



Role of CrRLK1L Cell Wall Sensors HERCULES1 and 2, THESEUS1, and FERONIA in Growth Adaptation Triggered by Heavy Metals and Trace Elements

Julia Richter¹, Marie Ploderer¹, Gaëlle Mongelard², Laurent Gutierrez² and Marie-Theres Hauser^{1*}

¹ Department of Applied Genetics and Cell Biology, University of Natural Resources and Life Sciences, Vienna, Vienna, Austria, ² Centre de Ressources Régionales en Biologie Moléculaire, Université de Picardie Jules Verne, Amiens, France

OPEN ACCESS

Edited by:

Gea Guerriero,
Luxembourg Institute of Science
and Technology, Luxembourg

Reviewed by:

Michael Deyholos,
University of British Columbia,
Canada
Frantisek Baluska,
University of Bonn, Germany

*Correspondence:

Marie-Theres Hauser
marie-theres.hauser@boku.ac.at

Specialty section:

This article was submitted to
Plant Nutrition,
a section of the journal
Frontiers in Plant Science

Received: 15 July 2017

Accepted: 25 August 2017

Published: 07 September 2017

Citation:

Richter J, Ploderer M, Mongelard G,
Gutierrez L and Hauser M-T (2017)
Role of CrRLK1L Cell Wall Sensors
HERCULES1 and 2, THESEUS1,
and FERONIA in Growth Adaptation
Triggered by Heavy Metals and Trace
Elements. *Front. Plant Sci.* 8:1554.
doi: 10.3389/fpls.2017.01554

Cell walls are not only a protective barrier surrounding protoplasts but serve as signaling platform between the extracellular environment and the intracellular physiology. Ions of heavy metals and trace elements, summarized to metal ions, bind to cell wall components, trigger their modification and provoke growth responses. To examine if metal ions trigger cell wall sensing receptor like kinases (RLKs) of the *Catharanthus roseus* RLK1-like (CrRLK1L) family we employed a molecular genetic approach. Quantitative transcription analyses show that *HERCULES1* (*HERK1*), *THESEUS1* (*THE1*), and *FERONIA* (*FER*) were differently regulated by cadmium (Cd), nickel (Ni), and lead (Pb). Growth responses were quantified for roots and etiolated hypocotyls of related mutants and overexpressors on Cd, copper (Cu), Ni, Pb, and zinc (Zn) and revealed a complex pattern of gene specific, overlapping and antagonistic responses. Root growth was often inversely affected to hypocotyl elongation. For example, both *HERK* genes seem to negatively regulate hypocotyl elongation upon Cd, Ni, Zn, and Pb while they support root growth on Cd, Cu, and Ni. The different *THE1* alleles exhibited a similar effect between roots and hypocotyls on Ni, where the loss-of-function mutant was more tolerant while the gain of function mutants were hypersensitive indicating that *THE1* is mediating Ni specific inhibition of hypocotyl elongation in the dark. In contrast hypocotyl elongation of the knock-out mutant, *fer-4*, was hypersensitive to Ni but exhibited a higher tolerance to Cd, Cu, Pb, and Zn. These data indicate an antagonistic action between *THE1* and *FER* in relation to hypocotyl elongation upon excess of Ni. *FER*s function as receptor for rapid alkalization factors (RALFs) was tested with the indicator bromocresol purple. While *fer-4* roots strongly acidified control and metal ion containing media, the etiolated hypocotyls alkalinized the media which is consistent with the already shorter hypocotyl of *fer-4*. No other *CrRLK1L* mutant exhibited this phenotype except of the *THE1*:GFP overexpressor on Ni suggesting that *THE1* might be involved in Ni induced and hypocotyl specific RALF signaling and growth regulating pathway. Overall, our findings establish a molecular link between metal ion stress, growth and the cell wall integrity sensors of the *CrRLK1L* family.

Keywords: cadmium, copper, lead, nickel, zinc, alkalization, root growth, hypocotyl elongation

INTRODUCTION

Heavy metals and trace elements in excess as well as deficiency in soils impose a major challenge to plant growth in general and crop productivity. From studies of metal ion tolerant and hyperaccumulating metallophytes as well as sensitive plants such as *Arabidopsis thaliana* we learnt that excess of metal ions induce a complex network of responses thoroughly reviewed in DalCorso et al. (2013) and Singh et al. (2016). The regulation of metal ion homeostasis involves increased biosynthesis of chelators as well as efflux and influx transporters essential for compartmentalization. Another strategy to prevent or reduce uptake of metals is by restricting metal ions to the cell wall. The cell wall, rich in functional carbohydrate originated carboxyl and hydroxyl as well as protein derived sulfhydryl and histidyl groups plays a key role in the immobilization of metal ions (Krzesłowska, 2011). Metal ion immobilization in the cell wall is mainly mediated by the pectic polysaccharide homogalacturonan (Pelloux et al., 2007). The capacity of binding is enhanced by the activity of cell wall-associated pectin-methylsterases (PMEs) or pectin-acetylsterase (PAEs) exposing free negatively charged carboxyl groups. They form salt bridges contributing to the mechanical strength of cell walls. Mainly calcium (Ca) is used in these so called “egg boxes” but other metals ions with often higher affinities such as aluminum (Al), copper (Cu), lead (Pb), zinc (Zn), cadmium (Cd), cobalt (Co), nickel (Ni), barium (Ba), strontium (Sr), manganese (Mn), magnesium (Mg), iron (Fe), chromium (Cr), and mercury (Hg) have been shown to bind to de-esterified pectins (Dronnet et al., 1996; Kartel et al., 1999; Meychik et al., 2011). Expression studies in diverse plants show that cell wall modifying enzymes were upregulated upon metal ion stress (Hassinen et al., 2007; Konlechner et al., 2013). Also PME activities and structural modifications of pectins change upon metal ion treatments and are related to metal ion tolerance and growth responses (Paynel et al., 2009; Douchiche et al., 2010; Weber et al., 2013; Yang et al., 2013; El-Moneim et al., 2014; Muschitz et al., 2015; Geng et al., 2017) and reviewed in Parrotta et al. (2015).

Pectin, highly de-esterified pectates and their degradation products are important components of the cell wall integrity pathways (Wolf et al., 2012; Voxeur and Höfte, 2016). It has been shown that pectins and oligogalacturonic acids bind to the extracellular domain of and activate WAK1 and WAK2, members of the *WALL ASSOCIATED KINASES* gene family (Decreux and Messiaen, 2005; Decreux et al., 2006; Kohorn et al., 2006). Furthermore, WAKs and WAK-LIKE receptors (WAKLs) are involved in the regulation of cell expansion and responses to metal ions (Lally et al., 2001; Wagner and Kohorn, 2001; Hou et al., 2005). Apart from WAK(L)s another pectin-associated kinase, proline-rich extensin-like receptor kinase 4 (PERK4), is involved in drought stress mediated growth responses (Bai et al., 2009). Also, several leucine-rich repeat (LRR) receptor like kinases (RLKs) are involved in cell wall integrity pathways related to pathogen signaling and are necessary for the synthesis of cell wall components upon high sucrose and NaCl conditions (Xu et al., 2008; Engelsdorf and Hamann, 2014).

In this study we focused on the *Catharanthus roseus* RLK1-like (CrRLK1L) protein family. The CrRLK1L family consists of 17 members which all share an extracellular domain homologous to the animal malectin protein with putative carbohydrate binding capacity. Therefore the CrRLK1L malectin-like domains might bind to oligo- or polysaccharides from cell wall polymers, by-products of cell wall degradation, or membrane-associated or secreted glycosylated proteins (Schallus et al., 2008; Boisson-Dernier et al., 2011). Recently it has been demonstrated that the CrRLK1L member, FERONIA (FER), is the receptor for peptides of the RAPID ALKALINIZATION FACTOR family (Haruta et al., 2014) while the ligands for the other members are still disclosed. CrRLK1L proteins play diverse roles during fertilization (Escobar-Restrepo et al., 2007; Boisson-Dernier et al., 2009; Miyazaki et al., 2009) but are also important during vegetative development and in plant pathogen interactions (Bai et al., 2014; Gachomo et al., 2014; Nissen et al., 2016; Stegmann et al., 2017).

Here, we focus on the four CrRLK1L members which regulate cell expansion and growth of seedlings: FER, THESEUS1 (THE1), HERCULES1 (HERK1), and HERK2 (Hématy et al., 2007; Guo et al., 2009a,b; Merz et al., 2017) and their role in growth responses on elevated concentrations of Cd, Cu, Ni, Pb, and Zn. With the help of loss- and gain-of-function alleles, a complex pattern of gene specific, overlapping and antagonistic reactions was revealed. Root growth was often inversely affected to hypocotyl elongation. Antagonistic roles of the two *HERK* genes and *FER* versus *THE1* were discovered in relation to root growth on Cu, in relation to hypocotyl elongation on Pb and Zn, and between *THE1* and *FER* on Ni. The effect of metal ions on the acidification ability was evaluated with bromocresol purple indicator medium. While *fer-4* roots strongly acidified control and metal ion containing media, etiolated hypocotyls alkalized the media which is consistent with the *fer-4* elongation defect. No other CrRLK1L mutant exhibited this phenotype except for the THE1:GFP overexpressor on Ni suggesting that THE1 might be involved in an Ni induced and hypocotyl specific rapid alkalization factor (RALF) signaling and growth regulating pathway. Overall, our findings establish a molecular link between metal ion stress, growth and the cell wall integrity sensors of the CrRLK1L family.

MATERIALS AND METHODS

Plant Material

Col-0 was used as wild type. T-DNA mutants were all in Col-0 background and provided either by the SALK collection (Alonso et al., 2003), *herk1* (SALK_008043), *herk2.1* (SALK_105055), *herk2.2* (SALK_107146), the SAIL collection (Sessions et al., 2002), *the1-4* (SAIL_683_H03), and the GABI-Kat collection (Kleinboelting et al., 2012), *fer-4* (GABI_GK106A06). The loss-of-function allele, *the1-6*, was isolated in a suppressor screen of *ctl1-1/pom* and described in Merz et al. (2017). The THE1:GFP overexpression line was described in Hématy et al. (2007).

Growth Conditions

Seeds were surface-sterilized in 5% sodium hypochlorite and rinsed three times with sterile deionized water and then transferred to nutrient agar medium plates containing 1/10 strength Hoagland salts, 1% (w/v) sucrose and 1% (w/v) agar (Duchefa). For metal ion treatments metal salts were added after autoclaving to final concentrations of 10 μM CdCl_2 , 5 μM CuSO_4 , 15 μM NiSO_4 , 100 μM $\text{Pb}(\text{NO}_3)_2$, and 100 μM ZnSO_4 . After 2 days of imbibition at 4°C in the dark, plates were vertically incubated in a growth chamber at 22°C with constant light (80 $\mu\text{mol m}^{-2} \text{s}^{-1}$). For measurements of etiolated hypocotyls, plates were wrapped in aluminum foil after exposure to light for 5 h. Isoxaben treatment for gene expression analyses was essentially done as described in Merz et al. (2017). Ni treatment for gene expression analyses was done for 6 h on 4 days after germination (dag) etiolated seedlings.

Bromocresol Purple (BCP) Plates

For pH assays sterilized seeds were placed on nutrient agar plates as described above supplemented with 150 μM BCP and metal ions in indicated concentrations. Plates were scanned on day 5 after germination.

Growth Analysis

The plates were scanned on days 3, 4, and 5 after germination for root growth and on day 5 after germination for hypocotyl measurements. The lengths were evaluated with the ImageJ software by freehand tracking.

RNA Isolation and cDNA Synthesis

Total RNA of 10–12 day old seedlings germinated and grown on control and metal ion supplemented medium were snap frozen and isolated using a LiCl/CTAB method. After grinding roughly 100 mg frozen seedlings 1 mL of pre-heated RNA extraction buffer (2% [w/v] hexadecyltrimethylammonium bromide, CTAB; 2% [w/v] polyvinylpyrrolidone, PVP; 100 mM Tris/HCl pH 8.0; 25 mM EDTA; 2 M NaCl; 0.5 g/L spermidine and 2.7% [v/v] 2-mercaptoethanol) was added, mixed and incubated at 65°C for 5 min. CTAB was removed through two times separation with 1 mL of ice-cold chloroform: isoamylalcohol (24:1) and centrifugation at 4°C. RNA in the supernatant was precipitated with 250 μL 10 M LiCl at 4°C for more than 1.5 h. After centrifugation and EtOH washes the pellet was dissolved in 20 μL RNase free water and stored at -80°C . RNA was quantified with the Qubit (Invitrogen) and the NanoDrop systems. RNA integrity was controlled on Agilent 2100 Bioanalyzer using RNA Nanochip. RNA samples were treated with DNaseI using the TURBO DNA-free™ kit (Ambion). cDNAs were synthesized from 4 μg total RNA using the Transcriptor reverse transcriptase (Roche) with 500 pmol of oligo(dT)₁₈ primer. The reaction was stopped at 85°C for 5 min without further treatment according to the manufacturer's instructions. cDNA was diluted 20 times with distilled water and tested by PCR using specific primers flanking an intron sequence to confirm the absence of genomic DNA contamination. cDNA synthesis for the

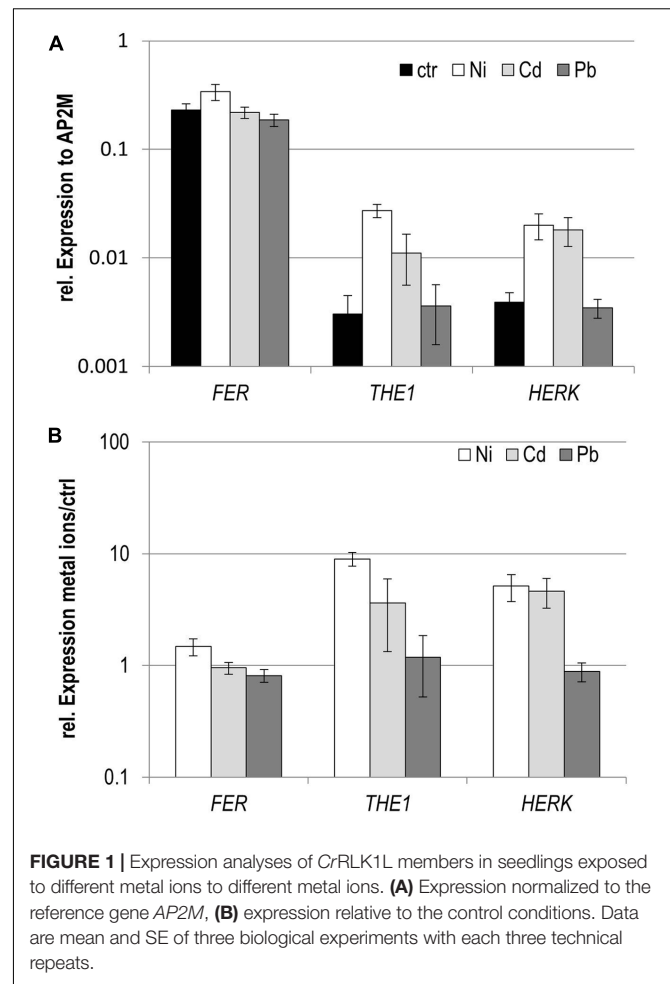


FIGURE 1 | Expression analyses of CrRLK1L members in seedlings exposed to different metal ions to different metal ions. **(A)** Expression normalized to the reference gene *AP2M*, **(B)** expression relative to the control conditions. Data are mean and SE of three biological experiments with each three technical repeats.

THE1 downstream gene expression was done as described in Merz et al. (2017).

Reverse Transcription (RT)-qPCR Data Analysis

AP2M (*ADAPTOR PROTEIN-2 MU-ADAPTIN*) has been validated to be the most stably expressed gene among eight tested and was used to normalize the RT-qPCR data (Gutierrez et al., 2012). CT and PCR efficiency (E) values were used to calculate expression using the formula $E_T^{(CT_{ctr} - CT_m)} / E_R^{(CT_{ctr} - CT_m)}$, where T is the target gene and R is the reference gene, CT is the crossing threshold value, m refers to cDNA from the metal ion treated seedlings, and ctr refers to cDNA from the control medium. All RT-qPCR results presented are means from three independent biological replicates and for each independent biological replicate, the relative transcript amount was calculated as the mean of three technical replicates, using the method for calculation of SE values in relative quantification recommended by Rieu and Powers (2009). RT-qPCR for the *THE1* downstream gene expression was done as described in Merz et al. (2017). Primers used for the RT-qPCRs are summarized in Supplementary Table 2.

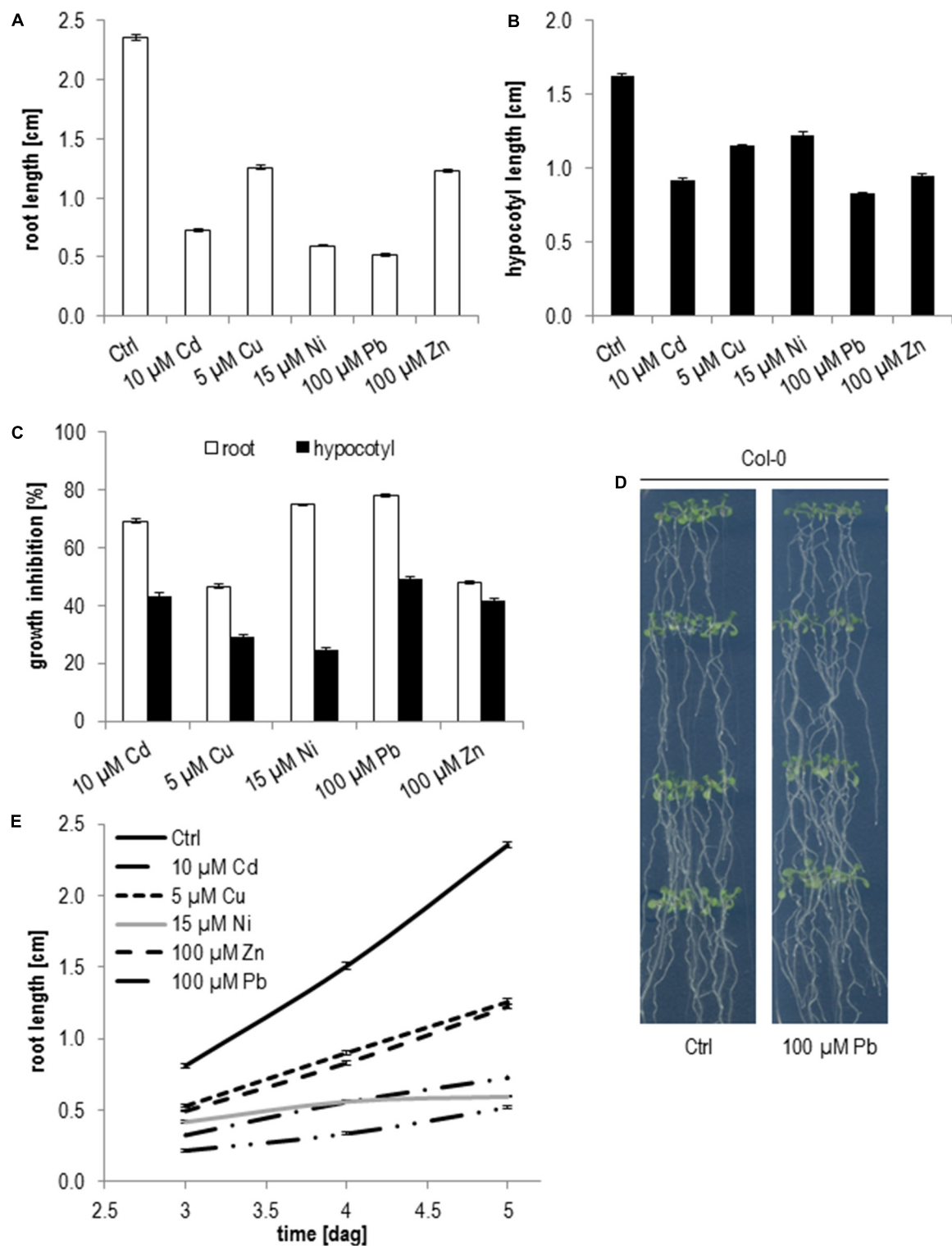


FIGURE 2 | Root growth and etiolated hypocotyl elongation of Col-0 seedlings upon elevated metal ion concentrations. **(A)** Root length 5 days after germination, **(B)** hypocotyl length after 5 days of germination in the dark, **(C)** growth inhibition relative to control medium, **(D)** root growth between days 3 and 5 after germination, **(E)** seedlings on control and Pb 11 days after germination. Shown are mean and SE of at least three biological replicates with 20 seedlings each.

RESULTS AND DISCUSSION

Members of the CrRLK1L Family Are Induced upon Metal Ion Exposure

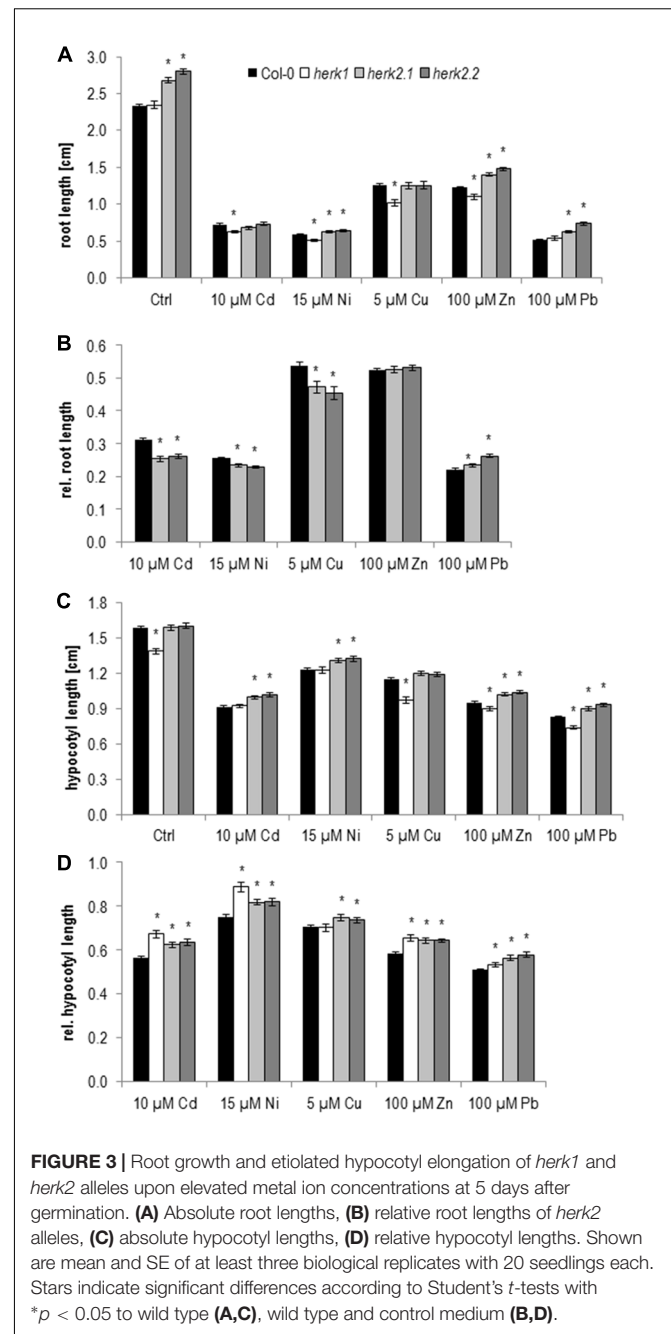
In an expression survey for genes responsive to the metal ions Cd, Ni, and Pb in seedlings, the CrRLK1L members *THE1*, *FER*, and *HERK1* exhibited a remarkable expression pattern. All three were induced by Ni, only slightly for *FER* but strongly for *HERK1* and *THE1* (Figure 1). Pb did not induce any transcriptional regulation whereas Cd triggered upregulation of *THE1* and *HERK1*.

Root and Etiolated Hypocotyl Growth Differ in Response to Metal Ion Medium

Growth and cell expansion responses of germinating *Arabidopsis* seedlings to elevated metal ion concentrations depend strongly on the composition of the growth medium and on the cation exchange capacity of the agar. The high cation concentration such as provided by the frequently used 1x Murashige and Skoog medium (Murashige and Skoog, 1962) interferes with the uptake of additional metal ions. On 1x MS medium 400 μ M Zn and 40 μ M Cd were needed to observe a slight root length reduction for Zn and a 30% reduction for Cd (Kobae et al., 2004). A comparable experiment with germinating seedlings on 1/10 Hoagland medium supplemented with metal ions needed only 50 μ M Zn for a 43.2% and 2 μ M Cd for a 57.6% root growth inhibition (Tennstedt et al., 2009). Since the MS medium is also Fe-saturated and Cu-deficient all metal ion experiments were performed on 1/10 Hoagland medium (Supplementary Table 1).

Root growth depends on two parameters, the rate of cell division and cell expansion, while dark stimulated hypocotyl growth depends solely on cell expansion. To separate the cell division from cell expansion effects, root growth was determined on seedlings germinating and grown for 5 days in the light and hypocotyl elongation was assessed on 5 dag dark-grown seedlings. In both sets of experiments identical 1/10 Hoagland media supplemented with either 10 μ M Cd, 5 μ M Cu, 15 μ M Ni, 100 μ M Pb, or 100 μ M Zn were used.

Generally, hypocotyl elongation is less sensitive to metal ions (Figures 2A–C). While on 10 μ M Cd root growth was reduced by 70% in relation to control medium the growth inhibition of hypocotyls was only 43%. Similar results were obtained for 5 μ M Cu with 46% root and 30% hypocotyl inhibition, 100 μ M Zn with 47% root and 41% hypocotyl inhibition, 15 μ M Ni with 74% root and 25% hypocotyl inhibition and 100 μ M Pb with 78% root and 49% hypocotyl inhibition (Figure 2C) indicating that these metal ions might influence cell division more severely than cell expansion. It might also be possible that cell expansion in the root is differently affected by these metal ions than in etiolated hypocotyls. The difference between root and hypocotyl growth is particularly true for Ni where root growth rate was continuously declining and nearly stopped 5 days



after germination (dag) (Figure 2D). Pb shows the opposite effect and root growth recovered starting at 5 dag and roots reached nearly the same length as wild type at 11 dag (Figure 2E).

Apart of the dissimilar effects of metal ions on cell division and cell expansion another explanation for the differing sensitivities between root and hypocotyl growth would be that metal ion transport to hypocotyls is less effective. Data on metal ion concentrations in separated roots and hypocotyls and quantifications of the cell division rate and cell sizes would resolve these possibilities. Although the results of these analyses would contribute to the understanding of the root/hypocotyl response

TABLE 1 | Summary of the significant growth and elongation responses in comparison to wild type.

Genotype	ctrl		Cd		Cu		Ni		Pb		Zn	
	r	h	r	h	r	h	r	h	r	h	r	h
<i>herk1</i>	=	↓	↓	↑	↓	=	↓	↑	=	↑	↓	↑
<i>herk2.1</i>	↑	=	↓	↑	↓	=	↓	↑	↑	↑	=	↑
<i>herk2.2</i>	↑	=	↓	↑	↓	=	↓	↑	↑	↑	=	↑
<i>the1-6</i>	↓	=	↓	=	=	=	↑	↑	=	↓	↓	↓
<i>the1-4</i>	=	↓	↑	=	↓	=	↑	↓	=	↑	=	↑
THE1:GFP	↑	↓	↑	↑	↓	↑	↓	↓	=	=	=	↑
<i>fer-4</i>	↓	↓	=	↑	↓	↑	=	↓	↑	↑	↓	↑

Data of roots and hypocotyls are indicated with *r* and *h*, respectively. In case growth on control medium was already significantly different between wild type and the CrRLK1L mutants the significances of the normalized data are employed.

differences, the focus of this study was to determine if members of the CrRLK1L family are involved in mediating growth responses to metal ions.

HERCULES1 and 2 Negatively Regulate Hypocotyl Elongation on Cd, Ni, Zn, and Pb While They Support Root Growth on Cd, Cu, and Ni

Previous growth assays with single mutants impaired in *HERK1* and *HERK2* expression did not show any obvious growth phenotype. But the mutants developed strong cell elongation defects in combination with the *the1-4* allele (Guo et al., 2009a,b). However *the1-4* turned out to be a hypermorphic allele. Thus the growth defects of the double and triple mutants indicate that *HERK1* and *HERK2* act antagonistically and not redundantly to *THE1* (Merz et al., 2017). On 1/10 Hoagland control medium, roots of *herk1* seedlings grow similarly to wild type in contrast to both *herk2* alleles, which exhibited a significantly faster root growth (Figure 3A and Table 1).

On metal ion supplemented medium root growth of *herk1* seedlings is stronger inhibited upon Cd, Ni, Cu, and Zn than wild type. Since the two *herk2* alleles already grow faster on control medium, analyzing the effect of metal ions on these mutants by only looking at the absolute root length data might be misleading. Therefore, their root growth on metal ions was normalized to control conditions (Figure 3B and Table 1). These analyses revealed that the apparent higher tolerance to Ni, and Zn diminished and only the reduced lead response remained while an enhanced Cd, Cu, and Ni sensitivity was revealed. In summary, both *HERK* genes are likely to be important to support root growth upon higher Cd, Cu, and Ni concentrations while the Zn response is *HERK1* specific and the Pb response is *HERK2* specific.

As with wild type the hypocotyl elongation in the mutants differs from root growth responses. Already on control medium *herk1* elongates significantly less than wild type and the two *herk2* alleles (Figure 3C and Table 1). Therefore, normalization to control conditions was necessary to reveal that hypocotyl elongation of *herk1* was less disturbed by Cd, Ni, Zn, and Pb than wild type (Figure 3D and Table 1). Identical

responses were found in the two *herk2* mutants (Figures 3C,D and Table 1). In conclusion, both *HERK* genes appear to mediate the inhibition of hypocotyl elongation upon Cd, Ni, Zn, and Pb while they support root growth on Cd, Cu, and Ni.

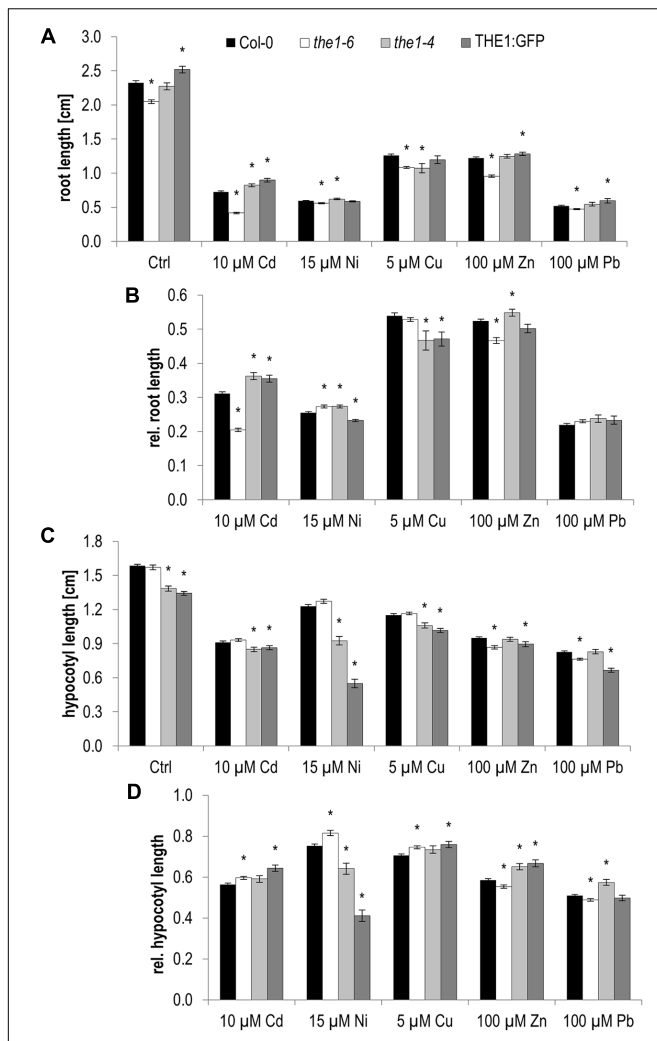
THESEUS1 Is Mediating Ni Specific Inhibition of Hypocotyl Elongation in the Dark without Cell Wall Damage Responses

THE1 signaling negatively affects cell expansion upon inhibition of cellulose synthesis (Hématy et al., 2007; Merz et al., 2017). Therefore *THE1* is the strongest candidate to uncover metal ion induced cell wall damage responses. Well-characterized loss-of-function and gain-of-function alleles were available to compare their responses to metal ions.

Already on control medium root growth of *THE1*-related knock-out and overexpression lines behaved inversely (Figure 4A and Table 1). While the loss-of-function mutant *the1-6* developed shorter roots, roots of the THE1:GFP overexpressor grew faster than wild type. Similar contrasting root growth responses were significant on Cd and Ni. On Cu only THE1:GFP and the hypermorphic allele *the1-4* developed shorter roots as did *the1-6* on Zn (Figure 4B and Table 1).

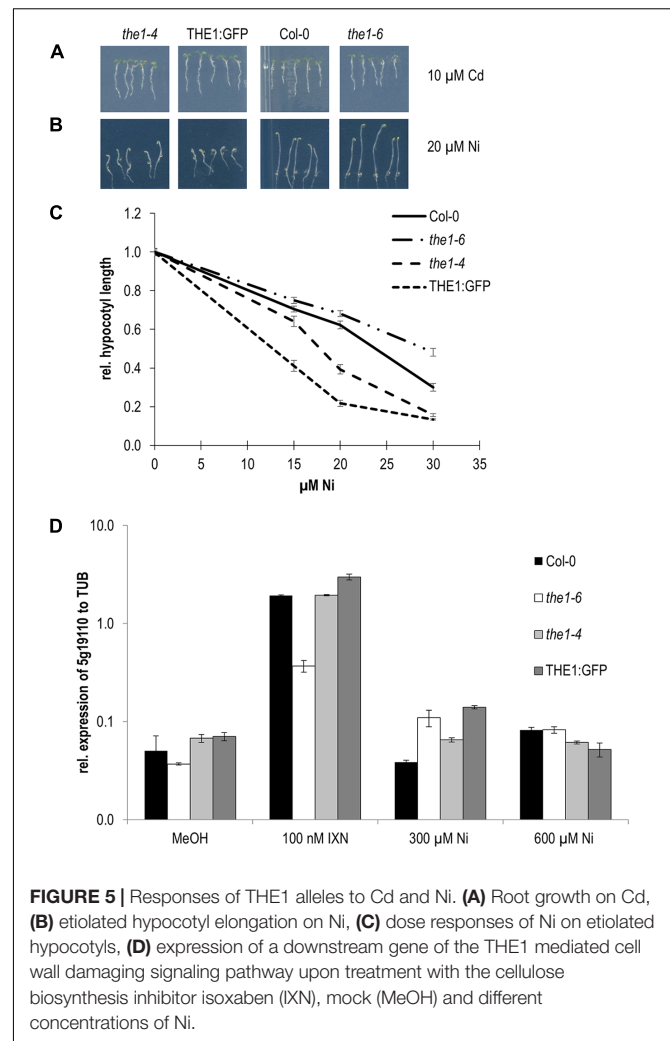
We previously showed that the hypermorphic *the1-4* expresses a transcript encoding a predicted membrane-associated truncated protein lacking the kinase domain. Differences between THE1:GFP and the hypermorphic *the1-4* allele, as seen on control, Ni and Zn, possibly relate to the missing cytoplasmic domain in *the1-4* which might be important for feedback regulation (Merz et al., 2017).

Hypocotyl elongation in the dark behaved different to roots for *the1-6* on control Cd, and Pb, for THE1:GFP on control, Cu and Zn and for *the1-4* on all conditions (Figures 4C,D and Table 1). While in roots the strongest effect was shown by *the1-6* on Cd, for hypocotyls it was on Ni by *the1-4* and THE1:GFP (Figure 5A). The opposing effects of the loss- and gain-of function alleles on hypocotyl elongation were even more distinct on 20 and 30 μ M Ni (Figures 5B,C). These results indicate that THE1 mediates



Ni specific inhibition of hypocotyl elongation in the dark and promotes Cd specific growth tolerance in roots.

As mentioned above *THE1* is known to inhibit cell elongation in hypocotyls upon cell wall damage. Therefore, we examined if Ni induces cell wall damage specific responses such as the expression of the downstream gene in the *THE1* signaling pathway, *EDGP/At5g19110*. The expression of *At5g19110* was strongly induced in the overexpressor and the hypermorphic alleles and did not respond in the loss-of-function allele, *the1-6* in chemically induced cell wall damage by treatment with the cellulose biosynthesis inhibitor isoxaben. Upon Ni treatment a very low expression in the different *THE1* alleles was quantified with no significant alterations to wild type **(Figure 5D)**. These data suggest that Ni perception by *THE1*



is independent of cell wall damage specific gene expression responses.

FERONIA Mediates Growth Inhibition of Hypocotyl on Cd, Cu, Pb, and Zn While it Promotes Growth on Ni

FER is the best studied member of the *CrRLK1L* gene family. *FER* has been identified due to its crucial function during pollen-tube ovule interaction (Escobar-Restrepo et al., 2007) but is also important for bacterial pathogen interaction (Keinath et al., 2010; Stegmann et al., 2017), mechanosensing (Shih et al., 2014) and in the antagonistic cross-talk between ABA and auxin in vegetative tissues (Yu et al., 2012). Mutants of *FER* exhibit diverse pleiotropic phenotypes. They are stunted, develop smaller leaves (Guo et al., 2009b) and shorter etiolated hypocotyls which are hypersensitive to ethylene and insensitive to brassinosteroids (Deslauriers and Larsen, 2010). Moreover *fer* mutants develop shorter root hairs (Duan et al., 2010; Du et al., 2016) and larger seeds (Yu et al., 2014) and exhibit a reduced gravitropic response

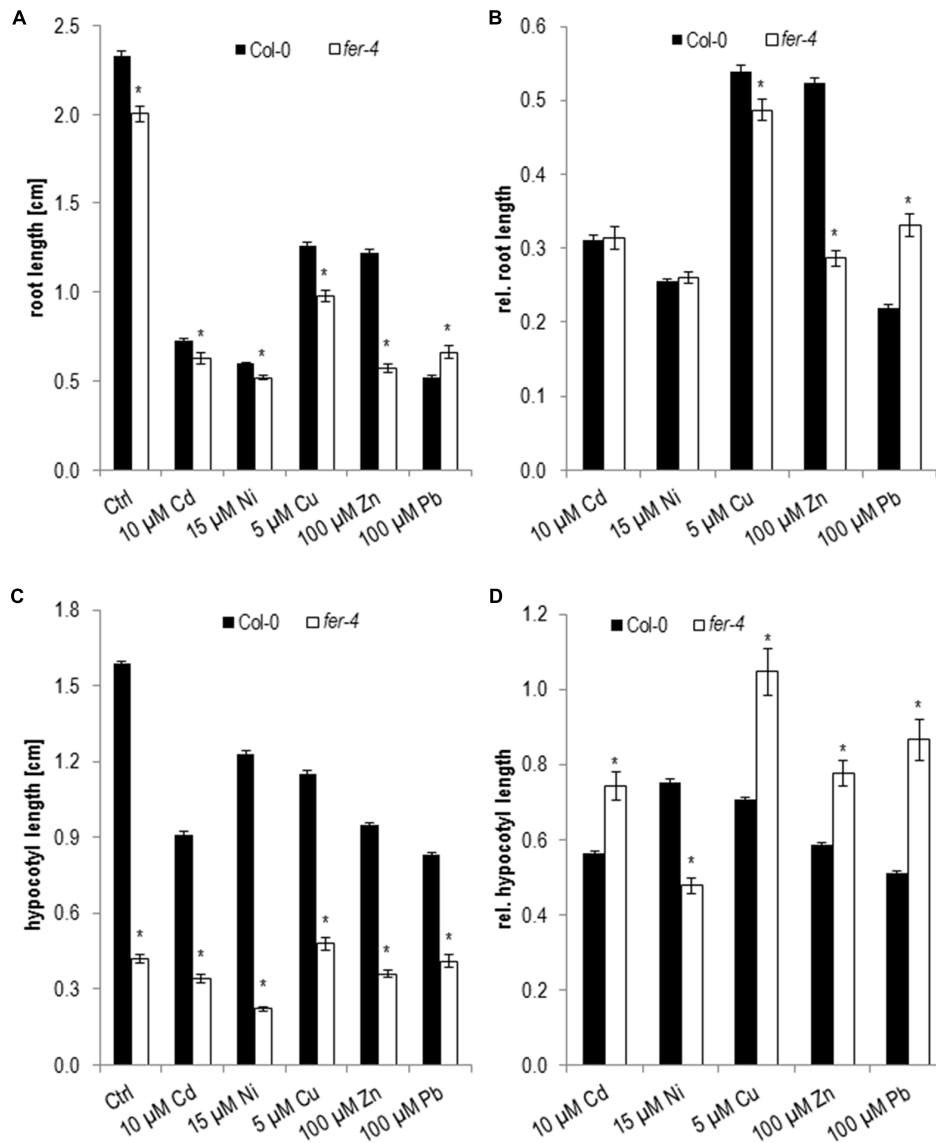


FIGURE 6 | Root growth and hypocotyl elongation of *fer-4* on medium supplemented with elevated metal ion concentrations at 5 days. **(A)** Absolute root length, **(B)** corresponding relative root lengths, **(C)** absolute hypocotyl lengths, **(D)** corresponding relative hypocotyl lengths. Shown are mean and SE of at least three biological replicates with 20 seedlings each. Stars indicate significant differences according to Student's *t*-tests with $*p < 0.05$ to wild type **(A,C)**, wild type and control medium **(B,D)**.

which correlates with less apoplasmic alkalization at the lower side of gravistimulated roots (Barbez et al., 2017).

In our growth analyses root lengths of 5 days old *fer-4* seedlings were significantly shorter on control medium and in most metal ion treatments except on Pb (Figure 6A and Table 1). However when root growth was normalized to control conditions *fer-4* was hypersensitive to Zn and Cu while it was more tolerant to Pb (Figure 6B and Table 1). In concordance with published data etiolated *fer-4* hypocotyl reached only 25% of the size of wild type (Figure 6C and Table 1). Thus the different effects of the metal ions were revealed after normalization to control conditions (Figure 6D and Table 1) and showed that *fer-4* is

hypersensitive to Ni and has a higher tolerance to all other tested metal ions.

Medium Alkalinization of Etiolated Seedlings Is Constitutive in *fer-4* Hypocotyls and Ni Specific in the THE1 Overexpressor

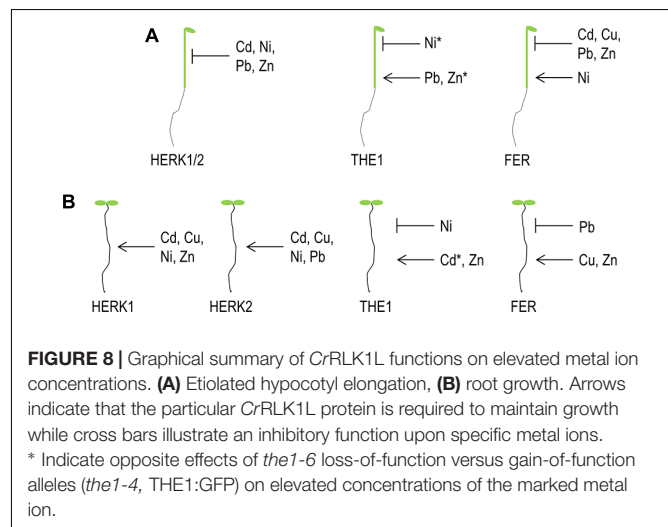
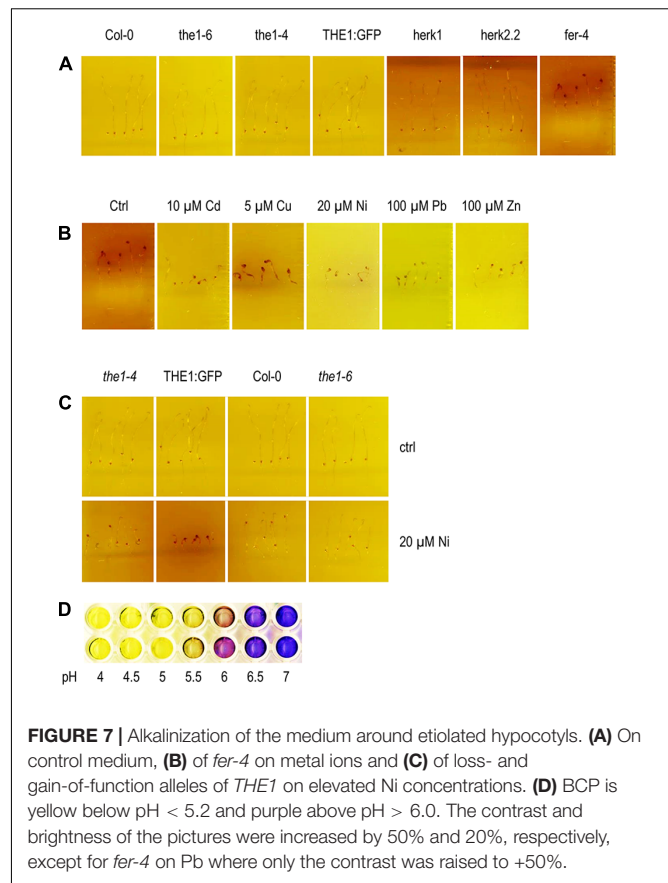
FER binds and is activated by secreted peptides of the RALF family (Haruta et al., 2014) triggering alkalization of the apoplast. This alkalization is thought to counteract the acidification crucial for cell expansion. Light grown *fer-4*

seedlings acidified a bath medium faster than wild type which is indicative for a higher plasma membrane H^+ -ATPase activity (Haruta et al., 2014).

Since growth of etiolated hypocotyl depends on rapid cell expansion we tested the acidification capacity of the *CrRLK1L* mutants on control medium and upon metal ion treatments (Figures 7A–C). All genotypes acidified the medium supplemented with the pH indicator bromocresolpurple (BCP) (Figure 7D). Only *fer-4* exhibited a remarkable alkalization around the hypocotyl while the root was still acidifying the medium (Figure 7B). This alkalization might be one of the reasons why etiolated hypocotyls of *fer-4* are short and indicate that the plasma membrane H^+ -ATPase activity is differently regulated from roots. The phenotype further suggests that *FER* is necessary for the acidification of etiolated hypocotyls. It is conceivable that in the absence of *FER RALF* expression is triggered and another receptor in the hypocotyl mediates the alkalization. The alkalization surrounding the *fer-4* etiolated hypocotyl was still visible on medium supplemented with metal ions. Hypocotyl mediated alkalization of the medium is strongest on control medium, followed by Cu, Ni, and only slightly on Cd and Zn. Since, the medium supplemented with Pb has a lower pH itself the alkalization takes longer. The only other genotype with reduced acidification capacity was the overexpressor *THE1:GFP* on Ni (Figure 7C and Supplementary Figure 1). Similar to *fer-4*, *THE1:GFP* developed shorter hypocotyls on Ni. It might be possible that overexpression of *THE1:GFP* increases the sensitivity to Ni induced hypocotyl specific RALFs and that *THE1* might be involved in their signaling pathway.

CONCLUSION

Our survey revealed a molecular link between metal ion stress, growth and the cell wall integrity sensors of the *CrRLK1L* family. By analyzing growth responses in roots and hypocotyls a complex network of patterns of gene specific, overlapping and antagonistic responses which are summarized in Table 1 and Figure 8 was uncovered. Few general patterns were extractable of the data: in all *HERK1*, *HERK2* and *FER* mutants root growth is affected inversely to hypocotyl elongation. This tendency is also true for the different *THE1* alleles but for example on Zn in the opposite direction. The hypocotyl of the gain-of-function and hypermorphic alleles of *THE1* behaved similar to *HERK1*, *HERK2*, and *FER* while the loss-of-function allele exhibited hypersensitivity toward Zn (Table 1). The difference of the metal ion mediated growth responses might arise as a result of the presence of different ligands in roots and fast expanding etiolated hypocotyls. One of the candidate ligands might be members of the large *RALF* gene family. *RALF1* has been shown to bind and activate *FER* leading to the phosphorylation and inhibition of the plasma membrane H^+ -ATPase *AHA2*, reduction of the apoplast/cell wall acidification and the inhibition of cell expansion genes (Haruta et al., 2014). It is possible that other *RALFs* with different expression and pro-peptide processing patterns are triggering similar reactions in conjunction with



other *CrRLK1Ls* (Wolf and Höfte, 2014). Other possible ligands related to the putative carbohydrate binding feature of the extracellular malectin-like domain might be metal ions loaded pectin/homogalacturonans or their degradation products, oligogalacturonans. Boisson-Dernier et al. (2011) proposed for the pollen specific *CrRLK1L* member, *ANXUR1* and 2 (*ANX1* and *ANX2*) a possible interaction between homogalacturonans. This hypothesis was based of the premature bursting of pollen

tubes in the double mutants of *anx1/anx2* and the pollen specific expression of pectin modifying enzymes which correlated with a specific de-/methylsterification at the pollen tube tip and thus mechanical property. A related hypothesis was presented by Wolf and Höfte (2014) proposing a feedback loop between RALF and pectin modifying enzymes. RALF induced alkalization of the cell wall would activate PME which upon removal of the methyl groups lead to cell wall acidification and cell expansion. Based on this hypothesis, metal ions might be complexed by demethylsterified pectins and stiffens the cell wall prematurely before cell expansion is completed. The puzzle why different metal ions trigger specific CrRLK1Ls and why and how their signaling outcome induces opposite effects on growth remains to be solved in the future.

AUTHOR CONTRIBUTIONS

JR and M-TH designed the study and JR, MP, and GM performed the experiments and JR, M-TH, and LG analyzed the data. M-TH and JR wrote the manuscript.

REFERENCES

- Alonso, J. M., Stepanova, A. N., Leisse, T. J., Kim, C. J., Chen, H. M., and Shinn, P. (2003). Genome-wide insertional mutagenesis of *Arabidopsis thaliana*. *Science* 301, 653–657. doi: 10.1126/science.1086391
- Bai, L., Ma, X., Zhang, G., Song, S., Zhou, Y., Gao, L., et al. (2014). A receptor-like kinase mediates ammonium homeostasis and is important for the polar growth of root hairs in *Arabidopsis*. *Plant Cell* 26, 1497–1511. doi: 10.1105/tpc.114.124586
- Bai, L., Zhang, G., Zhou, Y., Zhang, Z., Wang, W., Du, Y., et al. (2009). Plasma membrane-associated proline-rich extensin-like receptor kinase 4, a novel regulator of Ca²⁺ signalling, is required for abscisic acid responses in *Arabidopsis thaliana*. *Plant J.* 60, 314–327. doi: 10.1111/j.1365-313X.2009.03956.x
- Barbez, E., Dünser, K., Gaidora, A., Lendl, T., and Busch, W. (2017). Auxin steers root cell expansion via apoplastic pH regulation in *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. U.S.A.* 114, E4884–E4893. doi: 10.1073/pnas.1613499114
- Boisson-Dernier, A., Kessler, S. A., and Grossniklaus, U. (2011). The walls have ears: the role of plant CrRLK1Ls in sensing and transducing extracellular signals. *J. Exp. Bot.* 62, 1581–1591. doi: 10.1093/jxb/erq445
- Boisson-Dernier, A., Roy, S., Kritsas, K., Grobei, M. A., Jaciubek, M., Schroeder, J. I., et al. (2009). Disruption of the pollen-expressed FERONIA homologs ANXUR1 and ANXUR2 triggers pollen tube discharge. *Development* 136, 3279–3288. doi: 10.1242/dev.040071
- DalCorso, G., Manara, A., and Furini, A. (2013). An overview of heavy metal challenge in plants: from roots to shoots. *Metallomics* 5, 1117–1132. doi: 10.1039/c3mt00038a
- Decreux, A., and Messiaen, J. (2005). Wall-associated kinase WAK1 interacts with cell wall pectins in a calcium-induced conformation. *Plant Cell Physiol.* 46, 268–278. doi: 10.1093/pcp/pci026
- Decreux, A., Thomas, A., Spies, B., Brasseur, R., Van Cutsem, P., and Messiaen, J. (2006). In vitro characterization of the homogalacturonan-binding domain of the wall-associated kinase WAK1 using site-directed mutagenesis. *Phytochemistry* 67, 1068–1079. doi: 10.1016/j.phytochem.2006.03.009
- Deslauriers, S. D., and Larsen, P. B. (2010). FERONIA is a key modulator of brassinosteroid and ethylene responsiveness in *Arabidopsis* hypocotyls. *Mol. Plant* 3, 626–640. doi: 10.1093/mp/ssq015

FUNDING

The project was supported by a grant of the Austrian Science Fund FWF I 1725-B16 and the French National Research Agency project ANR-FWF I 1725-B16.

ACKNOWLEDGMENTS

We thank Francois Jobert for the design of the THE1, HERK1, and FER primers used in the RT-qPCR. We thank the seed stock distribution center Nottingham Arabidopsis Stock Centre (NASC), Kai Dünser for the *fer-4* and Herman Höfte and Marjolaine Martin for *herk1*, *herk2.1*, and *herk2.2* mutants.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fpls.2017.01554/full#supplementary-material>

- Douchiche, O., Soret-Morvan, O., Chaibi, W., Morvan, C., and Paynel, F. (2010). Characteristics of cadmium tolerance in ‘Hermes’ flax seedlings: contribution of cell walls. *Chemosphere* 81, 1430–1436. doi: 10.1016/j.chemosphere.2010.09.011
- Dronnet, V. M., Renard, C. M. G. C., Axelos, M. A. V., and Thibault, J. F. (1996). “Heavy metals binding by pectins: selectivity, quantification and characterisation,” in *Progress in Biotechnology*, ed. J.V.a.A.G.J. Voragen (Amsterdam: Elsevier), 535–540.
- Du, C., Li, X., Chen, J., Chen, W., Li, B., Li, C., et al. (2016). Receptor kinase complex transmits RALF peptide signal to inhibit root growth in *Arabidopsis*. *Proc. Natl. Acad. Sci. U.S.A.* 113, E8326–E8334. doi: 10.1073/pnas.1609626113
- Duan, Q., Kita, D., Li, C., Cheung, A. Y., and Wu, H.-M. (2010). FERONIA receptor-like kinase regulates RHO GTPase signaling of root hair development. *Proc. Natl. Acad. Sci. U.S.A.* 107, 17821–17826. doi: 10.1073/pnas.1005366107
- El-Moneim, D. A., Contreras, R., Silva-Navas, J., Gallego, F. J., Figueiras, A. M., and Benito, C. (2014). Pectin methylsterase gene and aluminum tolerance in *Secale cereale*. *Environ. Exp. Bot.* 107, 125–133. doi: 10.1016/j.envexpbot.2014.06.006
- Engelsdorf, T., and Hamann, T. (2014). An update on receptor-like kinase involvement in the maintenance of plant cell wall integrity. *Ann. Bot.* 114, 1339–1347. doi: 10.1093/aob/mcu043
- Escobar-Restrepo, J.-M., Huck, N., Kessler, S., Gagliardini, V., Gheyselinck, J., Yang, W.-C., et al. (2007). The FERONIA receptor-like kinase mediates male-female interactions during pollen tube reception. *Science* 317, 656–660. doi: 10.1126/science.1143562
- Gachomo, E. W., Jno Baptiste, L., Kefela, T., Saidel, W. M., and Kotchoni, S. O. (2014). The *Arabidopsis* CURVY1 (CVY1) gene encoding a novel receptor-like protein kinase regulates cell morphogenesis, flowering time and seed production. *BMC Plant Biol.* 14:221. doi: 10.1186/s12870-014-0221-7
- Geng, X., Horst, W. J., Golz, J. F., Lee, J. E., Ding, Z., and Yang, Z.-B. (2017). LEUNIG_HOMOLOG transcriptional co-repressor mediates aluminium sensitivity through PECTIN METHYLESTERASE46-modulated root cell wall pectin methylsterification in *Arabidopsis*. *Plant J.* 90, 491–504. doi: 10.1111/tbj.13506
- Guo, H., Li, L., Ye, H., Yu, X., Algreen, A., and Yin, Y. (2009a). Three related receptor-like kinases are required for optimal cell elongation in *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. U.S.A.* 106, 7648–7653. doi: 10.1073/pnas.0812346106

- Guo, H., Ye, H., Li, L., and Yin, Y. (2009b). A family of receptor-like kinases are regulated by BES1 and involved in plant growth in *Arabidopsis thaliana*. *Plant Signal. Behav.* 4, 784–786. doi: 10.1073/pnas.0812346106
- Gutierrez, L., Mongelard, G., Floková, K., Pácurar, D. I., Novák, O., Staswick, P., et al. (2012). Auxin controls *Arabidopsis* adventitious root initiation by regulating jasmonic acid homeostasis. *Plant Cell* 24, 2515–2527. doi: 10.1105/tpc.112.099119
- Haruta, M., Sabat, G., Stecker, K., Minkoff, B. B., and Sussman, M. R. (2014). A peptide hormone and its receptor protein kinase regulate plant cell expansion. *Science* 343, 408–411. doi: 10.1126/science.1244454
- Hassinen, V. H., Tervahauta, A. I., Halimaa, P., Plessl, M., Peraniemi, S., Schat, H., et al. (2007). Isolation of Zn-responsive genes from two accessions of the hyperaccumulator plant *Thlaspi caerulescens*. *Planta* 225, 977–989. doi: 10.1007/s00425-006-0403-0
- Hématy, K., Sado, P.-E., Van Tuinen, A., Rochange, S., Desnos, T., Balzergue, S., et al. (2007). A receptor-like kinase mediates the response of *Arabidopsis* cells to the inhibition of cellulose synthesis. *Curr. Biol.* 17, 922–931. doi: 10.1016/j.cub.2007.05.018
- Hou, X., Tong, H., Selby, J., Dewitt, J., Peng, X., and He, Z. H. (2005). Involvement of a cell wall-associated kinase, WAKL4, in *Arabidopsis* mineral responses. *Plant Physiol.* 139, 1704–1716. doi: 10.1104/pp.105.066910
- Kartel, M. T., Kupchik, L. A., and Veisov, B. K. (1999). Evaluation of pectin binding of heavy metal ions in aqueous solutions. *Chemosphere* 38, 2591–2596. doi: 10.1016/S0045-6535(98)00466-4
- Keinath, N. F., Kierszniowska, S., Lorek, J., Bourdais, G., Kessler, S. A., Shimosato-Asano, H., et al. (2010). PAMP (pathogen-associated molecular pattern)-induced changes in plasma membrane compartmentalization reveal novel components of plant immunity. *J. Biol. Chem.* 285, 39140–39149. doi: 10.1074/jbc.M110.160531
- Kleinboelting, N., Huep, G., Kloetgen, A., Viehoveer, P., and Weisshaar, B. (2012). GABI-Kat SimpleSearch: new features of the *Arabidopsis thaliana* T-DNA mutant database. *Nucleic Acids Res.* 40, D1211–D1215. doi: 10.1093/nar/gkr1047
- Kobae, Y., Uemura, T., Sato, M. H., Ohnishi, M., Mimura, T., Nakagawa, T., et al. (2004). Zinc transporter of *Arabidopsis thaliana* AtMTP1 is localized to vacuolar membranes and implicated in zinc homeostasis. *Plant Cell Physiol.* 45, 1749–1758. doi: 10.1093/pcp/pci015
- Kohorn, B. D., Kobayashi, M., Johansen, S., Friedman, H. P., Fischer, A., and Byers, N. (2006). Wall-associated kinase 1 (WAK1) is crosslinked in endomembranes, and transport to the cell surface requires correct cell-wall synthesis. *J. Cell Sci.* 119, 2282–2290. doi: 10.1242/jcs.02968
- Konlechner, C., Turktas, M., Langer, I., Vaculik, M., Wenzel, W. W., Puschenreiter, M., et al. (2013). Expression of zinc and cadmium responsive genes in leaves of willow (*Salix caprea* L.) genotypes with different accumulation characteristics. *Environ. Pollut.* 178, 121–127. doi: 10.1016/j.envpol.2013.02.033
- Krzyszowska, M. (2011). The cell wall in plant cell response to trace metals: polysaccharide remodeling and its role in defense strategy. *Acta Physiol. Plant.* 33, 35–51. doi: 10.1007/s11738-010-0581-z
- Lally, D., Ingmire, P., Tong, H. Y., and He, Z. H. (2001). Antisense expression of a cell wall-associated protein kinase, WAK4, inhibits cell elongation and alters morphology. *Plant Cell* 13, 1317–1331. doi: 10.1105/tpc.13.6.1317
- Merz, D., Richter, J., Gonneau, M., Sanchez-Rodriguez, C., Eder, T., Sormani, R., et al. (2017). T-DNA alleles of the receptor kinase THESEUS1 with opposing effects on cell wall integrity signaling. *J. Exp. Bot.* doi: 10.1093/jxb/erx263
- Meychik, N. R., Nikolaeva, Y. I., Komarynets, O. V., and Ermakov, I. P. (2011). Barrier function of the cell wall during uptake of nickel ions. *Russ. J. Plant Physiol.* 58, 409–414. doi: 10.1134/S1021443711030137
- Miyazaki, S., Murata, T., Sakurai-Ozato, N., Kubo, M., Demura, T., Fukuda, H., et al. (2009). ANXUR1 and 2, sister genes to FERONIA/SIRENE, are male factors for coordinated fertilization. *Curr. Biol.* 19, 1327–1331. doi: 10.1016/j.cub.2009.06.064
- Murashige, T., and Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiol. Plant.* 15, 473–497. doi: 10.1111/j.1399-3054.1962.tb08052.x
- Muschitz, A., Riou, C., Mollet, J. C., Gloaguen, V., and Faugeron, C. (2015). Modifications of cell wall pectin in tomato cell suspension in response to cadmium and zinc. *Acta Physiol. Plant.* 37:245. doi: 10.1007/s11738-015-22792000-y
- Nissen, K. S., Willats, W. G. T., and Malinovsky, F. G. (2016). Understanding CrRLK1L function: cell walls and growth control. *Trends Plant Sci.* 21, 516–527. doi: 10.1016/j.tplants.2015.12.004
- Parrotta, L., Guerriero, G., Sergeant, K., Cai, G., and Hausman, J.-F. (2015). Target or barrier? The cell wall of early- and later- diverging plants vs cadmium toxicity: differences in the response mechanisms. *Front. Plant Sci.* 6:133. doi: 10.3389/fpls.2015.00133
- Paynel, F., Schaumann, A., Arkoun, M., Douchiche, O., and Morvan, C. (2009). Temporal regulation of cell-wall pectin methylesterase and peroxidase isoforms in cadmium-treated flax hypocotyl. *Ann. Bot.* 104, 1363–1372. doi: 10.1093/aob/mcp254
- Pelloux, J., Rustérucci, C., and Mellerowicz, E. J. (2007). New insights into pectin methylesterase structure and function. *Trends Plant Sci.* 12, 267–277. doi: 10.1016/j.tplants.2007.04.001
- Rieu, I., and Powers, S. J. (2009). Real-time quantitative RT-PCR: design, calculations, and statistics. *Plant Cell* 21, 1031–1033. doi: 10.1105/tpc.109.066001
- Schallus, T., Jaechk, C., Fehér, K., Palma, A. S., Liu, Y., Simpson, J. C., et al. (2008). Malectin: a novel carbohydrate-binding protein of the endoplasmic reticulum and a candidate player in the early steps of protein N-glycosylation. *Mol. Biol. Cell* 19, 3404–3414. doi: 10.1091/mbc.E08-04-0354
- Sessions, A., Burke, E., Presting, G., Aux, G., McElver, J., Patton, D., et al. (2002). A high-throughput *Arabidopsis* reverse genetics system. *Plant Cell* 14, 2985–2994. doi: 10.1105/tpc.004630
- Shih, H.-W., Miller, N. D., Dai, C., Spalding, E. P., and Monshausen, G. B. (2014). The receptor-like kinase FERONIA is required for mechanical signal transduction in *Arabidopsis* seedlings. *Curr. Biol.* 24, 1887–1892. doi: 10.1016/j.cub.2014.06.064
- Singh, S., Parihar, P., Singh, R., Singh, V. P., and Prasad, S. M. (2016). Heavy metal tolerance in plants: role of transcriptomics, proteomics, metabolomics, and ionomics. *Front. Plant Sci.* 6:1143. doi: 10.3389/fpls.2015.01143
- Stegmann, M., Monaghan, J., Smakowska-Luzan, E., Rovenich, H., Lehner, A., Holton, N., et al. (2017). The receptor kinase FER is a RALF-regulated scaffold controlling plant immune signaling. *Science* 355, 287–289. doi: 10.1126/science.aal2541
- Tennstedt, P., Peisker, D., Böttcher, C., Trampczynska, A., and Clemens, S. (2009). Phytochelatin synthesis is essential for the detoxification of excess zinc and contributes significantly to the accumulation of zinc. *Plant Physiol.* 149, 938–948. doi: 10.1104/pp.108.127472
- Voxeur, A., and Höfte, H. (2016). Cell wall integrity signaling in plants: “To grow or not to grow that’s the question”. *Glycobiology* 26, 950–960. doi: 10.1093/glycob/cww029
- Wagner, T. A., and Kohorn, B. D. (2001). Wall-associated kinases are expressed throughout plant development and are required for cell expansion. *Plant Cell* 13, 303–318. doi: 10.1105/tpc.13.2.303
- Weber, M., Deinlein, U., Fischer, S., Rogowski, M., Geimer, S., Tenhaken, R., et al. (2013). A mutation in the *Arabidopsis thaliana* cell wall biosynthesis gene pectin methylesterase 3 as well as its aberrant expression cause hypersensitivity specifically to Zn. *Plant J.* 76, 151–164. doi: 10.1111/tpj.12279
- Wolf, S., Hématy, K., and Höfte, H. (2012). Growth control and cell wall signaling in plants. *Annu. Rev. Plant Biol.* 63, 381–407. doi: 10.1146/annurev-arplant-042811-105449
- Wolf, S., and Höfte, H. (2014). Growth control: a saga of cell walls, ROS, and peptide receptors. *Plant Cell* 26, 1848–1856. doi: 10.1105/tpc.114.125518
- Xu, S.-L., Rahman, A., Baskin, T. I., and Kieber, J. J. (2008). Two leucine-rich repeat receptor kinases mediate signaling, linking cell wall biosynthesis and ACC synthase in *Arabidopsis*. *Plant Cell* 20, 3065–3079. doi: 10.1105/tpc.108.063354

- Yang, X. Y., Zeng, Z. H., Yan, J. Y., Fan, W., Bian, H. W., Zhu, M. Y., et al. (2013). Association of specific pectin methylesterases with Al-induced root elongation inhibition in rice. *Physiol. Plant.* 148, 502–511. doi: 10.1111/ppl.12005
- Yu, F., Li, J., Huang, Y., Liu, L., Li, D., Chen, L., et al. (2014). FERONIA receptor kinase controls seed size in *Arabidopsis thaliana*. *Mol. Plant* 7, 920–922. doi: 10.1093/mp/ssu010
- Yu, F., Qian, L., Nibau, C., Duan, Q., Kita, D., Levasseur, K., et al. (2012). FERONIA receptor kinase pathway suppresses abscisic acid signaling in *Arabidopsis* by activating ABI2 phosphatase. *Proc. Natl. Acad. Sci. U.S.A.* 109, 14693–14698. doi: 10.1073/pnas.1212547109

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Richter, Ploderer, Mongelard, Gutierrez and Hauser. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.