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## Continuous reduction of tellurite to recoverable tellurium nanoparticles using an upflow anaerobic sludge bed (UASB) reactor

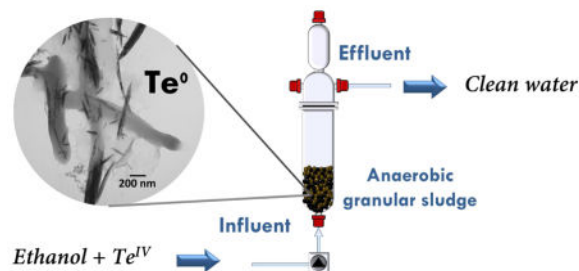
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### Abstract

According to the U.S. Department of Energy and the European Union, tellurium is a critical element needed for energy and defense technology. Thus methods are needed to recover tellurium from waste streams. The objectives of this study was to determine the feasibility of utilizing upflow anaerobic sludge bed (UASB) reactors to convert toxic tellurite ( $\text{Te}^{\text{IV}}$ ) oxyanions to non-toxic insoluble elemental tellurium ( $\text{Te}^0$ ) nanoparticles (NP) that are amendable to separation from aqueous effluents. The reactors were supplied with ethanol as the electron donating substrate to promote the biological reduction of  $\text{Te}^{\text{IV}}$ . One reactor was additionally amended with the redox mediating flavonoid compound, riboflavin (RF), with the goal of enhancing the bioreduction of  $\text{Te}^{\text{IV}}$ . Its performance was compared to a control reactor lacking RF. The continuous formation of  $\text{Te}^0$  NPs using the UASB reactors was found to be feasible and remarkably improved by the addition of RF. The presence of this flavonoid was previously shown to enhance the conversion rate of  $\text{Te}^{\text{IV}}$  by approximately 11-fold. In this study, we demonstrated that this was associated with the added benefit of reducing the toxic impact of  $\text{Te}^{\text{IV}}$  towards the methanogenic consortium in the UASB and thus enabled a 4.7-fold higher conversion rate of the chemical oxygen demand. Taken as a whole, this work demonstrates the potential of a methanogenic granular sludge to be applied as a bioreactor technology producing recoverable  $\text{Te}^0$  NPs in a continuous fashion.

### Graphical Abstract



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## Keywords

tellurite; tellurium nanoparticles; continuous bioreactors; redox mediators; riboflavin

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## 1. INTRODUCTION

Tellurium (Te) is a scarce and valuable element which is part of the chalcogen group. The concentration of this metalloid in the earth's crust ranges between 0.005 to 0.027 ppm (Wedepohl, 1995; Ba et al., 2010). Te is found in copper ores, as well as, associated with gold and silver in minerals, such as, calaverite ( $\text{AuTe}_2$ ) and silvanite ( $\text{AgAuTe}_4$ ) (Taylor, 1999; Chasteen et al., 2009). The most stable forms of Te in the environment are tellurate  $\text{TeO}_4^{2-}$  ((+6),  $\text{Te}^{\text{VI}}$ ), tellurite  $\text{TeO}_3^{2-}$  ((+4),  $\text{Te}^{\text{IV}}$ ), elemental  $\text{Te}^0$  (0) and telluride (-2) (Zannoni et al., 2008). Te is commercially obtained through different processes which include the leaching of the metalloid from the anode slimes produced during the electrolytic recovery of copper (Morrison, 1977; Taylor, 1999). Tellurite is the predominant soluble species of Te in leachates from copper and gold mining (Kyle et al., 2011; Fan et al., 2013).

The soluble oxyanions of Te, especially  $\text{Te}^{\text{IV}}$ , are highly toxic to most microorganisms. The observed toxic effects towards different microbial communities were found to be related to the cells growth conditions (either in planktonic form or as biofilms) and related to the oxidation state of the Te oxyanions. *Escherichia coli* and *Staphylococcus aureus* were more affected by  $\text{Te}^{\text{IV}}$  than by  $\text{Te}^{\text{VI}}$  (Turner et al., 2012). For both of these bacteria planktonic cells were more sensitive than biofilms. Te oxyanions inhibited distinct trophic groups in an anaerobic methanogenic mixed culture differently.  $\text{Te}^{\text{IV}}$  was very toxic to acetate- and hydrogen- consuming trophic groups; while,  $\text{Te}^{\text{VI}}$  was moderately toxic to the acetate consuming- and non-inhibitory to the hydrogen consuming- trophic groups of the same mixed culture (Ramos-Ruiz et al., 2016b). Thus the presence of these soluble oxyanions in the environment can potentially negatively affect anaerobic treatment processes such as the methanogenic phase in a landfill.

Due to the increasing number of reported applications of Te, this element has been regarded as a critical element by the US Department of Energy and by the European Union because a shortage of its supply might compromise the development of energy and defense related advanced technologies (de Boer and Lammertsma, 2013). In recent years, Te has been largely used as an additive in the rubber vulcanization process, as well as, an alloy to improve the thermo-electrical and opto-electronic properties of metal and glass. This metalloid plays a major role in the transition to clean energy technology. Te is currently used to produce CdTe thin-film photovoltaic cells, one of the most common types of solar panels commercially available (U.S. Department of Energy's National Renewable Energy Laboratory, 2013), as well as, to manufacture  $\text{Bi}_2\text{Te}_3$  thermoelectric modules (Amatya and Ram, 2011). Te has also been used to develop novel materials such as Te-based fluorescent quantum dots which have the potential to be used as probes in biosensors (Deng et al., 2007). To insure the proper supply of Te it is imperative to develop efficient processes to recover Te from mining residues and from leachates of decommissioned Te-containing products, such as CdTe solar panels in the future.

Biotechnology can provide an environmentally-friendly and cost-effective alternative to recover elemental  $\text{Te}^0$  from soluble Te in aqueous diluted streams since different microorganisms have been found to be capable of reducing a wide range of oxidized elements to their insoluble zerovalent forms (Korbekandi et al., 2009; Narayanan and Sakthivel, 2010). Several microorganisms are able to catalyze  $\text{Te}^{\text{VI}}/\text{Te}^{\text{IV}}$  reduction to  $\text{Te}^0$ , depositing nanoparticles inside and outside the cells. The bacteria *Bacillus selenitireducens* and *Sulfurospirillum barnesii* were reported to grow using  $\text{Te}^{\text{IV}}$  and  $\text{Te}^{\text{VI}}$  as electron acceptors, respectively (Baesman et al., 2007; Baesman et al., 2009). In a previous work, we studied the ability of an unadapted methanogenic consortium in granular sludge to reduce both Te oxyanions to  $\text{Te}^0$  in batch experiments. The methanogenic consortium was found to be capable of reducing both Te species utilizing the endogenous substrates in the sludge biomass as the electron donor, and the presence of an external source of electrons, such as  $\text{H}_2$ , had modest effects on increasing the reduction rates (Ramos-Ruiz et al., 2016a).  $\text{Te}^{\text{IV}}$  was reduced considerably faster than  $\text{Te}^{\text{VI}}$  (Ramos-Ruiz et al., 2016a). The Te oxyanion reduction rates were greatly enhanced by the addition of redox mediating compounds. Redox mediators are quinones or flavins that shuttle electrons from the cells to oxidized compounds (Van der Zee and Cervantes, 2009). 2-Hydroxy-1,4-naphthoquinone (lawsone) increased the rate of  $\text{Te}^{\text{IV}}$  reduction, as well as, the quantity of  $\text{Te}^0$  NPs formed outside the cells in batch tests where the facultative photosynthetic bacterium, *Rhodobacter capsulatus* (Borghese et al., 2014), was supplied with malate as the electron-donating substrate. Riboflavin (RF) accelerated the rate of  $\text{Te}^{\text{IV}}$  reduction by anaerobic methanogenic granular sludge up to 10.8- fold (Ramos-Ruiz et al., 2016a).

Among biological technologies, upflow anaerobic sludge blanket (UASB) reactors use anaerobic mixed cultures to treat wastewater in a continuous fashion. UASB reactors have proven to be effective to remove heavy metals, such as, cadmium (Cd) and zinc (Zn) from waste waters under sulfidogenic conditions (Goncalves et al., 2007). Furthermore, UASB reactors have been successfully used to study the recovery of the precious platinum group metal, palladium (Pd) (Pat-Espadas et al., 2016b). The present study was aimed at evaluating the potential to use UASB technology to reduce  $\text{Te}^{\text{IV}}$  to  $\text{Te}^0$  NP using a methanogenic microbial consortium in granular sludge provided with ethanol as an exogenous source of electron-donating substrate. The impact of RF to act as a redox mediator on facilitating the reduction process was tested by amending it to one of two UASB columns.

## 2. MATERIALS AND METHODS

### 2.1. Chemicals

$\text{Te}^{\text{IV}}$  as  $\text{Na}_2\text{TeO}_3$  and riboflavin (RF) were purchased from Sigma-Aldrich (St. Louis, MO, USA), and the Te standard (1000 ppm in 3%  $\text{HNO}_3$ ) was purchased from RICCA Chemical Company (Arlington, TX, USA).

### 2.2. Biomass source

An anaerobic granular biofilm obtained from a full scale up-flow anaerobic sludge blanket (UASB) reactor at *Mahou's* (beer brewery in Guadalajara, Spain) wastewater treatment plant, was used as the source of inoculum. This biomass contained 0.0657 g volatile

suspended solids (VSS)  $\text{g}^{-1}$  wet wt. The maximum methanogenic activities of the sludge were  $565.8 \pm 63.8$  mg methane expressed as chemical oxygen demand (COD- $\text{CH}_4$ )  $\text{gVSS}^{-1} \text{day}^{-1}$  and  $570.9 \pm 25.9$  mg COD- $\text{CH}_4$   $\text{gVSS}^{-1} \text{day}^{-1}$  for the assays utilizing acetate and hydrogen as substrate, respectively. The sludge was stored at  $4^\circ\text{C}$ .

### 2.3. Set-up of the continuous reactors

Two laboratory-scale upflow anaerobic granular sludge bed (UASB) reactors (270 mL) were run in parallel (see Fig S1 in supplementary information (SI)) in four different periods of time, based on the changes in  $\text{Te}^{\text{IV}}$  concentration in the influent as shown in Table S1 in SI, and which are briefly discussed below. Each reactor was inoculated to a final concentration of  $10 \text{ g VSS L}^{-1}$  of anaerobic granular biofilm. Both reactors were continuously operated for 146 days at room temperature ( $23 \pm 2^\circ\text{C}$ ) with hydraulic retention times (HRT) of  $\sim 0.6$  days. In order to study the effect of the redox mediator riboflavin (RF) on the reduction of  $\text{Te}^{\text{IV}}$ , one reactor (R1) was operated as a control without the addition of RF, while the mineral medium (M1, please refer to SI) used in the second reactor (R2) was spiked with different concentrations of the redox mediator during their continuous operation. The conditions of operation for both reactors during the four time periods are summarized in Table S1 in SI. Briefly, the first period of time, which comprises the initial 20 days of operation, was used as an acclimation stage and for this, the influents of both reactors were supplemented with basal media (M1, please see SI) lacking  $\text{Te}^{\text{IV}}$  and containing  $\sim 50 \text{ mg L}^{-1}$  ( $104.0 \text{ mg COD L}^{-1}$ ) of ethanol as substrate. In period II (day 20 to 50), both reactors were supplemented with mineral medium (M1 please see SI) containing  $10 \text{ mg L}^{-1}$  of  $\text{Te}^{\text{IV}}$ ,  $\sim 50 \text{ mg L}^{-1}$  of ethanol, and one reactor was amended with RF at a molar ratio  $\text{Te}:\text{RF}$  1:0.5. During period III (day 50 to 100) the concentration of  $\text{Te}^{\text{IV}}$  in the basal medium (M1 please see SI) was increased to  $20 \text{ mg L}^{-1}$  and two different levels of ethanol were used,  $\sim 50 \text{ mg L}^{-1}$  (days 50 to 75) and  $\sim 140 \text{ mg L}^{-1}$  ( $291.3 \text{ mg COD L}^{-1}$ ; days 75 to 100); two different  $\text{Te}:\text{RF}$  molar ratios were also supplied to R2 (1:0.25 for days 50 to 75, and 1:0.5 for days 75 to 100). In period IV (days 100 to 146), the concentration of  $\text{Te}^{\text{IV}}$  was increased to  $40 \text{ mg L}^{-1}$ , the amount of ethanol was also raised to  $\sim 280 \text{ mg L}^{-1}$  ( $582.7 \text{ mg COD L}^{-1}$ ) and the  $\text{Te}:\text{RF}$  molar ratio in R2 was 1:0.25.

Liquid samples obtained periodically from the influent and effluent streams of the reactors were centrifuged in centrifuge tubes at 12000 rpm for 10 min, and the supernatants were diluted into a 2% v/v  $\text{HNO}_3$  solution. The acidified samples were analyzed for total soluble Te as described in SI section. In order to account for potential losses of  $\text{Te}^0$  NPs in the effluents of the reactors, selected liquid samples were filtered through 25 nm membrane filters (EMD Millipore, Billerica, MA, USA); the filtrates were acidified to a final concentration of 2% v/v  $\text{HNO}_3$  and analyzed for dissolved Te using an ICP-OES as described in SI section. The presence of  $\text{Te}^0$  NPs was then estimated as the difference between the Te concentration from the centrifugation and filtration steps.

Samples of the influent and effluent ports were taken for the immediate measurement of pH according to standard methods, and of ethanol and acetate as described in SI section. The amount of ethanol and acetate (as a product of ethanol degradation), soluble Te, and the pH

of the influents and effluents of the bioreactors were used as indicators of the performance of the reactors.

#### 2.4. Activity kinetics batch bioassays

Two independent batch experiments were conducted to investigate the impact of the concentration of  $\text{Te}^{\text{IV}}$  and the effect of the biomass concentration on the rate of  $\text{Te}^{\text{IV}}$  reduction. These experiments were performed in 590 mL serum bottles (Wheaton, Millville, NJ, USA) amended with different amounts of granular sludge (from 0.19 to 3 g VSS  $\text{L}^{-1}$ ), 370 mL of a liquid mixture containing different amounts of two (0.5 or 5 g  $\text{Te}^{\text{IV}}$   $\text{L}^{-1}$ ) stock solutions to attain concentrations between  $1.2 \pm 0.2 - 74.6 \pm 14.3$  mg  $\text{Te}^{\text{IV}}$   $\text{L}^{-1}$  according of the purpose of the assay, mineral basal medium (M2, please see SI), and different stoichiometric excesses of hydrogen ( $\text{H}_2$ ) as the electron donor was supplied to the 220 mL of headspace proving 10.4 mmol  $\text{H}_2$   $\text{gas}$   $\text{L}^{-1}$   $\text{liq}$  (excesses ranging from 6.6- to 664-fold based on  $e^-$  equivalents). The experimental conditions utilized in these experiments are summarized in Table S2 in SI. Prior to the addition of  $\text{H}_2$  to the bottles, a gas mixture of  $\text{N}_2/\text{CO}_2$  (80:20 v/v) was bubbled through the liquid phase of the opened flasks for 3 min, and then after closing the bottles with a butyl rubber septum and an aluminum seal, the  $\text{N}_2/\text{CO}_2$  mixture was passed through the headspace of the bottles for an additional 4 min. using an inlet and outlet needle inserted at the top of the stoppers, to eliminate the remaining  $\text{O}_2$  and ensure anaerobic conditions in the experiments. After  $\text{O}_2$  was eliminated from the flasks,  $\text{H}_2$  was provided to the bottles as a gas mixture of  $\text{H}_2:\text{CO}_2$  (80:20 v/v) with an overpressure of 8 psi (0.54 atm) by inverting them and injecting the gas directly to the liquid phase, in order to attain the desired concentration (10.4 mmol  $\text{H}_2$   $\text{gas}$   $\text{L}^{-1}$   $\text{liq}$ ).

All the experiments were carried out as duplicate replicates and incubated in the dark at 30°C on a 105 rpm orbital shaker. Samples of the liquid phase were periodically withdrawn with a syringe to study the reduction of  $\text{Te}^{\text{IV}}$ . Afterwards, the samples were transferred to centrifugal filters (Amicon® ultra-4 3K, EMD Millipore, Billerica, MA, USA) and immediately centrifuged (Centrifuge 5804, Eppendorf, Enfield, CT, USA) at 4,500 rpm for 25 min. The filtrate was transferred to a 2% v/v  $\text{HNO}_3$  solution to be preserved before analyzing for soluble Te as described in SI section.

#### 2.5. Quantification of $\text{Te}^0$ NPs associated to the anaerobic sludge

On the last day of operation (day 146), the liquid media, the anaerobic granular sludge, and the glass beads at the bottom of the columns (used to distribute evenly the influent and to prevent any loss of granular sludge to the influent line), were separated. The granular sludge was then homogenized by manually stirring using a lab spatula, and three samples of  $1.505 \pm 0.028$  g of each reactor were digested using a mixture of 9 mL of concentrated  $\text{HNO}_3$  (70% wt.) and 3 mL of concentrated  $\text{HCl}$  (37% wt.) according to EPA standard procedures (Environmental Protection Agency, 2007) to quantify the total amount of Te associated to the biomass. Digested samples were diluted in DI water to reach a  $\text{HNO}_3$  concentration of 2% v/v and were analyzed for Te using an ICP-OES as described in SI section, and the Te content of the sample was extrapolated to obtain the Te associated to the full amount of biomass used in the reactors. To determine the amount of  $\text{Te}^0$  NPs formed outside the cells,  $16.29 \pm 1.12$  g of the homogenized anaerobic sludge of each reactor were transferred to 590

mL serum bottles (Wheaton, Millville, NJ, USA), along with 100 mL of the mineral medium (M1) used in the influent of both reactors. The bottles were then closed and shaken to separate the external colloidal material from the cells; the bottles were allowed to settle for 1.5 min to separate the coarser material as described in a previous work (Ramos-Ruiz et al., 2016a), and 4 samples of the liquid suspension containing colloidal material and soluble Te species were withdrawn. Two samples were digested and diluted as mentioned above, while the rest of the samples were transferred to centrifugal filters (Amicon® ultra-4 3KDa, EMD Millipore, Billerica, MA, USA) and immediately centrifuged (Centrifuge 5804, Eppendorf, Enfield, CT, USA) at 4,500 rpm for 25 min. After this step, the filtrates containing only dissolved Te species were transferred to a 2% v/v HNO<sub>3</sub> solution. All acidified samples were analyzed for total Te using an inductively coupled plasma-optical emission spectroscopy instrument (ICP-OES Optima 2100 DV, Perkin-Elmer TM, Shelton, CT) as described in SI. The amount of Te<sup>0</sup> NPs formed outside the cells was then calculated as the difference between the total amount of Te (colloidal and dissolved) obtained in the digestion and the dissolved Te estimated in the centrifugal filtration steps. The fraction of extracellular Te<sup>0</sup> NPs was then estimated after dividing the amount of Te<sup>0</sup> NPs formed outside the cells by the total amount of Te associated to the biomass.

### 3. RESULTS

#### 3.1. Kinetic Bioassays using H<sub>2</sub> as electron donor

Two batch experiments were set up to investigate the effect of the initial concentration of Te<sup>IV</sup> and of the initial biomass concentration on the reduction of Te<sup>IV</sup>. For the experiment designed to study the effect of the initial concentration of Te, a fixed amount of biomass (1.5 g VSS L<sup>-1</sup>) and six different concentrations of Te<sup>IV</sup> were provided. Figure S2A in SI shows the time course of Te reduction as a function of the initial concentration of Te<sup>IV</sup> supplied to the bottles. In the figure it can be observed that even at the highest concentration used in the experiment (74.6 ± 14.3 mg Te<sup>IV</sup> L<sup>-1</sup>) the sludge was able to carry out Te<sup>IV</sup> reduction; however, a lag phase of ~21 hours was observed in that treatment. On the other hand, a rapid loss of Te<sup>IV</sup> was observed starting at day 0 for the treatments with an initial Te<sup>IV</sup> concentration of 41.9 and 58.3 mg Te<sup>IV</sup> L<sup>-1</sup>. The three lowest initial Te<sup>IV</sup> concentrations tested had classic time course patterns expected for a first order reaction (rates progressively decreasing as the Te<sup>IV</sup> was consumed). The higher concentration treatments had first order patterns at incubation times of 21 h and beyond. Figure 1A shows the initial rates of Te<sup>IV</sup> reduction as a function of the initial concentration of the oxyanion (the rate was calculated after the lag phase for the highest concentration treatment). These rates increased proportionally with the initial concentration of the oxyanion. A strong dependence of the reduction rates to the initial Te concentration is evident (R<sup>2</sup>=0.9238), confirming the first order nature of the Te<sup>IV</sup> reduction kinetics.

The enzyme content and other reduced cofactors of the anaerobic biofilm are expected to be in direct relationship with the biomass concentration; therefore, it is expected that the Te reducing activity might be related to the anaerobic granular sludge concentration within the systems. To test the effect of the biomass concentration on Te<sup>IV</sup> reduction, 30 mg L<sup>-1</sup> of Te<sup>IV</sup> were supplied to the bottles along with five different amounts of biomass. Figure S2B



in SI shows the time course of Te reduction as a function of the sludge concentration. The rates of reduction increased with the corresponding increase of the biomass, the bottles containing  $0.75 \text{ g VSS L}^{-1}$  were able to reduce the full amount of Te in less than five days. The treatments with less biomass  $< 0.75 \text{ g VSS L}^{-1}$  showed slower reduction rates, and the treatment with the lowest amount of biomass ( $0.19 \text{ g VSS L}^{-1}$ ) had a lag phase of 3 days, after which the reduction occurred. Figure 1B summarizes the reduction rates as a function of the biomass concentration. These rates increase proportionally with the biomass concentration providing strong evidence a dependence of the rate on the biomass concentration ( $R^2=0.9764$ ).

### 3.2. Continuous reduction of $\text{Te}^{\text{IV}}$

Figures 2 and 3 present the time course of the concentrations of COD, expressed as  $\text{mg COD L}^{-1}$ , (including ethanol and acetate) and of soluble Te in the influent and effluent of the UASB reactors used in this study, respectively. The first period of time (days 0 to 20) was used as an acclimation stage for the sludge supplied only with ethanol in the influent. Steady state removal of ethanol was achieved in both reactors from the beginning of the period as can be seen in Figure 2. During the second period of operation (days 20 to 50), the influents of the reactors were spiked with  $10 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$ . Ethanol was mostly removed throughout this period of operation in both reactors; however, a decrease in COD removal of  $\sim 10\%$  (Figure S3) was observed in the reactor lacking RF after approximately 30 days of operation, but the decreased removal was recovered by day 40. As can be observed in Figure 3, the biomass was able to fully reduce  $\text{Te}^{\text{IV}}$  to  $\text{Te}^0$  in both reactors, since no soluble  $\text{Te}^{\text{IV}}$  was measured in the effluents. Important differences in the performances of the bioreactors became evident during the third period of operation (days 50 to 100) at which time the concentrations of  $\text{Te}^{\text{IV}}$  in the influents were increased to  $20 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$ . In the case of the reactor lacking riboflavin (R1), the COD removal decreased to  $\sim 40\%$  (Figure S3) around day 60. The decrease in the COD removal efficiency was reflected by an accumulation of acetate in this period as can be observed in Figure 2A. The reduction of  $\text{Te}^{\text{IV}}$  was also affected as evidenced by a higher effluent concentration of the oxyanion. Around day 70, only 80% of the  $\text{Te}^{\text{IV}}$  in the influent was reduced during this stage as can be observed in Figure 3A. However, when the concentration of ethanol in the influent was increased from 98 to  $285 \text{ mg COD L}^{-1}$  (day 75 to 100), the reduction of  $\text{Te}^{\text{IV}}$  fully recovered. The removal of COD also increased to 80–90% but, the degradation of ethanol was affected since a small accumulation of ethanol and acetate started to become evident just prior to day 100 (Figure 2A). The most noteworthy response of this reactor was observed during the fourth period of operation (days 100 to 150) when the concentration of  $\text{Te}^{\text{IV}}$  in the influent was augmented to  $40 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  and the concentration of ethanol was increased further. A remarkable loss of  $\text{Te}^{\text{IV}}$  reduction capacity was observed in R1 when only  $59.6 \pm 11.2\%$  of the Te of the influent was reduced, and a progressive trend in  $\text{Te}^{\text{IV}}$  reduction deterioration was observed (only 33% was reduced during the last day of operation, Figure 3A). The inhibition of the biomass in R1 was also evident due to the presence of a considerable amount of ethanol and acetate in the effluent. During this period the removal of COD was only  $29.4 \pm 12.0\%$  and a progressive trend in COD removal deterioration was also observed (the removal of COD decreased dramatically to 14.3% at the end of the operation time).

The reactor amended with riboflavin (R2) was able to sustain  $\text{Te}^{\text{IV}}$  reduction during the whole period of operation without further complications. The biomass was able to degrade ethanol and completely remove COD as can be observed in Figures 2B and 3B. During the fourth period of operation (days 100 to 150), R2 continued to completely reduce  $\text{Te}^{\text{IV}}$  (Figure 3B) presumably due to enhanced reductive capacity afforded by RF. Although inhibition of the biomass in this reactor was observed, it was remarkably lower compared to R1, the COD removal was sustained at  $77.8 \pm 6.1\%$ . (Figure S3 in SI).

### 3.3. Mass balances at the end of the operation period

At the end of the experiment (day 146), samples of the biomass obtained from the bioreactors were digested and analyzed to corroborate the amount of  $\text{Te}^{\text{IV}}$  retained as elemental Te NPs in the columns both, outside and inside of the cells. The distribution of Te between the phases of both reactors is summarized in Table 1.

In the reactor lacking RF (R1), the Te recovered by the digestion of the sludge along with the Te measured in the effluent of R1 accounted for 99.7% of the cumulative amount of  $\text{Te}^{\text{IV}}$  fed during the full time of operation. The amount of  $\text{Te}^0$  NPs lost to the effluent of the reactor was determined as described in section 2.5. The results showed that a non-significant amount of colloidal Te was present in the effluent of R1. The fraction of Te leaving the column in particulate form (colloidal fraction) varied from 0.6 to 11.5% of the total Te in the effluent, for the selected samples collected at different moments of the operation period; while, the remaining Te left the column completely dissolved. The small amount of colloidal material found in the effluent, as well as, in the digestion of the well homogenized sludge of R1 at the end of operation period, suggest a potential formation of intracellular material over extracellular precipitation.

In the case of R2, the Te recovered from the digestion of the sludge, the Te measured in the effluent of the reactor, and the amount found attached to the glass beads used at the base of reactor (intended to uniformly distribute the influent), accounted for 63.1% of the cumulative  $\text{Te}^{\text{IV}}$  fed to the reactor during the 146 days of operation. The incomplete recovery of Te by the digestion is potentially due to a focussed accumulation of Te at the bottom of the column where incomplete homogenization of the biomass may have occurred during sampling for the aforementioned digestion. To account for a potential release of  $\text{Te}^0$  NPs to the effluent of the reactor, the same procedure used in R1 was followed. The results showed that a more important amount of colloidal Te was present in the effluent of reactor R2 compared to that of R1. According to the results obtained from the filtrations of the effluent, it was calculated that between 13.8 to 84.5% of the total Te was present in particulate form in this stream while the rest was dissolved. The latter suggests an important formation of extracellular material compared to intracellular precipitation.

### 3.4. $\text{Te}^0$ NPs characterization

Samples of the biomass obtained from both bioreactors were analyzed by TEM and XRD to investigate the nature and shape of the particles of  $\text{Te}^0$  formed inside the columns. Figures S4 and S5 show the TEM images of the material produced internally and externally to the cells in the bioreactors.



In general, the  $\text{Te}^0$  NPs were closely associated to the biomass in the bioreactor R1 as shown in Figures S4C and S6 (in SI), while in the case of the reactor containing RF (R2), the NPs were found mostly dispersed and away from the cells (data not shown). With respect to the size of the NPs in both reactors, a highly heterogeneous material was found. In both reactors small individual needle shards with lengths ranging between 30 to 300 nm with diameters of  $\sim 10$  nm were observed randomly oriented. Rosettes, as those shown in Figures S4D and S6 (in SI) were only found in the reactor lacking RF. Bundles formed with orderly oriented rod shaped NPs were also discovered in both bioreactors; however, those produced in R1 (lengths ranging from 90 to 315 nm) were larger than those precipitated in R2 (lengths ranging from 140 to 190 nm) as shown in Figure S4 in SI (Panels A and B).

Internal accumulation of  $\text{Te}^0$  NPs was also corroborated in the cytoplasm and in the periplasmic membranes of the microorganisms in the biofilms coming from both reactors. More cellular damage and internal accumulation was evident in the biomass of R1 as is evident in Figure S5 in SI. Important membrane damage can be observed in the bottom right of Figure S5C as well as larger  $\text{Te}^0$  NPs inside the cells compared to the NPs of R2 (Figures S5A and B).

The XRD analyses showed that the solid product collected from the sludge of both bioreactors at the end of the operation time (day 146) corresponded to elemental  $\text{Te}^0$ . Figure S7 shows the XRD patterns for the aggregates found in both reactors. The marks indicate the positions of the expected peaks in the Te diffraction pattern which correspond to the characteristic diffraction peaks for crystalline  $\text{Te}^0$ . According to the XRD patterns, the samples coming from the reactor lacking riboflavin contained a larger amount of  $\text{Te}^0$  (either internalized or outside the cells) than those coming from the RF amended reactor (R2). These results support the idea that the material in the reactor with RF was mainly deposited in the lower part of the column.

## 4. DISCUSSION

### 4.1. Ethanol as the exogenous source of electrons

The addition of an external electron donor was intended to provide electrons needed to carry out  $\text{Te}^{\text{IV}}$  reduction in the continuous columns. In a previous work, we found that the sludge itself contains significant endogenous substrate to carry out the reduction of  $20 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  in batch experiment; however, the addition of an external electron donor, such as,  $\text{H}_2$  stimulated the reduction reaction of  $\text{Te}^{\text{IV}}$  (Ramos-Ruiz et al., 2016a). Therefore, the presence of an external electron donor ensures the adequate supply of electron equivalents required to reduce  $\text{Te}^{\text{IV}}$  in a continuous process, and at the same it might represent several benefits, such as, the increase of the  $\text{Te}^{\text{IV}}$  reduction rate. Based on the production of methane estimated from the decay of the biomass in the anaerobic granules, the amount of endogenous substrate in the sludge corresponds to 60 to 166 mg chemical oxygen demand (COD)  $\text{g}^{-1}$  VSS (Field et al., 2004; Tapia-Rodriguez et al., 2010). Considering the amount of sludge used to inoculate the bioreactors ( $10 \text{ g VSS L}_{\text{reactor}}^{-1}$  in a volume of 0.27 L), the endogenous capacity of the reactors was estimated to be between 0.162 to 0.4482 g COD. The total amount of  $\text{Te}^{\text{IV}}$  supplied to the reactors would require approximately of 0.23 g COD- $\text{Te}^{\text{IV}}$ . For this reason, addition of an excess external electron donor was needed,

especially when considering that there is competition with other processes for electrons such as methanogenesis.

Ethanol was used as the external electron donor to support the reduction of  $\text{Te}^{\text{IV}}$  throughout this work. Ethanol was selected since it represents a safer and more economical option compared to other electron donors which have proven to be effective in stimulating the reduction of Te oxyanion, such as  $\text{H}_2$  (Ramos-Ruiz et al., 2016a).  $\text{H}_2$  is one of the products of the anaerobic fermentation of ethanol by the acetogenic- bacteria, which are present in methanogenic anaerobic sludge, as can be noted in eq 1.



The effect of ethanol might be comparable to  $\text{H}_2$  since it is in fact the biogenic  $\text{H}_2$  that is the source of effective electrons. In batch experiments, where the reduction of  $20 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  was studied using different external electron donors,  $\text{H}_2$  increased the rate of  $\text{Te}^{\text{IV}}$  reduction 1.30-times with respect to the systems lacking external electron donor. The other product of ethanol fermentation, acetate, was found to be a poor electron donating substrate for  $\text{Te}^{\text{IV}}$  (Ramos-Ruiz et al., 2016a). Likewise compared to ethanol and  $\text{H}_2$ , acetate was also a poor electron-donor for the bioreduction of other oxidized compounds such as  $\text{As}^{\text{V}}$ ,  $\text{U}^{\text{VI}}$ , and  $\text{Pd}^{\text{II}}$  (Field et al., 2004; Tapia-Rodriguez et al., 2010; Pat-Espadas et al., 2016a) in anaerobic sludge.

To overcome the competition with other processes, ethanol was added in different excesses with respect to the amount stoichiometrically required to reduce the  $\text{Te}^{\text{IV}}$  in the influent of the columns during the four stages of operation (7-, 14-, and 20- times). The effectiveness of ethanol as electron donor, as well as, the importance of adding it in excess was evident during the third period of operation when the lowest ethanol excess was used (7- times). The electron-donor supply capacity of the anaerobic reactor was apparently depleted as a small accumulation of  $\text{Te}^{\text{IV}}$  in the effluent of the reactors was measured during this stage.  $\text{Te}^{\text{IV}}$  accumulation in the effluent disappeared when the ethanol supplied to the influent was increased at day 75 to achieve a 20-fold excess.

#### 4.2. Continuous reduction of $\text{Te}^{\text{IV}}$ oxyanion to recoverable $\text{Te}^0$ NPs

During periods II and III of operation, the non-adapted anaerobic methanogenic granular sludge used in this study was able to perform  $\text{Te}^{\text{IV}}$  reduction, up to a concentration of  $20 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  in the influent to elemental  $\text{Te}^0$  in a continuous fashion with reduction efficiencies ranging from 83% to 96% in the case of R1 (lacking RF), and of 99.5% in R2 (with RF). In the third period of operation, when the supply of electron equivalents was restricted due to the low excess of ethanol supplied to the reactors, an accumulation of  $\text{Te}^{\text{IV}}$  caused mild inhibition of the fermenting bacteria which did not recover after the reduction of  $\text{Te}^{\text{IV}}$  was restored around day 70 in R1. No toxic effect towards the fermenting bacteria was evident in R2 throughout periods II and III of operation. In the fourth period of operation, when the concentration of  $\text{Te}^{\text{IV}}$  was increased to  $40 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$ , the granular sludge in R1 lost its  $\text{Te}^{\text{IV}}$ -reducing capacity. The remarkable toxic effects of  $\text{Te}^{\text{IV}}$  towards the fermenting bacteria

was evident from an important accumulation of ethanol and acetate in the effluent of R1 around day 100. On the other hand, the presence of a low concentration of RF in R2 as a redox mediating agent allowed for the  $\text{Te}^{\text{IV}}$  reducing capacity of the sludge to remain intact during the full time of operation. As a consequence, only moderately toxic effects towards the ethanol fermenting bacteria were observed in R2.

To date, only few studies have assessed the recovery of  $\text{Te}^0$  NPs in a continuous mode (Basnayake et al., 2001; Rajwade and Paknikar, 2003). In one of the studies, a continuous stirred tank reactor (CSTR) was set up to assess the production of  $\text{Te}^0$  by means of  $\text{Te}^{\text{IV}}$  reduction using *Pseudomonas mendocina* MCM B-180 and sucrose as a substrate. The CSTR was able to sustain the complete reduction of  $100 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  in the influent (translated to a  $\text{Te}^{\text{IV}}$  load of  $1.40 \text{ mg Te}^{\text{IV}} \text{ h}^{-1} \text{ L}_{\text{reactor}}$ ) (Rajwade and Paknikar, 2003). These results correlate well with our findings with the reactor lacking RF in which  $20 \text{ mg Te}^{\text{IV}} \text{ L}^{-1}$  (representing a  $\text{Te}^{\text{IV}}$  load of  $1.53 \text{ mg Te}^{\text{IV}} \text{ h}^{-1} \text{ L}_{\text{reactor}}$ ) were effectively removed from the influent; however, the  $\text{Te}^{\text{IV}}$  reduction capacity of our continuous system was remarkably improved with the use of low concentrations of RF in a way that the reactor was able to carry out the reduction of a higher volumetric load of  $\text{Te}^{\text{IV}}$  ( $3.06 \text{ mg Te}^{\text{IV}} \text{ h}^{-1} \text{ L}_{\text{reactor}}$ ).

This type of technology has been also successfully used to investigate the recovery of some precious metals like palladium (Pd) using ethanol as substrate (Pat-Espadas et al., 2016b) for a maximum concentration of  $15 \text{ mg Pd}^{\text{II}} \text{ L}^{-1}$  in the influent of the reactor, achieving removal efficiencies of  $98.9 \pm 0.7\%$ . The removal of the chalcogen selenate ( $\text{Se}^{\text{VI}}$ ) was also investigated in a UASB reactor using a methanogenic consortium and lactate as the electron donating substrate. Complete removal of  $\text{Se}^{\text{VI}}$  at  $1.4 \text{ mg Se}^{\text{VI}} \text{ L}^{-1}$  in the influent was achieved in the reactor (Lenz et al., 2008). Our results suggest that UASB technology represents a viable option to recover  $\text{Te}^0$  NPs from solubilized Te obtained from decommissioned materials, such as, CdTe solar panels, and from mining aqueous waste streams. The reactor's performance might be importantly enhanced by the addition of a redox mediator, such as, RF.

### 4.3. Effect of riboflavin on $\text{Te}^{\text{IV}}$ reduction

Several advantages of the use of RF were observed during the operation of the continuous columns in the present study. This redox mediator (RM) lowered the toxic effects of  $\text{Te}^{\text{IV}}$  by forming poorly bioavailable  $\text{Te}^0$  NPs, and most importantly, the RF amended reactor was able to reduce twice the amount of Te compared to the one lacking RF. An increase in the rate of  $\text{Te}^{\text{IV}}$  reduction caused by the presence of the catalyst RF might explain these operational advantages.

The toxic effects of  $\text{Te}^{\text{IV}}$  towards the microorganisms in the granular sludge were remarkably lower in R2 compared to R1. The acetogenic- and acetoclastic- microorganisms in the methanogenic sludge were highly inhibited at the highest concentration of  $\text{Te}^{\text{IV}}$  tested of  $40 \text{ mg Te} \text{ L}^{-1}$  in R1; while, in R2, the RF amendment greatly lowered the toxic effects, which were only moderately impacting the acetoclastic- microorganisms (based on observing some residual acetate). The high toxicity of  $\text{Te}^{\text{IV}}$  towards methanogens in anaerobic granular sludge was recently reported (Ramos-Ruiz et al., 2016b). The acetoclastic methanogens were more sensitive towards  $\text{Te}^{\text{IV}}$  than the hydrogen consuming

methanogens. The estimated 50% inhibiting concentration ( $IC_{50}$ ) of the methanogenic acetoclastic activity was  $8.6 \text{ mg Te}^{IV} \text{ L}^{-1}$  (Ramos-Ruiz et al., 2016b).  $\text{Te}^{IV}$  has also been reported to be very toxic for most bacteria at very low concentrations as of  $1 \text{ mg L}^{-1}$  (Taylor, 1999). The minimal concentration required to eliminate a biofilm of *Escherichia coli* was  $1.8 \text{ mg Te}^{IV} \text{ L}^{-1}$  (Turner et al., 2012).

To the best of our knowledge, this is the first report of applying a redox mediator to enhance the continuous bioreduction of  $\text{Te}^{VI}$  oxyanions. Redox mediators, such as, RF, quinones, and humus, are compounds known to stimulate biological reactions by facilitating the transfer of electrons from cells to oxidized inorganic compounds (e.g. selenate and palladium<sup>II</sup>). This enables the direct reduction of inorganic compounds by the reduced form of the redox mediators (Van der Zee and Cervantes, 2009). In the previous work, we tested the effectiveness of four different RM compounds in the reduction of  $\text{Te}^{IV}$  (lawsone, AQDS, riboflavin and hydroxocobalamin) at two different Te/RM molar ratios (Ramos-Ruiz et al., 2016a). RF had the greatest impact on the reduction of the  $\text{Te}^{IV}$  increasing rates by 3.6- and 10.8- fold when the Te/RF molar ratios were held at 10:1 and 1:1, respectively. The effect of other RM compounds (lawsone, AQDS, menadione) on the reduction of  $\text{Te}^{IV}$  has also been reported before (Wang et al., 2011). Lawsone increased the rate of  $\text{Te}^{IV}$  reduction by 10-fold in a system using the bacterium *E. coli* with glucose supplied as electron donor (Wang et al., 2011). Lawsone almost doubled the  $\text{Te}^{IV}$  reduction rate for the photosynthetic bacterium *Rhodobacter capsulatus* when pyruvate was used as substrate. The stimulation however was found to be independent of the lawsone concentration (Borghese et al., 2014).

The catalytic properties of RF observed in this work might be explained by the fact that the standard redox potential ( $E^{0'}$  for pH 7) of the RM is between those of the electron donor and electron acceptor reactions ( $\text{H}_2$  oxidation and Te oxyanions reduction). The standard redox potentials for the chemical species involved in this reduction are  $2\text{H}^+/\text{H}_2$   $E^{0'} = -0.414 \text{ V}$ ;  $\text{RF}_{\text{ox}}/\text{RF}_{\text{red}}$   $E^{0'} = -0.208 \text{ V}$  (Bird et al., 2011); and  $\text{Te}^{IV}$  ( $\text{HTeO}_3^-/\text{Te}^0$ )  $E^{0'} = 0.196 \text{ V}$  (calculated from reported  $E^0$  values (Bouroushian, 2010)). The effectiveness of an electron shuttle is dependent of the energy of activation of their reduction and oxidation (Van der Zee et al., 2001). For these reasons, RF was suspected to decrease the energy of activation of the redox reactions, and as a consequence the  $\text{Te}^{IV}$  reduction reaction occurred faster.

## CONCLUSION

The feasibility of continuously converting  $\text{Te}^{IV}$  oxyanion to recoverable  $\text{Te}^0$  NPs with methanogenic granular sludge in a UASB fed ethanol as the electron donating substrate was demonstrated for the first time. The sludge used in this work was able to sustain the reduction of high loads of the highly toxic oxyanion  $\text{Te}^{IV}$ . Due to its redox mediating properties, RF greatly enhanced the performance of the UASB by increasing the rate of  $\text{Te}^{IV}$  reduction. The faster formation of non-toxic  $\text{Te}^0$  NPs due to the increase of the Te reduction rate resulted in a better detoxification; thus, indirectly RF improved  $\text{Te}^{IV}$  reduction. All these encouraging findings, might be used as the basis for the development of a safe, environmentally friendly and cost-effective large-scale process to recover  $\text{Te}^0$  from aqueous streams containing  $\text{Te}^{IV}$ .

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

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### Highlights

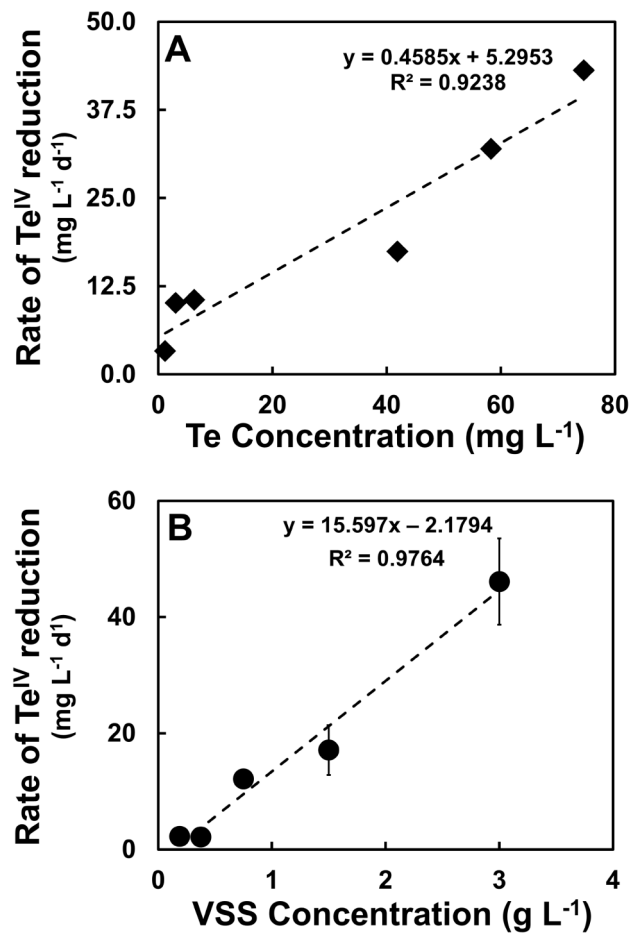
A continuous anaerobic bioreactor converted  $\text{Te}^{\text{IV}}$  to elemental  $\text{Te}^0$  nanoparticles

$\text{Te}^{\text{IV}}$  reduction rates depended on biomass and  $\text{Te}^{\text{IV}}$  concentrations

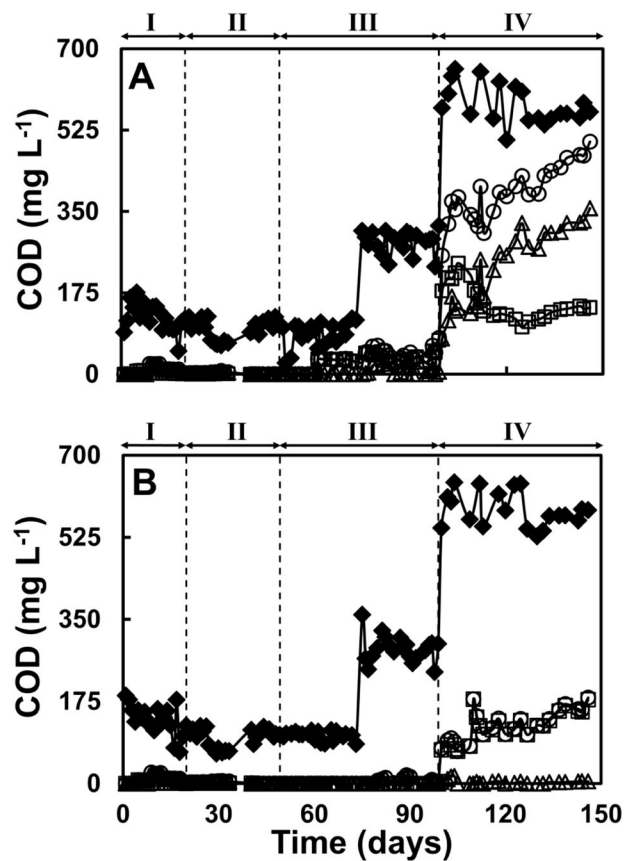
Ethanol used as an electron donating substrate promoted  $\text{Te}^{\text{IV}}$  reduction

Riboflavin used as redox mediator greatly improved  $\text{Te}^{\text{IV}}$  reduction

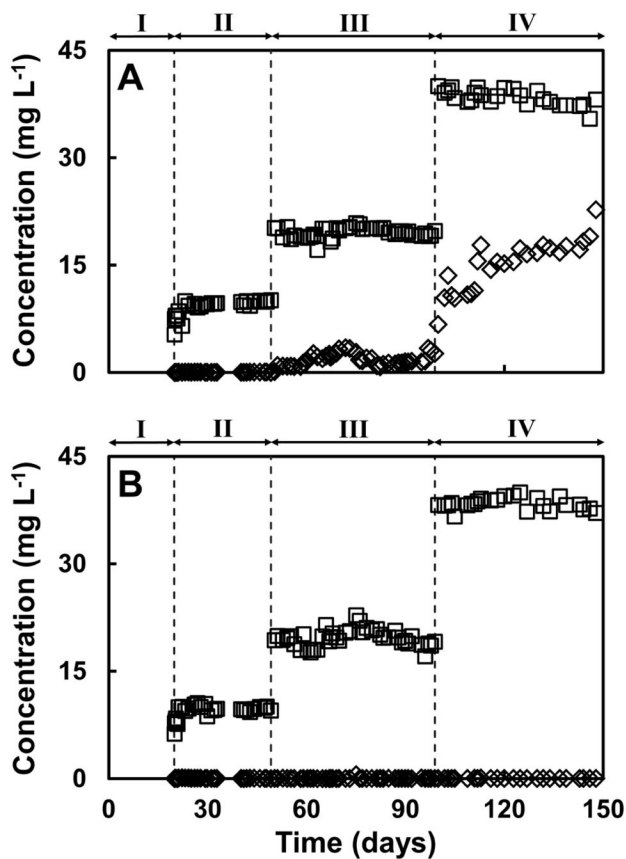
Riboflavin markedly lowered  $\text{Te}^{\text{IV}}$  toxicity to chemical oxygen demand removal



**Figure 1.** Rate of Te<sup>IV</sup> reduction as a function of the initial Te<sup>IV</sup> concentration (**Panel A**) and of the initial concentration of biomass in the bioassays (**Panel B**).



**Figure 2.** Performance of the reactors in terms of the time course of the COD concentration in the influents and effluents of both reactors. **Panel A**, reactor lacking RF (R1). **Panel B**, reactor amended with RF (R2). **Legends:** (◆), total COD in the influent; (○), total COD in the effluent; (□), acetate-COD in the effluent; (△), ethanol-COD in the effluent. The vertical lines indicate the time when the concentrations of  $\text{Te}^{\text{IV}}$  in the influents were increased.



**Figure 3.** Time course of  $\text{Te}^{\text{IV}}$  concentration in the influent ( $\square$ ) and effluent ( $\diamond$ ) of the continuous reactors during the time of operation. **Panel A**, reactor lacking riboflavin (R1). **Panel B**, reactor supplied with riboflavin (R2). The vertical lines indicate the time when the concentrations of Te in the influents were increased.

**Table 1**

Recovery of Te in different fractions at the end of the experiment.

	R1	R2
$\Sigma Q_{Te\ inf}$ (g)	1.63	1.69
$\Sigma Q_{Te\ eff}$ (g)	0.461	0.002
$\Sigma Q_{Te\ eff}/\Sigma Q_{Te\ inf}$ (%)	28.3	0.1
$\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff}$ (g)	1.17	1.69
$(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})/\Sigma Q_{Te\ inf}$ (%)	71.7	99.9
$X_{Te}$ (g)	1.165±0.0002	0.673±0.024
Te <sub>in glass beads</sub> (g)	-	0.391±0.007
$\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff} - X_{Te} - Te_{in\ glass\ beads}$ (g)	-	0.619
$X_{Te}/(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})$ (%)	99.6	39.8
$(X_{Te} + Te_{in\ glass\ beads})/(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})$ (%)	99.6	63.0
Coll <sub>Te</sub> (g)	0.105±0.001	0.028±5.85×10 <sup>-5</sup>
UF <sub>Te</sub> (g)	4.175×10 <sup>-3</sup> ±4.38×10 <sup>-5</sup>	9.44×10 <sup>-5</sup> ±2.30×10 <sup>-6</sup>
Coll <sub>Te</sub> -UF <sub>Te</sub> (g)	0.101±0.001	0.028±5.62×10 <sup>-6</sup>
Te <sup>0</sup> NP/(X <sub>Te</sub> ) (%)	8.68±0.10	39.3*

\* This percentage was estimated assuming that the Te associated to the beads at bottom of R2 was formed externally to the cells.

Cumulative Te fed to the column ( $\Sigma Q_{Te\ inf}$ )

Cumulative Te discharge with effluent ( $\Sigma Q_{Te\ eff}$ )

Fraction of cumulative Te discharge with effluent compared to total Te fed to the column ( $\Sigma Q_{Te\ eff}/\Sigma Q_{Te\ inf}$ )

Estimated cumulative Te retained in the column ( $\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff}$ )

Fraction of cumulative Te retained in the column compared to total Te fed to the column ( $(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})/\Sigma Q_{Te\ inf}$ )

Te found retained by the sludge ( $X_{Te}$ )

Total Te found associated to glass beads ( $Te_{in\ glass\ beads}$ )

Total Te lost to the glass beads of the reactors ( $\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff} - X_{Te} - Te_{in\ glass\ beads}$ )

Total Te recovered by sludge digestion ( $X_{Te}/(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})$ )

Total Te recovered by digestion and attached to glass beads ( $(X_{Te} + Te_{in\ glass\ beads})/(\Sigma Q_{Te\ inf} - \Sigma Q_{Te\ eff})$  (%)

Total Te associated to the sludge (see section 2.6, soluble and in suspension) after 1.5 min settling (Coll<sub>Te</sub>)

Total soluble Te associated to the sludge in suspension after 1.5 min settling (see section 2.6, Te passing through 3KDa filter (UF<sub>Te</sub>))

Te<sup>0</sup> NPs formed extracellularly associated to the sludge (Coll<sub>Te</sub>-UF<sub>Te</sub>)

Fraction of Te<sup>0</sup> found as extracellular material (Te<sup>0</sup> NP/(X<sub>Te</sub>))