicals in September 2016.^{4,17} The Antibody Society, in collaboration with key stakeholders, developed proposals to revise the INN system to provide scientifically sound, distinguishing names for therapeutic antibodies in current and future development.

Two potential solutions were discussed. The first was to drop the use of the source infix and sequence alignments to categorize antibodies altogether. The second was to improve the current system, for example by generating a new expanded 'engineered' source infix which should take current and future developments in antibody generation technologies into account. Although no general consensus was reached at the workshop, dropping the source infix was a favored solution.¹⁷ Removing the source infix would, as a side effect, create more flexibility in the assignment of INNs. This is important as it was noted that, due the large increase in applications for biologicals, it is becoming increasingly difficult to design new distinguishable INNs.¹⁷

After considering all options, the WHO announced it was decided at the 64th consultation on INN for Pharmaceutical Substances held April 4–7, 2017 in Geneva, Switzerland that they will discontinue the use of the source infix in antibody INNs.¹⁸ The make-up of previous and new antibody INN nomenclature rules are summarized in Fig. 3.

Antibody INNs: Beyond the source infix

The target infix (substem A) is determined by the target (molecule, cell or organ) class.¹⁷ The information provided by a single syllable can only in very general terms describe the intended target for an antibody, especially as an antibody's target molecule is often expressed on multiple cells in multiple

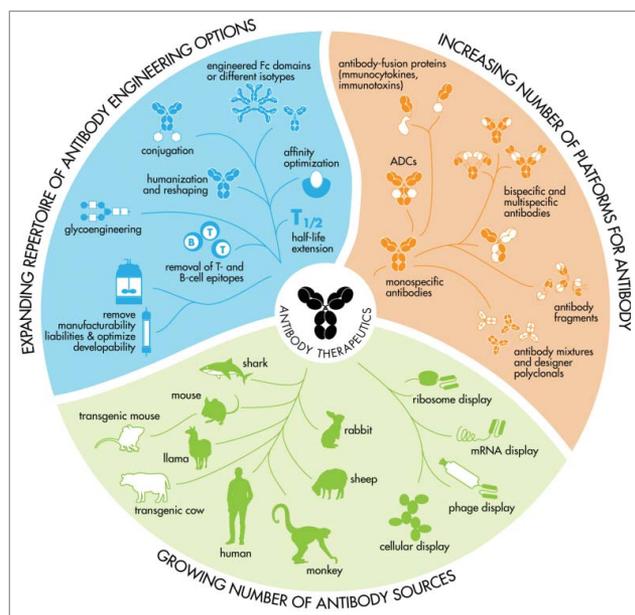


Figure 2. An expanding toolbox for the generation of therapeutic antibodies that meet modern biopharmaceutical requirements. Therapeutic antibodies can be generated in many ways and capturing an antibody's source in a single syllable is therefore no longer possible.

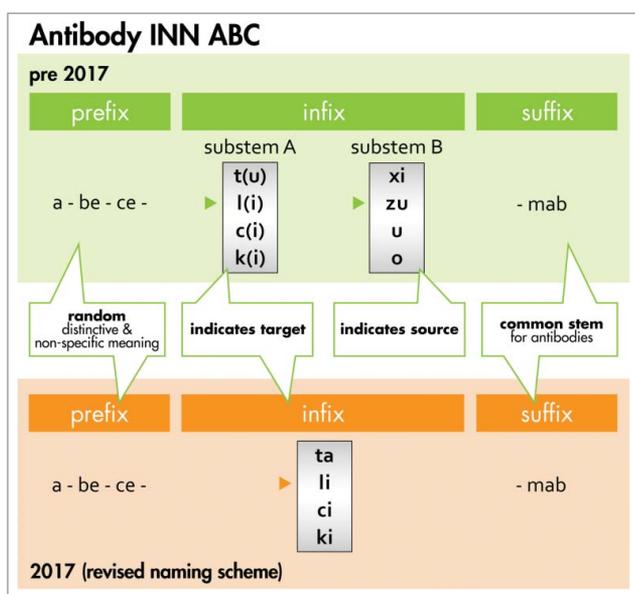


Figure 3. Antibody INN ABC. The general naming scheme for antibody INN before 2017 is compared with the new system. Prior to 2017, the random prefix was followed by a target infix (substem A) of which -t(u)- for tumor, -l(i)- for immunomodulatory, -c(i)- for cardiovascular, and -k(i)- for interleukin represented major classes. The source infix (substem B) indicated the source of which -xi- for chimeric, -zu- for humanized and -u- for human represented major classes (see the Bioreview (2014)²⁷ for complete listing). In the new scheme, the source infix designating the species has been discontinued as recommended by the INN expert group during the 64th INN Consultation.^{18, 60} To avoid confusion with earlier schemes, -ta- now designates tumor antigen. Furthermore, -ba- designates bacterial, -ami- serum amyloid protein(SAP)/amyloidosis, -ci- cardiovascular, -fung- fungal, -gros- skeletal muscle mass-related growth factors and receptors, -ki- interleukin, -li- immunomodulating, -ne- neural, -os- bone, -toxa- toxin and -vi- viral. The source infix -vet- for veterinary use antibodies is retained and added to the 'target' infix list. The suffix -mab represents the common stem for antibody therapeutics.¹⁸

organs. In addition, since therapeutic antibodies may be developed for additional or different indications after the INN has been assigned, the INN is not always consistent with the disease target. For example, rituximab includes the -tu- target infix consistent with its oncology but not inflammatory disease indications. Discontinuation of the target infix was discussed as a means of generating further flexibility in generating clearly distinct INNs for antibodies.¹⁷ Overall, arguments for discontinuing the target infix are less compelling and urgent than for the source infix. Optimizing or potentially discontinuing the use of the target infix should therefore be addressed in future discussions.

Antibody INNs may sometimes contain a second word¹⁷ that is added to conjugates of antibodies with other molecules such as a toxin, chelator or chemical (e.g., the ADCs shown in Table 1). If the antibody also contains a radiolabel, the isotope is listed at the start of the INN (i.e., name of the isotope, element and isotope number followed by the generic antibody name). The INN does not contain a specific designation to specify that a substance is a fusion protein, although this is currently under discussion. Antibodies with identical amino acid sequence (or containing minor processing differences) but distinct glycosylation are provided with a new INN containing a Greek letter as a second word, e.g., daclizumab beta (Table 1). This antibody was generated from a new cell line using a distinct manufacturing process generating carbohydrate species that are more homogeneously fucosylated than the originator product Zenapax[®] (daclizumab), thereby reducing IgG

Fc receptor IIIa binding and antibody-dependent cell-mediated cytotoxicity.^{17,19} Finally, the second word 'pegol' indicates PEGylated antibodies, e.g., certolizumab pegol.

The stem *-mab* has been used in INNs for all antibody-containing substances. However, the intention to introduce INNs for antibody-fusion proteins consisting of a single word containing the stem *-fusp* has been discussed. Although general consensus was not reached, it was decided to test the *-fusp* stem on 2 outstanding requests.¹⁷

For recombinantly expressed polyclonal antibodies (also known as designer polyclonals), each antibody in the mixture will usually require a separate INN. However, this may not be appropriate for recombinant polyclonal antibodies manufactured using single batch production strategies. In a USAN, the suffix *-pab*²⁰ may then be used, such as in rozrolimupab, which comprised 25 different recombinant anti-rhesus D antibodies.²¹ A suffix for such recombinant polyclonal drug substances is not available for INNs, but in view of future development should likely be considered.

Beyond INN

In some cases, a prefix may be added to specific antibody products to avoid medication errors and facilitate pharmacovigilance.²² The FDA, for example, included the prefix *ado-* to the ADC *ado-trastuzumab emtansine* (Kadcyla) to distinguish it from the non-drug conjugated *trastuzumab*. This is to mitigate the risk that the name is misread or mislabeled and to avoid administration of the wrong drug, which could lead to serious adverse events.^{22,23} Further use of prefixes for specific drugs should be considered as additional conjugates with the same antibody are being developed (e.g., *trastuzumab duocarmazine* (Table 1)).

The addition of a Biological Qualifier (BQ) to the INN has also been extensively discussed by WHO, the USAN Council and the FDA. It is envisioned that each biologic product, including originator products, related biologic products and biosimilars, would obtain an additional suffix. The WHO proposed the use of a random 4-letter code supplemented with a 2-digit checksum as a BQ, which should be used in conjunction with the INN to uniquely identify an antibody product to promote world-wide pharmacovigilance.²⁴ The BQ would furthermore help the accurate identification of antibody products by health care providers and patients and avoid unintended substitution. Although the BQ is considered an important issue, details are still being worked out while a pilot program with regulators is being discussed.¹⁷ The FDA recently released an Industry Guidance document proposing the use of a 4-letter suffix as a biologic qualifier.^{22,25} Three biosimilar antibody products have so far been issued such a suffix, these include *adolimumab-atto* (Amjevita), *infliximab-dyyb* (Inflectra) and *infliximab-abda* (Renflexis). A further alignment between regulatory authorities will be essential to achieve consistency and maximal leverage of the approach.

Future perspectives

The WHO decision to discontinue the use of the source infix in antibody INNs is an important step forward and addresses the

concerns and objections raised by The Antibody Society on behalf of many academic and industry scientists in the antibody field.² The new naming scheme accepted at the 64th Consultation has swiftly been implemented by WHO as we have learned that it has already been adopted in INNs currently proposed to applicants. Since the shortcomings of the 2014 source definitions extend to the Additional Information provided in the INN description (e.g., as in RL77 discussed here), we urge the WHO INN expert group to completely retire the use of categorizing therapeutic antibodies for source by using sequence alignments. The WHO announced that the INN description is envisioned to contain more extensive information on the antibody's origin and that information regarding the species on which the antibody's sequence is based will continue to be included in the definition for antibody INN.¹⁸ We fully support the inclusion of more precise information of the antibody's origin and generation in the INN description. However, continuing the practice of describing antibodies as chimeric, humanized and human in the INN description on the basis of ambiguous and inconsistent definitions, in spite of discontinuing the source infix in the actual INN, would be a very poor solution that should be reconsidered. Going forward, we request the WHO to consider omitting the source infix from antibody INNs currently under discussion (e.g., INNs in the 2016 Proposed List PL116),²⁶ as well as to review anomalies in previously assigned INNs. Most importantly, discontinuing the use of sequence alignments to determine an antibody's origin in the INN description would allow the most complete resolution of the issue.

Methods

Database analysis of antibody INNs

The antibody names in the Recommended INN: List 77³ were examined. The assigned VH and VL reference genes or alleles were searched in IMGT/mAb-DB^{34,35} using the INN as "General Query" and examined in the IMGT/2D structure-DB card for the INNs using the link provided in the Table on the respective IMGT/mAb-DB result page. The saved IMGT/DomainGapAlign results were accessed using the links provided at the bottom in the box designating the V-domain of the heavy and light chain, respectively. This link provides the top 5 "Closest reference gene and allele(s) from the IMGT V domain directory." The top hit corresponds to the reference gene and species listed in RL77.³ The top hit and percentage homology are noted in Table 1 in this manuscript. Next, a new IMGT/DomainGapAlign search against the current databased was performed by scrolling to the top of the page and executing "Align and IMGT-gap my sequence(s)." Searches were performed against the database available in weeks 17 and 18 (2017). Antibodies for which a different top hit relative to the species stated in RL77 was found are highlighted in red in Table 1 and further details are provided in Table 2. Patent applications and patents can be accessed via <https://worldwide.espacenet.com/>.

Web-based materials

All links and searches in this manuscript were checked for accuracy at the time of writing. Since links may become

inoperative or linked information may be retired or changed, relevant copies are being kept on file at The Antibody Society and may be accessed in the "member's only" area on The Antibody Society's website (<http://www.antibodysociety.org>) or directly requested from the authors.

Disclosure of potential conflicts of interest

The authors are employees (PWHIP and PJC) or are associated (AP) with companies that have a commercial interest in therapeutic antibody products and antibody engineering technologies.

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Appendix. The INN source infix explained

The WHO provides International Nonproprietary Names (INNs) to therapeutic antibodies.^{27,28} A complete and current list of INNs for therapeutic antibodies approved or under regulatory review in the US and EU can be found on The Antibody Society's website.²⁹ The INN is composed of a random, unique prefix of several syllables, a first infix (substem A), which is defined by the target, and second infix (substem B), which is defined by the source, and the suffix (stem) *-mab*.²⁷ The source infixes were developed during 1991–1993,³⁰ and, although definitions were refined over the years, the delineations between

the various categories remained the same until 2014 (see Fig. 1).^{28,31} The most common infix indicating the source are: *-o-* for all mouse sequence-derived antibodies, *-u-* for all human sequence-derived antibodies, *-xi-* for antibodies in which the mouse-derived variable region were combined with a human constant region (variable domain exchange) and *-zu-* for antibodies in which the mouse-derived complementarity determining regions were combined with a human variable region framework and human constant region. The origin of the antibody or the technology used to generate the therapeutic antibody defined the infix unequivocally in the early years.

Progress in antibody technologies, however, has increasingly blurred boundaries between the various source categories, resulting in an antibody landscape consisting of a continuum of sequences. Through these advances, therapeutic antibodies with the characteristics that are required for modern biopharmaceuticals can be generated with sequences derived from a myriad of *in vitro* and *in vivo* technologies, distinct animal species and transgenic animals or even fully synthetic sources. These further changes include the introduction of defined point mutations for optimizing binding, the mitigation of manufacturability and developability liabilities (such as replacing amino acids prone to undesired post-translation modifications) and the removal of T-cell epitopes (to lower the antibody's immunogenicity risk profile). Finally, many technologies to optimize the therapeutic antibodies' functionality are being applied (see Fig. 2). The existing nomenclature system, therefore, was becoming outdated, which was a challenge that the WHO INN expert group attempted to address by developing novel infix definitions.

The WHO updated definitions for the source infix released in 2014²⁷ handled the distinction between chimeric antibodies and humanized antibodies in a novel and unprecedented way. The new definitions included a sequence alignment procedure whereby the source infixes were now defined by the sequence of the end product and not their factual origin. In 2015, the INN expert group clarified that alignments need to be performed by using the IMGT/DomainGapAlign tool.^{10,11} The *-xi-* or *-zu-* infix is determined via the alignment of the amino acid sequence encoded by the V-gene only (i.e., FR1 through FR3 for the heavy chain (VH) and FR1 through CDR3 for the light chain (VL)) to the IMGT sequence reference database. The top hit(s) determine the closest species. For a humanized antibody, the first hit for "Closest reference gene and allele(s) from the IMGT V domain" in the DomainGapAlign result therefore should be a human sequence. A first hit on the list with any other species designates a chimeric antibody. Although this sounds easy in theory, execution is less straightforward. In cases where multiple sequences have similar scores, such as for example where 'human' is second or third to 'macaque' with only small deviations in the percentage homology score, the designation 'humanized' may still be provided by WHO. However, since precise cut-offs have not been disclosed, it is no longer possible for stakeholders to unambiguously predict the source designation that will be provided in the INN. The procedure therefore lacks transparency and consistency. Remarkably, no definitions for determining the human *-u-* source infix have been provided, and its designation therefore still hinges on the origin of the antibody's variable domains in a human sequence library of some sort, and relies on information provided

by the INNs applicant. This creates a necessity for an arbitrary definition regarding whether an antibody with a fully human sequence derived from 1) a library of natural genes, 2) a library of synthetic genes, 3) from transgenic animals, or 4) from human patients would automatically receive the -u- infix, and potentially blurs the boundary between the -u- and -zu- infix.

Although INNs are used world-wide, several countries use a separate system of non-proprietary names, one example of which is the United States Adapted Names (USAN). The USAN council is administered by the American Medical Association (AMA), and includes members of several agencies, e.g., the Food and Drug Administration (FDA). Interestingly, the post-2014 assignment of source infixes has also generated debate between the INN expert group and the USAN Council. In contrast to the INN top hit procedure explained above, the source designation for a USAN is defined by an 85% sequence cut-off (with +85% human sequence content designating a humanized antibody and a <85% content a chimeric).^{32,33} This discrepancy has further confused the field.² During the 62nd open consultation on INN, the FDA and INN expert group

representatives acknowledged the difference and indicated that harmonization is essential because therapeutic antibodies in development may have either an INN or a USAN and about half have both.⁴ We note that all USAN for therapeutic antibodies issued in 2017 thus far are also registered with INNs. In contrast, of the 27 INNs published in 2017, only 9 also carry a USAN.^{3,32}

The flaws in determining “humanness” of antibody sequences by alignment approaches were analyzed and debated by us and others elsewhere.² Overall, the notion that an antibody’s origin can be captured in a single syllable has lost its validity due to the increasing complexity of the antibody landscape (Fig. 2). Moreover, the once useful information that was carried in this one syllable can become outright misleading. The cut-off value of a USAN according to human sequence content at precisely 85% is highly questionable, as is an INN assigned according to the top homology hit being a macaque or almost identical human sequence. The source designation in antibody INNs therefore became a highly disputed issue, and its resolution was awaiting an urgent deployment.