



Determination of action potential wavelength restitution in *Scn5a*^{+/-} mouse hearts modelling human Brugada syndrome

Gary Tse^{1,*}, Sheung Ting Wong², Vivian Tse³, Jie Ming Yeo²

¹School of Biomedical Sciences, Li Ka Shing Faculty of Medicine, University of Hong Kong, Hong Kong, China

²Faculty of Medicine, Imperial College, London, UK

³Department of Physiology, McGill University, Canada

J Geriatr Cardiol 2017; 14: 595–596. doi:10.11909/j.issn.1671-5411.2017.09.011

Keywords: Action potential duration; Conduction; Depolarization; Restitution; Repolarization; Wavelength

Brugada syndrome is a primary electrical disorder of the heart, predisposing affected individuals to potentially lethal, ventricular tachy-arrhythmias.^[1–7] A number of mechanisms have been identified as being important increasing the risk of these rhythms.^[8] Wavelength (λ) restitution has been suggested to predict the onset of action potential duration (APD) alternans in mouse *Scn5a*^{+/-} hearts modelling Brugada syndrome.^[9] Classical APD restitution analysis yielded mixed success in its ability to predict the onset of APD alternans and arrhythmogenicity. APD restitution relates APD to the previous diastolic interval (DI). APD restitution gradients > 1 is associated with the emergence of APD alternans,^[10] and increased arrhythmogenicity in a number of different genetic and pharmacological mouse models, such as Brugada syndrome, long QT syndrome type 3 and hypokalaemia.^[11–13] Matthews and colleagues previously demonstrated a non-linear relationship between APD alternans and APD restitution gradient and underestimated the extent of APD alternans, suggesting that it may partly underlie its lack of success in predicting arrhythmogenicity.^[14] Another reason is that effective refractory period (ERP) can be altered independently of APD.^[15]

The lack of predictive power of APD restitution led Matthews and colleagues to devise a novel λ restitution analysis by recording monophasic action potential (MAP) recordings in wild-type and *Scn5a*^{+/-} hearts during dynamic pacing, which introduced a stepwise increase in pacing rate.^[9] The MAP method is an *ex vivo* recording technique that has widely been used to study whole heart electrophysiology in Langendorff systems. Activation latencies and APDs were derived from the MAPs obtained from the ventricles, with conduction velocity (θ) approximated by the reciprocal of activation latency, θ^{-1} . This in turn enabled the

calculation of λ^{-1} , which was approximated by $\theta^{-1} \times \text{APD}$, with the explicit assumption that ERP was equal to APD.

Whilst we do not doubt the important role of wavelength in determining arrhythmogenicity, the method chosen by Matthews and colleagues may not be accurate in estimating wavelength for the following reasons. Firstly, the discordance between APD and ERP are apparent from the data generated by the authors' own group, but this has not been highlighted. Specifically, Martin and colleagues showed that APD is longer than ERP in wild-type hearts, whereas it is shorter than ERP *Scn5a*^{+/-} hearts.^[16]

Secondly, estimation of wavelength using the authors' method requires accurate measurements of APD. Yet, the group's data on APD values have been highly discrepant, as can be seen in their own studies on Brugada syndrome.^[17,18] For example, in the left ventricular (LV) epicardium, APD₇₀ and APD₅₀ were not significantly altered by quinidine.^[17] However, the authors later found that these were increased by quinidine.^[18] In the LV endocardium, APD₉₀, APD₇₀ and APD₅₀ were decreased by quinidine.^[17] Their later study found that these were increased by quinidine.^[18] Given these discrepancies, λ did not appear to be accurately determined.

Together, the current evidence clearly shows that λ reduction,^[19] and increased APD restitution^[20] are important mechanism by which cardiac arrhythmias are generated and maintained. However, the role of λ restitution is unclear, but more accurate methods of determining this parameter experimentally need to be devised before a more definite conclusion can be reached.

References

- 1 Tse G, Lai TH, Yeo JM, *et al.* Mechanisms of electrical activation and conduction in the gastrointestinal system: lessons from cardiac electrophysiology. *Front Physiol* 2016; 7: 182.

*Correspondence to: gary.tse@doctors.org.uk

- 2 Tse G, Lai ET, Tse V, Yeo JM. Molecular and electrophysiological mechanisms underlying cardiac arrhythmogenesis in diabetes mellitus. *J Diabetes Res* 2016; 2848759.
- 3 Tse G, Lai ET, Yeo JM, Yan BP. Electrophysiological mechanisms of Bayés syndrome: insights from clinical and mouse studies. *Front Physiol* 2016; 7: 188.
- 4 Tse G, Lai ETH, Yeo JM, *et al.* Mechanisms of electrical activation and conduction in the gastrointestinal system: lessons from cardiac electrophysiology. *Front Physiol* 2016; 7.
- 5 Tse G, Lai ET, Lee AP, *et al.* Electrophysiological mechanisms of gastrointestinal arrhythmogenesis: lessons from the heart. *Front Physiol* 2016; 7: 230.
- 6 Tse G, Yeo JM. Conduction abnormalities and ventricular arrhythmogenesis: The roles of sodium channels and gap junctions. *Int J Cardiol Heart Vasc* 2015; 9: 75–82.
- 7 Tse G. Mechanisms of Cardiac Arrhythmias. *J Arrhythm* 2015; 32: 75–81.
- 8 Tse G. Novel conduction-repolarization indices for the stratification of arrhythmic risk. *J Geriatr Cardiol* 2016; 13: 811–812.
- 9 Matthews GD, Guzadhur L, Sabir IN, *et al.* Action potential wavelength restitution predicts alternans and arrhythmia in murine *Scn5a*(+/-) hearts. *J Physiol* 2013; 591: 4167–4188.
- 10 Nolasco JB, Dahlen RW. A graphic method for the study of alternation in cardiac action potentials. *J Appl Physiol* 1968; 25: 191–196.
- 11 Sabir IN, Li LM, Grace AA, Huang CL. Restitution analysis of alternans and its relationship to arrhythmogenicity in hypokalaemic Langendorff-perfused murine hearts. *Pflugers Arch* 2008; 455: 653–666.
- 12 Hothi SS, Booth SW, Sabir IN, *et al.* Arrhythmogenic substrate and its modification by nicorandil in a murine model of long QT type 3 syndrome. *Prog Biophys Mol Biol* 2008; 98: 267–280.
- 13 Tse G, Wong ST, Tse V, Yeo JM. Restitution analysis of alternans using dynamic pacing and its comparison with S1S2 restitution in heptanol-treated, hypokalaemic Langendorff-perfused mouse hearts. *Biomed Rep* 2016; 4: 673–680.
- 14 Matthews GD, Guzadhur L, Grace A, Huang CL. Nonlinearity between action potential alternans and restitution, which both predict ventricular arrhythmic properties in *Scn5a*+/- and wild-type murine hearts. *J Appl Physiol* 2012; 112: 1847–1863.
- 15 Tse G, Tse V, Yeo JM. Ventricular anti-arrhythmic effects of heptanol in hypokalaemic, Langendorff-perfused mouse hearts. *Biomed Rep* 2016; 4: 313–324.
- 16 Martin CA, Grace AA, Huang CLH. Refractory dispersion promotes conduction disturbance and arrhythmias in a *Scn5a*(+/-) mouse model. *Pflugers Archiv* 2011; 462: 495–504.
- 17 Stokoe KS, Balasubramaniam R, Goddard CA, *et al.* Effects of flecainide and quinidine on arrhythmogenic properties of *Scn5a*+/- murine hearts modelling the Brugada syndrome. *J Physiol* 2007; 581: 255–275.
- 18 Martin CA, Zhang Y, Grace AA, Huang CL. Increased Right Ventricular Repolarization Gradients Promote Arrhythmogenesis in a Murine Model of Brugada Syndrome. *J Cardiovasc Electrophysiol* 2010; 21: 1153–1159.
- 19 Wilde AA, Postema PG, Di Diego JM, *et al.* The pathophysiological mechanism underlying Brugada syndrome: depolarization versus repolarization. *J Mol Cell Cardiol* 2010; 49: 543–553.
- 20 Tse G, Wong STT, Lee YT, *et al.* Cardiac dynamics: alternans and arrhythmogenesis. *J Arrhythm* 2016; 32: 411–417.