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## From the draining lymph node to the liver: The induction and effector mechanisms of malaria-specific CD8<sup>+</sup> T cells

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### Abstract

Parasitic protozoa cause considerable disease in humans and, due to their intracellular life cycle, induce robust CD8<sup>+</sup> T cell responses. A greater understanding of the factors that promote and maintain CD8<sup>+</sup> T cell-mediated immunity against these pathogens is likely needed for the development of effective vaccines. Immunization with radiation-attenuated sporozoites, the infectious stage of the malaria parasite transmitted by mosquitoes, is an excellent model to study these questions as CD8<sup>+</sup> T cells specific for a single epitope can completely eliminate parasite infection in the liver. Furthermore, live, radiation-attenuated parasites represent the 'gold standard' for malaria vaccination. Here, we will highlight recent studies aimed at understanding the factors required for the induction, recruitment, and maintenance of effector and memory CD8<sup>+</sup> T cells against malaria liver stages.

### Keywords

*Plasmodium*; CD8<sup>+</sup> T cell responses; dendritic cells (DCs); T cell memory; *in vivo* imaging; vaccines

### Introduction

CD8<sup>+</sup> T cells are critical for the control of intracellular pathogens, including viral, bacterial, and protozoan parasites. To date, most of our knowledge regarding the antigen presentation requirements, generation, and maintenance of effector and memory CD8<sup>+</sup> T cells is based on non-infectious or viral models that fail to account for the complexity and antigenic diversity of protozoan parasites. This knowledge gap is significant considering that parasitic protozoa — *Plasmodium* spp., *Toxoplasma gondii*, *Leishmania* spp., and *Trypanosoma cruzi*— pose

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major threats to human health. Owing to their ability to invade and replicate within host cells, intracellular pathogens are susceptible to CD8<sup>+</sup> T cell-mediated elimination. Thus, CD8<sup>+</sup> T cells are important for protective immunity against these pathogens; however, the degree of protection conferred by these cells depends on the species and the developmental stage of the parasite. Importantly, protective immunity against malaria liver stages is mediated, to a significant extent, by CD8<sup>+</sup> T cells specific for the *Plasmodium* circumsporozoite (CS) antigen [1,2]. Here, we review how CD8<sup>+</sup> T cell responses against malaria parasites are initiated and sustained following a natural course of infection while drawing parallels to other intracellular pathogens. Furthermore, we will discuss the implications of these findings on the development of whole parasite vaccines.

## Early host-pathogen interactions in the skin

Many pathogens enter their mammalian host through the skin, a complex organ that is critical for both physical protection and host defense. In accordance with its key role in immune surveillance, the skin supports a diverse community of resident and migratory immune cells including neutrophils, macrophages, mast cells, dendritic cells (DCs), and lymphocytes [3,4]. Host-pathogen interactions in the skin have a tremendous impact on disease outcome and protective immunity. Consequently, the ‘skin stage of malaria’ has garnered considerable attention within recent years. Malaria infection begins when a female *Anopheles* mosquito injects *Plasmodium* sporozoites into the skin of its host during blood feeding. After their deposition in the skin, sporozoites glide rapidly (~1–2 μm/second) before exiting the dermis via blood or lymphatic vessels [5]. The exquisite motility of malaria sporozoites appears to limit degradation by skin-resident phagocytes while promoting progression from the skin site of inoculation to the liver site of infection [6]. Although some sporozoites enter the bloodstream and access the liver within minutes after their inoculation, many take hours to exit the skin [7] and a small proportion (~0.5–5%) remain and develop into exoerythrocytic forms [8,9]. The prolonged residence and development of parasites in the skin likely provides ample chemoattractant signals for innate leukocytes. Neutrophils are rapidly recruited to the skin after needle or mosquito bite inoculation of infectious sporozoites and sustain significantly high numbers in the skin and skin-draining lymph nodes (DLNs) for up to 24 hours post-inoculation [6,10]. Following the first wave of neutrophil recruitment, inflammatory monocytes populate the skin and DLNs [10]. Despite the dramatic neutrophilic response in these organs after skin deposition of sporozoites, neutrophil depletion appears to have no impact on the development of a protective CD8<sup>+</sup> T cell response [10]. Interestingly, neutrophils also infiltrate the skin after sham injection, needle inoculation of salivary gland extract from arthropod vectors, and uninfected mosquito and sand fly bites [10,11]. The early neutrophilic response generated under these conditions is likely a byproduct of a host response aimed at wound repair and sterilization as neutrophils were recently shown to be recruited to the skin after sterile laser damage [12].

Although neutrophils and inflammatory monocytes can contribute to adaptive immunity [13,14], DCs are critically involved in both the detection of pathogens in the periphery as well as the activation and differentiation of T cells in lymphoid organs [15]. Skin-derived DCs are a heterogeneous population of cells that differs in their ability to present antigens to

CD4<sup>+</sup> and CD8<sup>+</sup> T cells [15,16] and can be broadly defined into the following three subsets: langerin-positive CD103<sup>+</sup> dermal DCs, langerin-negative CD11b<sup>+</sup> CD103<sup>-</sup> dermal DC, and langerin-positive CD103<sup>-</sup> Langerhans cells (LCs) [15]. After intradermal (ID) injection of sporozoites, ~20% of skin-deposited sporozoites were found to be closely associated with CD11b<sup>+</sup> myeloid cells in the skin [10]. However, we recently demonstrated a nonessential role for Langerhans cells and langerin<sup>+</sup> dermal DCs in sporozoite antigen presentation to CD8<sup>+</sup> T cells using a mouse model system that allowed for the selective depletion of these DC subsets [17]. In addition, we did not observe appreciable migration of skin DCs to the DLNs after sporozoite injection into the dermis by mosquito bite or needle inoculation, nor did we detect a difference in CD8<sup>+</sup> T cell priming after chemical inhibition of DC migration to the DLNs [18]. Nevertheless, the immunological significance of malaria parasites that remain and undergo partial development within the skin is largely unknown. It is possible that the inflammatory response induced by parasites in the skin may exert “remote control” over the composition of leukocytes in the DLNs as described following cutaneous inflammation with Complete Freund’s Adjuvant and Keyhole Limpet Hemocyanin [19]. In support of this, a Th1 chemokine/cytokine profile was observed in the DLNs 24 hours after needle inoculation of *P. berghei* sporozoites [10]. Additionally, parasite development in the skin may provide late liver stage antigens and, thus contribute to a broader anti-malaria CD8<sup>+</sup> T cell response [20,21].

### Innate and adaptive immunity to malaria parasites in the DLNs

While many pathogens initiate infection in the skin of their mammalian host, several pathogens—*Plasmodium spp.*, *Schistosma mansoni*, *L. major*, and *Toxoplasma gondii*—additionally gain access to the DLNs [5,7,22–24], raising key questions about the fate of parasites in these organs as well as their contribution to adaptive immunity. These questions are especially pertinent in the context of a malaria infection as a significant proportion (~15–20%) of skin-deposited sporozoites migrate to the DLNs [5,7,25,26]. Using a model mimicking natural exposure to sporozoite-infected mosquitoes, we previously demonstrated that CD8<sup>+</sup> T cells specific for the malaria CS protein, a leading vaccine candidate, were primed by DCs in the DLNs of mice [25]. Remarkably, IFN- $\gamma$  producing CD8<sup>+</sup> T cells were detected in the DLNs within 48 hours after immunization by irradiated *P. yoelii*-infected mosquito bites, whereas significant responses in the spleen, liver, and liver-DLNs were not observed until 72 hours post-immunization. Furthermore, animals that had their DLNs removed prior to sporozoite immunization generated poor CS-specific CD8<sup>+</sup> T cell responses [25]. In agreement with these findings, Obeid and colleagues demonstrated that CD8<sup>+</sup> T cell responses generated in the DLNs were sufficient for sterile protection against live sporozoites [27]. The DLNs were also found to be the primary site of prolonged antigen presentation [28]. In these studies, antigen persistence required CD11c<sup>+</sup> LN-resident DCs and macrophages and was critical for the optimal expansion of protective CD8<sup>+</sup> T cell responses directed against the CS protein [28]. Given the importance of the DLNs in the generation and maintenance of protective immunity against malaria sporozoites, our laboratory and others have worked to characterize host-parasite interactions in this organ.

Sporozoites that gain access to the DLN are first found in the subcapsular sinus [5,10,17], a region of the LN populated by CD169<sup>+</sup> macrophages specialized in trapping pathogens and

initiating adaptive immune responses (reviewed in [29]). Additionally, sporozoites and parasite-derived antigens were found in association with CD11c<sup>+</sup> DCs 5 to 8 hours after sporozoite migration to the DLN [5,17]. However, the contribution of these interactions to CD8<sup>+</sup> T cell priming was unknown until recently. Using genetically manipulated malaria parasites, as well as antibody-mediated immobilization of sporozoites, we determined that sporozoite access to the DLNs is required for robust CD8<sup>+</sup> T cell responses [17] (Fig. 1). In further support of a limited role for migratory skin-DCs, static and dynamic *in vivo* imaging revealed the formation of CD8<sup>+</sup> T cell-DC clusters (a surrogate for antigen presentation) in the draining LN paracortex as early as 8 hours after sporozoite immunization. These early interactions between LN-resident DCs and CD8<sup>+</sup> T cells correlated with T cell activation because we observed the up-regulation of CD69, an early activation marker, and the production of IFN- $\gamma$  by CS-specific CD8<sup>+</sup> T cells at 8 and 16 hours post-inoculation, respectively. In addition, we provided direct evidence of antigen presentation by LN-resident CD8 $\alpha$ <sup>+</sup> DCs to CD8<sup>+</sup> T cells using histo-cytometry, an analytical microscopy method that allows for the quantification of cellular interactions *in situ* [30,31]. Moreover, malaria-specific CD8<sup>+</sup> T cell responses were significantly reduced in *Batf3*<sup>-/-</sup> mice that possess substantial defects in their numbers of CD8 $\alpha$ <sup>+</sup> DCs [32,33]. It is worth noting that other studies have demonstrated a requirement for CD8 $\alpha$ <sup>+</sup> DCs in the presentation of *Plasmodium* blood and liver stage antigens to CD8<sup>+</sup> T cells [34–37]. In summary, the innate and adaptive immune responses elicited by malaria sporozoites that migrate to the DLNs are essential for the induction of CD8<sup>+</sup> T cell-mediated immunity.

### Antigen presentation requirements of sporozoite-specific CD8<sup>+</sup> T cells

While it is clear that DCs are critical for the induction of CD8<sup>+</sup> T cells, it is well appreciated that DC maturation is needed for the optimal priming of these cells [38]. DC maturation, their migration to secondary lymphoid organs, and cross-presentation of exogenous antigens is greatly enhanced following their recognition of microbial molecular patterns by pattern recognition receptors (PRRs) such as toll-like receptors (TLRs) and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs). TLRs recognize a diverse range of microbe-derived lipids, carbohydrates, peptides, and nucleic acid structures and elicit different signaling cascades depending on the engagement of the following intracellular adaptor proteins: myeloid differentiation factor 88 (MyD88), MyD88-adaptor-like (MAL), Toll/IL-1 receptor (TIR) domain-containing adaptor inducing IFN- $\beta$  (TRIF), and TRIF-related adaptor molecule (TRAM) [39]. Like TLRs, NLRs respond to pathogen-associated molecular patterns (PAMPs), but are additionally capable of responding to endogenous molecules released during stress or infection such as ATP, reactive oxygen species, and potassium efflux [40]. The oligomerization of certain NLRs results in the assembly of the inflammasome, a large macromolecular complex that is critical for the maturation of the proinflammatory cytokines interleukin-1 beta (IL-1 $\beta$ ) and IL-18 [40]. Although TLR and inflammasome agonists from *P. falciparum* and *P. berghei* blood stage parasites have been characterized [41,42], a sporozoite-derived PAMP, nor its contribution to protective CD8<sup>+</sup> T cell responses, has not been described. To address this question, we used transgenic *P. berghei* parasites expressing SIINFEKL, the H-2K<sup>b</sup>-restricted epitope of ovalbumin (*P. berghei* CS<sup>5M</sup>) described previously [43], and a panel of mice deficient in components of

PRR signaling. *Myd88*<sup>-/-</sup> *Trif*<sup>-/-</sup> and *Ipa*<sup>-/-</sup> *Asc*<sup>-/-</sup> mice were both protected against live sporozoite challenge after immunization by irradiated *P. berghei* CS<sup>5M</sup>-infected mosquito bites. In addition, there was no difference in the magnitude or function of endogenous sporozoite-specific CD8<sup>+</sup> T cells primed in the absence of TLR or inflammasome (NLRP1, NLRP3, and IPAF)-dependent signaling [18]. Therefore, it is possible that protective CD8<sup>+</sup> T cell responses against malaria parasites require PRR-mediated signaling through compensatory and redundant mechanisms or alternative PRRs.

An additional consideration is how DCs acquire, process, and present malaria antigens to naïve CD8<sup>+</sup> T cells. DCs are especially adept at processing exogenous antigens derived from apoptotic, necrotic, or pathogen-infected cells in a major histocompatibility complex class (MHC) I-restricted manner via cross-presentation [44]. In contrast to *T. gondii* and *L. major*, *Plasmodium* sporozoites do not productively invade DCs, and instead, cross-presentation is required for the induction of CD8<sup>+</sup> T cell responses directed against molecular determinants in the CS protein [25,45]. In subsequent studies, Cockburn *et al.* demonstrated that DCs cross-present malaria antigens to CD8<sup>+</sup> T cells through an endosome-to-cytosol pathway [43]; however, the mechanism by which sporozoite antigens are deposited in the cytosol of DCs is largely unknown. Furthermore, the TAP-dependent pathway was also shown to be critical for the presentation of liver stage antigens by infected hepatocytes [43]. Therefore, both the induction and effector phases of anti-malaria CD8<sup>+</sup> T cells require TAP-mediated delivery of CS epitopes from the cytosol to the ER in DCs and hepatocytes, respectively.

### Trafficking and survival of tissue-specific CD8<sup>+</sup> T cells

In order to continue their life cycle in the mammalian host, sporozoites must invade and replicate within hepatocytes. Importantly, infected hepatocytes present parasite-derived antigens to CD8<sup>+</sup> T cells and are susceptible to CD8<sup>+</sup> T cell-mediated killing [1,25,46]. Because parasites remain in the liver for only a short time, intrahepatic CD8<sup>+</sup> T cells are probably key factors in protection against malaria. Therefore, a better understanding of the mechanisms regulating CD8<sup>+</sup> T cells trafficking and survival in the liver is critical for malaria vaccine design. During activation, effector T cells can be “selectively imprinted” to express certain homing receptors that regulate their migration to peripheral organs [47]. While this process has been shown to be important for the preferential homing of effector CD8<sup>+</sup> T cells to barrier tissues such as the intestine and skin (reviewed in [48]), it is unclear whether parasite-specific T cells are instructed for ‘liver-homing’ shortly after their priming in lymphoid organs. After sporozoite immunization, CS-specific effector CD8<sup>+</sup> T cells were found in various peripheral organs, such as the liver, lungs and kidneys [49], suggesting that the initial homing pattern of CD8<sup>+</sup> T cells is not a stringent process. The promiscuous trafficking of activated CS-specific CD8<sup>+</sup> T cells is likely due to their simultaneous expression of several inflammatory chemokine receptors [50–52].

Although the initial trafficking of effector CD8<sup>+</sup> T cell appears to be an indiscriminate process, the local persistence of memory CD8<sup>+</sup> T cells is clearly dependent on specific environmental cues, e.g. chemokines and cytokines. For example, the transmembrane chemokine CXCL16 is abundantly expressed in the liver sinusoids and was shown to be crucial for the maintenance of hepatic NKT cells during steady-state [53]. Importantly,

CXCR6, the chemokine receptor which recognizes CXCL16, was found to be significantly up-regulated by liver-resident CD8<sup>+</sup> T cells several weeks after sporozoite immunization [51,52]. These findings hinted at a potential role for CXCR6 in the retention of these cells. Indeed, adoptive transfer studies using *Cxcr6*<sup>-/-</sup> CD8<sup>+</sup> T cells demonstrated that CXCR6 was dispensable for the early recruitment, but important for the long-term survival, of memory CD8<sup>+</sup> T cells in the livers of sporozoite-immunized mice [52]. A similar phenotype was recently observed in an experimental model of *Listeria monocytogenes* [54]. Though it is unclear how CXCR6 expression regulates the survival of CD8<sup>+</sup> T cells in the liver, one possible mechanism is through the up-regulation of anti-apoptotic molecules such as BCL2. In support of this notion, *Cxcr6*<sup>-/-</sup> effector cells exhibited lower expression of BCL2 after malaria liver stage infection [52]. It is worth noting that studies in other models have shown that anti-apoptotic genes are highly expressed by resident CD8<sup>+</sup> T cells in the skin and the brain [55,56]. Another nonexclusive possibility is that homeostatic cytokines, such as IL-7 and IL-15, mediate the lifespan of tissue-resident CD8<sup>+</sup> T cells. Recently, the survival of liver-resident CD8<sup>+</sup> T cells was shown to require IL-15 after immunization with *P. berghei* sporozoites [57].

### Parasite-specific CD8<sup>+</sup> T cells: A unique liver-resident memory population

While it is clear that environmental cues regulate the survival of parasite-specific memory CD8<sup>+</sup> T cells in the liver, it was previously unknown whether this population was maintained through local proliferation or through the migration of memory cells from the periphery [58,59]. In the latter scenario, the CD8<sup>+</sup> T cells recovered from the liver would exhibit an overlapping phenotype and molecular signature with those recovered from the lymphoid organs. To explore these possibilities, we used TCR transgenic mice specific for the H-2K<sup>d</sup>-restricted *P. yoelii* epitope SYVSAEQI [60] and compared the gene transcription profiles of the clonal CD8<sup>+</sup> T cell population from different tissues [51]. Memory CD8<sup>+</sup> T cells recovered from the spleen were mostly CD62L<sup>Hi</sup> and CCR7<sup>Hi</sup>, a phenotype of the central memory population (T<sub>cm</sub>). In contrast, memory CD8<sup>+</sup> T cells residing in the liver were CD62L<sup>Lo</sup> and CCR7<sup>Lo</sup>, suggesting that these T cells are predominantly effector cells (T<sub>em</sub>) that have entered non-lymphoid tissues from the circulation. A comparative transcriptome analysis of resident memory CD8<sup>+</sup> T cells in the spleen versus the liver revealed striking differences between these T cell populations [51]. A large number of genes involved in transcription, cell cycle, trafficking, and signaling were differentially regulated in the liver-resident memory CD8<sup>+</sup> T cell population and these cells underwent more vigorous homeostatic proliferation than their counterparts in the spleen. Importantly, liver-resident memory CD8<sup>+</sup> T cells were homogeneously CD69<sup>Hi</sup> and expressed lower levels of the transcription factors Eomes and KLF2, consistent with the tissue-resident (T<sub>rm</sub>) phenotype recently reported [61]. KLF2 is known to regulate the expression of S1PR1 and CD62L, two crucial molecules involved in lymphocyte recirculation. Subsequently, the loss of KLF2 was found to prevent the recirculation of CD8<sup>+</sup> T cells and promote the establishment of a tissue-resident memory population [62]. Hence, the KLF2<sup>Lo</sup> memory population observed in the livers of sporozoite-immunized mice is likely distinct from the circulating T<sub>em</sub> population, suggesting that this population resides in the liver and is not frequently renewed by lymphoid migrants (Fig. 1).



## Protection mediated by local T cells: Direct or indirect killing?

Tissue-resident memory T cells (Trm) are critical for host resistance [63,64] and have been shown to provide better protection than other memory populations against vaccinia and influenza viruses [63,65]. Although virus-specific Trm can recruit circulating memory T cells [66], recruited cells likely contribute a minor role to protection against malaria parasites because of the brief period in which CD8<sup>+</sup> T cells can limit their development in the liver (2 days in mice). For this reason, host resistance against malaria liver stages is probably mediated by tissue-resident CD8<sup>+</sup> T cells. Indeed, we observed that mice with reduced numbers of liver-resident memory CD8<sup>+</sup> T cells were less protected against live sporozoite challenge, despite having normal numbers of memory CD8<sup>+</sup> T cells in the spleen [52].

Given the short window in which liver-stage parasites can be eliminated, T cells must be poised for an immediate antimicrobial response. It is well established that CD8<sup>+</sup> T cells recognize pathogen-derived peptides presented by MHC I molecules on infected cells. Therefore, the elimination of target cells is generally considered to be a cell-contact dependent process. In fact, mouse studies using bone marrow chimeras revealed a requirement for direct peptide recognition between effector CD8<sup>+</sup> T cells and hepatocytes as CD8<sup>+</sup> T cells were unable to eliminate cells bearing a non-cognate MHC class I molecule [25]. Recently, the elimination of parasitized hepatocytes by effector CD8<sup>+</sup> T cells was visualized using intravital imaging [46]. Strikingly, large clusters of CD8<sup>+</sup> T cells formed around infected hepatocytes (Fig. 1). The initiation of these T cell clusters was strictly dependent on cognate interactions as CD8<sup>+</sup> T cells specific for an irrelevant antigen did not initiate clustering around hepatocytes. More importantly, T cell clustering required G-protein coupled signaling and appeared to be crucial for the effective elimination of parasitized hepatocytes because the protective capacity of effector cells was greatly reduced following inhibition of T cell clustering with pertussis toxin [46]. Together, our *in vivo* imaging studies demonstrated that the killing of target cells is a prolonged process that requires the recruitment of multiple T cells to the infected hepatocyte. In addition to direct recognition of infected hepatocytes, malaria-specific CD8<sup>+</sup> T cells mediate protection through the production of several effector cytokines, including IFN- $\gamma$  and TNF $\alpha$ , and to some extent perforin/granzyme B [67,68]. Liver-resident memory CD8<sup>+</sup> T cells were found to express higher levels of granzyme B and IFN- $\gamma$  transcripts than their counterparts in other organs [51]. However, it is likely that these pathways work redundantly because CD8<sup>+</sup> T cells lacking IFN- $\gamma$  could still limit parasite growth [69] and mice lacking perforin and FasL mounted a protective response against *Plasmodium* parasites (reviewed in [70,71]).

Within the last few years, studies have highlighted the role of CD8<sup>+</sup> T cells not only as efficient cytolytic T cells, but as ‘early sensors’ of foreign pathogens in lymphocytic choriomeningitis virus, herpes simplex virus, and vaccinia virus infection models. In these studies, IFN- $\gamma$  produced by tissue-resident T cells triggered local anti-viral responses and facilitated protection against irrelevant viral antigens [72,73]. In light of these findings, we tested whether sporozoite-specific CD8<sup>+</sup> T cells could eliminate parasitized hepatocytes through bystander killing independent of, or in addition to, the direct killing of target cells. To address these questions, mice were co-infected with two different *P. berghei* parasites in

equal ratios, with only one of the strains carrying a CD8<sup>+</sup> T cell epitope recognized by TCR transgenic cells that were adoptively transferred to these mice. In contrast to what was observed with viral pathogens, malaria-specific CD8<sup>+</sup> T cells failed to eliminate the irrelevant ‘bystander’ parasites [74] (Fig. 1). These results suggest that sterile immunity likely requires the location and killing of every infected cell in a cell-contact dependent manner. Moreover, they further support the notion that CD8<sup>+</sup> T cells recognize parasite-derived peptides directly on the surface of hepatocytes.

Because parasite-infected cells are eliminated in a cell-contact dependent manner involving multiple T cells, it is not surprising that the number of memory CD8<sup>+</sup> T cells needed for protective immunity against malaria parasites far exceeds what is required for bacterial and viral infections (~100–1000 fold) [75]. A sufficiently high number of anti-parasite CD8<sup>+</sup> T cells, particularly in the liver, is critical for early pathogen control since one schizont that escapes immunosurveillance can give rise to thousands of merozoites. Thus, early pathogen control in the liver is key to host resistance. These stringent requirements, in part, explain the challenges to achieving sterile protection against malaria parasites.

### CD8<sup>+</sup> T cell responses to blood-stage malaria

After an obligatory developmental stage within hepatocytes, *Plasmodium* parasites develop into merozoites and infect erythrocytes. Since erythrocytes lack the expression and processing machinery of MHC I molecules, CD8<sup>+</sup> T cells are not generally thought to play a major role in controlling malaria blood stages. Nevertheless, studies have shown that CD8<sup>+</sup> T cell depletion results in the delayed clearance of *P. chabaudi* blood stages in mice [76] and blood-stage specific CD8<sup>+</sup> T cell responses have been described in rodents [77] and in *P. falciparum*-infected humans [78]. These studies suggest that CD8<sup>+</sup> T cells, to some extent, contribute to protective immunity against blood-stage infection; however, CD8<sup>+</sup> T cells have also been implicated in severe host pathology in experimental models. For a more detailed review of this topic, we direct the reader to the article written by Renia and colleagues in this special issue.

### Implications for malaria vaccine effort

It is well established that live, but not dead, malaria sporozoites can induce protective CD8<sup>+</sup> T cell responses after ID and intravenous (IV) routes of immunization [25,79]. Our recent studies demonstrating a requirement for direct parasite access to the DLNs for CD8<sup>+</sup> T cell priming [17] provide an explanation for why ID injection of dead sporozoites, which do not reach the DLNs [5], elicit poor CD8<sup>+</sup> T cell responses [25]. However, the requirement for live sporozoites following an IV route of immunization is less obvious. To elaborate, sporozoites injected IV have direct access to the spleen and we have previously shown that CD8<sup>+</sup> T cells are first primed here after IV immunization [60]. Thus, it is possible that motile sporozoites deliver antigens that are efficiently presented by DCs in the lymphoid organs, e.g. shed or particulate antigens. Another possibility is that sporozoite traversal through host cells deposits antigens in the cytosol while triggering a cytokine microenvironment that favors CD8<sup>+</sup> T cell priming. As with dead sporozoites, antibody-treated sporozoites injected ID do not reach the DLNs and fail to prime robust CS-specific



CD8<sup>+</sup> T cell responses [17,43]. These findings may have direct implications for vaccine efforts as they indicate that it might be difficult to generate CD8<sup>+</sup> T cell responses in individuals with high-titer anti-CS antibodies.

The superior immunogenicity of live versus dead parasites is not unique to malaria, but was also observed in vaccination models of *L. infantum chagasi* and *S. mansoni* [80,81]. Furthermore, an experimental model of *T. gondii* infection recently revealed that antigens derived from direct infection, and not through phagocytosis of heat-killed or invasion-blocked parasites, were critical for optimal CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses [82]. In addition to providing a diverse array of antigens, live attenuated parasites more efficiently access tissue and cellular compartments required for the induction of long-lasting immunity. Therefore, it appears that the strategies important for parasite infection—cell traversal and cell invasion—may also be critical for immune activation, and should be incorporated into whole parasite vaccine approaches.

Although live, radiation-attenuated parasites represent the ‘gold standard’ for malaria vaccination and form the rationale for a whole parasite vaccine [83,84], there are several technical limitations to this approach (reviewed in [85]). The recent success of the IV administered live, attenuated *P. falciparum* sporozoite vaccine is encouraging [83]; however, it remains to be seen whether large-scale IV vaccination is feasible and if this vaccine can provide long-term protection in humans. In addition to whole parasite vaccine approaches, vaccines designed to induce CD8<sup>+</sup> T cell-mediated immunity through the use of attenuated viral vectors or synthetic constructs have yielded encouraging results in mouse studies but have yet to demonstrate significant efficacy in human trials. Finally, the observation that sterile protection against *Plasmodium* parasites requires a large population of intrahepatic memory CD8<sup>+</sup> T cells raises several questions as to whether these numbers can be maintained in vaccinated humans. The demonstration of prolonged antigen presentation, and the subsequent activation of naive recent thymic emigrants [28], is promising because it suggests a potential mechanism by which the memory CD8<sup>+</sup> T cell population may be renewed after vaccination with non-replicating organisms. Because it is clear that CD8<sup>+</sup> T cells exert their protective function in the liver, further research is required to understand how to recruit and maintain these cells in this organ.

## Conclusions

CD8<sup>+</sup> T cells are critical for protective immunity against several medically important pathogens, especially the malaria parasite *Plasmodium* spp. Through the use of experimental models of malaria infection, transgenic animals, and high-resolution imaging we have gained valuable insight into the factors regulating the activation, survival, and effector functions of malaria-specific CD8<sup>+</sup> T cells. Despite these advances, it is clear that protective immunity against malaria parasites is more stringent than for other pathogens, requiring the location and elimination of every infected cell in a cell-contact dependent manner. Therefore, an important future direction will be to translate the information obtained from experimental models to the development of effective vaccines in humans.

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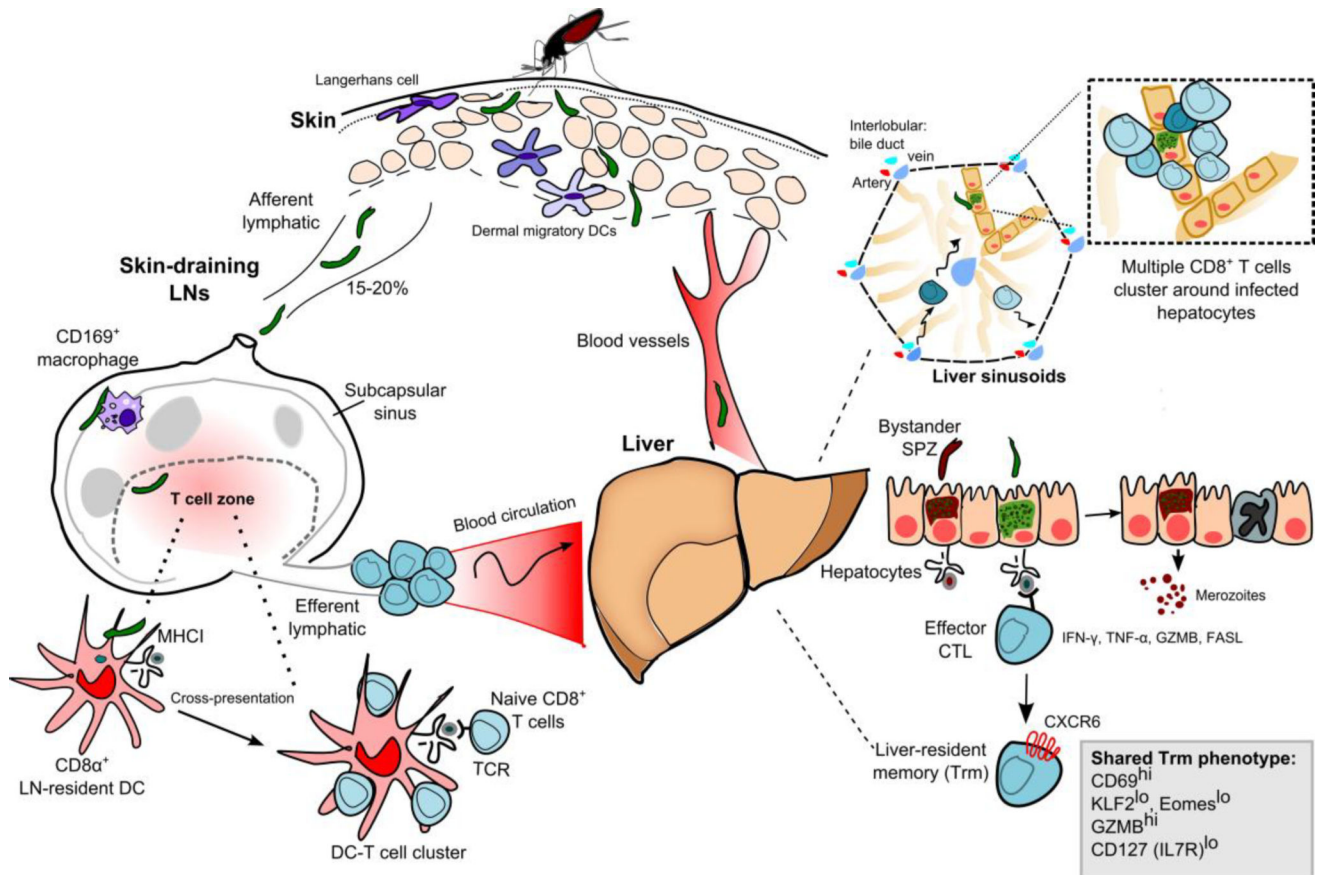
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**Fig. 1.**

Anopheline mosquitoes inject *Plasmodium* sporozoites (SPZ) into the skin of their mammalian host, a tissue populated by epidermal Langerhans cells and dermal DCs. Sporozoites must enter the bloodstream to continue their development in the liver; however, a proportion of inoculated sporozoites (~15–20%) reach the DLN. Sporozoites that migrate to the DLN are first found in the subcapsular sinus in association with CD169<sup>+</sup> macrophages. Within the T-cell zone, CD8α<sup>+</sup> LN-resident DCs uptake and cross-present SPZ-derived antigens to naive CD8<sup>+</sup> T cells. Following their activation in the DLN, malaria-specific CD8<sup>+</sup> T cells enter the blood circulation and reach peripheral tissues, including the liver. Antigen-specific CD8<sup>+</sup> T cells patrol along the liver sinusoids before forming large clusters of multiple effector T cells around an infected hepatocyte. The elimination of parasite-infected hepatocytes by CD8<sup>+</sup> T cells is a cell-contact dependent process that requires the recognition of cognate parasite-derived peptides presented by MHC I molecules on infected hepatocytes. Liver-resident memory CD8<sup>+</sup> T cells exhibit a tissue-resident memory T cell phenotype (Trm) and depend on the chemokine receptor CXCR6 for their long-term maintenance in this organ.