Agricultural Sciences. In the article "Genes from mycoparasitic fungi as a source for improving plant resistance to fungal pathogens" by Matteo Lorito, Sheridan L. Woo, Irene Garcia Fernandez, Gabriella Colucci, Gary E. Harman, José A. Pintor-Toro, Edgardo Filippone, Simona Muccifora, Christopher B. Lawrence, Astolfo Zoina, Sadik Tuzun, and Felice Scala, which appeared in number 14, July 7, 1998, of *Proc. Natl. Acad. Sci. USA* (**95,** 7860–7865), the following correction should be noted. The third author's name was erroneously written as Irene Garcia Fernandez. It should be Irene Garcia.

Immunology. In the article "Differential thymic selection outcomes stimulated by focal structural alteration in peptide/major histocompatibility complex ligands" by Yoseph Ghendler, Mai-kun Teng, Jin-huan Liu, Torsten Witte, Ju Liu, Ki Seok Kim, Petra Kern, Hsiu-Ching Chang, Jia-huai Wang, and Ellis L. Reinherz, which appeared in number 17, August 18, 1998, of *Proc. Natl. Acad. Sci. USA* (**95,** 10061–10066), the following correction should be noted. In Fig. 1*C*, the bars for L4 should be striped instead of black. The figure and legend appear below.

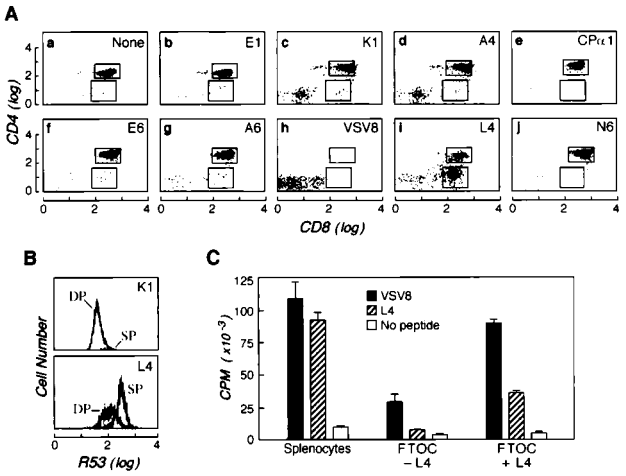

FIG. 1. The L4 peptide induces positive selection of N15tg thymocytes. FTOC was performed by using N15tg/RAG-$2^{-/-}/\beta_2m^{-/-}$ (H-2^b) thymic lobes in media containing 5 μg/ml human β_2m with or without 10 μM of the indicated peptides. After 7 days, thymocytes were released from the lobes by pressing through a steel mesh, counted, and triple-stained with PE-conjugated anti-CD4, Red613-conjugated anti-CD8, and mAb R53 (anti-N15 β chain clonotype) plus FITC-conjugated anti-rat IgG (19). (*A*) The CD8 versus CD4 staining profile of total thymocytes is shown. In this representative experiment the yield of $CD8^+$ SP cells after L4 incubation was 13.5×10^4 cells compared with $0.3–2.2 \times 10^4$ cells in FTOC incubated in the absence of exogenous peptides or in the presence of the other indicated peptides. The total thymocyte number recovered from VSV8-exposed FTOC was significantly lower (10^5 cells per lobe) than from FTOC incubated with any of the other peptides ($4–6 \times 10^5$ cells per lobe). (*B*) The histograms of the N15 TCRβ chain expression on DP and $CD8^+$ SP thymocytes derived from FTOC incubated with K1 or L4 peptides are shown. Note that the K1 histogram represents data similar to that obtained with the other peptides (except VSV8). The $CD8^+$ SP thymocytes that mature on L4 express a higher level of the TCR than the DP thymocytes harvested from the same lobe. (*C*) Thymocytes selected on L4 are functionally responsive to VSV8. Thymocytes from the organ cultures described above [cultured with (+) or without (−) L4] or fresh splenocytes from N15tg/RAG-$2^{-/-}$ mouse were assayed for their proliferative response to irradiated EL-4 cells, in the present of rIL-2 and 10 nM VSV8 or 10 μM L4 or no peptide. After 48 h, each well was pulsed for 18 h with [^{3}H]thymidine, harvested on filter discs, and counted. The proliferative responses for the peptides are shown. Results are mean values of triplicate samples with SD noted.

Medical Sciences. In the article "CREB binding protein is a required coactivator for Smad-dependent, transforming growth factor β transcriptional responses in endothelial cells" by James N. Topper, Maria R. DiChiara, Jonathan D. Brown, Amy J. Williams, Dean Falb, Tucker Collins, and Michael A. Gimbrone, Jr., which appeared in number 16, August 4, 1998, of *Proc. Natl. Acad. Sci. USA* **95,** 9506–9511), the following correction should be noted. The graphics, but not the corresponding legends, of Figs. 4 and 6 have been inadvertantly transposed. The corrected figures and corresponding legends are shown below.

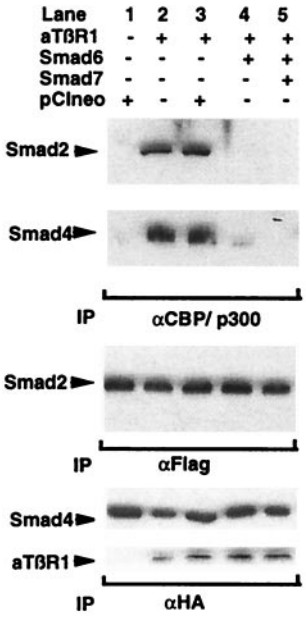

FIG. 4. Smads 2 and 4 can interact with CBP *in vivo*. Cos-7 cells were transfected with the indicated combinations of epitope-tagged activated TBF-β type-1 receptor, Smad expression construct, or empty expression vector. The cells were then lysed and immunoprecipitated with an anti-CBP/P300 antisera. Coprecipitating Smad proteins were detected by Western blot. The upper two panels demonstrate that significant amounts of both Smad2 and Smad4 protein coimmunoprecipitate with CBP/P300 in the presence of the activated receptor. These interactions are inhibited by the simultaneous expression of either Smad6 or Smad7 but not by cotransfection of an empty expression vector (pCIneo). The bottom panels confirm comparable Smad2, Smad4, and activated receptor expression by immunoprecipitation and Western blotting with antisera against the epitopes fused to these proteins.

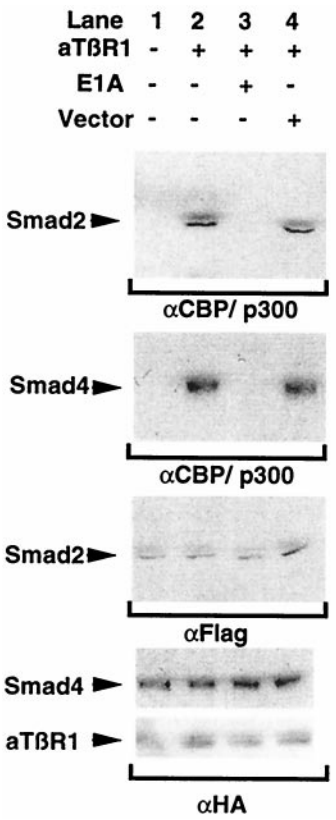

FIG. 6. 12S E1A inhibits the association of activated Smad2 and Smad4 with CBP *in vivo.* Cos-7 cells were transfected with expression constructs expressing epitope-tagged (HA, Flag) versions of Smad2 and Smad4 and the indicated combinations of activated TBF-β type-1 receptor and 12S E1A expression vector. The cells were subsequently lysed and subjected to immunoprecipitation with anti-CBP/p300 antisera, and levels of coimmunoprecipitating Smad proteins were determined by Western blot. The bottom panels confirm comparable Smad2, Smad4, and activated receptor expression.