

Submit a Manuscript: http://www.f6publishing.com

World J Gastroenterol 2017 September 7; 23(33): 6164-6171

DOI: 10.3748/wjg.v23.i33.6164

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

ORIGINAL ARTICLE

Retrospective Study

Ketogenic diet poses a significant effect on imbalanced gut microbiota in infants with refractory epilepsy

Gan Xie, Qian Zhou, Chuang-Zhao Qiu, Wen-Kui Dai, He-Ping Wang, Yin-Hu Li, Jian-Xiang Liao, Xin-Guo Lu, Su-Fang Lin, Jing-Hua Ye, Zhuo-Ya Ma, Wen-Jian Wang

Gan Xie, He-Ping Wang, Zhuo-Ya Ma, Wen-Jian Wang, Department of Respiratory Medicine, Shenzhen Children's Hospital, Shenzhen 518026, Guangdong Province, China

Jian-Xiang Liao, Xin-Guo Lu, Su-Fang Lin, Jing-Hua Ye, Department of Pediatric Neurology, Shenzhen Children's Hospital, Shenzhen 518026, Guangdong Province, China

Qian Zhou, Chuang-Zhao Qiu, Wen-Kui Dai, Yin-Hu Li, WeHealthGene Institute, Shenzhen 518129, Guangdong Province, China

Author contributions: Dai WK designed the study and Wang WJ managed the project; Zhou Q and Qiu CZ interpreted the data; Xie G and Zhou Q wrote the manuscript; Qiu CZ and Li YH conducted bioinformatics analysis; Wang HP and Ye JH collected sample information; Liao JX, Lu XG, Lin SF and Ma ZY contributed to the study design and patients' diagnoses; all authors read and approved the final manuscript. Xie G, Zhou Q and Qiu CZ contributed equally to this work.

Supported by the Innovation Fund of Science and Technology Commission of Shenzhen Municipality, China, No. JCYJ-20150403100317071.

Institutional review board statement: The study was reviewed and approved by the Institutional Review Board of Shenzhen Children's Hospital.

Conflict-of-interest statement: The authors declare that they have no competing interests.

Data sharing statement: Sequencing data are available from the NCBI Sequence Read Archive (SRA) database (Accession number: SRP100388).

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and

the use is non-commercial. See: http://creativecommons.org/ licenses/by-nc/4.0/

Manuscript source: Unsolicited manuscript

Correspondence to: Wen-Jian Wang, Doctor, Director, Department of Respiratory Medicine, Shenzhen Children's Hospital, No. 7019, Yitian Road, Shenzhen 518026, China. dhbk2005@163.com Telephone: +86-755-83936101 Fax: +86-755-83009800

Received: April 24, 2017 Peer-review started: May 4, 2017 First decision: June 5, 2017 Revised: June 9, 2017 Accepted: July 12, 2017 Article in press: July 12, 2017 Published online: September 7, 2017

Abstract

AIM

To investigate whether patients with refractory epilepsy and healthy infants differ in gut microbiota (GM), and how ketogenic diet (KD) alters GM.

METHODS

A total of 14 epileptic and 30 healthy infants were recruited and seizure frequencies were recorded. Stool samples were collected for 16S rDNA sequencing using the Illumina Miseq platform. The composition of GM in each sample was analyzed with MOTHUR, and intergroup comparison was conducted by R software.

RESULTS

After being on KD treatment for a week, 64% of epileptic infants showed an obvious improvement, with a 50% decrease in seizure frequency. GM structure in epileptic



infants (P1 group) differed dramatically from that in healthy infants (Health group). Proteobacteria, which had accumulated significantly in the P1 group, decreased dramatically after KD treatment (P2 group). *Cronobacter* predominated in the P1 group and remained at a low level both in the Health and P2 groups. *Bacteroides* increased significantly in the P2 group, in which *Prevotella* and *Bifidobacterium* also grew in numbers and kept increasing.

CONCLUSION

GM pattern in healthy infants differed dramatically from that of the epileptic group. KD could significantly modify symptoms of epilepsy and reshape the GM of epileptic infants.

Key words: Ketogenic diet; Cronobacter; Seizures; Gut microbiota; Epilepsy

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Many infants with epilepsy are refractory to current antiepileptic drugs, and ketogenic diet (KD) could help to moderate seizure frequency as an alternative treatment. A large number of reports have demonstrated that gut microbiota (GM) can affect children's neurodevelopment. Concurrently, GM could be dramatically affected by diet. KD could rapidly alter GM and alleviate seizure frequency in infants with refractory epilepsy. The GM structure of epileptic infants - comprising large numbers of pathogens, such as *Streptococcus* - differed from that of healthy controls. After KD therapy, GM of epileptic patients changed significantly, with fewer pathogens and more beneficial bacteria.

Xie G, Zhou Q, Qiu CZ, Dai WK, Wang HP, Li YH, Liao JX, Lu XG, Lin SF, Ye JH, Ma ZY, Wang WJ. Ketogenic diet poses a significant effect on imbalanced gut microbiota in infants with refractory epilepsy. *World J Gastroenterol* 2017; 23(33): 6164-6171 Available from: URL: http://www.wjgnet. com/1007-9327/full/v23/i33/6164.htm DOI: http://dx.doi. org/10.3748/wjg.v23.i33.6164

INTRODUCTION

Pediatric epilepsy is widespread, with complications including cognitive impairment, delayed neurodevelopment and loss of bodily control^[1,2]. Disequilibrium between excitation and depression of the central nervous system is acknowledged as the main factor in epilepsy incidence^[3]. Prior reports have identified increased inflammatory reactions and pro-inflammatory cytokines, such as interleukin (IL)-6, IL-17 and interferon, in the cerebrospinal fluid (CSF)^[4]. Anti-epileptic drugs (AEDs) and surgery are the main conventional treatments for infants with epilepsy^[5].

However, there are still 30% of epileptic infants who suffered from therapeutic futility and recurrent attacks.

A growing number of reports indicated that KD is a promising therapeutic alternative for infants with refractory epilepsy, as it has been shown to ameliorate their clinical symptoms, including the frequency of seizures^[6-10]. It remains unclear exactly how this occurs. Several reports implicated changed neurotransmitters after KD therapy, including γ -aminobutyric acid (GABA), monoamines and glutamate^[7,11]. Dahin *et al*^[12] and Freeman *et al*^[13] also identified increased ketone bodies (KBs) and decreased dopamine and serotonin^[12,13]. However, Sariego-Jamardo *et al*^[14] found little change of neurotransmitters, pterins and amino acids in the CSF of KD responders as opposed to non-responders. These discrepant findings suggested a need for the further elucidation of the mechanisms of KD therapy.

Several studies showed that diet posed a significant effect on GM^[8,15]. A high-fat diet induced selective enrichment of bile-metabolizing microbiota, such as Bacteroides^[16], whilst high-fiber foods promoted the accumulation of plant-polysaccharide fermenting microbial organisms, including Prevotella and Clostridium^[16]. A number of reports implicated involvement of GM in enteric nervous system, blood-brain barrier and glial cell development, all of which were pivotal to behavioral control and cognitive progression^[17,18]. GM could produce neurotransmitters and gut hormones directly^[19] or indirectly by producing signaling molecules to regulate host cells^[20]. GM-derived short-chain fatty acids (SCFAs) could stimulate enterochromaffin cells to produce serotonin^[21]. Wikoff *et al*^[22] also documented decreased serotonin in peripheral serum in the absence of GM. Moreover, Clostridium sporogenes and Ruminococcus gnavus promoted decarboxylation of tryptophan to tryptamine, which modulated mood and appetite through amine-associated receptors^[23]. Based on the involvement of GM in the gut-brain axis, increasing reports demonstrated imbalanced GM in neurogenic diseases (NDs), including autism-spectrum disorder, Parkinson's disease, and depression^[24]. However, GM dysbiosis in childhood epilepsy remains unexplored.

Previous studies declared that short-term dietary intake could rapidly alter human GM^[8,15]. In this study, we performed a comparison between diseased infants (before and after KD treatment) and healthy controls, to explore if and how GM of infants with refractory epilepsy differed with that of age-matched healthy subjects. We also evaluated the therapeutic effect of KD on refractory epilepsy and the changes in GM after treatment. It is hoped that this research will help to bridge some gaps in the current understanding of refractory childhood epilepsy.

MATERIALS AND METHODS

Sample collection

We enrolled 14 pediatric patients with refractory



epilepsy (aged 1.95 ± 3.10 years, 11 male and 3 female) in Shenzhen Children's Hospital, according to the following inclusion criteria: Convulsion more than four times per week after treatment with \geq 3 AEDs; no antibiotic exposure for at least 1 mo; no known genetic metabolic disorders or severe systemic illnesses; and successive KD therapy for at least 1 wk. KD was provided by Zeneca (Shenzhen, China), including Qitong ketogenic liquid milk (3.4 g protein, 8.0 g lipid and 0.6 g carbohydrate per 100 g milk), Qitong ketogenic cookies and Qitong ketogenic set-meal packages^[25].

Healthy subjects (aged up to 3 years, 15 male and 15 female) were also recruited based on the following criteria: No antibiotic exposure for at least 1 mo before this study, no disease symptoms for at least 1 mo following recruitment, and no history of seizures (Supplementary Table 1). Fisher's exact test was used to evaluate the effect of gender and age on GM composition.

DNA extraction, library construction and sequencing

The genomic DNA of microbiota was extracted from stool samples using the Power Soil DNA Isolation Kit (Mo Bio Laboratories, Carlsbad) following the manufacture's protocol. The hypervariable V3-V4 region of the 16S rRNA gene was amplified using PCR kit (TransGenAP221-02, Peking), and DNA products were quantified by gel electrophoresis and Qubit (Thermo Fisher, Singapore). After library construction, the qualified libraries were sequenced using the Illumina MiSeq Sequencing platform (Illumina, San Diego).

Taxonomy classification and diversity detection

After filtration, overlapped paired reads were assembled as tags with FLASH (v1.2.11), and clustered to operational taxonomic units (OTUs) through USEARCH (v7.0.1090)^[26]. Representative OTUs were mapped against the Greengenes database (v201305)^[27] and classified with RDP classifier (v2.2)^[28]. The diversity of microbiota was calculated with MOTHUR (v1.31.2)^[29].

Principal component analysis and statistical analysis

PCA was performed with R software (v3.2.5). Wilcoxon rank-sum test was used to compare GM in diseased infants and healthy controls (Health group). Comparative analysis between the epileptic infants before (P1 group) and after treatment (P2 group) was conducted by Wilcoxon signed-rank test. Linear discriminant analysis Effect Size (LEfSe) analysis was used to identify microbial species which were apparently enriched in a specific group.

RESULTS

Data output and patients' characteristics

The average number of high-quality sequencing reads produced for each sample was 117196 (range, 31900



Figure 1 Gut microbial diversity of the three groups. Distribution of Shannon index (evenness) is shown. Red, blue, and green represent the Health, P1 and P2 groups, respectively. The gut microbiota (GM) of the healthy infants was more stable than that of the other two groups.

to 305190). The number of assembled tags averaged 22800, with a range from 12655 to 27337. Both gender and age had no significant effect on GM (P = 0.069 and 0.234, respectively).

GM of healthy individuals differs dramatically with that of diseased infants

Shannon index analysis indicated higher GM diversity in healthy infants, in comparison with infants with refractory epilepsy (Figure 1, Supplementary Table 2). PCA of GM profile also identified that healthy infants could be clearly distinguished from patients (Figure 2, Supplementary Table 3). The phylum Firmicutes predominated in patients (45.82%) and was unchanged after KD therapy (47.00%) (Supplementary Table 4). Bacteroidetes accounted for 53.01% of GM in healthy infants, followed by Firmicutes (34.38%). After KD treatment, Bacteroidetes increased from 26.75% to 38.71%. Actinobacteria was enriched in healthy infants (8.49%) and occupied a lower percent in patients (2.38% before treatment and 2.92% after treatment). Proteobacteria was highly accumulated in infants with refractory epilepsy (24.34%) and decreased dramatically after KD therapy (10.77%). At the genus level, Cronobacter was dominant in the patients (23.30% vs 0.00% in the healthy group). By contrast, healthy subjects harbored more than twice Bacteroides (42.68%) than infants with refractory epilepsy (17.93%). Prevotella and Bifidobacterium also accumulated in the healthy group (7.25% and 7.84%, respectively) (Supplementary Table 5).

KD therapy ameliorates epilepsy and GM of patients started to improve

After a week of KD therapy, 3 (21%) patients were seizure-free and 6 (43%) had a 50% to 90% decrease of seizure frequency (Supplementary Table 1). The





Figure 2 Principal component analysis. Each plot in the principal component analysis (PCA) graph stands for a sample. Red, blue and green colors represent the Health, P1 and P2 groups, respectively.



Figure 3 Gut microbiota structures in the Health, P1 and P2 groups at the genus level. SVG package (version 1.1) was used to produce the paragraph. The size of the circle representing each genus was determined by the relative abundance of the three groups, and the width of line linking the P1, P2 and Health groups indicates the relative abundance of each group.

remaining 5 (36%) infants experienced no significant improvement in seizure control (Supplementary Table 1). GM of the P2 group was more similar to that

of the Health group, by comparison with P1 group (Figures 3 and 4). After KD treatment, *Bacteroides* increased significantly, by 24.42%. *Prevotella* also



Figure 4 Significantly enriched gut microbiota components in the Health, P1 and P2 groups. LEfSe analysis was applied to detect the gut microbiota (GM) components in the three groups. Red, green, and blue represent the Health, P1 and P2 groups, respectively. The LDA score was set as \leq 2. The enrichment degree is proportional to the LDA score.



increased in the P1 group from 0.37% to 1.85% after KD treatment (Figure 3 and Supplementary Table 5). *Cronobacter* decreased sharply in after-treatment patients, from 23.3% to 10.44 % (Figures 3 and 4 and Supplementary Table 5). KD exposure also induced a decrease in *Erysipelatoclostridium* (by 8.67% in the P1 group and 4.89% in the P2 group); it represented just 0.64% in healthy infants (Figures 3 and 4 and Supplementary Table 5). *Streptococcus*, *Alistipes*, *Ruminiclostridium*, *Barnesiella* and *Enterococcus* also decreased after KD therapy (Figures 3 and 4 and Supplementary Table 5).

DISCUSSION

KD is increasingly used for the treatment of refractory epilepsy in childhood, but the mechanism remains unclear. Previous reports indicated that GM played an important role in the gut-brain axis^[24], and was affected significantly by intake of high-fat food^[16]. This study focused on differed GM structures between healthy and epileptic infants, as well as altered GM in patients after one week of KD treatment. The results pointed to an imbalanced GM in patients and a significant improvement after KD therapy.

Proteobacteria comprises a variety of notorious pathogens, such as *Escherichia*, *Salmonella* and *Vibrio*. It accounted for 24.34% in pediatric patients and decreased dramatically after KD treatment. Bacteroidetes was dominant in healthy infants and increased largely in after-treatment patients.

We identified accumulated Bacteroides in healthy subjects as well as in patients after treatment. Bacteroides was reported to digest and metabolize highfat food and to regulate the secretion of IL-6 and IL-17 in dendritic cells (DCs), a process strongly associated with seizure severity of epileptic patients^[4,16]. However, patients-enriched Cronobacter decreased dramatically after KD therapy. Prior reports demonstrated that there were multiple virulence determinants of Cronobacter, including Cronobacter plasminogen activator and ferric ion transporter protein, which paly a detrimental role in human health^[30-32]. Prevotella is a robust producer of SCFAs^[33], which could protect the intestinal mucosa and function as neurotransmitters. Previous reports also indicated that SCFAs mediated nervous impulse and mitigated Parkinson's disease^[33,34]. Similarly, we identified increased Prevotella in the Health and P2 group, when compared with the P1 group. Some other genera also offer clues to epilepsy recovery, such as Erysipelatoclostridium, Blautia, Bifidobacterium and Streptococcus. Bifidobacterium was well known to be beneficial to health^[35], and *Streptococcus*, a common pathogen, played a role especially in respiratory diseases^[36]. Although GM imbalance in diseased infants was identified and GM improved after KD treatment, more exploration was needed to elucidate the contribution of a healthy GM to epilepsy onset/recovery.

This study revealed that KD can mitigate the sym-

ptoms of epilepsy and correct an imbalanced GM in epileptic infants. However, further analysis is needed to unravel how GM may be involved in epilepsy onset/ recovery.

There are some limitations that need to be clarified. First, 16S rDNA analysis identified microbes at the genus level, which makes it difficult to unravel different microbes at the species or function level. Second, it would be more useful to evaluate the efficacy of KD treatment and its effect on the GM if this could be done with a longer period of follow-up. Third, an animal model might be applied to demonstrate whether GM imbalance could induce epilepsy associated symptoms. Considering these limitations, we are planning to perform metagenomic analysis on GM of healthy and epileptic infants. This will provide more insights into distinct metabolic networks in imbalanced GM.

In conclusion, we found that GM of infants with refractory epilepsy differed dramatically from that of healthy infants. Epileptic patients harbored significantly enriched pathogens and decreased beneficial bacteria. Although this study provides new insight into the involvement of GM in pediatric refractory epilepsy, the gap between KD and epilepsy recovery is still huge. To uncover the mechanism and pathogens involved in refractory infantile epilepsy, further research should underscore functional gene networks in GM.

ACKNOWLEDGMENTS

We thank the staff of WeHealthGene who contributed to the project, but whose names are not included in the author list.

COMMENTS

Background

Infants with refractory epilepsy could not be cured by several anti-epileptic drugs (AEDs) and ketogenic diet (KD) was increasingly used as an alternative therapy to refractory epilepsy. High-fat diet was reported to pose a significant impact on gut microbiota (GM), which could regulate neural systems.

Research frontiers

Previous reports demonstrated that GM could affect neural systems by secreting metabolites as neurotransmitters. In parallel, the gut-brain axis is a research hot spot in biomedicine, including the study of autism, Parkinson's disease, and depression.

Innovations and breakthroughs

This study showed that the GM pattern of diseased infants differs significantly from that of healthy controls. The decreased number of dominant pathogens and significantly increased number of beneficial bacteria after KD treatment offer new insight into KD therapy for epilepsy.

Applications

This study found several types of bacteria altered in the GM, suggesting that these bacteria could be monitored as biomarkers to provide an important reference for epilepsy treatment.

Terminology

GM, which consists of many kinds of bacteria including pathogens, commensals,



and probiotics, plays an important role in the human body.

Peer-review

The authors have performed important research in pediatric epilepsy. They discovered that the composition of the GM in healthy and diseased infants was significantly different, specifically in healthy infants as opposed to those with refractory epilepsy. Bacterial patterns were dramatically changed after KD therapy, and this was associated with a reduction in the frequency of seizures. These findings should enhance our knowledge of the relationship between epilepsy and GM and provide new insight into the clinical treatment of epilepsy.

REFERENCES

- Nordli DR Jr. Epileptic encephalopathies in infants and children. J Clin Neurophysiol 2012; 29: 420-424 [PMID: 23027099 DOI: 10.1097/WNP.0b013e31826bd961]
- 2 Nickels KC, Zaccariello MJ, Hamiwka LD, Wirrell EC. Cognitive and neurodevelopmental comorbidities in paediatric epilepsy. *Nat Rev Neurol* 2016; 12: 465-476 [PMID: 27448186 DOI: 10.1038/ nrneurol.2016.98]
- Goldberg EM, Coulter DA. Mechanisms of epileptogenesis: a convergence on neural circuit dysfunction. *Nat Rev Neurosci* 2013; 14: 337-349 [PMID: 23595016 DOI: 10.1038/nrn3482]
- 4 **Mao LY**, Ding J, Peng WF, Ma Y, Zhang YH, Fan W, Wang X. Interictal interleukin-17A levels are elevated and correlate with seizure severity of epilepsy patients. *Epilepsia* 2013; **54**: e142-e145 [PMID: 23944193 DOI: 10.1111/epi.12337]
- 5 Vigevano F, Arzimanoglou A, Plouin P, Specchio N. Therapeutic approach to epileptic encephalopathies. *Epilepsia* 2013; 54 Suppl 8: 45-50 [PMID: 24571117 DOI: 10.1111/epi.12423]
- 6 Kossoff EH, Zupec-Kania BA, Amark PE, Ballaban-Gil KR, Christina Bergqvist AG, Blackford R, Buchhalter JR, Caraballo RH, Helen Cross J, Dahlin MG, Donner EJ, Klepper J, Jehle RS, Kim HD, Christiana Liu YM, Nation J, Nordli DR Jr, Pfeifer HH, Rho JM, Stafstrom CE, Thiele EA, Turner Z, Wirrell EC, Wheless JW, Veggiotti P, Vining EP; Charlie Foundation, Practice Committee of the Child Neurology Society; Practice Committee of the Child Neurology Society; International Ketogenic Diet Study Group. Optimal clinical management of children receiving the ketogenic diet: recommendations of the International Ketogenic Diet Study Group. *Epilepsia* 2009; **50**: 304-317 [PMID: 18823325 DOI: 10.1111/j.1528-1167.2008.01765.x]
- 7 Lima PA, Sampaio LP, Damasceno NR. Neurobiochemical mechanisms of a ketogenic diet in refractory epilepsy. *Clinics* (Sao Paulo) 2014; 69: 699-705 [PMID: 25518023 DOI: 10.6061/ clinics/2014(10)09]
- 8 van der Louw E, van den Hurk D, Neal E, Leiendecker B, Fitzsimmon G, Dority L, Thompson L, Marchió M, Dudzińska M, Dressler A, Klepper J, Auvin S, Cross JH. Ketogenic diet guidelines for infants with refractory epilepsy. *Eur J Paediatr Neurol* 2016; 20: 798-809 [PMID: 27470655 DOI: 10.1016/j.ejpn.2016.07.009]
- 9 Neal EG, Chaffe H, Schwartz RH, Lawson MS, Edwards N, Fitzsimmons G, Whitney A, Cross JH. The ketogenic diet for the treatment of childhood epilepsy: a randomised controlled trial. *Lancet Neurol* 2008; 7: 500-506 [PMID: 18456557 DOI: 10.1016/ S1474-4422(08)70092-9]
- 10 Noebels JL. The Voltage-Gated Calcium Channel and Absence Epilepsy. In: Noebels JL, Avoli M, Rogawski MA, Olsen RW, Delgado-Escueta AV, editors. Jasper's Basic Mechanisms of the Epilepsies. 4th ed. Bethesda (MD), 2012: 1049-1066
- 11 Ruskin DN, Masino SA. The nervous system and metabolic dysregulation: emerging evidence converges on ketogenic diet therapy. *Front Neurosci* 2012; 6: 33 [PMID: 22470316 DOI: 10.3389/fnins.2012.00033]
- 12 Dahlin M, Mansson Je Fau Amark P, Amark P. CSF levels of dopamine and serotonin, but not norepinephrine, metabolites are influenced by the ketogenic diet in children with epilepsy. *Epilepsy Res* 2012; 99: 132-138 [DOI: 10.1016/j.eplepsyres.2011.11.003]
- 13 Freeman JM, Kossoff EH, Hartman AL. The ketogenic diet: one

decade later. *Pediatrics* 2007; **119**: 535-543 [PMID: 17332207 DOI: 10.1542/peds.2006-2447]

- 14 Sariego-Jamardo A, García-Cazorla A, Artuch R, Castejón E, García-Arenas D, Molero-Luis M, Ormazábal A, Sanmartí FX. Efficacy of the Ketogenic Diet for the Treatment of Refractory Childhood Epilepsy: Cerebrospinal Fluid Neurotransmitters and Amino Acid Levels. *Pediatr Neurol* 2015; **53**: 422-426 [PMID: 26476148 DOI: 10.1016/j.pediatrneurol.2015.07.013]
- 15 Wu J, Zhang Y, Yang H, Rao Y, Miao J, Lu X. Intestinal Microbiota as an Alternative Therapeutic Target for Epilepsy. *Can J Infect Dis Med Microbiol* 2016; 2016: 9032809 [PMID: 27882059 DOI: 10.1155/2016/9032809]
- Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, 16 Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M, Borruel N, Casellas F, Fernandez L, Gautier L, Hansen T, Hattori M. Havashi T. Kleerebezem M. Kurokawa K. Leclerc M. Levenez F, Manichanh C, Nielsen HB, Nielsen T, Pons N, Poulain J, Qin J, Sicheritz-Ponten T, Tims S, Torrents D, Ugarte E, Zoetendal EG, Wang J, Guarner F, Pedersen O, de Vos WM, Brunak S, Doré J; MetaHIT Consortium, Antolín M, Artiguenave F, Blottiere HM, Almeida M, Brechot C, Cara C, Chervaux C, Cultrone A, Delorme C, Denariaz G, Dervyn R, Foerstner KU, Friss C, van de Guchte M, Guedon E, Haimet F, Huber W, van Hylckama-Vlieg J, Jamet A, Juste C, Kaci G, Knol J, Lakhdari O, Layec S, Le Roux K, Maguin E, Mérieux A, Melo Minardi R, M'rini C, Muller J, Oozeer R, Parkhill J, Renault P, Rescigno M, Sanchez N, Sunagawa S, Torrejon A, Turner K, Vandemeulebrouck G, Varela E, Winogradsky Y, Zeller G, Weissenbach J, Ehrlich SD, Bork P. Enterotypes of the human gut microbiome. Nature 2011; 473: 174-180 [PMID: 21508958 DOI: 10.1038/nature09944]
- 17 Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Tóth M, Korecka A, Bakocevic N, Ng LG, Kundu P, Gulyás B, Halldin C, Hultenby K, Nilsson H, Hebert H, Volpe BT, Diamond B, Pettersson S. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med* 2014; 6: 263ra158 [PMID: 25411471 DOI: 10.1126/scitranslmed.3009759]
- 18 Collins J, Borojevic R, Verdu EF, Huizinga JD, Ratcliffe EM. Intestinal microbiota influence the early postnatal development of the enteric nervous system. *Neurogastroenterol Motil* 2014; 26: 98-107 [PMID: 24329946 DOI: 10.1111/nmo.12236]
- 19 Matsumoto M, Kibe R, Ooga T, Aiba Y, Kurihara S, Sawaki E, Koga Y, Benno Y. Impact of intestinal microbiota on intestinal luminal metabolome. *Sci Rep* 2012; 2: 233 [PMID: 22724057 DOI: 10.1038/srep00233]
- 20 Batterham RL, ffytche DH, Rosenthal JM, Zelaya FO, Barker GJ, Withers DJ, Williams SC. PYY modulation of cortical and hypothalamic brain areas predicts feeding behaviour in humans. *Nature* 2007; 450: 106-109 [PMID: 17934448 DOI: 10.1038/ nature06212]
- 21 Yano JM, Yu K, Donaldson GP, Shastri GG, Ann P, Ma L, Nagler CR, Ismagilov RF, Mazmanian SK, Hsiao EY. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* 2015; 161: 264-276 [PMID: 25860609 DOI: 10.1016/j.cell.2015.02.047]
- 22 Wikoff WR, Anfora AT, Liu J, Schultz PG, Lesley SA, Peters EC, Siuzdak G. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc Natl Acad Sci USA* 2009;106: 3698-3703 [PMID: 19234110 DOI: 10.1073/pnas.0812874106]
- 23 Williams BB, Van Benschoten AH, Cimermancic P, Donia MS, Zimmermann M, Taketani M, Ishihara A, Kashyap PC, Fraser JS, Fischbach MA. Discovery and characterization of gut microbiota decarboxylases that can produce the neurotransmitter tryptamine. *Cell Host Microbe* 2014; 16: 495-503 [PMID: 25263219 DOI: 10.1016/j.chom.2014.09.001]
- Sharon G, Sampson TR, Geschwind DH, Mazmanian SK. The Central Nervous System and the Gut Microbiome. *Cell* 2016; 167: 915-932 [PMID: 27814521 DOI: 10.1016/j.cell.2016.10.027]
- 25 **Suo C**, Liao J, Lu X, Fang K, Hu Y, Chen L, Cao D, Huang T, Li B, Li C. Efficacy and safety of the ketogenic diet in Chinese children.

Seizure 2013; 22: 174-178 [PMID: 23273808 DOI: 10.1016/ j.seizure.2012.11.014]

- 26 Edgar RC. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat Methods* 2013; 10: 996-998 [PMID: 23955772 DOI: 10.1038/nmeth.2604]
- 27 DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, Huber T, Dalevi D, Hu P, Andersen GL. Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol* 2006; 72: 5069-5072 [PMID: 16820507 DOI: 10.1128/AEM.03006-05]
- 28 Wang Q, Garrity GM, Tiedje JM, Cole JR. Naive Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Appl Environ Microbiol* 2007; 73: 5261-5267 [PMID: 17586664 DOI: 10.1128/AEM.00062-07]
- 29 Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ, Sahl JW, Stres B, Thallinger GG, Van Horn DJ, Weber CF. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl Environ Microbiol* 2009; **75**: 7537-7541 [PMID: 19801464 DOI: 10.1128/AEM.01541-09]
- 30 Jackson EE, Forsythe SJ. Comparative study of Cronobacter identification according to phenotyping methods. *BMC Microbiol* 2016; 16: 146 [PMID: 27401027 DOI: 10.1186/s12866-016-0768-6]
- 31 Bowen AB, Braden CR. Invasive Enterobacter sakazakii disease in

infants. *Emerg Infect Dis* 2006; **12**: 1185-1189 [PMID: 16965695 DOI: 10.3201/eid1208.051509]

- 32 Singh N, Raghav M, Narula S, Tandon S, Goel G. Profiling of Virulence Determinants in Cronobacter sakazakii Isolates from Different Plant and Environmental Commodities. *Curr Microbiol* 2017; 74: 560-565 [PMID: 28258294 DOI: 10.1007/s00284-017-1219-9]
- 33 Scheperjans F, Aho V, Pereira PA, Koskinen K, Paulin L, Pekkonen E, Haapaniemi E, Kaakkola S, Eerola-Rautio J, Pohja M, Kinnunen E, Murros K, Auvinen P. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov Disord* 2015; 30: 350-358 [PMID: 25476529 DOI: 10.1002/mds.26069]
- 34 Miletta MC, Petkovic V, Eblé A, Ammann RA, Flück CE, Mullis PE. Butyrate increases intracellular calcium levels and enhances growth hormone release from rat anterior pituitary cells via the G-proteincoupled receptors GPR41 and 43. *PLoS One* 2014; 9: e107388 [PMID: 25310566 DOI: 10.1371/journal.pone.0107388]
- 35 Ghouri YA, Richards DM, Rahimi EF, Krill JT, Jelinek KA, DuPont AW. Systematic review of randomized controlled trials of probiotics, prebiotics, and synbiotics in inflammatory bowel disease. *Clin Exp Gastroenterol* 2014; 7: 473-487 [PMID: 25525379 DOI: 10.2147/CEG.S27530]
- 36 Krzyściak W, Pluskwa KK, Jurczak A, Kościelniak D. The pathogenicity of the Streptococcus genus. Eur J Clin Microbiol Infect Dis 2013; 32: 1361-1376 [PMID: 24141975 DOI: 10.1007/ s10096-013-1914-9]

P- Reviewer: Daniel F, Prakash N S- Editor: Qi Y L- Editor: Wang TQ E- Editor: Xu XR







Published by Baishideng Publishing Group Inc

7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.f6publishing.com/helpdesk http://www.wjgnet.com





© 2017 Baishideng Publishing Group Inc. All rights reserved.