

# Getting in charge of $\beta$ -synuclein fibrillation

DOI 10.1074/jbc.H117.780528

Meytal Landau<sup>1</sup>

From the Department of Biology, Technion-Israel Institute of Technology, Haifa 3200003, Israel

Edited by Wolfgang Peti

The synuclein family has long been associated with Parkinson's disease and dementia. Although the self-assembly of  $\alpha$ -synuclein ( $\alpha$ S) into oligomers and amyloid fibrils is well established, the aggregation propensity of other members of the family and their role in disease is still under debate. Moriarty *et al.* now suggest that the pH switching that occurs between different cellular environments could control  $\beta$ -synuclein ( $\beta$ S) aggregation via altering its charge distribution, thus opening new possible roles for  $\beta$ S in Parkinson's and other neurodegenerative diseases.

Amyloids are known to be involved in more than 20 fatal diseases. Yet, despite decades of research, there are still many open questions regarding the mechanisms of their formation, their resulting structure, and the basis by which they cause toxicity. This is mostly due to the aggregative and highly polymorphic nature of amyloids that challenges mechanistic and structural investigations. In particular, Parkinson's disease is closely linked to the amyloid-forming protein  $\alpha$ S.<sup>1</sup> However, there are other members of the synuclein family that possibly also contribute to disease progression. In particular,  $\beta$ S shares 62% sequence identity with  $\alpha$ S, and there are known  $\beta$ S mutations associated with dementia (Fig. 1) (1). However, the functional role of  $\beta$ S seems to vary widely from study to study, and many basic details of its structure and activity remain unclear. In their paper, Moriarty *et al.* (2) now demonstrate a possible explanation for these diverse outcomes, showing that  $\beta$ S fibrillation is regulated by a pH switch, suggesting new fibrillation mechanisms and a more complex *in vivo* role than formerly anticipated.

Previous studies have demonstrated that both  $\alpha$ S and  $\beta$ S are composed of three domains: an N-terminal amphipathic domain, a central hydrophobic core domain known as the non-amyloid component (NAC), originally reported to be deposited with amyloid- $\beta$  in the brains of Alzheimer's disease patients, and an acidic C-terminal domain (Fig. 1). The NAC domain, and in particular a central hydrophobic segment within the domain, constitutes the critical determinant of  $\alpha$ S fibrillation and toxicity (3). This segment is missing in  $\beta$ S, and as a result,

$\beta$ S was anticipated to lack aggregation propensity. Indeed,  $\beta$ S was suggested to inhibit  $\alpha$ S fibrillation and ameliorate disease (4). However, in other studies, fibrillation of  $\beta$ S has been observed and associated with neurodegeneration (5), and the presence of external substances such as environmental pollutants enhanced  $\beta$ S aggregation (6).

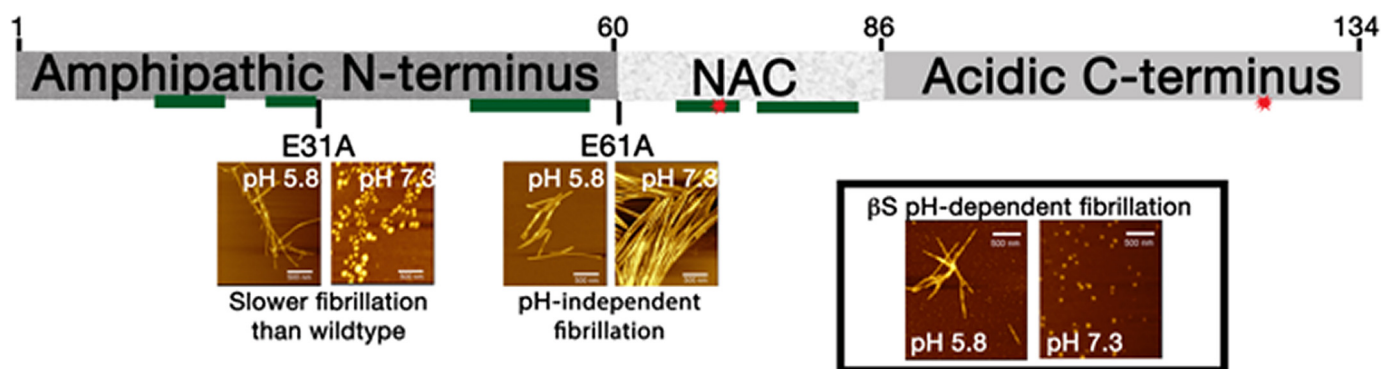
Given that synucleins are found in several different cellular compartments where the pH varies and that previous studies showed impacts of changing pH on amyloid protein fibrillation rates, Moriarty *et al.* (2) tested this effect and found that  $\beta$ S formed fibrils at a slightly acidic pH found along the endosomal pathway (pH 5.8), but not at pH 7.3. The importance of the different domains of  $\beta$ S in fibrillation and pH regulation was investigated by constructing eight chimeras, in which the three domains of  $\alpha$ S and  $\beta$ S were swapped. Fibril formation and kinetics were monitored via atomic-force microscopy (AFM) and binding to the amyloid-indicator dye thioflavin T. The chimeras revealed that all three domains regulate fibrillation of  $\beta$ S, although in markedly different ways. An inhibitory effect on fibrillation was attributed to the C-terminal domain, which has the lowest sequence identity (~30%) to  $\alpha$ S. In contrast, the N-terminal domain appeared critical for  $\beta$ S fibrillation. Swapping  $\beta$ S and  $\alpha$ S N-terminal domains (90% sequence identity) induced a relatively small effect on fibrillation rate in most chimeras. However, the chimera containing the N-terminal domain of  $\alpha$ S with the NAC and C-terminal domains of  $\beta$ S showed no fibrillation, even at acidic pH, which may indicate interdependent relations between the different domains. The role of the N-terminal domain in regulating fibrillation was also supported by computational modeling in which  $\beta$ S was threaded onto the solid-state NMR-based structure of  $\alpha$ S (7) and simulated using Rosetta Symmetry (8). The simulations suggested that the N-terminal domain is imperative for the formation and stability of a Greek-key-like architecture observed for  $\alpha$ S (2). NMR analysis of the  $\beta$ S monomer also suggested interactions between the N-terminal and NAC domains. Moreover, a glutamate to alanine alteration at position 31 (E31A) in the N-terminal domain delayed fibrillation of  $\beta$ S (Fig. 1), supporting the involvement of this domain in aggregation (2).

What is the role of the NAC domain? The chimeras constructed by Moriarty *et al.* reinforced the critical role of this domain in  $\alpha$ S fibrillation regardless of the tested pH and the identity of the other domains. Moreover, the chimeras containing the NAC domain of  $\alpha$ S exhibited faster fibrillation than all their equivalent chimeras containing the NAC domain of  $\beta$ S. Importantly, the chimeras that included the NAC domain of  $\beta$ S fibrillated only at acidic pH, suggesting that a pH switch is likely dependent on this domain. Accordingly, mutation of the first

The author declares that she has no conflicts of interest with the contents of this article.

<sup>1</sup> Supported by the I-CORE Program of the Planning and Budgeting Committee, the Israel Science Foundation, Center of Excellence in Integrated Structural Cell Biology Grant 1775/12, and Israel Science Foundation Grant 560/16. To whom correspondence should be addressed. E-mail: mlandau@technion.ac.il.

<sup>1</sup> The abbreviations used are:  $\alpha$ S,  $\alpha$ -synuclein;  $\beta$ S,  $\beta$ -synuclein;  $\gamma$ S,  $\gamma$ -synuclein; NAC, non-amyloid component; AFM, atomic force microscopy.



**Figure 1. The pH-controlled fibrillation of  $\beta$ -synuclein.** The three domains of  $\beta$ S are indicated. The approximate location of two mutations associated with dementia (P123H and V70M) (1) are indicated with red stars. Segments predicted to be amyloidogenic according to the 3D-profile method (10) are indicated by green rectangles below the domains. AFM images shown for wild-type  $\beta$ S (boxed on the right) and its mutants are taken from Ref. 2.

residue at the NAC domain, which is occupied by a glutamate residue in both  $\alpha$ S and  $\beta$ S, into an alanine (E61A) abolished the pH switch and resulted in rapid and robust fibrillation of  $\beta$ S at both neutral and acidic pH. Because  $\beta$ S lacks the critical hydrophobic stretch that is present in the NAC domain of  $\alpha$ S (3), fibrillation might be nucleated via alternative core segments, either in the NAC or N-terminal domains. Interestingly, an amyloid-like structure was determined for a 10-residue segment from the N-terminal domain of  $\alpha$ S that included the A53T mutation observed in families with inherited forms of Parkinson's disease (9). The equivalent segment in  $\beta$ S,  $^{47}$ GVVQGVASVA $^{56}$ , is predicted to be highly amyloidogenic according to the 3D-profile method (Fig. 1) (10), with a higher amyloid-forming propensity score than that of the equivalent  $\alpha$ S wild-type segment. It puts forward the possible involvement of this region in nucleating fibrillation of  $\beta$ S.

The results of Moriarty *et al.* suggest a possible biological and pathophysiological role of  $\beta$ S fibrils in specific microenvironments and inside organelles. The authors postulated that the pH switch stems from rearrangement of charge distribution involving altered protonation of side chains that regulate fibrillation propensities via "polar zippers." It is striking that residue Glu-61, which appears to be critical for controlling the pH switch, is conserved in  $\alpha$ S, which already fibrillates at both pH values. This highlights the complex regulation of these two synuclein homologs and supports the possibility of different nucleation regions. Regardless of whether the Greek-key-like architecture of  $\alpha$ S is preserved in  $\beta$ S, it is very likely that the regions forming  $\beta$ -sheet interactions differ between  $\alpha$ S and  $\beta$ S. Thus, elucidating the high-resolution structure of the  $\beta$ S fibril core will provide important information into how this polymorphism may occur. What about  $\gamma$ -synuclein ( $\gamma$ S), another member of the family? Fibrillation should be expected because it shows high sequence conservation between the N-terminal and NAC domains with  $\alpha$ S (67% sequence identity) but has a significantly shorter and variable C-terminal domain. Yet, previous results show lack of fibrillation (4), which might be related to the microenvironment tested, as shown for  $\beta$ S, or to a critical inhibitory role of the short C-terminal domain. Overall, this manuscript further demonstrates the complexity of the

amyloid fold and its vast polymorphism. It will be intriguing to figure out how this complex regulation of fibrillation evolved in a specific protein family and whether this was a dedicated process or a mishap leading to human diseases.

## References

- Ohtake, H., Limprasert, P., Fan, Y., Onodera, O., Kakita, A., Takahashi, H., Bonner, L. T., Tsuang, D. W., Murray, I. V., Lee, V. M., Trojanowski, J. Q., Ishikawa, A., Idezuka, J., Murata, M., Toda, T., Bird, T. D., Leverenz, J. B., Tsuji, S., and La Spada, A. R. (2004) Beta-synuclein gene alterations in dementia with Lewy bodies. *Neurology* **63**, 805–811
- Moriarty, G. M., Olson, M. P., Atieh, T. B., Janowska, M. K., Khare, S. D., and Baum, J. (2017) A pH-dependent switch promotes  $\beta$ -synuclein fibril formation via glutamate residues. *J. Biol. Chem.* **292**, 16368–16379
- Giasson, B. I., Murray, I. V., Trojanowski, J. Q., and Lee, V. M. (2001) A hydrophobic stretch of 12 amino acid residues in the middle of  $\alpha$ -synuclein is essential for filament assembly. *J. Biol. Chem.* **276**, 2380–2386
- Uversky, V. N., Li, J., Souillac, P., Millett, I. S., Doniach, S., Jakes, R., Goedert, M., and Fink, A. L. (2002) Biophysical properties of the synucleins and their propensities to fibrillate: Inhibition of  $\alpha$ -synuclein assembly by  $\beta$ - and  $\gamma$ -synucleins. *J. Biol. Chem.* **277**, 11970–11978
- Taschenberger, G., Toloe, J., Tereshchenko, J., Akerboom, J., Wales, P., Benz, R., Becker, S., Outeiro, T. F., Looger, L. L., Bähr, M., Zweckstetter, M., and Kügler, S. (2013)  $\beta$ -Synuclein aggregates and induces neurodegeneration in dopaminergic neurons. *Ann. Neurol.* **74**, 109–118
- Yamin, G., Munishkina, L. A., Karymov, M. A., Lyubchenko, Y. L., Uversky, V. N., and Fink, A. L. (2005) Forcing nonamyloidogenic  $\beta$ -synuclein to fibrillate. *Biochemistry* **44**, 9096–9107
- Tuttle, M. D., Comellas, G., Nieuwkoop, A. J., Covell, D. J., Berthold, D. A., Kloepper, K. D., Courtney, J. M., Kim, J. K., Barclay, A. M., Kendall, A., Wan, W., Stubbs, G., Schwieters, C. D., Lee, V. M. Y., George, J. M., and Rienstra, C. M. (2016) Solid-state NMR structure of a pathogenic fibril of full-length human  $\alpha$ -synuclein. *Nat. Struct. Mol. Biol.* **23**, 409–415
- DiMaio, F., Leaver-Fay, A., Bradley, P., Baker, D., and André, I. (2011) Modeling symmetric macromolecular structures in Rosetta3. *PLoS One* **6**, e20450
- Rodriguez, J. A., Ivanova, M. I., Sawaya, M. R., Cascio, D., Reyes, F. E., Shi, D., Sangwan, S., Guenther, E. L., Johnson, L. M., Zhang, M., Jiang, L., Arbing, M. A., Nannenga, B. L., Hattne, J., Whitelegge, J., Brewster, A. S., Messerschmidt, M., Boutet, S., Sauter, N. K., Gonen, T., and Eisenberg, D. S. (2015) Structure of the toxic core of  $\alpha$ -synuclein from invisible crystals. *Nature* **525**, 486–490
- Goldschmidt, L., Teng, P. K., Riek, R., and Eisenberg, D. (2010) Identifying the amyloids, proteins capable of forming amyloid-like fibrils. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 3487–3492