



High-Quality Whole-Genome Sequences of the Oligo-Mouse-Microbiota Bacterial Community

Debora Garzetti,^{a,b} Sandrine Brugiroux,^a Boyke Bunk,^c Rüdiger Pukall,^c
Kathy D. McCoy,^d Andrew J. Macpherson,^d Bärbel Stecher^{a,b}

Max von Pettenkofer Institute of Hygiene and Medical Microbiology, Ludwig-Maximilians-University of Munich, Munich, Germany^a; German Center for Infection Research (DZIF), Partner Site Munich, Munich, Germany^b; Leibniz Institute DSMZ—German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany^c; Maurice Müller Laboratories, Department of Clinical Research (DKF), UVCN, University Hospital, Bern, Switzerland^d

ABSTRACT The Oligo-Mouse-Microbiota (Oligo-MM¹²) is a community of 12 mouse intestinal bacteria to be used for microbiome research in gnotobiotic mice. We present here the high-quality whole genome sequences of the Oligo-MM¹² strains, which were obtained by combining the accuracy of the Illumina platforms with the long reads of the PacBio technology.

In a recent study, we described a defined intestinal community of 12 murine strains, termed Oligo-Mouse-Microbiota (Oligo-MM¹²), which permanently colonize gnotobiotic mice over several generations and provide colonization resistance against *Salmonella enterica* serovar Typhimurium (1). This bacterial consortium has been thoroughly characterized by biochemical and molecular methods, and the individual strains have been deposited at the German Culture Collection of Microorganisms and Cell Cultures (DSMZ) (Table 1). The genomes of the 12 bacteria were previously sequenced and assembled via different techniques and algorithms (1–3). Since the Oligo-MM¹² strains are being used by an increasing number of research groups (1, 3–5), the multitude of genome sequences precludes the possibility of a meaningful exchange of data within the scientific community. Thus, there is a strong need for availability and constant update of the Oligo-MM¹² reference genomes.

It is well recognized that sequences from the Illumina platforms have low error rates, with systematic errors being mainly situated at the end of the reads, but are too short for an efficient complete genome assembly (6). On the contrary, the long reads generated by PacBio sequencing are less accurate and contain random errors (6). Aiming to create a set of reference genomes, in this study we present the high-quality genome sequences of the Oligo-MM¹² bacteria, which were assembled by a hybrid approach combining Illumina and PacBio sequences (Table 1).

As previously described (1), the complete genome sequence of *Acutalibacter muris* KB18 was obtained on the PacBio RSII platform and assembled using the RS_HGAP_Assembly.3 protocol (default parameters). Error correction was then performed by mapping Illumina reads onto the finished genome with the Burrows–Wheeler Alignment tool (7), with subsequent variant calling using CLC Genomics Workbench version 7.0.4. Here, Illumina MiSeq reads (1) of the remaining 11 bacterial genomes were assembled onto their respective PacBio complete genomes (2) by applying a reference-guided approach using SPAdes (8), with a minimum contig length of 500 bp. Assemblies were evaluated with QUILT (Quality Assessment Tool for genome assemblies) (9), and the final genomes were automatically annotated using RAST (Rapid Annotations using Subsystems Technology) (10). In future studies, genetic variation, genome evo-

Received 20 June 2017 Accepted 11 August 2017 Published 19 October 2017

Citation Garzetti D, Brugiroux S, Bunk B, Pukall R, McCoy KD, Macpherson AJ, Stecher B. 2017. High-quality whole-genome sequences of the Oligo-Mouse-Microbiota bacterial community. *Genome Announc* 5:e00758-17. <https://doi.org/10.1128/genomeA.00758-17>.

Copyright © 2017 Garzetti et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Debora Garzetti, garzetti@mvp.uni-muenchen.de.

TABLE 1 Assembly information and accession numbers of the Oligo-MM¹² genomes

Oligo-MM strain	Total length (bp)	No. of contigs	No. of genes	DSM no.	Accession no.
[<i>Clostridium</i>] <i>innocuum</i> I46	4,468,984	1	4,629	26113	CP022722
<i>Bacteroides caecimuris</i> I48	4,800,416	19	4,225	26085	NHMU00000000
<i>Lactobacillus reuteri</i> I49	2,063,604	3	2,006	32035	NHMT00000000
<i>Enterococcus faecalis</i> KB1	3,025,555	1	2,942	32036	CP022712
<i>Acutalibacter muris</i> KB18	3,802,813	1	3,990	26090	CP021422
<i>Bifidobacterium animalis</i> subsp. <i>animalis</i> YL2	2,021,926	2	1,732	26074	NHMR00000000
<i>Muribaculum intestinale</i> YL27	3,306,969	1	2,786	28989	CP021421
<i>Flavonifractor plautii</i> YL31	3,813,655	5	3,924	26117	NHMQ00000000
[<i>Clostridium</i>] <i>clostridioforme</i> YL32	7,157,460	16	7,735	26114	NHTR00000000
<i>Akkermansia muciniphila</i> YL44	2,737,167	1	2,731	26127	CP021420
<i>Turicimonas muris</i> YL45	2,887,709	20	2,754	26109	NHMP00000000
<i>Blautia coccooides</i> YL58	5,128,482	1	5,230	26115	CP022713

lution, and functional genomics, among other research applications, of the Oligo-MM¹² community can be assessed by high-quality analyses.

Accession number(s). The assembled whole-genome sequences of the Oligo-MM¹² strains have been deposited in DDBJ/ENA/GenBank under the accession numbers given in Table 1.

ACKNOWLEDGMENTS

We thank Cathrin Spröer, Nicole Heyer, and Simone Severitt for sequencing of the KB18 PacBio genome.

This work was supported by the German Center for Infection Research (DZIF), the Center for Gastrointestinal Microbiome Research (CEGIMIR), and the German Research Foundation (DFG). The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

REFERENCES

- Brugiroux S, Beutler M, Pfann C, Garzetti D, Ruscheweyh HJ, Ring D, Diehl M, Herp S, Lötscher Y, Hussain S, Bunk B, Pukall R, Huson DH, Münch PC, McHardy AC, McCoy KD, Macpherson AJ, Loy A, Clavel T, Berry D, Stecher B. 2016. Genome-guided design of a defined mouse microbiota that confers colonization resistance against *Salmonella enterica* serovar Typhimurium. *Nat Microbiol* 2:16215. <https://doi.org/10.1038/nmicrobiol.2016.215>.
- Uchimura Y, Wyss M, Brugiroux S, Limenitakis JP, Stecher B, McCoy KD, Macpherson AJ. 2016. Complete genome sequences of 12 species of stable defined moderately diverse mouse microbiota 2. *Genome Announc* 4(5):e00951-16. <https://doi.org/10.1128/genomeA.00951-16>.
- Lagkouvardos I, Pukall R, Abt B, Foesele BU, Meier-Kolthoff JP, Kumar N, Bresciani A, Martínez I, Just S, Ziegler C, Brugiroux S, Garzetti D, Wenning M, Bui TP, Wang J, Hugenholtz F, Plugge CM, Peterson DA, Hornef MW, Baines JF, Smidt H, Walter J, Kristiansen K, Nielsen HB, Haller D, Overmann J, Stecher B, Clavel T. 2016. The Mouse intestinal Bacterial Collection (miBC) provides host-specific insight into cultured diversity and functional potential of the gut microbiota. *Nat Microbiol* 1:16131. <https://doi.org/10.1038/nmicrobiol.2016.131>.
- Studer N, Desharnais L, Beutler M, Brugiroux S, Terrazos MA, Menin L, Schürch CM, McCoy KD, Kuehne SA, Minton NP, Stecher B, Bernier-Latmani R, Hapfelmeier S. 2016. Functional intestinal bile acid 7 α -dehydroxylation by *Clostridium scindens* associated with protection from *Clostridium difficile* infection in a gnotobiotic mouse model. *Front Cell Infect Microbiol* 6:191. <https://doi.org/10.3389/fcimb.2016.00191>.
- Li H, Limenitakis JP, Fuhrer T, Geuking MB, Lawson MA, Wyss M, Brugiroux S, Keller I, Macpherson JA, Rupp S, Stolp B, Stein JV, Stecher B, Sauer U, McCoy KD, Macpherson AJ. 2015. The outer mucus layer hosts a distinct intestinal microbial niche. *Nat Commun* 6:8292. <https://doi.org/10.1038/ncomms9292>.
- Loman NJ, Pallen MJ. 2015. Twenty years of bacterial genome sequencing. *Nat Rev Microbiol* 13:787–794. <https://doi.org/10.1038/nrmicro3565>.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25:1754–1760. <https://doi.org/10.1093/bioinformatics/btp324>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUASt: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.