

# Bioprospecting endophytic diazotrophic *Lysinibacillus sphaericus* as biocontrol agents of rice sheath blight disease

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**Abstract** The present study tried to explore the possible in vitro biocontrol mechanisms of *Lysinibacillus sphaericus*, a diazotrophic endophyte from rice against the rice sheath blight pathogen *Rhizoctonia solani*. The in vivo biocontrol potential of the isolate and the induction of systemic resistance under greenhouse conditions have also been experimented employing different treatments with positive control carbendazim, the chemical fungicide. The endophytic isolate showed 100% growth inhibition of the fungal pathogen via volatile organic compound production and was positive for the production of siderophores, bio-surfactants, HCN, and ammonia. Under greenhouse conditions, foliar and soil application of *L. sphaericus* significantly decreased the percentage of disease incidence. All bacterized treatments are superior to chemical fungicide treatment. Application of *L. sphaericus* in single and combination treatments induces systemic resistance as evident from the significant accumulation of defense enzymes such as peroxides, polyphenol oxides and phenylalanine ammonia in addition to the increase of phenolic compounds. The results biologically prospect endophytic diazotroph *L. sphaericus* as a potent plant growth promoter with excellent biocontrol efficiency.

**Keywords** *Lysinibacillus sphaericus* · Diazotrophs · ISR · Biocontrol · Siderophore · *Rhizoctonia solani*

## Introduction

Sheath blight (ShB) of rice, caused by *Rhizoctonia solani*, accounts for significant yield losses worldwide. Strong sources of genetic resistance are not available for ShB and the disease is currently managed through the use of chemical fungicides (Kumar et al. 2011). Fungicidal management of ShB often gives inconsistent results and is not economical. Indiscriminate use of fungicides and chemical fertilizers to increase rice yields creates several concerns related to environmental hazards, pathogen resistance, leaching losses and the destruction of beneficial microflora. Furthermore, overfertilization of synthetic fertilizers, especially N fertilizers, and intensive tillering are considered as major predisposing factors of the rice sheath blight (Slaton et al. 2003). Even though biological control of the rice sheath blight by various plant growth-promoting rhizobacteria (PGPR) has been well studied, several limitations have been found associated with the use of such PGPRs mainly through exposure to various biotic and abiotic stresses (Malfanova et al. 2011).

The current research focuses on developing endophytic bacteria as potent biocontrol agents, since they can elicit antagonism at the site of infection due to colonization abilities (Hardoim et al. 2008). An endophyte is an endosymbiont that lives inside the host plants without causing any disease. In addition to the basic biocontrol mechanisms like the production of siderophores, HCN, volatile and nonvolatile organic compounds and lytic enzymes, endophytic bacteria are thought to be capable of triggering plant defense mechanisms by a phenomenon known as induced systemic resistance (ISR) (Choudhary and Johri 2009). ISR by endophytic bacteria is known to be triggered by various signaling molecules followed by pathogen invasion. Plants respond to infectious agents by

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recognizing an array of these microbial signals. Siderophores, lipopolysaccharides, exopolysaccharides, volatiles, flagella, salicylic acid, pyocyanins, etc. are among the reported ISR determinants of endophytic and rhizospheric bacteria (Gupta et al. 2012). These molecular proteins are sufficient to stop infections before pathogen establishment (Akram et al. 2015). Therefore, systemic invasions and colonization by competent endophytes with antagonistic potential can play a major role in the induction of systemic resistance.

The ShB disease incidence of rice occurs only in the late tillering stage and early immunization through free-living bacteria fails to offer disease resistance (Kumar et al. 2011). One of the major predisposing factors for ShB incidence is increased use of N fertilizer. Hence, it is important to develop a nitrogen-fixing endophyte of rice plants with biocontrol activity against ShB disease. Various endophytic bacteria have been reported as biocontrol agents (Liu et al. 2009). Ji et al. (2014) reported the suppression of Sh B disease by a rice endophytic diazotrophic *Bacillus subtilis* under gnotobiotic conditions. The present study made an attempt to study the putative in vitro biocontrol mechanisms of endophytic diazotrophic bacteria *Lysinibacillus sphaericus* against the sheath blight pathogen *R. solani* and to assess the in vivo effects of *L. sphaericus* in the suppression of rice sheath blight disease under greenhouse conditions.

## Materials and methods

### Isolation of endophytic nitrogen-fixing bacteria from rice plants

The isolation of endophytic bacteria from rice plants was carried out employing the procedures of Hallmann et al. (2006) with major modifications. Surface-sterilized samples were transferred to Dobereiner's semisolid N-free Bromothymol blue (BTB) agar for the selective isolation of nitrogen-fixing bacteria and incubated at  $28 \pm 2$  °C for 7 days. The isolates from the positive N-free semisolid media were streaked onto solid N-free Dobereiner's media amended with 1 M  $\text{NH}_4\text{Cl}$  (Gyaneshwar et al. 2001) to get isolated colonies. The selected endophytes were subjected to ARA in a Bruker Gas Chromatograph model GC 450 with flame ionization detector (FID) using nitrogen-free Fahraeus broth (NFFB).

### Identification of the isolates

Morphological, cultural and biochemical characteristics of the selected antagonistic endophyte L1, isolated from surface-sterilized leaf tissue were analyzed according to

Bergey's Manual of Systematic Bacteriology. The 16S rDNA typing of the isolate was done using 16S rDNA universal primers: 10–30 F: 50-GAGTTTGATCCT GGCTCAG-30 and 530R: 50-G (AT) ATTACCGCGGC GGCTG-30. The genomic DNA of the isolate was extracted using DNeasy Tissue Kit (Qiagen, Germany) according to the instructions of the manufacturer. The purified amplicon was sequenced using ABI3730xl Genetic Analyzer (Applied Biosystems, USA), and the sequences (16S rDNA) were then used for similarity searches using BLAST (Basic Local Alignment Search Tool) program in the NCBI Gene Bank ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)) DNA database for identifying the bacterial strains.

The *nif H* gene amplification was done according to the procedure described by Balachandar et al. (2006). The genomic DNA was extracted and amplified for *nif H* using colony PCR techniques. *nif* Gene was amplified using the following primer sequences: *nif H* for-5' TAYGG-NAARGGNGGHATYGGYATC and *nif H*-rev ATRTRTTNG CNGC RTAVABBGCC ATCAT (Poly et al. 2001). The evolutionary history of 16S rDNA analysis and *nif* gene amplification was inferred individually using the neighbor-joining method (Tamura et al., 2004). The tree was drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. Phylogenetic analyses were conducted in MEGA4 (Tamura et al. 2007).

### In vitro evaluation of endophytic bacteria for biocontrol of *Rhizoctonia solani*

The rice sheath blight pathogen *Rhizoctonia solani* Kuhn was obtained from the culture collection center, Department of Plant Pathology, Kerala Agricultural University, Mannuthy, Thrissur, Kerala. The pathogen was maintained by regular subculturing on potato dextrose agar (PDA) slants and kept at 4 °C.

The in vitro antagonistic effect of *L. sphaericus* was determined using a dual culture plate assay on nutrient agar: PDA agar. The agar well diffusion assay was performed using the culture filtrate of the overnight-grown NB culture of *Lysinibacillus sphaericus*. The overnight culture was centrifuged at  $18,558 \times g$  for 10 min and the cell free supernatant was used for determining the antifungal activity using the well diffusion assay. A 5 mm mycelial disc of 5-day-old test pathogen was placed in the center of the set nutrient agar: PDA agar plates. Two wells, each about 5 mm in diameter, were made by punching the agar with a sterile steel cork borer on either side of the pathogen inoculums and 100  $\mu\text{l}$  of the culture filtrate was incorporated into the wells. The plates were incubated for 7 days at  $28 \pm 2$  °C and the inhibitory activity of the culture filtrate was expressed as the percent growth inhibition, compared

to the control kept with the sterile supernatant of centrifuged nutrient broth alone incorporated in the wells. The experiment was repeated thrice with three replications.

### In vitro analysis of putative biocontrol mechanisms

Putative biocontrol mechanisms of the selected antagonistic endophyte L1 was assessed in vitro. Qualitative detection of siderophores was done by ferric chloride test according to Suryakala et al. (2004). Quantitative estimation was also done by employing the procedure described by Adhikari et al. (2013). The quantity of siderophores synthesized was expressed as  $\mu\text{mol}$  benzoic acid  $\text{mL}^{-1}$  of the cell free culture filtrate (CFCF). Hydrocyanic acid (HCN) production by the isolates was tested by the method of Miller and Higgins (1970). The production of volatile organic compounds was observed by the paired bottom plate method and that of nonvolatile and/or diffusible organic compounds was done by using CFCF of the isolate studied, adopting well diffusion assay as described elsewhere. Observations were made for the absence of sclerotium germination and the percentage of reduction in the growth of *R. solani* in the control and test plates.

The in vitro  $\beta$  1,3 glucanase activity was measured according to the procedure described by Rajendran et al. (2007) using cell-free culture filtrate. The amount of reducing sugar was calculated from the glucose standards and the activity of  $\beta$  1,3 glucanase was expressed in  $\text{nkat mL}^{-1}$ . One  $\text{nkat}$  corresponds to 1  $\text{nmol}$  of glucose equivalents per seconds. In vitro chitinase activity was also measured using the culture filtrate prepared in the above section. The chitinase activity was expressed as equivalents of *N*-acetyl glucosamine (GlcNAc, Hi Media) kept as standard.

### Biocontrol efficiency under greenhouse conditions

The biocontrol efficiency of *L. sphaericus* against *R. solani* was assessed under greenhouse conditions, using various treatments (Table 3). Rice seeds of crop variety ponmani were obtained from the State Seed Farm, Okkal, Perumbavoor, run by Kerala Agricultural University. The experiment was arranged in a completely randomized design (CRD) and was repeated thrice with three replications per treatment.

### Pot preparation

Soil from rice farming fields (loamy texture, pH 4.68; electrical conductivity,  $0.031 \text{ ds M}^{-1}$ ; organic carbon, 1.92%; available nitrogen,  $128 \text{ kg ha}^{-1}$ ; available phosphorus,  $4.30 \text{ kg ha}^{-1}$ ; available potassium,  $85.12 \text{ kg ha}^{-1}$ ; dehydrogenase activity  $1.36 \mu\text{g TPF soil day}^{-1}$ ) was air

dried and passed through a 2 mm sieve. Fertilizer recommendations as per POP, Kerala Agricultural University, include urea at the rate of  $123 \text{ kg ha}^{-1}$ , Raj phosphate at the rate of  $260 \text{ kg ha}^{-1}$ , muriate of potash at the rate of  $87 \text{ kg ha}^{-1}$  and lime or dolomite  $240 \text{ kg ha}^{-1}$ . Fertilizers at the rate mentioned were weighed separately for each pot and mixed with soil and filled in pots ( $27 \times 25 \text{ cm}$ ) at the rate of 10 kg per pot. N was applied at half rates from the recommended dose and in three equal splits: as basal dose at tillering and final dose after panicle initiation. The P and K fertilizers were applied entirely as per recommendations as basal dose. Lime was applied at two split doses before sowing and 1 month after sowing.

### Preparation of *L. sphaericus* inoculum and mode of application in greenhouse experiments

Rice seeds of the variety ponmani were surface sterilized with 2% sodium hypochlorite for 3 min and washed with sterile distilled water five times. Seeds were divided into approximately two equal parts; part A and part B. Seeds from part A were selected for bacterization with endophytic isolate *L. sphaericus* and fungicide Bavistin 50DF (carbendazim 50%DF manufactured by Biostadt India Ltd) treatment. Surface-sterilized seeds of part B were meant for seedling germination in nursery beds and control experiments. Nursery beds were prepared for all treatments other than seed treatment. For this, randomly selected surface-sterilized seeds from part B were dipped in pre-sterilized tap water in sterile Petri plates for pre-germination in warm condition for 3 days. Pre-germinated seeds were allowed to grow in nursery beds for the next 18 days by maintaining an optimum water regime under proper climatic conditions in a greenhouse.

For seed treatment, randomly selected surface-sterilized seeds from part A were soaked in 24-h-old NB culture ( $2 \times 10^8 \text{ cfu mL}^{-1}$ ) of *L. sphaericus* and incubated for 24 h at room temperature. Thereafter, seeds were air dried in the laminar airflow chamber for 30 min. The seeds were directly transferred to the already prepared potted soil. Four 5 cm digs were made in each pot and each dig received randomly selected five bacterized seeds. Seed-treated pots were prepared separately for combination treatments T7–T12. The positive chemical control carbendazim was also treated with seeds at the rate of  $5 \text{ g kg}^{-1}$  of seeds (Rajendran and Samiyappan 2008).

For soil inoculation, also 24-h-old 5 mL standard inoculum ( $2 \times 10^8 \text{ cfu mL}^{-1}$ ) of *L. sphaericus* was used. 21-day-old seedlings were uprooted from the nursery bed and washed several times with sterilized water and transplanted onto each pit in the pots. Another set of 21-day-old seedlings from the nursery was transplanted to furrowed pots in a similar manner without inoculation. This setup was meant for foliar spray treatments. For foliar spray

treatments, 10 mL of 24-h-old inoculum ( $2 \times 10^8$  cfu mL<sup>-1</sup>) was diluted with 90 mL of tap water and used for an application. Foliar treatment was given on 30 days of plant age, prior to challenge inoculation with the pathogen. Foliar application of fungicide carbendazim was also done in a similar manner at a concentration of 0.1% at similar plant ages.

For the combination of seed treatments, the second set of seed-treated plants received 5 ml standard inoculum in the soil at 21 days of plant age (T7) in addition to seed treatment (T1). In T9, the 30-day-old plants received a foliar treatment in addition to the seed treatment. For treatments T11, the soil-inoculated plants received a foliar application on the 30th day of plant age. Carbendazim was also applied in a similar manner for treatments T8, T10 and T12, respectively. The plants were watered twice a day to maintain optimum soil moisture regime and kept under greenhouse conditions with ambient irradiance, temperature and humidity. The humidity in the green was maintained at around RH 80%. Control plants did not receive any biotization or chemical treatment at any stages of plant growth.

### Pathogen inoculation

The ShB was artificially introduced by challenge inoculation on the 45th day of plant age, according to the methods described by Kumar et al. (2011) with minor modifications. Sclerotia of fungal pathogen *R. solani*, grown on PDA plates as described, served as a source of pathogen inoculum. The leaf sheaths of 45-day-old plants at 2 cm above water level were carefully opened and inoculated with sclerotia at the rate of one sclerotium per plant. After inoculation, the pots were covered with polyethylene sheeting for 5 days to maintain high humidity (>95%), which favored disease development. The minimum and maximum temperatures were  $19 \pm 2$  and  $30 \pm 2$  °C, respectively, during the experiment period. The lesion length was measured after 15 days of challenge inoculation and lesion size was calculated by multiplying with length and width of lesions.

The percent sheath blight intensity (SBI) was determined using the following formula:

$$\begin{aligned} \% \text{ Of disease intensity (DI)} \\ &= 100 \times \text{total lesion size/total plant height.} \end{aligned}$$

$$\begin{aligned} \% \text{ Of disease suppression (DS)} \\ &= 100 - \text{treatment /control} \times 100. \end{aligned}$$

### Effect of *L. sphaericus* in the induction of defense enzymes against *R. solani*

The role of endophytic diazotrophic *L. sphaericus* in induced systemic resistance (ISR) of rice plants against rice

sheath blight pathogen *R. solani* was assessed under greenhouse conditions. Plant seeds treated with chemical fungicide Bavistin 50DF (carbendazim 50%DF manufactured by Biostadt India Ltd) were kept as a positive standard chemical control for comparison. Plants developed from uninoculated seedlings from the same nursery bed prepared for biocontrol efficiency were used as healthy untreated controls. Plants belonging to the same criteria, but challenge inoculated with the fungal pathogen *R. solani* on the 45th day of plant age, served as pathogen control (T8). Here also, the pots were arranged in CRD (completely randomized design) and the experiments repeated thrice with three replications. The pots and plants were prepared as per the methods described for biocontrol efficiency. Greenhouse conditions and watering were also maintained under the same conditions. The mode of pathogen inoculation also followed a similar methodology.

### Sample collection and extraction

Leaf samples from each experiment were carefully excised at different time intervals, viz., 24, 48 and 72 h, after pathogen inoculation, homogenized with liquid nitrogen in a pre-chilled mortar and pestle and kept at  $-80$  °C until analysis. The increased activity of enzymes such as peroxidase (PO), polyphenyl oxidase (PPO), phenylalanine ammonia-lyase (PAL), chitinase and  $\beta$  1,3 glucanase were tested spectrophotometrically using a UV-Vis spectrophotometer. In addition, the effect of *L. sphaericus* in the induction of total phenolic content was also measured. All experiments were done according to the procedures described by Rajendran and Swamiyappan (2008).

### Assay of peroxidase (PO)

Leaf samples from each treatment were homogenized in 3 mL 0.1 M phosphate buffer solution (pH 7.0) at 4 °C. The homogenate was centrifuged at  $20,070 \times g$  at 4 °C for 15 min and the supernatant was used as enzyme source. The reaction mix consisted of 0.05 mL pyrogallol, 0.5 mL enzyme extract and 0.5 mL 1% hydrogen peroxide. The reaction mix was mixed well and incubated at room temperature. The changes in absorbance at 420 nm were recorded at 30 s intervals for 3 min. The enzyme activity was expressed as changes in the absorbance of the reaction mixture  $\text{min}^{-1} \text{g}^{-1}$  fresh weight.

### Assay of polyphenol oxidase (PPO)

One gram of leaf samples from each treatment was homogenized in 2 mL of 0.1 M sodium phosphate buffer at pH 6.5 and centrifuged at  $20,070g$  for 15 min at 4 °C. The supernatant was used as the enzyme source. The reaction

mix included 200  $\mu\text{L}$  of enzyme extract and 1.5 mL of 0.1 M sodium phosphate buffer at pH 6.5. The reaction was allowed to start by adding 200  $\mu\text{L}$  of 0.01 M catechol and the enzyme activity was measured at 495 nm as changes in absorbance  $\text{min}^{-1} \text{g}^{-1}$  fresh weight.

#### Assay of phenylalanine ammonia-lyase (PAL)

The assay mixture for PAL consisted of 100  $\mu\text{L}$  enzyme extract, 500  $\mu\text{L}$  of 50 mM Tris-HCl (pH 8.8) and 600  $\mu\text{L}$  of 1 mM L-phenylalanine. The reaction mix was incubated at room temperature for 60 min and was stopped by adding 2 N HCl. After that, 1.5 mL of toluene was added and vortexed for 30 s and centrifuged at  $78\times g$  for 5 min. The toluene fraction containing the *trans*-cinnamic acid was separated and measured at 290 nm against a toluene blank. The enzyme activity was calculated using a standard graph plotted against different concentrations of cinnamic acid in toluene and expressed as  $n$  moles *trans*-cinnamic acid  $\text{min}^{-1} \text{gm}^{-1}$ .

#### Assay of $\beta$ 1,3 glucanase activity

$\beta$  1,3 glucanase activity was assayed by the laminarin-dinitrosalicylate method. One gram of leaves was extracted from each treatment with 5 mL of 0.05 M sodium acetate buffer (pH 5.0) by grinding at 4 °C using a mortar and pestle. The extract was centrifuged at  $20,070\times g$  for 15 min at 4 °C and the supernatant was used as the enzyme source. The reaction mixture consisted of 62.5  $\mu\text{L}$  of 4% laminarin and 62.5  $\mu\text{L}$  of enzyme extract. The reaction was carried out at 40 °C for 10 min and stopped by adding 375  $\mu\text{L}$  of dinitrosalicylic reagent and heating for 5 min in a boiling water bath. The resulting colored solution was diluted with 4.5 mL of distilled water, vortexed and the absorbance was measured at 500 nm. The enzyme activity was expressed as  $\mu\text{g}$  glucose released/min/g fresh weight.

#### Assay of chitinase activity

Chitinase enzyme activity was assayed spectrophotometrically using colloidal chitin (see Appendix) as substrate. One gram of leaf sample from each treatment was homogenized in 2 mL of 0.1 M sodium citrate buffer (pH 5.0). The homogenate was centrifuged at  $7840\times g$  for 15 min at 5 °C, and 0.4 mL supernatant was added to the reaction mixture consisting of 10  $\mu\text{L}$  of 0.1 M sodium acetate buffer (pH 4) and 0.1 mL of colloidal chitin. After incubation of 2 h at 37 °C, the reactions were stopped by centrifugation at  $78\times g$  for 3 min. 0.3 mL of the supernatant was pipetted into the glass reagent tube containing 30  $\mu\text{L}$  of 1 M potassium phosphate buffer (pH 7) and incubated with 20  $\mu\text{L}$  of 3% (v/v) snail gut enzyme for 1 h.

After 1 h, the mixture was brought to pH 8.9 by adding 70  $\mu\text{L}$  0.1 M sodium borate buffer (pH 8.9). The mixture was incubated in a boiling water bath for 3 min, followed by rapid cooling in ice. The mixture was incubated for 20 min at 37 °C immediately after the addition of 2 mL DMAB. The absorbance was measured at 585 nm. *N*-acetyl glucosamine (GlcNAc) (HI Media) was used as the standard and the enzyme activity was expressed as GlcNAc equivalent $^{-1} \text{g}^{-1}$  fresh weight.

#### Assay of total phenolic content

One gram of leaf tissue from each treatment was homogenized in 10 mL of 80% methanol (HiMedia) using a mortar and pestle. It was mixed well and agitated in a water bath shaker for 15 min at 70 °C. 1 mL of this methanolic extract was added to a mixture of 5 mL distilled water and 250  $\mu\text{L}$  1 N Folin-Ciocalteu reagent. The solution was kept at 25 °C for 3 min. After incubation, 1 mL of the saturated solution of sodium carbonate and 1 mL of distilled water were added to the reaction mix and incubated for 1 h at 25 °C. The absorbance of the blue color that developed was measured at 725 nm. The total soluble phenol was calculated from standard graph drawn and expressed as catechol  $\text{g}^{-1}$  fresh weight.

#### Statistical analysis

Data from repeated laboratory and greenhouse studies were analyzed separately and subjected to analysis of variance (ANOVA) (SPSS Software) and the least significant difference test (LSD) was performed to separate the group means when ANOVAs were significant at  $P \leq 0.05$ . The means were compared by Duncan's multiple range test using the statistical package SAS version 8.3 (SAS Institute Inc., Cary, NC, the USA). The alphabets, given as superscripts in the tables denote ranking of treatments based on Duncan's multiple range test.

## Results

### Identification of the endophytic diazotroph

The isolate L1 was identified as *L. sphaericus* based on its morphological, biochemical and genotypic characteristics. BLAST analysis revealed 99.9% sequence similarity with *L. sphaericus*. The 16S rRNA amplicon was deposited in the National Center for Biotechnology Information (NCBI) GenBank under Accession Number KJ 872548. The *nif* genes were shown to be *nif* H specific by BLAST analysis and sequence similarity search in NCBI GenBank. The

sequences of *nif* H amplicons were deposited in the NCBI GenBank with Accession Number KR133403.

### Putative in vitro biocontrol mechanisms

The endophytic diazotrophic isolate *L. sphaericus* was able to produce siderophores ( $17.85 \pm 0.04 \mu\text{mol/mL}$  Culture filtrate). The isolate showed 100 and 76% of growth inhibition in volatile and nonvolatile organic compound production, respectively. HCN production was positively detected by the change in color of filter paper dipped in picric acid solution. In vitro production of  $\beta$  1,3 glucanase and chitinase by the antagonist *L. sphaericus* was  $0.08 \pm 0.02 \text{ mg mL}^{-1}$  glucose and  $2.2 \pm 0.03 \text{ mg mL}^{-1}$  *N*-acetyl glucosamine (Table 1). The positive reaction in qualitative drop collapsing test was confirmed quantitatively by a higher emulsification index (E24) of  $9.09 \pm 0.004$  and  $7.4 \pm 0.04$  in both the medium tested, proving the production of biosurfactants (Table 2).

### Induction of defense enzymes

Studies on the induction of defense enzymes revealed that upon pathogen inoculation, higher amounts of defense enzymes were expressed in bacterized rice plants in comparison to non-bacterized plants. In all the experiments,

**Table 1** Bioactive metabolites produced by the selected endophytic diazotroph *Lysinibacillus sphaericus* under in vitro conditions

In vitro analysis of bioactive metabolites (Plate assay)	Results
Siderophores	$17.85 \mu\text{mol benzoic acid mL}^{-1}$ CFCF
Ammonia	Positive
Biosurfactants	Positive
HCN	Positive
Volatile organic compound production	100% Mycelial inhibition
Nonvolatile organic compound production	76.9% Mycelial inhibition
In vitro $\beta$ 1,3 glucanase	Negative
In vitro chitinase	Negative

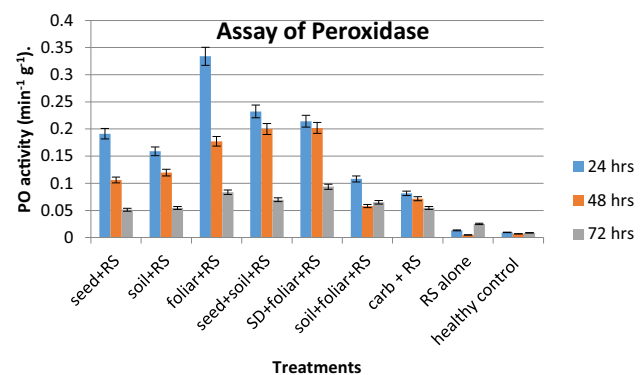
**Table 2** Emulsification index analysis for assessing the biosurfactant production ability of the selected isolate *Lysinibacillus sphaericus*

Isolate	Type of oil	Emulsification index (E24) <sup>a</sup>
<i>Lysinibacillus sphaericus</i>	Cooking oil	$9.09 \pm 0.004$
	Engine oil	$7.43 \pm 0.04$
Control	Cooking oil	$2.33 \pm 0.23$
	Engine oil	$5.22 \pm 0.15$

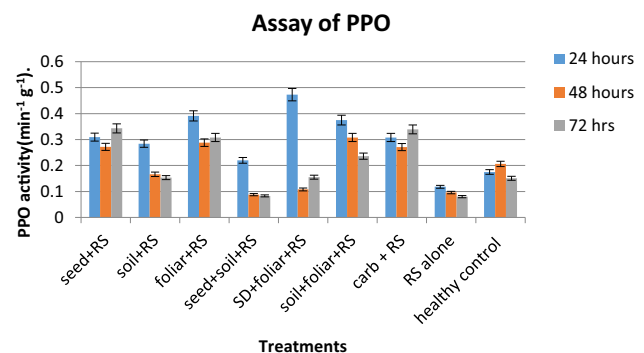
<sup>a</sup> Values are mean of three replications  $\pm$  SD

foliar treatments resulted in the highest enzyme production during 24 h past inoculation both in single and combination treatments. In general, the pattern of enzyme induction was found to reach the maximum during 24 h of time intervals and thereafter declined gradually. The PO activity was found to vary according to the time intervals of sampling. Plants which received bacterial application as foliar treatment (T3) showed the highest PO activity, followed by combination treatments T4 and T5. Plants treated with fungicide carbendazim also showed induction of PO activity (Fig. 1). The PO activity was found to decrease toward 72 h after challenge inoculation with fungal pathogen and in plants without any treatment.

The PPO enzyme activity was found to increase at 24 h after challenge inoculation in all treatments considerably. The activity tends to be decreased 48 and 72 h after challenge inoculation. The highest PPO activity at 24 h was shown by T3 ( $0.39 \text{ min}^{-1} \text{ g}^{-1}$ ) and is statistically at par with treatment T6 ( $0.38 \text{ min}^{-1} \text{ g}^{-1}$ ). At 72 h, the treatments T1 and T7, which received seed treatment of *L. sphaericus* and carbendazim, showed significant hike in PPO activity (Fig. 2) The highest PAL activity at all time intervals was



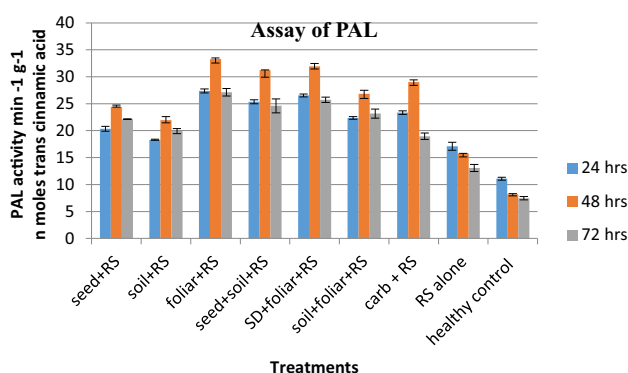
**Fig. 1** Changes in peroxidase activity ( $\text{min}^{-1} \text{ g}^{-1}$ ) by different treatments of endophytic *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions



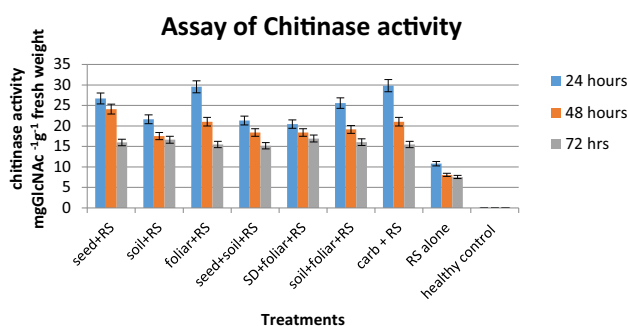
**Fig. 2** Changes in poly phenol oxidase (PPO  $\text{min}^{-1} \text{ g}^{-1}$ ) activity by different treatments of endophytic bacteria *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions

given by foliar treatments alone and in combination with seed and soil (Fig. 3). Plants treated with fungicide carbendazim also showed induction of PAL similar to bacterized plants in the time course, but at lower levels. Uninoculated healthy controls showed a linear decrease in PAL activity at 24, 48 and 72 h.

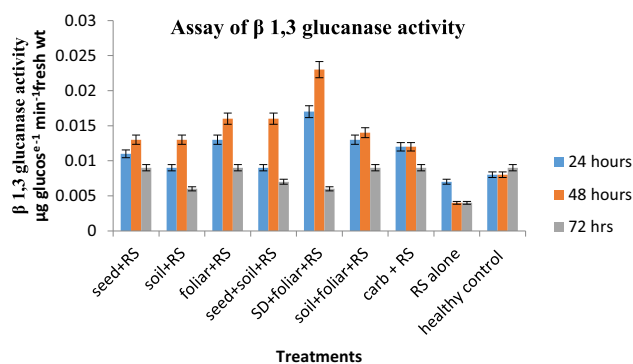
Induction of chitinase enzyme by endophytic bacteria in rice plants challenged with *R. solani* did not show much variation among different treatments under greenhouse conditions. Increase in chitinase activity was detected in appreciable amounts both 24 and 48 h after treatment and then declined (Fig. 4). The highest chitinase enzyme activity at 24 h after treatment was shown by rice plants treated with (T7) chemical fungicide carbendazim (30.12 mgGlcNAc<sup>-1</sup> g<sup>-1</sup> fresh weight), followed by foliar treatment (T3) (29.43 mgGlcNAc<sup>-1</sup> g<sup>-1</sup> fresh weight). All the treatments, including the positive control carbendazim, showed very minute level of induction of  $\beta$  1,3 glucanase enzyme activity at 24, 48 and 72 h of sampling (Fig. 5). Very low and statistically similar values were noticed in both the control plants, viz., disease control (T8) which did not receive any treatment and healthy control plants without



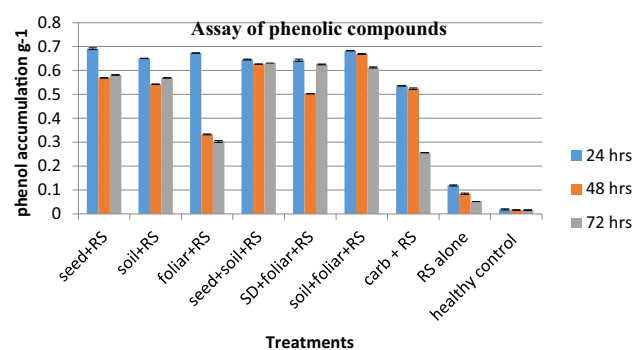
**Fig. 3** Changes in phenyl alanine lyase (PAL) activity by different treatments of endophytic bacteria *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions



**Fig. 4** Changes in chitinase activity by different treatments of endophytic bacteria *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions



**Fig. 5** Changes in  $\beta$  1,3 glucanase activity by different treatments of endophytic bacteria *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions



**Fig. 6** Changes in total phenol accumulation (g<sup>-1</sup>) by different treatments of endophytic bacteria *L. sphaericus* in rice plants challenged with *R. solani* under greenhouse conditions

challenge inoculation (T9). In general, an increase in enzyme activity was shown by all treatments at 48 h from that of 24 h after challenge inoculation. The enzyme activity was found to be declining at 72 h after challenge inoculation. In addition, accumulation of phenols was found to start after 24 h of treatment and remained more or less constant in all treatments challenge inoculated with *R. solani*, except for foliar treatments. In foliar treatments, phenolic accumulation was shown to be decreased toward 72 h of treatment and all results were significantly higher than those of the control plants (Fig. 6).

### Biocontrol efficiency under greenhouse conditions

Rice diazotrophic endophyte *L. sphaericus* effectively controlled the rice sheath blight disease with less variation in % DI and % DS. Treatment T3, which received an application of *L. sphaericus* in the form of a single foliar spray, showed the highest % DI and % DS (5.20 and 84.75%, respectively), which is found to be statistically on par with treatment T2 with soil applications of *L. sphaericus* (6.36 and 81.35%, respectively). The positive control

**Table 3** Control of rice sheath blight disease under greenhouse conditions by the selected endophytic isolate *Lysinibacillus sphaericus* in comparison to a registered fungicide, carbendazim

Treatments	Treatment details	Lesion height (cm)	Plant height (cm)	% DI	% DS
T1	SD ( <i>L. sphaericus</i> )	4.83 <sup>def</sup>	64.63 <sup>c</sup>	7.48 <sup>de</sup>	78.06 <sup>ab</sup>
T2	SO ( <i>L. sphaericus</i> )	4.17 <sup>f</sup>	65.10 <sup>b</sup>	6.36 <sup>e</sup>	81.35 <sup>a</sup>
T3	FS ( <i>L. sphaericus</i> )	3.37 <sup>c</sup>	64.70 <sup>c</sup>	5.20 <sup>e</sup>	84.75 <sup>a</sup>
T4	SD + SO ( <i>L. sphaericus</i> )	4.33 <sup>ef</sup>	65.40 <sup>b</sup>	6.63 <sup>e</sup>	80.56 <sup>a</sup>
T5	SD + FS ( <i>L. sphaericus</i> )	4.33 <sup>ef</sup>	67.40 <sup>a</sup>	6.43 <sup>e</sup>	81.14 <sup>a</sup>
T6	SO + FS ( <i>L. sphaericus</i> )	4.37 <sup>ef</sup>	67.13 <sup>a</sup>	6.50 <sup>e</sup>	80.94 <sup>a</sup>
T7	SD (carbendazim)	10.07 <sup>b</sup>	62.10 <sup>e</sup>	16.21 <sup>b</sup>	52.49 <sup>d</sup>
T8	SO (carbendazim)	7.85 <sup>bc</sup>	62.07 <sup>e</sup>	12.65 <sup>bc</sup>	62.80 <sup>cd</sup>
T9	FS (carbendazim)	6.93 <sup>cde</sup>	62.10 <sup>e</sup>	11.15 <sup>cd</sup>	67.41 <sup>bc</sup>
T10	SD + SO (carbendazim)	7.47 <sup>bcd</sup>	62.40 <sup>de</sup>	11.98 <sup>bc</sup>	64.83 <sup>c</sup>
T11	SD + FS (carbendazim)	7.40 <sup>bcd</sup>	62.67 <sup>d</sup>	11.81 <sup>cd</sup>	68.16 <sup>bc</sup>
T12	SO + FS (carbendazim)	6.93 <sup>cde</sup>	62.30 <sup>de</sup>	11.13 <sup>cd</sup>	67.25 <sup>bc</sup>
T13	Disease control	18.20 <sup>a</sup>	53.37 <sup>f</sup>	34.10 <sup>a</sup>	0.00 <sup>e</sup>
LSD at 5%		4.38	12.07	2.73	0.47

\* All values are means from three repeated experiments with three replications and means in a column followed by the same letter(s) are not significantly different according to Duncan's multiple range test ( $P = 0.05$ ) at 5% level of significance

carbendazim showed only 67.4 and 64.83% DS and 11.15 and 12.65% DI, respectively, for the same treatments. This shows the superiority of endophytic *L. sphaericus* as efficient colonizers and the ineffectiveness of chemical fungicides.

The application of *L. sphaericus* in combination treatments did not show much variation. The highest % DS among treatments in combination was given by T4, i.e., seed plus soil (80.56%). The treatments T5 and T6 showed 81.14 and 80.94%, respectively. Control plants which did not receive any biological or chemical treatment showed 34.07% of DS, confirming the potential of endophytic diazotrophic *L. sphaericus* in the suppression of rice sheath blight effectively (Table 3).

## Discussion

Beneficial plant microbial interactions offer great opportunities for sustainable agriculture. Eco-friendly management of crop diseases has been practised for many years extensively using PGPRs. As competent endophytes are evolved from potent root colonizers among PGPRs, the use of endophytic bacteria gains significant attention in effective and localized biocontrol (Hardoim et al. 2008; Gupta et al. 2012). The production of various allelochemicals and induced systemic resistance are the major mechanisms of action involved in biological control. Many reports are available with biocontrol abilities of endophytic bacteria isolated from diverse hosts, mainly belonging to *Bacillus*

sp. (Yang et al. 2011; Wang et al. 2009; Rajendran and Swamiyappan 2008). Ji et al. (2014) reported the antagonistic effects of endophytic diazotroph *Bacillus subtilis* against *R. solani* in rice. In their exclusive review, Gupta et al. (2012) describe that volatile organic compound production plays a major role in the biocontrol of plant pathogens via induction of systemic resistance. In the present study, 100% inhibition of mycelial growth could be observed in the volatile organic compound production. *Bacillus* sp. has been found to produce exceptional biocontrol activities due to volatile organic compound production (Solanki et al. 2015; Norman et al. 2011). Strobel (2006) reported the biocontrol efficiency of a volatile organic compound, 2,3-butanediol from endophytic *Bacillus* spp.

Microbial biosurfactants have been known to play a significant role in antifungal activities. Biocontrol activity of biosurfactants is due to detergent-like activity in dissolving cell wall lipid membrane, resulting in the loss of cell integrity and viability. Ongena et al. (2005) reported surfactants as elicitors of induced systemic resistance in plants by bacteria against phytopathogens. Microorganisms use the ability of hydrolytic enzyme production, such as cellulase, amylase, pectinase, chitinase,  $\beta$  1, 3 glucanase and xylanase, primarily as the mechanism of biocontrol. By producing these lytic enzymes, they can cleave the cell walls of fungal pathogens and cause mycelial destruction. Many plant growth-promoting endophytic bacteria are reported to produce in vitro lytic enzymes (Amareesan et al. 2014). As far as endophytes are considered, the production



of lytic enzyme plays other important roles in vertical spreading and secondary cell wall lysis of host plants, enabling colonization and endophytic establishment (Hurek and Hurek 2003).

Siderophore production by diazotrophic bacteria is a key phenomenon of metal chelation under iron-deficient condition, since the nitrogen-fixing enzyme nitrogenase requires both iron and molybdenum for activity (Gupta et al. 2012). Siderophores produced by biocontrol bacteria have a higher affinity for iron than the siderophores produced by fungal pathogens, allowing the antagonistic bacteria to scavenge most of the available iron and thereby prevent a proliferation of fungal pathogens. Ji et al. (2014) recently reported the endophytic diazotrophs *Klebsiella pneumoniae*, *B. subtilis* and *Microbacterium* sp., which showed antagonism toward *R. solani* as siderophore producers. Ammonia production by bacteria plays many roles in plant physiology and plant growth promotion and is produced mainly through nitrite ammonification in diazotrophic bacteria (Geetha et al. 2014). It acts as an antagonistic factor against phytopathogens also in combination with HCN and siderophore production (Allu et al. 2014). Haroim et al. (2008) reviewed that host plants play a major role in the selection of a bacterium to be an endophyte. Therefore, it is clear that the production of siderophores, ammonia and lytic enzymes by the diazotrophic isolate *L. sphaericus* is beneficial to the plant and to the bacterium itself, revealing its endophytic and symbiotic lifestyle.

Inducing a plant's own defense mechanism by prior application of a biological agent is an emerging concept and strategy in plant defense mechanism. Induced resistance is a physiological "status of improved defensive capacity" elicited by specific environmental stimuli, whereby the plant's innate defenses are potentiated against subsequent biotic challenges (van Loon 2000). Various signals from bacteria can act as ISR elicitors, including LPS, flagella, siderophores, antibiotics, volatile organic compounds and quorum signaling (van Loon et al. 2006). This is proved by the trends of increased accumulation of all defense enzymes, PR proteins, and phenolic compounds after 24 h of pathogen inoculation in the bacterized plants in ISR studies of *L. sphaericus*.

The role of PO, PPO and PAL in ISR studies has been documented by various authors against *R. solani* and other fungal pathogens in rice and other diverse crops (Chitrasree et al. 2011; Rajendran and Samiyappan 2008; Jayaraj et al. 2004). PAL is the key biosynthetic enzyme for a variety of plant secondary metabolites and plays a major role in phenolic compound accumulation (Zdor and Anderson 1992). The present study showed increased cinnamic acid accumulation in all treatments in a considerable manner. Cinnamic acid production actively occurs under pathogen

stress and also acts as a precursor for the synthesis of phenolic compound ferulic acid from phenylalanine through the phenyl propanoid pathway catalyzed by PAL. In this study, total phenolic accumulation was also found to vary in accordance with PAL accumulation. The PAL accumulation in healthy control plants was higher compared to phenolic accumulation. This can be justified by an argument that phenolic compounds are involved in anti-fungal activities too. In healthy plants, signals for phenolic accumulation may be absent due to lack of challenge inoculation and bacterization. These highlight and confirm the role of introducing endophytic bacteria *L. sphaericus* in induction of defense mechanisms.

Chitinase accumulation was found to be greater in response to pathogen invasion and might contribute to increased plant resistance. Similar results are reported by (Shrestha et al. 2008; Rajendran and Samiyappan 2008; Radiacommare et al. 2004) in rice plants. This is in contradiction to the in vitro experiments where chitinase production was absent in plate assay and estimations. This may be due to the expression of plant genes for chitinase. It is also clear from the data that in the uninoculated disease control, chitinase level was lower compared to inoculated and challenged plants. Hence, it can be assumed that the in vivo production of chitinase enzyme happened due to the combined action of *L. sphaericus* and plant genome. Furthermore, enhanced chitinase accumulation was found after 48 and 72 h. Since the chemical carbendazim acts as a common elicitor for all defense enzymes, this change in accumulation of chitinase in the bacteria-treated plants might be due to selective response to fungal chitins. In contrast,  $\beta$  1,3 glucanase levels were found to be very low compared to chitinase levels. Since the fungal cell wall of *R. solani* is made up of chitin alone and no glucan, the plant might have expressed chitinase only (Shrestha et al. 2008). (Leeman et al. 1995) reported the need of a 2 days' time interval between antagonistic application and challenge inoculation for the execution of ISR. However, small amounts of defense enzymes and PR proteins can be found in association with all treatments on healthy uninoculated plants. Shrestha et al. (2008) describe this may be due to the exposure of plants to low levels of stress under most types of growth conditions.

The greenhouse experiment results emphasized the superiority of endophytic microbial application over harmful chemical fungicide carbendazim. The statistically significant results found in this study nominate the replacement of chemical disease control by eco-friendly and cost-effective use of potential endophytic diazotroph *L. sphaericus*. All the treatments with *L. sphaericus* in single or combination mode found statistically similar and highest disease suppression by foliar application followed by soil application. This may be because foliar treatment was

given as the nearest application prior to challenge inoculation, resulting in enhanced proliferation and metabolic activities in the leaf cells. There are possibilities for translocation of bacteria from leaf tissues of the nearest aerial part stem. All these might have contributed to effective disease suppression. There are reports on effective biocontrol of *R.solani* by *Bacillus subtilis* as a foliar application (Liu et al. 2009).

Berg and Hallmann (2006) reported that the expression of antifungal mechanisms by endophytic bacteria is strain specific. A number of *Bacillus* sp. have been described as biocontrol agents toward *R. solani*, but the majority of them belong to soil or rhizosphere groups characterized by limited internal colonization. Several strains of the species *Bacillus amyloliquefaciens*, *B. subtilis*, *B. pasteurii*, *B. cereus*, *B. pumilus*, *B. mycoides*, and *B. sphaericus* have been reported to possess significant reductions in the incidence or severity of various diseases on a diversity of hosts in pot and field experiments (Kumar et al. 2011; Choudhary and Johri 2009). *L. sphaericus* which belongs to *Bacillus* sp., formerly known as *B. sphaericus* (Ahmed et al. 2007), are well-known commercial mosquitocidal agents also (Berry 2012). This is the first report on the in vitro and greenhouse biocontrol ability and induced systemic resistance of endophytic diazotroph *L. sphaericus* against *R. solani*. As it is an endophytic spore-forming bacillus and a well-known commercially available pesticidal agent, research on better formulations and field experiments can explore it as a propitious candidate for the eco-friendly management of rice sheath blight and sustainable agriculture.

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#### Compliance with ethical standards

**Conflict of interest** The authors hereby declare that there is no conflict of interest.

#### Appendix: Preparation of colloidal chitin agar

Dissolve 10 g of chitin from shrimp cells (HiMedia) slowly in 400 mL con. HCl by stirring at 4 °C and incubate the mixture in a water bath at 37 °C until viscosity disappears. To this, add 4 L sterile distilled water and leave overnight at 4 °C. Decant the supernatant slowly, collect the precipitate on a filter paper and wash extensively with sterile distilled water to attain a neutral pH. Dissolve the colloidal chitin in 250 mL distilled water prior to use.

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