

Targeting MUC1-C suppresses polycomb repressive complex 1 in multiple myeloma

Ashujit Tagde^{1,*}, Tahireh Markert^{1,*}, Hasan Rajabi¹, Masayuki Hiraki¹, Maroof Alam¹, Audrey Bouillez¹, David Avigan¹, Kenneth Anderson¹ and Donald Kufe¹

¹ Dana-Farber Cancer Institute, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA

* These authors have contributed equally to this work

Correspondence to: Donald Kufe, email: donald_kufe@dfci.harvard.edu

Keywords: MUC1-C, multiple myeloma, BMI1, RING1, RING2

Received: June 30, 2017

Accepted: July 16, 2017

Published: August 10, 2017

Copyright: Tagde et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

The polycomb repressive complex 1 (PRC1) includes the BMI1, RING1 and RING2 proteins. BMI1 is required for survival of multiple myeloma (MM) cells. The MUC1-C oncoprotein is aberrantly expressed by MM cells, activates MYC and is also necessary for MM cell survival. The present studies show that targeting MUC1-C with (i) stable and inducible silencing and CRISPR/Cas9 editing and (ii) the pharmacologic inhibitor GO-203, which blocks MUC1-C function, downregulates BMI1, RING1 and RING2 expression. The results demonstrate that MUC1-C drives BMI1 transcription by a MYC-dependent mechanism. MUC1-C thus promotes MYC occupancy on the BMI1 promoter and thereby activates BMI1 expression. We also show that the MUC1-C→MYC pathway induces RING2 expression. Moreover, in contrast to BMI1 and RING2, we found that MUC1-C drives RING1 by an NF-κB p65-dependent mechanism. Targeting MUC1-C and thereby the suppression of these key PRC1 proteins was associated with downregulation of the PRC1 E3 ligase activity as evidenced by decreases in ubiquitylation of histone H2A. Targeting MUC1-C also resulted in activation of the PRC1-repressed tumor suppressor genes, PTEN, CDNK2A and BIM. These findings identify a heretofore unrecognized role for MUC1-C in the epigenetic regulation of MM cells.

INTRODUCTION

Mucin 1 (MUC1) is a cell membrane heterodimeric complex that is aberrantly expressed in primary multiple myeloma (MM) cells and MM cell lines [1-5]. The transmembrane MUC1 C-terminal (MUC1-C) subunit of the heterodimer functions as an oncoprotein that is necessary for the proliferation and survival of MM cells [5-8]. In this respect, the MUC1-C cytoplasmic domain is an intrinsically disordered 72-amino acid structure that has the plasticity to act as a node and intersect with multiple signaling pathways linked to self-renewal, inflammation and transformation [9, 10]. MUC1-C activates the proinflammatory IKK → NF-κB pathway, binds directly to NF-κB p65 and promotes the induction of NF-κB target

genes, including MUC1 itself in an autoinductive loop [11-13]. The MUC1-C cytoplasmic domain also binds directly to β-catenin, inhibits β-catenin degradation and activates the WNT/β-catenin/TCF4 pathway [14, 15]. Studies in MM cells have demonstrated that MUC1-C increases occupancy of β-catenin on the MYC promoter and drives MYC transcription [8]. Moreover, analysis of microarray datasets from primary MM cells showed that MUC1 expression positively correlates with that of MYC [8]. By extension, silencing MUC1-C in MM cells results in downregulation of MYC and thereby MYC target genes [8]. MM cells are addicted to MYC [16-18]. Thus, targeting MUC1-C with the downregulation of MYC explains, at least in part, why MM cells are dependent on MUC1-C for their proliferation and survival

[5-8]. Of potential importance for targeting MUC1-C as a treatment for MM, the MUC1-C cytoplasmic domain includes a CQC motif that is essential for MUC1-C homodimerization, nuclear localization and function [6-8]. For these reasons, a cell-penetrating peptide, designated GO-203, has been developed that targets the MUC1-C CQC motif, inhibits MUC1-C homodimerization, nuclear import and function, and is effective in inducing MM cell death [6-8].

The polycomb repressive complex 1 (PRC1) includes the ring domain-containing BMI1, RING1 and RING2 proteins [19, 20]. BMI1 and RING1 bind to the catalytic RING2 subunit, and both contribute to the RING2 ubiquitin E3 ligase function [21]. BMI1 is also necessary for maintaining integrity of the complex [22]. In the prevailing hierarchical model, PRC1 is recruited to sites of H3K27 trimethylation (H3K27me3) generated by the polycomb repressive complex 2 (PRC2) [23], which includes enhancer of zeste homolog 2 (EZH2) and suppressor of zeste 12 homolog (SUZ12) components [24]. In turn, PRC1 catalyzes the mono-ubiquitination of histone H2A on K119 and promotes repression of *homeobox (HOX)* genes, among others [19-22, 25]. Of the PRC1 subunits, BMI1 has been linked to the self-renewal of normal stem cells and the tumorigenic potential of cancer stem-like cells (CSCs) [26-31]. BMI1 contributes to self-renewal and stemness by repressing the *CDNK2A* locus, which encodes the p16^{INK4a} and p14^{ARF} tumor suppressors [26, 28]. In carcinoma cells, BMI1 has also been linked to downregulation of the PTEN tumor suppressor [28, 32, 33]. Additionally, in MM cells, BMI1 suppresses the expression of multiple proapoptotic proteins, including BIM, and in this way is essential for MM self-renewal [34]. BMI1 also activates the WNT pathway by repressing the Dickkopf (DKK) family of WNT inhibitors [35]. Repression of DKK proteins contributes to activation of the *MYC* gene and thereby a positive-feedback loop linking the WNT pathway to induction of *BMI1* expression [35]. BMI1 is thus a potentially important target for the treatment of MM; however, to date, there are no clinically available BMI1 inhibitors [29]. Therefore, targeting upstream effectors that drive expression of BMI1 and other PRC1 components represents an attractive approach for reprogramming PRC1-mediated gene repression in MM.

The present studies demonstrate that MUC1-C drives expression of BMI1, RING1 and RING2 in MM cells. We show that MUC1-C activates the *BMI1* promoter by a MYC-mediated mechanism. In addition, we report that MUC1-C induces (i) RING2 by MYC-dependent signaling, and (ii) RING1 by activation of the NF- κ B p65 pathway. In concert with these findings, targeting MUC1-C results in suppression of H2A ubiquitylation and induction of the PTEN, p14^{ARF} and BIM tumor suppressors in MM cells.

RESULTS

MUC1-C induces BMI1 expression in MM cells

MUC1-C was stably silenced in MM cells to investigate whether this oncoprotein is involved in the regulation of BMI1 expression. In studies of RPMI8226 cells, MUC1-C silencing was associated with substantial downregulation of BMI1 mRNA and protein (Figure 1A, left and right). Similarly, silencing MUC1-C in OPM-2, KMS-12-E and U266 cells resulted in marked decreases in BMI1 expression (Figure 1B, left and right; Supplementary Figures 1A and 1B). In extending these observations to primary MM cells from 2 patients with refractory disease, we also found that targeting MUC1-C significantly reduces BMI1 levels (Figure 1C, left and right; Supplementary Figure 1C). To confirm these results, we stably transduced RPMI8226 cells to express a control Tet-CshRNA or a Tet-MUC1shRNA. DOX treatment of RPMI8226/Tet-MUC1shRNA, but not RPMI8226/Tet-CshRNA, cells was associated with significant downregulation of BMI1 mRNA and protein (Figure 1D, left and right). In concert with these results, the enforced overexpression of MUC1-C in OPM-2 cells was associated with an increase in BMI1 expression (Figure 1E, left and right), clearly indicating that MUC1-C and not the shed MUC1-N subunit is responsible for driving BMI1 expression.

The MUC1-C subunit consists of a 72-amino acid (aa) cytoplasmic domain with a CQC motif that is necessary and sufficient for the formation of MUC1-C homodimers and for MUC1-C-mediated transformation (Figure 2A). Accordingly, the cell-penetrating peptide inhibitor, GO-203, was developed to block the MUC1-C CQC motif in MM and other cancer cells (Figure 2A). Notably, treatment of RPMI8226 and OPM-2 cells with GO-203, but not the control peptide CP-2, induced a marked decrease in BMI1 mRNA and protein (Figure 2B and 2C, left and right). Downregulation of BMI1 expression was also observed in studies of GO-203-treated KMS-12-PE and NCIH929 cells (Supplementary Figure 2A and 2B). Furthermore, targeting MUC1-C with CRISPR/Cas9 gene editing was associated with a marked reduction in the BMI1 expression (Figure 2D, left and right; Supplementary Figure 2C). These findings collectively indicated that MUC1-C activates BMI1 expression in MM cells.

MUC1-C drives *BMI1* transcription by a MYC-dependent mechanism

MUC1-C induces *MYC* expression in MM cells by activating the WNT/ β -catenin/TCF-4 pathway [8]. The *BMI1* promoter contains a consensus E-Box (CACGTG)

at position -177 to -182 upstream to the transcription start site (Figure 3A), invoking the possibility that MUC1-C induces BMI1 expression through the MYC pathway in MM cells. To address this potential mechanism, we assessed the effects of targeting MYC on BMI1 expression. RPMI8226 cells were thus transduced to stably express a Tet-inducible MYC shRNA (Tet-MYCshRNA) or a Tet-inducible control shRNA (Tet-CshRNA). Treatment of RPMI8226/Tet-MYCshRNA cells with DOX resulted in the marked suppression of MYC and BMI1 levels as compared to that in DOX-treated RPMI8226/Tet-CshRNA cells (Figure 3B, left, middle and right). These results were confirmed in DOX-treated OPM-2/Tet-CshRNA and OPM-2/Tet-MYCshRNA cells (Figure 3C, left, middle and right). In extending these observations, we found that treatment of RPMI8226 cells with the BET bromodomain inhibitor, JQ1, was associated with the marked suppression of MYC, MUC1-C and BMI1 expression (Figure 3D, left, middle and right). Similar results were obtained in studies

of JQ1-treated OPM-2 cells (Figure 3E, left, middle and right), supporting the notion that the MUC1-C → MYC pathway induces BMI1 expression in MM cells.

MUC1-C activates the *BMI1* promoter by enhancing MYC occupancy

To determine whether MUC1-C drives *BMI1* transcription by a MYC-mediated mechanism, we transfected MM cells to express a pBMI1-Luc reporter that includes the MYC binding site (Figure 4A). Using this approach, we found that activation of pBMI1-Luc is decreased in RPMI8226/MUC1shRNA, as compared to RPMI8226/CshRNA, cells (Figure 4B). Silencing MUC1-C in OPM-2 cells was also associated with suppression of pBMI1-Luc activity (Figure 4C). Mutation of the MYC binding site (CACGTG→CGCGTG) in Mut-pBMI1-Luc (Figure 4A) resulted in downregulation of

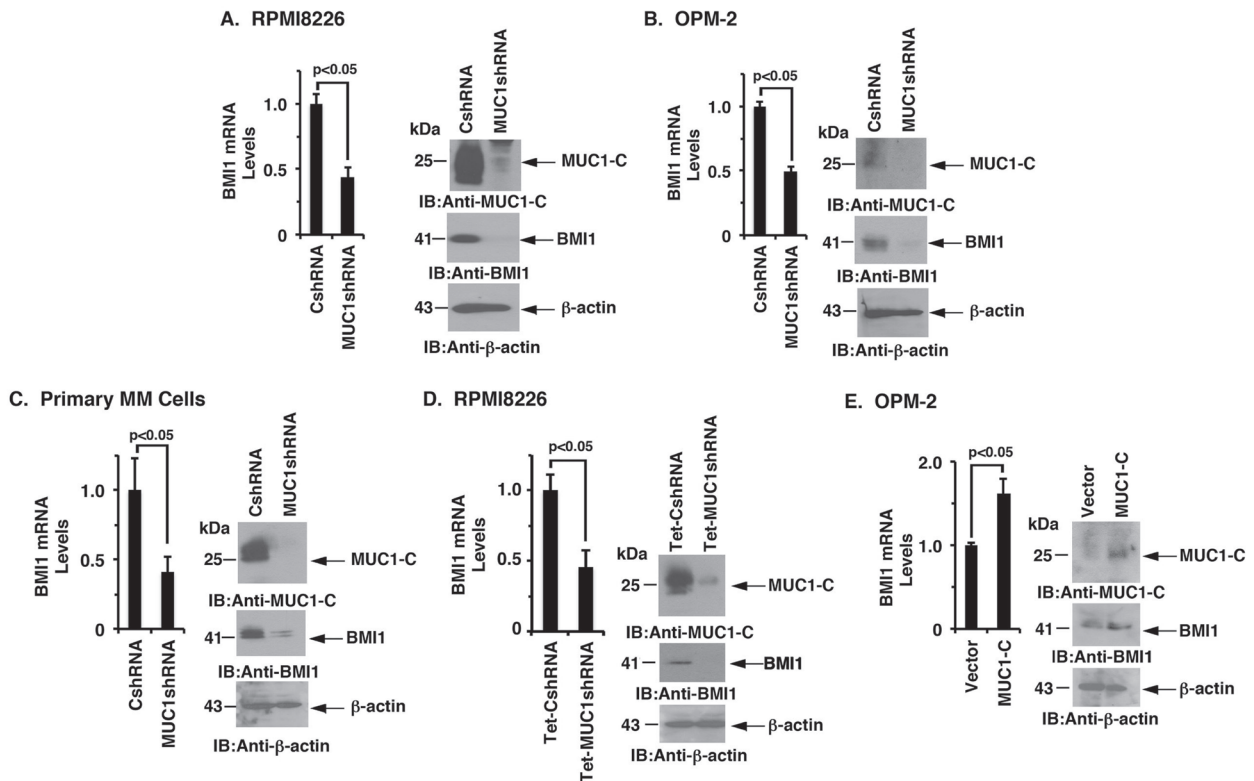


Figure 1: Targeting MUC1-C downregulates BMI1 expression. A.-C. RPMI8226 (A), OPM-2 (B), and primary MM (C) cells from Patient #1 stably expressing a control shRNA (CshRNA) or a MUC1 shRNA were analyzed for BMI1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). **D.** RPMI8226 cells were stably transduced to express a Tetracycline-inducible control shRNA (Tet-CshRNA) or a MUC1shRNA (Tet-MUC1shRNA). Cells treated with 200 ng/ml DOX for 5 d were analyzed for BMI1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels compared with that obtained for control DOX-treated Tet-CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). **E.** OPM-2 cells transiently transduced to express an empty vector or one expressing MUC1-C were analyzed for BMI1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the vector cells (assigned a value of 1). Lysates from were immunoblotted with the indicated antibodies (right).

reporter activity (Figure 4D; Supplementary Figure 3A). Moreover, JQ1 treatment of RPMI8226 and OPM-2 cells significantly reduced activation of the pBMI1-Luc reporter (Figure 4E; Supplementary Figure S3B). In concert with involvement of the MUC1-C → MYC pathway, ChIP analysis of the *BMI1* promoter in RPMI8226 (Figure 4F) and OPM-2 (Supplementary Figure 3C) cells demonstrated that silencing MUC1-C is associated with decreases in MYC occupancy.

MUC1-C induces RING1 and RING2 expression

PRC1 catalytic activity is dependent on BMI1, RING1 and RING2 [19-21]. Interestingly and as found for BMI1, targeting MUC1-C significantly reduced RING1 and RING2 levels in RPMI8226 (Figure 5A) and OPM-2 (Figure 5B), KMS-12-PE (Supplementary Figure 4A, left and right), U266 (Supplementary Figure 4B, left and right) and NCIH929 (Supplementary Figure S4C) cells. Targeting MUC1-C with (i) DOX-inducible silencing

(Figure 5C), (ii) CRISPR/Cas9 gene editing (Figure 5D), and (iii) pharmacologic inhibition with GO-203 (Figure 5E) was also associated with the downregulation of both RING1 and RING2. Additionally, overexpression of MUC1-C in OPM-2 cells was associated with upregulation of RING1 and RING2 expression (Supplementary Figure 4D, indicating that, like BMI1, MUC1-C drives RING1 and RING2 expression.

MUC1-C differentially activates RING1 and RING2

Notably, little is known about the regulation of RING1 and RING2. However, based on the identification of a MUC1-C → MYC → BIM1 pathway, we asked if MUC1-C induces the expression of RING1 and RING2 by a similar mechanism. In this context, inhibiting MYC with JQ1 was associated with marked downregulation of RING2 and a modest decrease in RING1 expression in RPMI8226 (Figure 6A, left and right) and OPM-

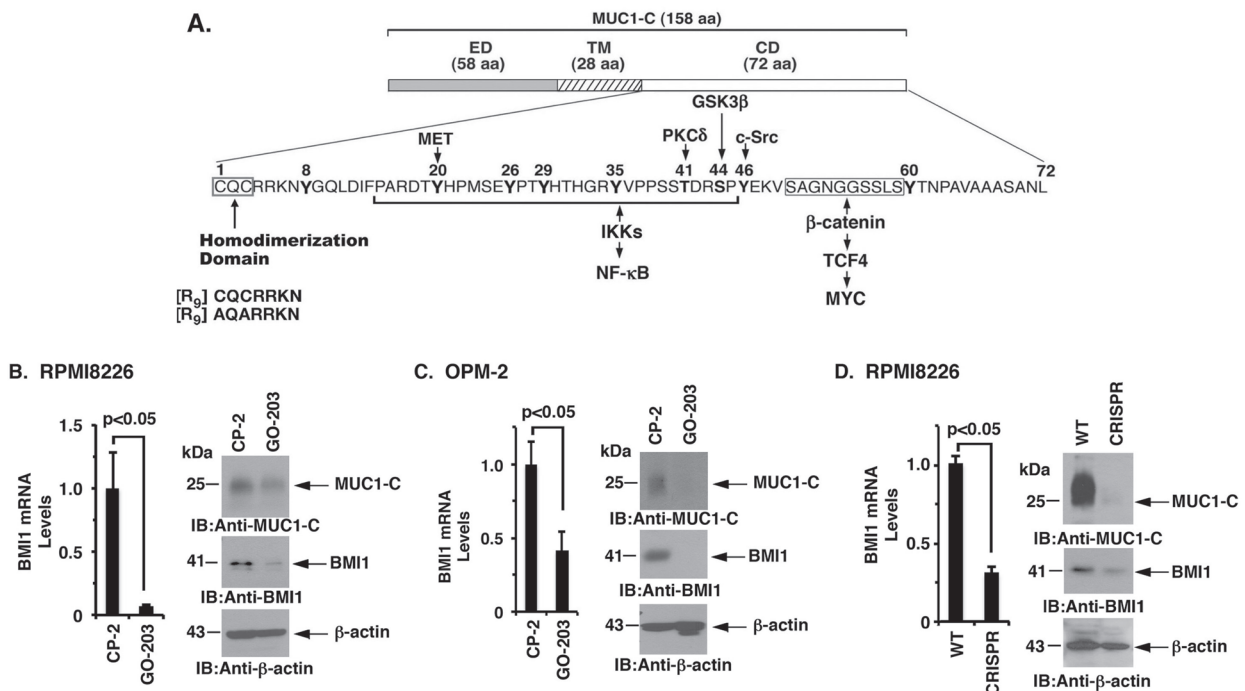


Figure 2: Targeting the MUC1-C cytoplasmic domain downregulates BMI1 expression. A. Schema of the MUC1-C subunit with the 58-aa extracellular domain (ED), the 28-aa transmembrane domain (TM) and the 72-aa cytoplasmic domain (CD). Highlighted is the CQC motif, which is necessary and sufficient for MUC1-C homodimerization, and is targeted by GO-203 and not the control peptide CP-2. Also highlighted are interactions of the MUC1-C cytoplasmic domain with effectors of the NF-κB and MYC pathways. B.-C. RPMI8226 (B) and OPM-2 (C) cells were treated with 5 μM CP-2 or GO-203 for 48 h. The cells were analyzed for BMI1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the CP-2-treated cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). D. RPMI8226 cells were silenced for MUC1 using CRISPR/Cas9 gene editing. The parental WT and CRISPR cells were analyzed for BMI1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the WT cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right).

2 (Supplementary Figure 5A) cells. DOX-inducible silencing of MYC in RPMI8226 (Figure 6B, left and right) and OPM-2 (Supplementary Figure 5B) cells also decreased RING2, and to a lesser extent RING1, mRNA and protein levels. Notably, the *RING2*, but not the *RING1*, promoter contains putative E-boxes for potential MYC binding (Figure 6C). Moreover and in support of these findings, targeting MUC1-C decreased MYC occupancy on the *RING2* promoter (Figure 6C, left and right). MUC1-C also activates NF- κ B p65 in association with epigenetic regulation [36]. In this regard, silencing NF- κ B p65 in RPMI8226 (Figure 6D, left and right) and OPM-2 (Supplementary Figure 5C) cells was associated with significant decreases in RING1, but not RING2,

mRNA and protein. Moreover, BAY-11-7085 treatment of RPMI8226 (Figure 6E, left and right) and OPM-2 (Supplementary Figure 5D) cells suppressed RING1 and not RING2 expression. Furthermore, analysis of *RING1* promoter revealed consensus sites for potential NF- κ B p65 binding (Figure 6F). Targeting MUC1-C also decreased NF- κ B p65 occupancy on the *RING1* promoter (Figure 6F, left and right). These results collectively supported a model in which MUC1-C drives (i) RING2 by a MYC-mediated mechanism and (ii) RING1 predominantly by the NF- κ B p65 pathway.

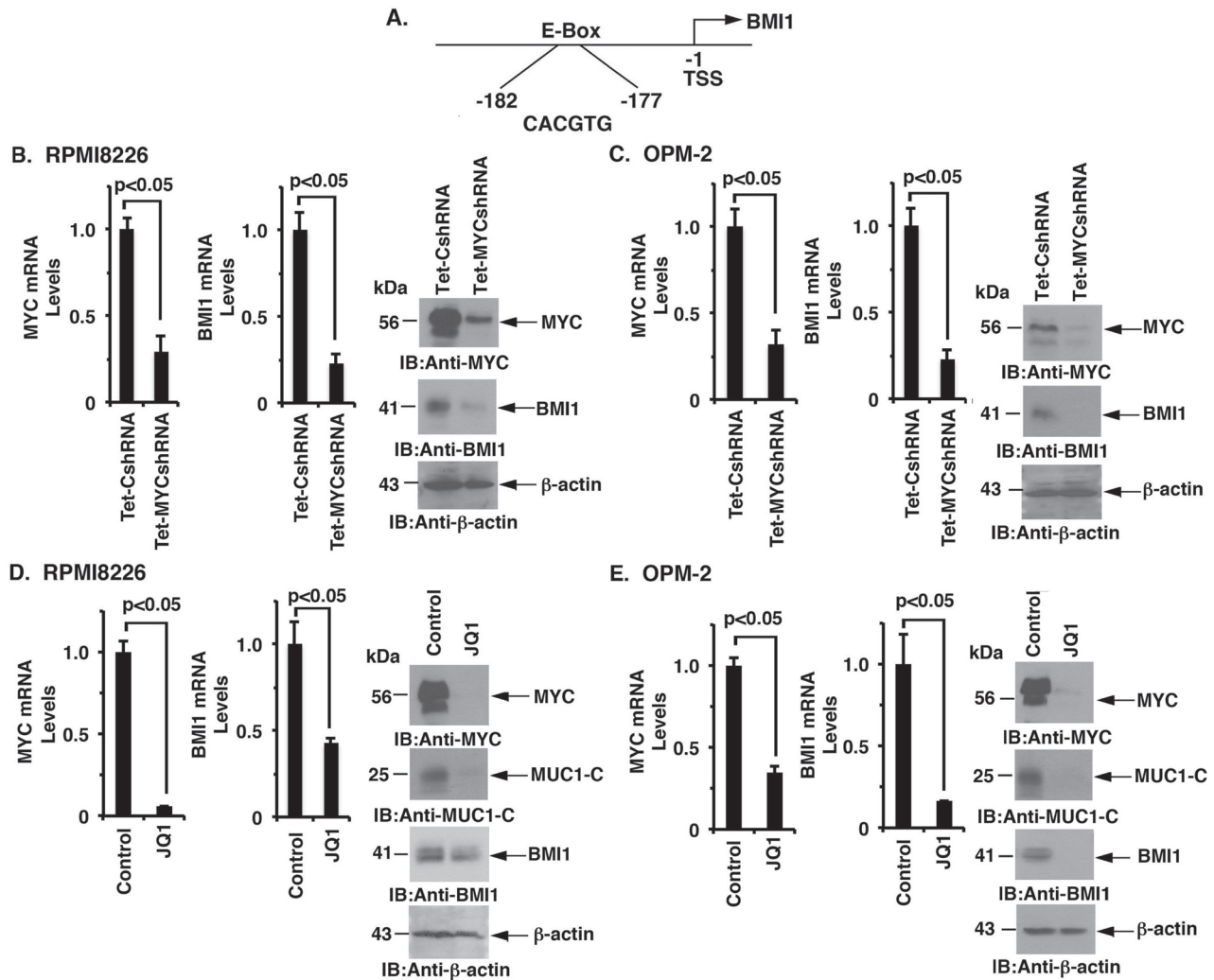


Figure 3: MUC1-C regulates BMI1 expression by a MYC-dependent mechanism. A. Schema of the BMI1 promoter with highlighting of a MYC binding site (CACGTG) at positions -177 to -182. B.-C. RPMI8226/Tet-CshRNA and RPMI8226/Tet-MYCshRNA (B) and OPM-2/Tet-CshRNA and OPM-2/Tet-MYCshRNA (C) cells treated with DOX for 5 d were analyzed for MYC and BMI1 mRNA levels by qRT-PCR (left and middle). The results (mean \pm SD of 3 determinations) are expressed as relative mRNA levels compared with that obtained for the Tet-CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). D.-E. RPMI8226 (D) and OPM-2 (E) cells treated with JQ1 or vehicle control for 48 h were analyzed for MYC and BMI1 mRNA levels by qRT-PCR (left and middle). The results (mean \pm SD of 3 determinations) are expressed as relative mRNA levels compared with that obtained for the control cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right).

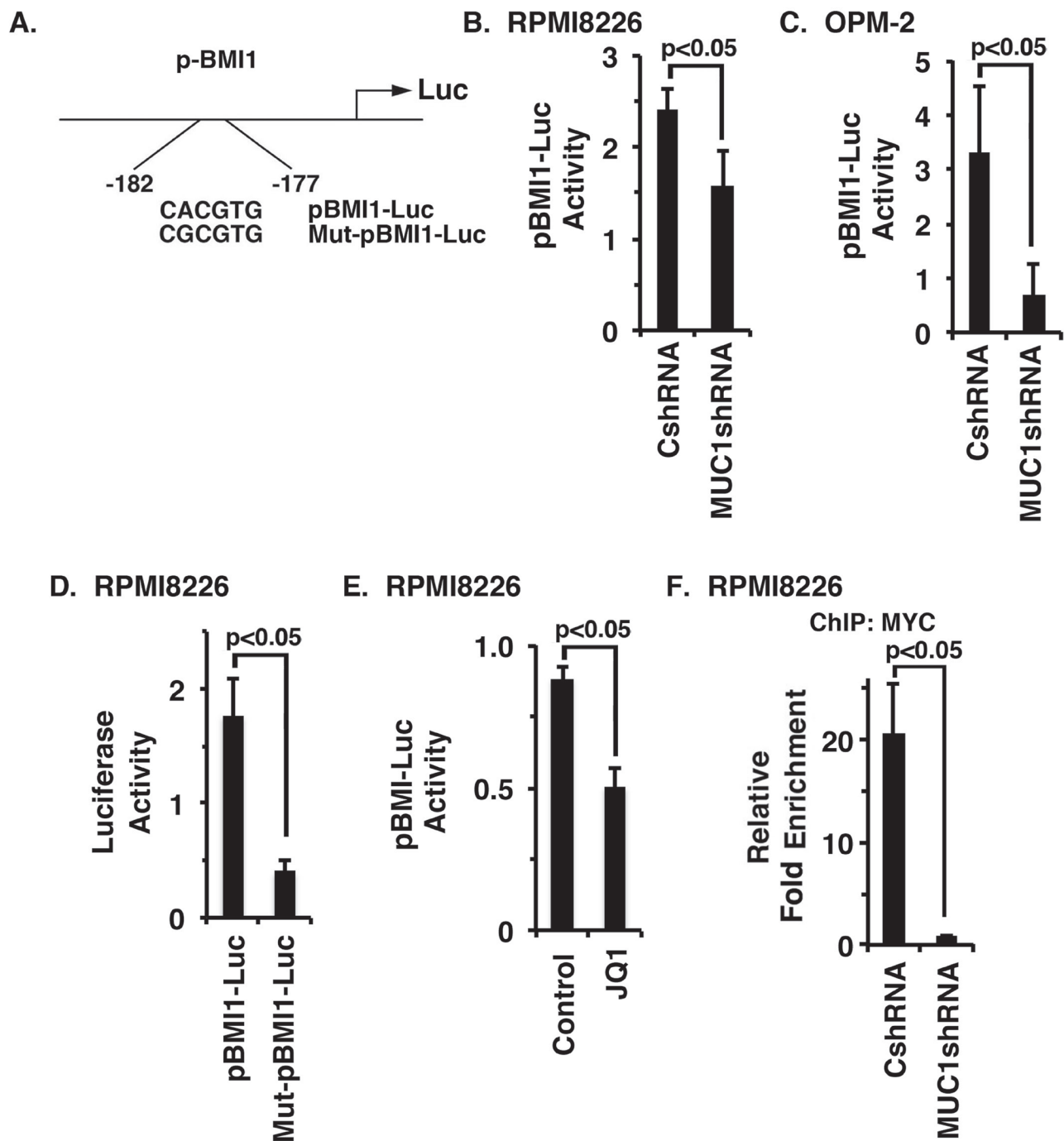


Figure 4: MUC1-C activates BMI1 gene transcription by a MYC-dependent mechanism. A. Schema of the pBMI1-Luc reporter with highlighting of the wild-type (WT) and mutated MYC binding site. B.-C. The designated RPMI8226 (B) and OPM-2 (C) cells expressing a CshRNA or MUC1shRNA were transfected with pGL3-Luc or pBMI1-Luc and SV40-Renilla-Luc as an internal control. Luciferase activity was measured at 24 h after transfection. The results (mean±SE of 3 determinations) are expressed as the relative luciferase activity compared with that obtained with cells expressing pGL3-Luc. D. RPMI8226 cells were transfected with (i) pGL3-Luc, (ii) pBMI1-Luc or Mut-pBMI1-Luc and (iii) SV40-Renilla-Luc. Luciferase activity was measured at 24 h after transfection. The results (mean±SE of 3 determinations) are expressed as the relative luciferase activity compared with that obtained with cells expressing pGL3-Luc. E. RPMI8226 cells were treated with JQ1 or vehicle control for 48 h and then transfected with the pGL3-Luc or pBMI1-Luc and SV-40-Renilla-Luc. Luciferase activity was measured at 24 h after transfection. The results (mean±SE of 3 determinations) are expressed as the relative luciferase activity compared with that obtained with cells expressing pGL3-Luc. F. Soluble chromatin from the RPMI8226/CshRNA and RPMI8226/MUC1shRNA cells was precipitated with anti-MYC or a control IgG antibody. The final DNA samples were amplified by qPCR with pairs of primers (Supplementary Table 2) for the MYC binding site in the *BMI1* promoter. The results (mean±SE of 3 determinations) are expressed as the relative fold enrichment compared with that obtained for the IgG control (assigned a value of 1).

Targeting MUC1-C derepresses PTEN, p14^{ARF} and BIM expression

PRC1 possesses H2A-K119 ubiquitin E3 ligase activity and catalyzes the ubiquitylation of H2A [32]. As expected from the suppression of BMI1, RING1 and RING2, we also found that targeting MUC1-C downregulates H2A ubiquitylation in RPMI8226 and OPM-2 cells (Figure 7A, left and right). PRC1 has been linked to the repression of certain tumor suppressor genes, such as *PTEN* and *INK4a/ARF (CDKN2A)*, in carcinoma cells [26, 28, 32]. By extension, we found that silencing MUC1-C in RPMI8226 (Figure 7B, left and right) and OPM-2 (Supplementary Figure S6A) cells is associated with the derepression of PTEN expression. Targeting MUC1-C with CRISPR/Cas9 was also associated with significant upregulation of PTEN levels (Supplementary

Figure 6B). In addition, targeting MUC1-C resulted in the induction of p14^{ARF} (Figure 7C, left and right; Supplementary Figure S6C). Other work has shown that BMI1 suppresses the proapoptotic BIM protein in MM cells [34]. Along these lines, silencing MUC1-C and thereby downregulation of BMI1 was associated with the upregulation of BIM (Figure 7D, left and right; Supplementary Figure S6D). These findings thus support a model in which (i) MUC1-C drives expression of BMI1, RING1 and RING2, and (ii) targeting MUC1-C suppresses PRC1 function in MM cells.

DISCUSSION

The findings that MUC1-C is invariably expressed in MM cells [1-4] and is necessary for their growth and survival [6-8] have supported MUC1-C

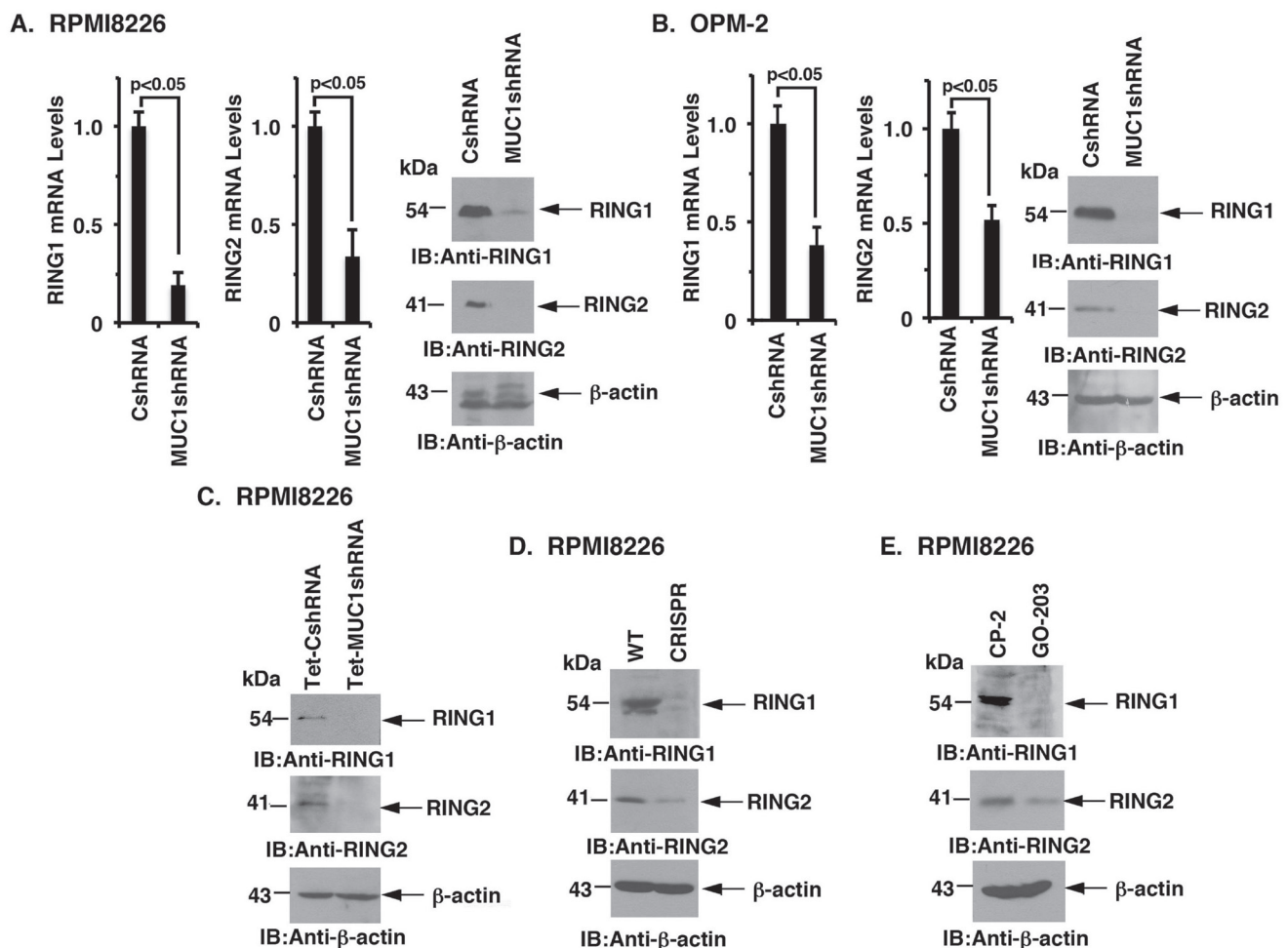


Figure 5: Targeting MUC1-C downregulates RING1 and RING2 expression. A.-B. The designated RPMI8226 (A) and OPM-2 (B) cells expressing a CshRNA or MUC1shRNA were analyzed for RING1 and RING2 mRNA levels by qRT-PCR (left and middle). The results (mean \pm SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). C.-E. Lysates from DOX-treated RPMI8226/Tet-CshRNA and RPMI8226/Tet-MUC1shRNA (C), RPMI8226/WT and RPMI8226/CRISPR (D), and CP-2- and GO-203-treated RPMI8226 cells (E) were immunoblotted with the indicated antibodies.

as a potential target for MM treatment. In addition, the recent demonstration that MUC1-C drives MYC in MM cells provided additional support for the importance of MUC1-C in the pathogenesis of MM [8]. In this respect,

there is abundant evidence that MYC is essential for the progression and survival of MM cells [16-18, 37]. What was not clear, however, is how the MUC1-C → MYC pathway functions in promoting the malignant

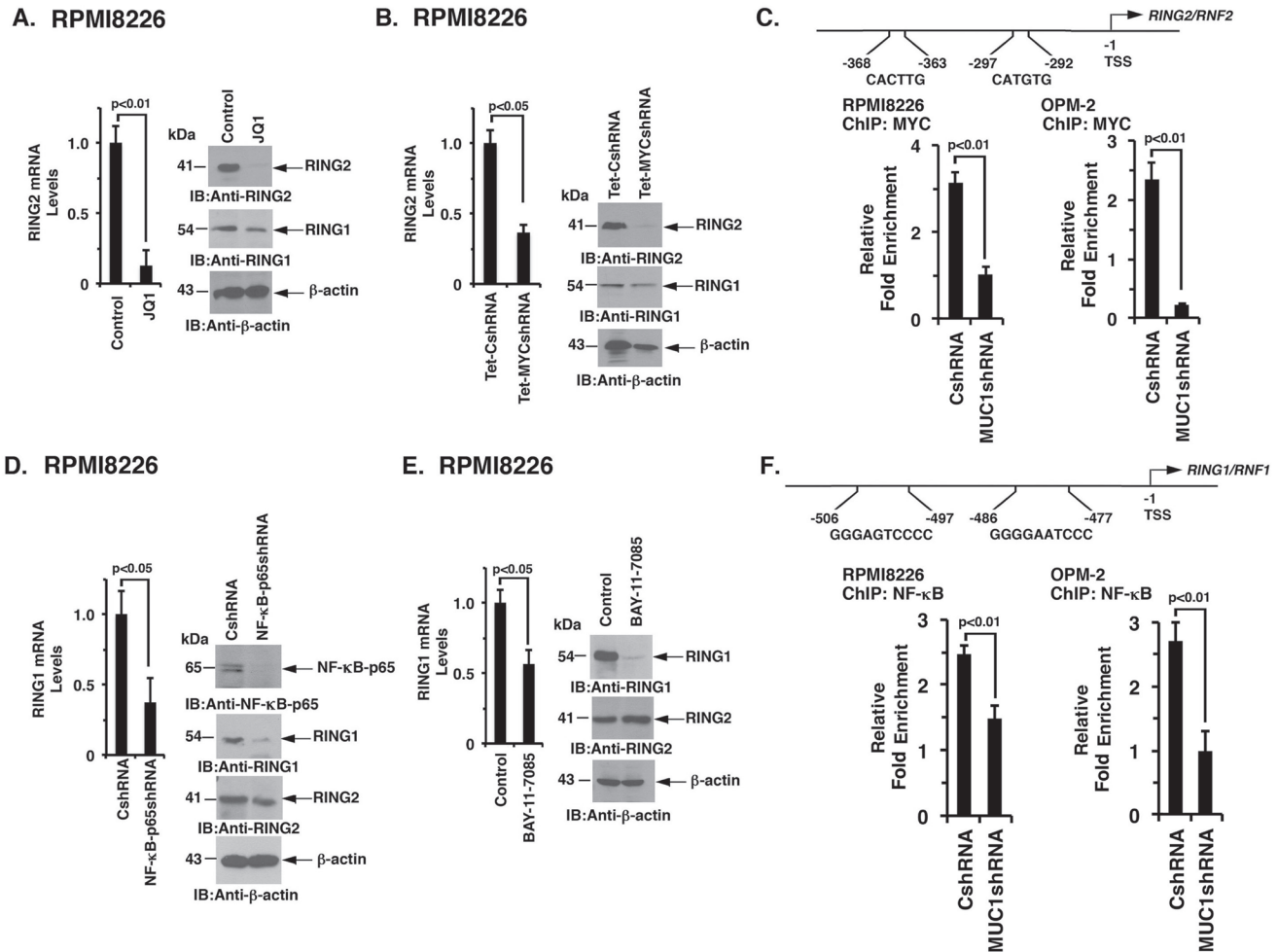


Figure 6: MUC1-C regulates RING2 and RING1 by MYC- and NF-κB p65-dependent mechanisms, respectively.

A. RPMI8226 cells treated with JQ1 or vehicle control for 48 h were analyzed for RING2 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the vehicle treated cells (assigned a value of 1, left). Lysates were immunoblotted with the indicated antibodies (right). **B.** RPMI8226/Tet-CshRNA and RPMI8226/Tet-MYCshRNA cells treated with DOX for 5 d were analyzed for RING2 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the Tet-CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). **C.** Schema of the *RING2* promoter with highlighting of putative MYC binding sites. Soluble chromatin from the RPMI8226/CshRNA and RPMI8226/MUC1shRNA (left) and OPM-2/CshRNA and OPM-2/MUC1shRNA (right) cells was precipitated with anti-MYC or a control IgG antibody. The final DNA samples were amplified by qPCR with pairs of primers (Supplementary Table S2) encompassing the MYC binding sites in the *RING2* promoter. The results (mean±SE of 3 determinations) are expressed as the relative fold enrichment compared with that obtained for the IgG control (assigned a value of 1). **D.** RPMI8226/CshRNA and RPMI8226/NF-κBshRNA were analyzed for RING1 mRNA levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the CshRNA cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). **E.** RPMI8226 cells treated with the 5 μM BAY-11-7085 or vehicle control for 24 h were assessed for RING1 levels by qRT-PCR (left). The results (mean±SD of 3 determinations) are expressed as relative mRNA levels as compared with that obtained for the vehicle treated cells (assigned a value of 1). Lysates were immunoblotted with the indicated antibodies (right). **F.** Schema of the *RING1* promoter with highlighting of putative NF-κB p65 binding sites. Soluble chromatin from the RPMI8226/CshRNA and RPMI8226/MUC1shRNA (left) and OPM-2/CshRNA and OPM-2/MUC1shRNA (right) cells was precipitated with anti-NF-κB p65 or a control IgG antibody. The final DNA samples were amplified by qPCR with pairs of primers (Supplementary Table 2) encompassing the NF-κB p65 binding sites in the *RING1* promoter. The results (mean±SE of 3 determinations) are expressed as the relative fold enrichment compared with that obtained for the IgG control (assigned a value of 1).

and RING2, we asked whether the MUC1-C → MYC pathway also drives expression of these additional PRC1 components. We found that, like BMI1, targeting MYC is associated with marked downregulation of RING2. In concert with a potential role for MUC1-C → NF-κB p65 signaling in regulating PRC1 function, we also found that targeting MUC1-C and NF-κB p65 suppresses RING1 expression. ChIP studies further demonstrated that MUC1-C promotes (i) MYC occupancy on the *RING2* promoter, and (ii) NF-κB p65 occupancy on the *RING1* promoter. Other studies will be needed to address the observation that targeting MYC is associated with partial downregulation of RING1 expression, which may be related to cross-talk between MUC1-C and MYC signaling. Our findings nonetheless underscore the notion that MUC1-C drives the expression of multiple components of the PRC1 complex, albeit by at least two distinct mechanisms, and thereby can integrate the regulation of BMI1, RING1 and RING2 in MM cells (Figure 7E). Our findings further support the premise that MUC1-C activates PRC1 function in epigenetic regulation. Indeed, targeting MUC1-C was associated with a marked decrease in PRC1-mediated H2A ubiquitylation and the induction of the PRC1-targeted tumor suppressor genes, *PTEN* and *INK4a/ARF* (Figure 7E). In concert with the demonstration that BIM1 suppresses the *BIM* gene in MM cells [34], we also found that targeting MUC1-C is associated with induction of BIM expression (Figure 7E). Our findings have thus identified a previously unrecognized role for MUC1-C in MM cells by activating the PRC1 complex and thereby the repression of certain TSGs.

With regard to translational relevance, PTC-209 is a small molecule inhibitor of BMI1 expression that has shown preclinical activity against colorectal and lung adenocarcinomas [29, 45]; however, to our knowledge, there are no clinically available BMI1 inhibitors. Our results indicate that targeting MUC1-C represents an alternative strategy for suppressing the expression of BMI1, as well as RING1 and RING2. In this regard, the MUC1-C inhibitor GO-203 has completed a Phase I trial in patients with advanced solid tumors and, based on the findings that GO-203 is highly synergistic with decitabine [46], is presently under study in a Phase II trial in combination with decitabine for patients with AML (NCT02204085). Therefore, the present results provide the basis for targeting MUC1-C and thereby reprogramming of the MM epigenome alone or in combination with other agents.

MATERIALS AND METHODS

Cell culture

RPMI8266, OPM-2, KMS-12-PE, U266 and NCIH929 (ATCC, Manassas, VA, USA) cells were cultured in RPMI1640 medium supplemented with 10% heat-inactivated fetal bovine serum (FBS), 100 U/mL penicillin, 100 μg/mL streptomycin and 2 mM L-glutamine. Primary CD138+ MM cells from 2 patients with refractory disease were isolated by Ficoll gradient separation using CD138 MicroBeads (Miltenyi Biotec, Somerville, MA, USA) and maintained in long-term culture in IMDM medium supplemented with 20% heat-inactivated FBS as described [8, 47, 48, 49]. Cells were treated with (i) doxycycline (DOX; Sigma, St. Louis, MO, USA), (ii) MUC1-C inhibitor GO-203 ([R]₉-CQCRRKN) or inactive control peptide CP-2 ([R]₉-AQARRKN), (iii) BRD4 inhibitor JQ1 [50], and (iv) NF-κB p65 inhibitor BAY-11-7085 (Santa Cruz Biotechnology, Dallas, TX, USA) or vehicle control dimethylsulfoxide (DMSO; Sigma).

MUC1, MYC and NF-κB p65 silencing

MM cell lines and primary cells were infected with lentiviral vectors expressing a MUC1 shRNA, NF-κB p65 shRNA or scrambled control shRNA vector (Sigma) [8, 46]. Cells were selected and maintained in puromycin. For DOX-inducible MUC1 and MYC silencing, MUC1shRNA (MISSION shRNA; Sigma, TRCN0000122938), MYCshRNA (MISSION shRNA; Sigma, TRCN0000039642) or a control scrambled CshRNA (Sigma) was inserted into the pLKO-Tet-puro vector (Addgene, Cambridge, MA, USA, Plasmid #21915). Viral vectors were produced in HEK293T cells as described [51, 52]. Cells expressing Tet-shRNA vectors were selected in puromycin. MUC1 was also silenced with CRISPR/Cas9 gene editing as described [8].

Real-time quantitative reverse-transcription PCR (qRT-PCR)

Total RNA was isolated using the RNeasy Mini Kit (Invitrogen, Carlsbad, CA, USA). Complementary DNA (cDNA) was synthesized using the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA) [8]. The cDNA samples were amplified using the SYBR Green qRT-PCR assay kit and the ABI Prism 7000 sequence detector (Applied Biosystems). Primers used for qRT-PCR analysis are listed in Supplementary Table 1. The results were analyzed using the $\Delta\Delta$ cycle threshold (DDCt) method as described [53].

Statistical significance was determined by the Student's t test.

Immunoblot analysis

Cells were lysed in NP-40 buffer containing protease inhibitor cocktail (ThermoScientific, Waltham, MA, USA). Immunoblotting was performed with anti-MUC1-C (NeoMarkers, Fremont, CA, USA), anti-BMI1, anti-RING1, anti-RING2, anti-H2Aub1, anti-H2A, anti-PTEN, anti-p14^{ARF} (Cell Signaling Technology, Danvers, MA, USA), anti-NF- κ B p65, anti-BIM (Santa Cruz Biotechnology), anti-MYC (Abcam, Cambridge, MA, USA) and anti- β -actin (Sigma).

Chromatin immunoprecipitation (ChIP) assays

Soluble chromatin was precipitated with anti-MYC (Abcam) or a control nonimmune IgG (Santa Cruz Biotechnology). For real-time ChIP quantitative polymerase chain reactions (qPCRs), the SYBR green system was used with the ABI Prism 7000 sequence detector (Applied Biosystems). Data are reported as relative fold enrichment as described [13, 36]. Primers used for qPCR are listed in Supplementary Table 2 [44].

Promoter-reporter assays

Cells were transfected with an empty pGL3 luciferase (Luc) reporter vector, a pGL3-pBMI1-Luc promoter-reporter or a mutated Mut-pBMI1-Luc vector provided by Dr. Goberdhan Dimri [35, 44] and SV-40-*Renilla*-Luc in the presence of LipofectamineTM 3000 Reagent (Invitrogen). After 24-48 h, the transfected cells were lysed using passive lysis buffer. The lysates were analyzed using the dual luciferase assay system (Promega, Madison, WI, USA).

Abbreviations

MUC1, mucin 1; MUC1-C, MUC1 C-terminal subunit; MM, multiple myeloma; PcG, polycomb group proteins, PRC1, polycomb repressive complex 1; BMI1, B cell-specific Moloney murine leukemia virus integration site 1; RING, ring finger protein; PRC2, polycomb repressive complex 2; EZH2, enhancer of zeste homolog 2; SUZ12, suppressor of zeste 12 homolog; DNMT, DNA methyltransferase; TSG, tumor suppressor gene; DOX, doxycycline.

CONFLICTS OF INTEREST

DK has ownership interest (including patents) and is a consultant/advisory board member of Genus Oncology.

No potential conflicts of interest were disclosed by the other authors.

GRANT SUPPORT

Research reported in this publication was supported by the National Cancer Institute of the National Institutes of Health under award numbers P50CA100707, R01CA097098 and R01CA166480.

REFERENCES

1. Treon SP, Mollick JA, Urashima M, Teoh G, Chauhan D, Ogata A, Raje N, Hilgers JH, Nadler L, Belch AR, Pilariski LM, Anderson KC. Muc-1 core protein is expressed on multiple myeloma cells and is induced by dexamethasone. *Blood*. 1999; 93: 1287-98.
2. Paydas S, Sahin B, Gonlusen G, Hazar B, Zorludemir S. MUC1 expression in plasmacytoma. *Leuk Res*. 2001; 25: 221-5.
3. Cloosen S, Gratama J, van Leeuwen EB, Senden-Gijsbers BL, Oving EB, von Mensdorff-Pouilly S, Tarp MA, Mandel U, Clausen H, Germeraad WT, Bos GM. Cancer specific Mucin-1 glycoforms are expressed on multiple myeloma. *Br J Haematol*. 2006; 135: 513-6. doi: 10.1111/j.1365-2141.2006.06331.x.
4. Baldus SE, Palmén C, Thiele J. MUC1 (EMA) expressing plasma cells in bone marrow infiltrated by plasma cell myeloma. *Histol Histopathol*. 2007; 22: 889-93. doi: 10.14670/HH-22.889.
5. Kawano T, Ahmad R, Nogi H, Agata N, Anderson K, Kufe D. MUC1 oncoprotein promotes growth and survival of human multiple myeloma cells. *Int J Oncology*. 2008; 33: 153-9.
6. Yin L, Ahmad R, Kosugi M, Kufe T, Vasir B, Avigan D, Kharbanda S, Kufe D. Survival of human multiple myeloma cells is dependent on MUC1 C-terminal transmembrane subunit oncoprotein function. *Mol Pharm*. 2010; 78: 166-74. doi: 10.1124/mol.110.065011.
7. Yin L, Kosugi M, Kufe D. Inhibition of the MUC1-C oncoprotein induces multiple myeloma cell death by downregulating TIGAR expression and depleting NADPH. *Blood*. 2012; 119: 810-6. doi: 10.1182/blood-2011-07-369686.
8. Tagde A, Rajabi H, Bouillez A, Alam M, Gali R, Bailey S, Tai YT, Hideshima T, Anderson K, Avigan D, Kufe D. MUC1-C drives MYC in multiple myeloma. *Blood*. 2016; 127: 2587-97. doi: 10.1182/blood-2015-07-659151.
9. Raina D, Agarwal P, Lee J, Bharti A, McKnight C, Sharma P, Kharbanda S, Kufe D. Characterization of the MUC1-C cytoplasmic domain as a cancer target. *PLoS One*. 2015; 10: e0135156. doi: 10.1371/journal.pone.0135156.
10. Kufe D. Mucins in cancer: function, prognosis and therapy. *Nature Reviews Cancer*. 2009; 9: 874-85. doi: 10.1038/

nrc2761.

11. Ahmad R, Raina D, Trivedi V, Ren J, Rajabi H, Kharbanda S, Kufe D. MUC1 oncoprotein activates the I κ B kinase β complex and constitutive NF- κ B signaling. *Nat Cell Biol.* 2007; 9: 1419-27. doi: 10.1038/ncb1661.
12. Ahmad R, Raina D, Joshi MD, Kawano T, Kharbanda S, Kufe D. MUC1-C oncoprotein functions as a direct activator of the NF- κ B p65 transcription factor. *Cancer Res.* 2009; 69: 7013-21. doi: 10.1182/blood-2011-07-369686.
13. Takahashi H, Jin C, Rajabi H, Pitroda S, Alam M, Ahmad R, Raina D, Hasegawa M, Suzuki Y, Tagde A, Bronson RT, Weichselbaum R, Kufe D. MUC1-C activates the TAK1 inflammatory pathway in colon cancer. *Oncogene.* 2015; 34: 5187-97. doi: 10.1038/onc.2014.442.
14. Huang L, Chen D, Liu D, Yin L, Kharbanda S, Kufe D. MUC1 oncoprotein blocks GSK3 β -mediated phosphorylation and degradation of β -catenin. *Cancer Res.* 2005; 65: 10413-22. doi: 10.1182/blood-2011-07-369686.
15. Rajabi H, Ahmad R, Jin C, Kosugi M, Alam M, Joshi M, Kufe D. MUC1-C oncoprotein induces TCF7L2 transcription factor activation and promotes cyclin D1 expression in human breast cancer cells. *J Biol Chem.* 2012; 287: 10703-13. doi: 10.1074/jbc.M111.323311.
16. Holien T, Vatsveen TK, Hella H, Waage A, Sundan A. Addiction to c-MYC in multiple myeloma. *Blood.* 2012; 120: 2450-3. doi: 10.1182/blood-2011-08-371567.
17. Delmore JE, Issa GC, Lemieux ME, Rahl PB, Shi J, Jacobs HM, Kasttrit E, Gilpatrick T, Paranal RM, Qi J, Chesi M, Schinzel AC, McKeown MR, et al. BET bromodomain inhibition as a therapeutic strategy to target c-Myc. *Cell.* 2011; 146: 904-17. doi: 10.1016/j.cell.2011.08.017.
18. Mertz JA, Conery AR, Bryant BM, Sandy P, Balasubramanian S, Mele DA, Bergeron L, Sims RJ 3rd. Targeting MYC dependence in cancer by inhibiting BET bromodomains. *Proc Natl Acad Sci USA.* 2011; 108: 16669-74. doi: 10.1073/pnas.1108190108.
19. Wang H, Wang L, Erdjument-Bromage H, Vidal M, Tempst P, Jones RS, Zhang Y. Role of histone H2A ubiquitination in Polycomb silencing. *Nature.* 2004; 431: 873-8. doi: 10.1038/nature02985.
20. de Napoles M, Mermoud JE, Wakao R, Tang YA, Endoh M, Appanah R, Nesterova TB, Silva J, Otte AP, Vidal M, Koseki H, Brockdorff N. Polycomb group proteins Ring1A/B link ubiquitylation of histone H2A to heritable gene silencing and X inactivation. *Dev Cell.* 2004; 7: 663-76. doi: 10.1016/j.devcel.2004.10.005.
21. Cao R, Tsukada Y, Zhang Y. Role of Bmi-1 and Ring1A in H2A ubiquitylation and Hox gene silencing. *Mol Cell.* 2005; 20: 845-54. doi: 10.1016/j.molcel.2005.12.002.
22. Wei J, Zhai L, Xu J, Wang H. Role of Bmi1 in H2A ubiquitylation and Hox gene silencing. *J Biol Chem.* 2006; 281: 22537-44. doi: 10.1074/jbc.M600826200.
23. Blackledge NP, Rose NR, Klose RJ. Targeting Polycomb systems to regulate gene expression: modifications to a complex story. *Nat Rev Mol Cell Biol.* 2015; 16: 643-9. doi: 10.1038/nrm4067.
24. Bracken AP, Dietrich N, Pasini D, Hansen KH, Helin K. Genome-wide mapping of Polycomb target genes unravels their roles in cell fate transitions. *Genes Dev.* 2006; 20: 1123-36. doi: 10.1101/gad.381706.
25. Ohtsubo M, Yasunaga S, Ohno Y, Tsumura M, Okada S, Ishikawa N, Shirao K, Kikuchi A, Nishitani H, Kobayashi M, Takihara Y. Polycomb-group complex 1 acts as an E3 ubiquitin ligase for Geminin to sustain hematopoietic stem cell activity. *Proc Natl Acad Sci U S A.* 2008; 105: 10396-401. doi: 10.1073/pnas.0800672105.
26. Park IK, Morrison SJ, Clarke MF. Bmi1, stem cells, and senescence regulation. *J Clin Invest.* 2004; 113: 175-9. doi: 10.1172/JCI20800.
27. Richly H, Aloia L, Di Croce L. Roles of the Polycomb group proteins in stem cells and cancer. *Cell Death Dis.* 2011; 2: e204. doi: 10.1038/cddis.2011.84.
28. Siddique HR, Saleem M. Role of BMI1, a stem cell factor, in cancer recurrence and chemoresistance: preclinical and clinical evidences. *Stem Cells.* 2012; 30: 372-8. doi: 10.1002/stem.1035.
29. Kreso A, van Galen P, Pedley NM, Lima-Fernandes E, Frelin C, Davis T, Cao L, Baiazitov R, Du W, Sydorenko N, Moon YC, Gibson L, Wang Y, et al. Self-renewal as a therapeutic target in human colorectal cancer. *Nat Med.* 2014; 20: 29-36. doi: 10.1038/nm.3418.
30. Lessard J, Sauvageau G. Bmi-1 determines the proliferative capacity of normal and leukaemic stem cells. *Nature.* 2003; 423: 255-60. doi: 10.1038/nature01572.
31. Rizo A, Olthof S, Han L, Vellenga E, de Haan G, Schuringa JJ. Repression of BMI1 in normal and leukemic human CD34(+) cells impairs self-renewal and induces apoptosis. *Blood.* 2009; 114: 1498-505. doi: 10.1182/blood-2009-03-209734.
32. Song LB, Li J, Liao WT, Feng Y, Yu CP, Hu LJ, Kong QL, Xu LH, Zhang X, Liu WL, Li MZ, Zhang L, Kang TB, et al. The polycomb group protein Bmi-1 represses the tumor suppressor PTEN and induces epithelial-mesenchymal transition in human nasopharyngeal epithelial cells. *J Clin Invest.* 2009; 119: 3626-36. doi: 10.1172/JCI39374.
33. Yang MH, Hsu DS, Wang HW, Wang HJ, Lan HY, Yang WH, Huang CH, Kao SY, Tzeng CH, Tai SK, Chang SY, Lee OK, Wu KJ. Bmi1 is essential in Twist1-induced epithelial-mesenchymal transition. *Nat Cell Biol.* 2010; 12: 982-92. doi: 10.1038/ncb2099.
34. Jagani Z, Wiederschain D, Loo A, He D, Mosher R, Fordjour P, Monahan J, Morrissey M, Yao YM, Lengauer C, Warmuth M, Sellers WR, Dorsch M. The Polycomb group protein Bmi-1 is essential for the growth of multiple myeloma cells. *Cancer Res.* 2010; 70: 5528-38. doi: 10.1158/0008-5472.CAN-09-4229.
35. Cho JH, Dimri M, Dimri GP. A positive feedback loop regulates the expression of polycomb group protein BMI1

- via WNT signaling pathway. *J Biol Chem.* 2013; 288: 3406-18. doi: 10.1074/jbc.M112.422931.
36. Rajabi H, Tagde A, Alam M, Bouillez A, Pitroda S, Suzuki Y, Kufe D. DNA methylation by DNMT1 and DNMT3b methyltransferases is driven by the MUC1-C oncoprotein in human carcinoma cells. *Oncogene.* 2016; 35: 6439-45. doi: 10.1038/onc.2016.180.
37. Chesi M, Robbiani DF, Sebag M, Chng WJ, Affer M, Tiedemann R, Valdez R, Palmer SE, Haas SS, Stewart AK, Fonseca R, Kremer R, Cattoretti G, et al. AID-dependent activation of a MYC transgene induces multiple myeloma in a conditional mouse model of post-germinal center malignancies. *Cancer Cell.* 2008; 13: 167-80. doi: 10.1016/j.ccr.2008.01.007.
38. Jacobs JJ, Scheijen B, Voncken JW, Kieboom K, Berns A, van Lohuizen M. Bmi-1 collaborates with c-Myc in tumorigenesis by inhibiting c-Myc-induced apoptosis via INK4a/ARF. *Genes Dev.* 1999; 13: 2678-90.
39. De Vos J, Thykjaer T, Tarte K, Ensslen M, Raynaud P, Requirand G, Pellet F, Pantesco V, Reme T, Jourdan M, Rossi JF, Orntoft T, Klein B. Comparison of gene expression profiling between malignant and normal plasma cells with oligonucleotide arrays. *Oncogene.* 2002; 21: 6848-57. doi: 10.1038/sj.onc.1205868.
40. Chng WJ, Kumar S, Vanwier S, Ahmann G, Price-Troska T, Henderson K, Chung TH, Kim S, Mulligan G, Bryant B, Carpten J, Gertz M, Rajkumar SV, et al. Molecular dissection of hyperdiploid multiple myeloma by gene expression profiling. *Cancer Res.* 2007; 67: 2982-9. doi: 10.1158/0008-5472.CAN-06-4046.
41. Zhan F, Barlogie B, Arzoumanian V, Huang Y, Williams DR, Hollmig K, Pineda-Roman M, Tricot G, van Rhee F, Zangari M, Dhodapkar M, Shaughnessy JD Jr. Gene-expression signature of benign monoclonal gammopathy evident in multiple myeloma is linked to good prognosis. *Blood.* 2007; 109: 1692-700. doi: 10.1182/blood-2006-07-037077.
42. Rajabi H, Alam M, Takahashi H, Kharbada A, Guha M, Ahmad R, Kufe D. MUC1-C oncoprotein activates the ZEB1/miR-200c regulatory loop and epithelial-mesenchymal transition. *Oncogene.* 2014; 33: 1680-9. doi: 10.1038/onc.2013.114.
43. Dimri M, Kang M, Dimri GP. A miR-200c/141-BMI1 autoregulatory loop regulates oncogenic activity of BMI1 in cancer cells. *Oncotarget.* 2016; 7: 36220-34. doi: 10.18632/oncotarget.8811.
44. Hiraki M, Maeda T, Bouillez A, Alam M, Tagde A, Hinohara K, Suzuki Y, Markert T, Miyo T, Komura K, Ahmad R, Rajabi H, Kufe D. MUC1-C activates BMI1 in human cancer cells. *Oncogene.* 2017; 36:2791-2801. doi: 10.1038/onc.2016.439.
45. Yong KJ, Basseres DS, Welner RS, Zhang WC, Yang H, Yan B, Alberich-Jorda M, Zhang J, de Figueiredo-Pontes LL, Battelli C, Hetherington CJ, Ye M, Zhang H, et al. Targeted BMI1 inhibition impairs tumor growth in lung adenocarcinomas with low CEBPalpha expression. *Sci Transl Med.* 2016; 8: 350ra104. doi: 10.1126/scitranslmed.aad6066.
46. Tagde A, Rajabi H, Stroopinsky D, Gali R, Alam M, Bouillez A, Kharbada S, Stone R, Avigan D, Kufe D. MUC1-C induces DNA methyltransferase 1 and represses tumor suppressor genes in acute myeloid leukemia. *Oncotarget.* 2016; 7: 38974-87. doi: 10.18632/oncotarget.9777.
47. Lokhorst HM, Lamme T, de Smet M, Klein S, de Weger RA, van Oers R, Bloem AC. Primary tumor cells of myeloma patients induce interleukin-6 secretion in long-term bone marrow cultures. *Blood.* 1994; 84: 2269-77.
48. Tassone P, Neri P, Burger R, Savino R, Shammam M, Catley L, Podar K, Chauhan D, Masciari S, Gozzini A, Tagliaferri P, Venuta S, Munshi NC, et al. Combination therapy with interleukin-6 receptor superantagonist Sant7 and dexamethasone induces antitumor effects in a novel SCID-hu *In vivo* model of human multiple myeloma. *Clin Cancer Res.* 2005; 11: 4251-8. doi: 10.1158/1078-0432.CCR-04-2611.
49. Deola S, Scaramuzza S, Birolo RS, Cerngul M, Ficara F, Dando J, Voena C, Vai S, Monari M, Pogliani E, Corneo G, Peccatori J, Selleri S, et al. Molecular purging of multiple myeloma cells by ex-vivo culture and retroviral transduction of mobilized-blood CD34+ cells. *J Transl Med.* 2007; 5: 35. doi: 10.1186/1479-5876-5-35.
50. Bouillez A, Rajabi H, Pitroda S, Jin C, Alam M, Kharbada A, Tagde A, Wong K, Kufe D. Inhibition of MUC1-C suppresses MYC expression and attenuates malignant growth in KRAS mutant lung adenocarcinomas. *Cancer Res.* 2016; 76: 1538-48. doi: 10.1158/0008-5472.CAN-15-1804.
51. Hiraki M, Suzuki Y, Alam M, Hinohara K, Hasegawa M, Jin C, Kharbada S, Kufe D. MUC1-C stabilizes MCL-1 in the oxidative stress response of triple-negative breast cancer cells to BCL-2 inhibitors. *Scientific Reports.* 2016; 6: 26643. doi: 10.1038/srep26643.
52. Hasegawa M, Takahashi H, Rajabi H, Alam M, Suzuki Y, Yin L, Tagde A, Maeda T, Hiraki M, Sukhatme V, Kufe D. Functional interactions of the cystine/glutamate antiporter, CD44v and MUC1-C oncoprotein in triple-negative breast cancer cells. *Oncotarget.* 2016; 7: 11756-69. doi: 10.18632/oncotarget.7598.
53. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods.* 2001; 25: 402-8. doi: 10.1006/meth.2001.1262.