

Identification of tumor antigens in malignant mesothelioma

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Abstract. Serological analysis of recombinant tumor cDNA expression library (SEREX) is a powerful and widely used method to explore the cancer immune environment. In the present study, immunoscreening of normal testicular tissues and malignant mesothelioma (MM) cancer MSTO-211H cell line cDNA libraries with sera from 5 MM patients led to the isolation of 16 independent antigens, which were designated 'Korea Pusan-Malignant Mesothelioma' (KP-MM)-1 to -16. In total, 3/16 antigens were identified using the results of previous SEREX analyses, and 13 were newly identified. Of these, KP-MM-8, which was subsequently identified as amyotrophic lateral sclerosis 2 chromosome region candidate 11, was shown to be tissue-restricted. Reverse transcription-polymerase chain reaction demonstrated KP-MM-8 to be expressed strongly only in the normal testis, and weakly in the spleen, prostate, ovary, heart and skeletal muscle. In addition, KP-MM-8 mRNA was identified in MM cell lines, and in various other cancer cell lines, including MM (3/4), lung cancer (5/7), melanoma (5/7) and liver cancer (5/5) cell lines. Additionally, 2/16 antigens (KP-MM-2 and KP-MM-6) exclusively reacted with sera from cancer patients. However, KP-MM-8 reacted with 1 of 8 MM sera. Notably, 8/8 patients with MM and 8/8 normal individuals exhibited antibodies reactive to KP-MM-5, which was identified as cell division cycle 25B, a known oncogene. Overall, this data suggests that KP-MM-8 may be considered as a cancer/testis-like antigen and KP-MM-5 as an immunogenic tumor antigen in MM patients.

Introduction

Malignant mesothelioma (MM) is a rare but rapidly fatal cancer that can develop from mesothelial cells in the pleura, peritoneum, pericardium and tunica vaginalis (1,2). The latency period of MM is exceptionally long and patients diagnosed with MM generally have poor prognoses (3), with an average life expectancy of less than a year, dependent on the tissue in which it originates (4,5). It has been reported that asbestos is the main carcinogen associated with MM (6,7); however, the molecular mechanism responsible for the pathogenesis of MM has not been elucidated.

Standard therapies for MM include chemotherapy, surgery and radiotherapy, but more effective strategies are required to improve the result of conventional therapeutic methods or cure this cancer. Immunotherapy may be a useful treatment (7,8); however, despite research progress, immunotherapy using tumor-associated antigens has been barely attempted in MM. Tumor antigens are classified into several categories, including differentiation antigens (9), mutated gene products (10), overexpressed oncogenes (11) and cancer/testis (CT) antigens (12,13). Of these, CT antigens are considered a promising tumor antigen group, since these proteins are expressed in the normal testis and tumor tissues (14,15).

Various methods have been devised to identify this group of tumor antigens. Serological analysis of recombinant tumor cDNA expression library (SEREX) is one such method, which has been shown to be powerful and effective (15). This technique screens cDNA expression libraries using the sera of cancer patients, and identifies cancer antigens recognized by autologous serum IgG (16). At present, >2,500 tumor antigens, including melanoma-associated antigen (MAGEA) (13), synaptonemal complex protein 1 (17), NY-ESO-1 (18), NY-SAR-35 (19) and KP-CoT-23 (20) have been identified using SEREX. Certain antigens, such as MAGE-A3 and NY-ESO-1, have been evaluated for immunogenicity and for their immunotherapeutic efficacies (21). In addition, CT antigens are expected to be utilized as biomarkers for diagnosis and prognosis (21,22).

In the present study, SEREX methodology was applied to define the spectrum of immunogenic proteins in MM, with

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focus on the CT-like antigens KP-MM-8 and KP-MM-5, which demonstrated high seroreactivity in patients with MM.

Materials and methods

Human sera and cell lines. Human sera from 8 healthy donors and 8 MM patients were obtained between February 2008 and December 2013, from the Department of Pathology and Environmental Health Center for Asbestos, Pusan National University Hospital (Yongsan, Korea) subsequent to diagnosis and staging. Human MM NCI-H226, NCI-H2452, NCI-H28 and MSTO-211H cell lines, human lung cancer A-427 and A549 cell lines, and the human melanoma cancer A375 cell line were obtained from the American Type Culture Collection (Manassas, VA, USA). Human colon cancer SNU-C1, SNU-C2A, SNU-C4 and SNU-C5 cell lines, human ovarian cancer SNU-8 and SNU-840 cell lines were obtained from the Korean Collection for Type Cultures (Jeongeup, Korea). All cell lines were maintained in RPMI-1640 medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum medium (Gibco; Thermo Fisher Scientific, Inc.), 2 mM L-glutamine, 100 units/ml penicillin and 100 µg/ml streptomycin. Cells were cultured at 37°C in a humidified 5% CO₂ atmosphere. The present study was performed with approval from the Ethical Committee of Pusan National University Yongsan Hospital (Yongsan, Korea).

Total RNA extraction from cell lines. Total RNA of human malignant mesothelioma NCI-H226, NCI-H2452, NCI-H28 and MSTO-211H cell lines, human lung cancer A-427 and A549 cell lines, human colon cancer SNU-C1, SNU-C2A, SNU-C4 and SNU-C5 cell lines and human ovarian cancer SNU-8 and SNU-840 cell lines were isolated using a RNA isolation kit (RNeasy Maxi kit; Qiagen, Hilden, Germany). Amounts of RNA isolated were assessed at 260 nm using a spectrophotometer (Ultrospec 2000; Pharmacia Biotech; GE Healthcare, Chicago, IL, USA). Total RNAs from: Human colon cancer HCT-15, SK-CO-1, SW-1116 and KM12/L4A cell lines; human lung cancer SK-LC-14, SK-LC-19, SK-LC-5, NCI-H23 and NCI-H740 cell lines; human melanoma cancer A375/C5, A375/P, A375/SM, SK-MEL-19, SK-MEL-128, SK-MEL-21 and SK-MEL-37 cell lines; human ovarian cancer OV-CAR-3, and SK-OV-3 cell lines; human breast cancer SK-OV-1, BT-474, MDA-MB-453, MCF-7, MDA-MB-231 and SK-BR-5 cell lines; human renal cancer SK-RC-18 and SK-RC-59 cell lines; human leukemia CEM/S and K-562 cell lines; human rhabdomyosarcoma A-204 cell lines; human thyroid cancer KAT-18 cell lines; and human liver cancer SK-HEP-1, SNU-354, SNU-423, SNU-449 and SNU-475 cell lines were obtained from the Ludwig Institute for Cancer Research (Memorial Sloan-Kettering Cancer Center, New York, NY, USA). Normal tissue total RNA was purchased from Clontech Laboratories, Inc. (Mountain View, CA, USA, cat. nos. 636742 and 636743).

Preparation of the cDNA library and sera. Poly(A)⁺ RNA from normal testis was purchased from Clontech Laboratories Inc. mRNA of the MSTO-211H cell line was extracted using

the PolyATtract mRNA Isolation System (Promega Corp., Madison, WI, USA). Poly(A)⁺ RNA (5 µg) was used to construct a cDNA library in ZAP Express vector (Stratagene; Agilent Technologies, Inc., Santa Clara, CA, USA), according to the manufacturer's instructions. The cDNA library contained approximately one million recombinants and was used for immunoscreening without amplification.

Sera from 8 healthy individuals and 8 patients with MM were diluted with 0.2% skimmed milk to 1:200. To remove serum antibodies that react with *Escherichia coli*/bacteriophage-associated antigens, sera were absorbed with *E. coli*/bacteriophage lysates, as previously described (20).

Immunoscreening of the cDNA library. Immunoscreening of the cDNA library was performed as described by Song *et al.* (20). Briefly, *E. coli* XL1 blue MRF cells (Stratagene; Agilent Technologies, Inc.) were transfected with the recombinant phages, plated at a density of ~5,000 pfu/150 mm plate (NZCYM-IPTG agar), and incubated for 8 h at 37°C. The cells were then transferred to nitrocellulose filters (PROTRAN BA 85; 0.45 µm; Schleicher & Schuell, Keene, NH, USA). The filters were incubated with patient sera, which had been preabsorbed with *E. coli*-phage lysate, at a dilution of 1:200, at room temperature overnight. The serum-reactive clones were detected using alkaline phosphatase-conjugated secondary antibody (Abcam, Cambridge, UK; cat. no., ab6859; dilution, 1:2,000) incubated for 1 h at room temperature and then visualized by incubation with 5-bromo-4-chloro-3-indolylphosphate/nitroblue tetrazolium. Subsequent to screening, isolated positive clones were removed from plates and preserved in suspension medium buffer with 25 µl chloroform.

Positive phages were mixed with a helper phage to co-infect XL-1 Blue MRF cells, and then rescued into pBlue-script phagemid forms by *in vivo* excision. Excised phagemids were transformed into host bacteria (XL0LR) to multiply for plasmid extraction and stock. The size of the cDNA inserted was determined by double restriction enzyme digestion with *EcoRI* and *XhoI*. The cDNA was sequenced commercially (Macrogen, Inc., Seoul, Korea).

Reverse transcription-polymerase chain reaction (RT-PCR). The cDNA preparations used as templates for RT-PCR reactions were prepared using 1 µg total RNA isolated from all cell lines and the Superscript First Strand Synthesis kit (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA). Primers were as follows: Amyotrophic lateral sclerosis 2 (juvenile) chromosome region, candidate 11 (ALS2CR11) forward, 5'-TCTGCAGGCTGGAGGTGGAA-3' and reverse, 5'-CCTTTCCCGGGTTGTTGCT-3'. The conditions used for RT-PCR were initial activation for 5 min at 94°C, 35 amplification cycles of denaturation for 30 sec at 94°C, and annealing for 30 sec at 63°C, extension for 1 min at 72°C, followed by a final elongation for 10 min at 72°C. A total of 20 µl of PCR product were loaded onto a 1.2% agarose gel and electrophoresis was performed at room temperature at 100 V 30 min using 1X Tris-acetate-EDTA buffer. The gels were stained with ethidium bromide solution diluted with distilled water to a ratio of 2:10⁴ for 10 min, and washed with H₂O. Data were analyzed by Gel Doc™ + Imager with Image Lab™ software v.5.2.1 (Bio-Rad, Richmond, CA, USA).

Table I. Malignant mesothelioma antigens identified by SEREX.

Antigen	Gene name	UniGene cluster ^a	Library source	Serum source ^b	Redundancy, n	Previously identified by SEREX ^c
KP-MM-1	IFT74	Hs.145402	Testis	M4	1	No
KP-MM-2	TMF1	Hs.267632	Testis	M4	2	Yes
KP-MM-3	BRAP	Hs.530940	Testis	M4	1	Yes
KP-MM-4	RPLP0	Hs.546285	MSTO-211H	M8	1	Yes
KP-MM-5	CDC25B	Hs.153752	Testis	M8	2	No
KP-MM-6	ZNF429	Hs.572567	Testis	M49	1	No
KP-MM-7	ZNF320	Hs.369632	Testis	M49	1	No
KP-MM-8	ALS2CR11	Hs.335788	Testis	M8	1	No
KP-MM-9	DNAJC2	Hs.558476	Testis	M8	1	No
KP-MM-10	NUB1	Hs.647082	Testis	M10	1	No
KP-MM-11	CCDC11	Hs.658630	Testis	M1	2	No
KP-MM-12	ZNF93	Hs.723768	Testis	M49	1	No
KP-MM-13	ZNF234	Hs.235992	Testis	M49	1	No
KP-MM-14	ZNF595	Hs.709469	Testis	M49	1	No
KP-MM-15	CCDC40	Hs.202542	Testis	M49	1	No
KP-MM-16	ZNF85	Hs.37138	Testis	M49	1	No

^aUniGene cluster of isolated antigens (<http://www.ncbi.nlm.nih.gov/>). ^bSera from malignant mesothelioma patients: M1, a patient diagnosed with malignant mesothelioma; M4, a patient diagnosed with malignant peritoneal mesothelioma; M8, a patient diagnosed with malignant pleural mesothelioma; M10, a patient diagnosed with malignant pleural mesothelioma; and M49, a patient diagnosed with malignant peritoneal mesothelioma. ^cSequences were compared with those in the SEREX database of the Ludwig Institute for Cancer Research ([http://ludwigsun5.unil.ch/CancerImmuno meDB/](http://ludwigsun5.unil.ch/CancerImmuno%20meDB/)). 'Yes' and 'No' indicate whether the antigen matched or did not match an antigen in the SEREX database, respectively.

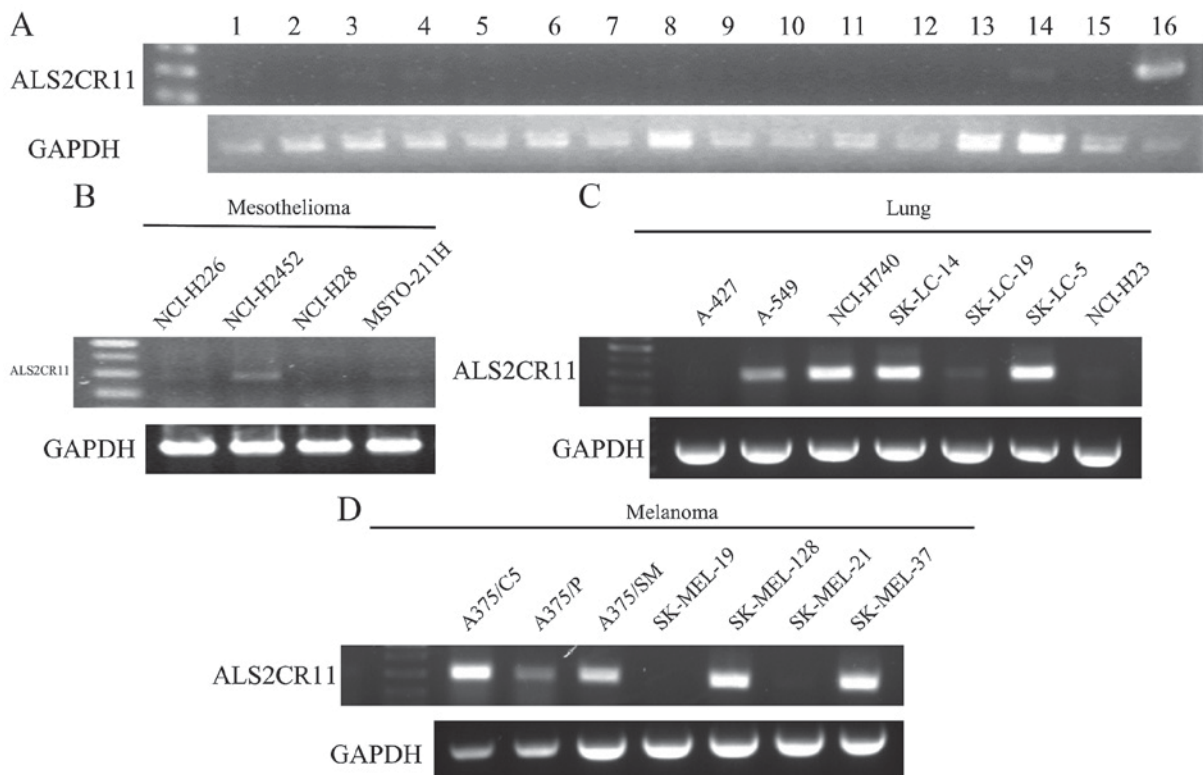


Figure 1. Reverse transcription-polymerase chain reaction analysis of ALS2CR11 expression in (A) normal tissues, as follows: 1, spleen; 2, thymus; 3, prostate; 4, ovary; 5, small intestine; 6, colon; 7, leukocyte; 8, heart; 9, brain; 10, placenta; 11, lung; 12, pancreas; 13, liver; 14, skeletal muscle; 15, kidney; and 16, testis. ALS2CR11 expression is also shown in (B) mesothelioma cancer, (C) lung cancer and (D) melanoma cell lines. cDNA templates were normalized using GAPDH as shown in the bottom panel. ALS2CR11, amyotrophic lateral sclerosis 2 (juvenile) chromosome region, candidate 11.

Seroreactivity by plaque assay. Selected immunoreactive clones were tested for reactivity against sera from individual MM patients and healthy donors using a plaque assay. λ -ZAP clone without an insert was co-plated and included in each assay as a negative control. KP-MM-5 was tested for reactivity against 10-fold serially diluted serum (1:10³ to 1:10⁶) using the same plaque assay.

Results

Identification of MM antigens by SEREX. To identify tumor antigens in MM, immunoscreening of a testis cDNA library and MSTO-211H cDNA library was performed using the sera of patients with MM. In total, 31 immunogenic clones were identified in this manner, and 16 of the 31 genes were verified as tumor antigen candidates and termed KP-MM-1 to KP-MM-16 (Table I). Of the 16, 3 were previously identified by SEREX in other tumor types, and 13 antigens did not match entries in the cancer immunome database, and were therefore considered new antigens.

KP-MM-8 as a CT-like antigen. Preliminary *in silico* mRNA expression profile analysis and characterization of the gene products identified in the present study were undertaken based on the tissue distributions of expressed sequence tags and serial analysis of gene expression tags in the Cancer Genome Anatomy Database (<http://cgap.nci.nih.gov/>) and the information contained in the GeneCards database (<http://www.genecards.org/>). One antigen, KP-MM-8, was identified as a CT-like antigen. KP-MM-8 was identified as ALS2CR11 by a nucleotide BLAST search.

Conventional RT-PCR was performed using mRNA from normal tissues to examine the expression of ALS2CR11 (Fig. 1A). RT-PCR showed that ALS2CR11 was strongly expressed only in the normal testis, and weakly in the spleen, prostate, ovary, heart and skeletal muscle. However, the expression in various cancer cell lines was relatively high. The expression of this antigen was positive in 3 of 4 MM cell lines, consisting of the NCI-H226, NCI-H2452 and MSTO-211H cell lines (Fig. 1B). In addition, it was expressed in various cancer cell lines, including lung cancer (5/7) (Fig. 1C), melanoma (5/7) (Fig. 1D) and liver cancer (5/5) cell lines, but was not expressed in colon cancer, renal cancer or leukemia cell lines (Table II). This result suggests that ALS2CR11 may be a putative CT like antigen.

Seroreactivity of isolated mesothelioma antigens. Seroreactivity testing of 7 phage clones, consisting of KP-MM-2, KP-MM-5, KP-MM-6, KP-MM-7, KP-MM-8, KP-MM-9 and KP-MM-12, was performed using the sera of 8 MM patients and 8 healthy donors by phage plaque assay (Fig. 2). The CT-like antigen KP-MM-8 was only positive in one MM serum (M8), and negative in all other 7 MM sera and to all normal sera. In total, 2 of the 8 MM patients had antibodies reactive to KP-MM-6-containing phagemids, whereas the 8 normal sera showed no reactivity against KP-MM-6 (Table III). Notably, all MM patients and normal individuals showed high reactivity to the KP-MM-5 antigen. Representative phage plaque assay for KP-MM-5 from the sera of M8 and normal individual N1, showed high-titer reactivity

Table II. Summary of ALS2CR11 mRNA expression as determined by reverse transcription-polymerase chain reaction.

Cancer cell line	Frequency, n
Mesothelioma	3/4
Colon cancer	0/8
Lung cancer	5/7
Melanoma	5/7
Ovarian cancer	2/4
Breast cancer	1/6
Renal cancer	0/2
Leukemia	0/2
Rhabdomyosarcoma	1/1
Thyroid cancer	1/1
Liver cancer	5/5

ALS2CR11, amyotrophic lateral sclerosis 2 (juvenile) chromosome region, candidate 11.

Table III. Detection of selected antigens against sera obtained from malignant mesothelioma patients and normal individuals.

KP-MM-antigen	No. of cases	
	Mesothelioma	Normal
KP-MM-2	3/8	0/8
KP-MM-5	8/8	8/8
KP-MM-6	2/8	0/8
KP-MM-7	1/8	0/8
KP-MM-8	1/8	0/8
KP-MM-9	2/8	2/8
KP-MM-12	1/8	0/8

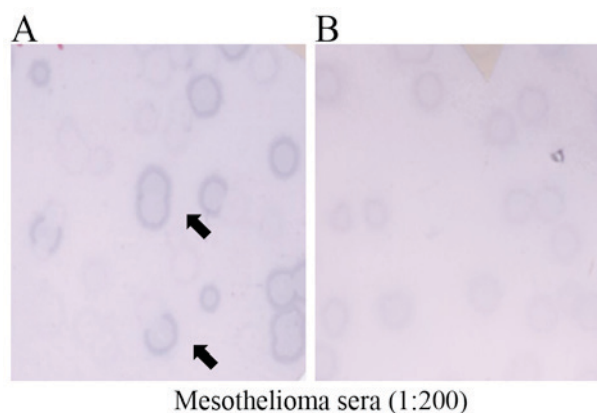


Figure 2. Representative phage plaque assays of the sera of the mesothelioma patients (A) M49 and (B) M11 for KP-MM-7. λ -ZAP phages without insert were mixed with KP-MM-7 clones and acted as internal negative controls. Mixtures were transfected into *E. coli*, and plaques were blotted onto nitrocellulose membranes and incubated with the sera of mesothelioma patients (dilution, 1:200). Spots were evaluated as (A) positive when clones were clearly distinguishable from control phage and as (B) negative if not. Arrows indicate positive clones.

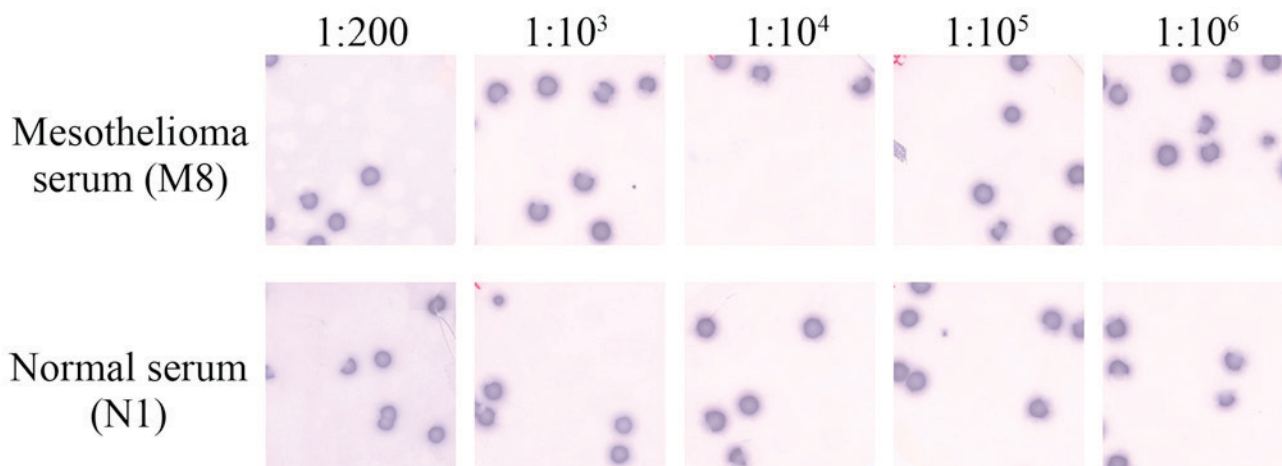


Figure 3. Seroreactivity of the malignant mesothelioma patient M8 and the normal healthy individual N1 for KP-MM-5 (CDC25B). Serum dilutions are indicated. λ -ZAP phages without an insert were mixed with test clones and acted as internal negative controls; these are visible as a background to positive clones. Assays were scored positive only if test clones were clearly distinguishable from control phages.

(1:10⁶) (Fig. 3). The presence of consistently high-titer antibodies against KP-MM-5 suggested it to be an immunogenic tumor antigen in MM patients. However, normal sera also showed extremely strong reactivity for KP-MM-5; this observation requires further investigation.

Discussion

Previously, it was reported that several major CT antigens are expressed in MM (23); however, it is theorized that numerous tumor antigens remain unidentified, and that even identified tumor antigens require further functional study. To identify new CT antigens, MM was studied using the SEREX technique, and as a result, 31 sequences were isolated and 16 antigens were identified as independent tumor-specific antigens.

Notably, 6 of the 16 identified genes (KP-MM-6, KP-MM-7, KP-MM-12, KP-MM-13, KP-MM-14 and KP-MM-19) belonged to the zinc finger gene family. Associations between the ZNF gene and cancer properties are not well understood, although it has been reported that certain ZNF genes have potential as biological markers of cancer (24). Thus, additional investigation of the functional roles of ZNF gene during the pathogenesis of cancers, including MM, is required.

KP-MM-8 was identified in a testis library that reacted to mesothelioma serum, and BLAST search identified this gene as ALS2CR11, a member of the ALS2CR multigene family, which is associated with amyotrophic lateral sclerosis (25). Nevertheless, the present study identifies the ALS2CR11 gene as putative CT antigen, and little is known about the contribution made by this gene to carcinogenesis.

In the present study, ALS2CR11 was found to have CT-like antigen features, particularly a restricted mRNA expression pattern. Considering the mRNA expression profile of ALS2CR11 in the GeneCards database and the present RT-PCR results, ALS2CR11 appears to be expressed in various cancer cell lines and in few normal tissues, which is a noteworthy feature of CT antigens.

However, seroreactivity testing showed KP-MM-8 reacted with only 1 MM serum, although this result was based on a limited number of patient sera samples. Additional investigation is required with more samples to determine whether ALS2CR11 functions as a CT antigen, and to identify the molecular basis for its contribution to tumorigenesis.

KP-MM-5 was identified in a testis library that reacted with mesothelioma serum. This clone was identified as cell division cycle (CDC) 25B, and CDC25 phosphatases activate cyclin-dependent kinases (CDKs) and targets of the checkpoint kinase (CHK) 1/CHK2-mediated checkpoint pathway (26). Numerous studies have been conducted on members of the CDC25 protein phosphatase family, and it has been reported that CDC25B plays an important role in tumorigenesis (27,28). Specifically, CDC25B activates M phase promoting factor and promotes G2/M transition, and the overexpression of CDC25B contributes to excessive cell division (29).

Presence of high-titer reactive antibodies against KP-MM-5 suggests that KP-MM-5 is an immunogenic tumor antigen in MM patients. However, normal sera showed strong reactivity for KP-MM-5. This result requires additional investigation, and suggests that numerous aspects of the actions of CDC25B have yet to be identified.

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References

1. Raptopoulos V: Peritoneal mesothelioma. *Crit Rev Diagn Imaging* 24: 293-328, 1985.
2. Cunha P, Luz Z, Seves I, Sousa C, Skiappa, Ribeiro L, Marques C and Oliveira M: Malignant peritoneal mesothelioma-diagnostic and therapeutic difficulties. *Acta Med Port* 15: 383-386, 2002 (In Portuguese).
3. Ahmed I, Ahmed Tipu S and Ishtiaq S: Malignant mesothelioma. *Pak J Med Sci* 29: 1433-1438, 2013.

4. Røe OD and Stella GM: Malignant pleural mesothelioma: History, controversy and future of a manmade epidemic. *Eur Respir Rev* 24: 115-131, 2015.
5. Mirabelli D, Roberti S, Gangemi M, Rosato R, Ricceri F, Merler E, Gennaro V, Mangone L, Gorini G, Pascucci C, *et al*: Survival of peritoneal malignant mesothelioma in Italy: A population-based study. *Int J Cancer* 124: 194-200, 2009.
6. Rudd RM: Malignant mesothelioma. *Br Med Bull* 93: 105-123, 2010.
7. Robinson BW and Lake RA: Advances in malignant mesothelioma. *N Engl J Med* 353: 1591-1603, 2005.
8. Kondola S, Manners D and Nowak AK: Malignant pleural mesothelioma: An update on diagnosis and treatment options. *Ther Adv Respir Dis* 10: 275-288, 2016.
9. Coulie PG, Brichard V, Van Pel A, Wölfel T, Schneider J, Traversari C, Mattei S, De Plaen E, Lurquin C, Szikora JP, *et al*: A new gene coding for a differentiation antigen recognized by autologous cytolytic T lymphocytes on HLA-A2 melanomas. *J Exp Med* 180: 35-42, 1994.
10. Labrecque S, Naor N, Thomson D and Matlashewski G: Analysis of the anti-p53 antibody response in cancer patients. *Cancer Res* 53: 3468-3471, 1993.
11. Disis ML, Calenoff E, McLaughlin G, Murphy AE, Chen W, Groner B, Jeschke M, Lydon N, McGlynn E, Livingston RB, *et al*: Existent T-cell and antibody immunity to HER-2/neu protein in patients with breast cancer. *Cancer Res* 54: 16-20, 1994.
12. Boon T, Coulie PG and Van den Eynde B: Tumor antigens recognized by T cells. *Immunol Today* 18: 267-268, 1997.
13. Chen YT, Güre AO, Tsang S, Stockert E, Jäger E, Knuth A and Old LJ: Identification of multiple cancer/testis antigens by allogeneic antibody screening of a melanoma cell line library. *Proc Natl Acad Sci USA* 95: 6919-6923, 1998.
14. van der Bruggen P, Traversari C, Chomez P, Lurquin C, De Plaen E, Van den Eynde BJ, Knuth A and Boon T: A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. *J Immunol* 178: 2617-2621, 2007.
15. Caballero OL and Chen YT: Cancer/testis (CT) antigens: Potential targets for immunotherapy. *Cancer Sci* 100: 2014-2021, 2009.
16. Sahin U, Türeci O, Schmitt H, Cochlovius B, Johannes T, Schmits R, Stenner F, Luo G, Schobert I and Pfreundschuh M: Human neoplasms elicit multiple specific immune responses in the autologous host. *Proc Natl Acad Sci USA* 92: 11810-11813, 1995.
17. Türeci O, Sahin U, Zwick C, Koslowski M, Seitz G and Pfreundschuh M: Identification of a meiosis-specific protein as a member of the class of cancer/testis antigens. *Proc Natl Acad Sci USA* 95: 5211-5216, 1998.
18. Chen YT, Scanlan MJ, Sahin U, Türeci O, Güre AO, Tsang S, Williamson B, Stockert E, Pfreundschuh M and Old LJ: A testicular antigen aberrantly expressed in human cancers detected by autologous antibody screening. *Proc Natl Acad Sci USA* 94: 1914-1918, 1997.
19. Lee SY, Obata Y, Yoshida M, Stockert E, Williamson B, Jungbluth AA, Chen YT, Old LJ and Scanlan MJ: Immunomic analysis of human sarcoma. *Proc Natl Acad Sci USA* 100: 2651-2656, 2003.
20. Song MH, Ha JM, Shin DH, Lee CH, Old L and Lee SY: KP-CoT-23 (CCDC83) is a novel immunogenic cancer/testis antigen in colon cancer. *Int J Oncol* 41: 1820-1826, 2012.
21. Suri A, Jagadish N, Saini S and Gupta N: Targeting cancer testis antigens for biomarkers and immunotherapy in colorectal cancer: Current status and challenges. *World J Gastrointest Oncol* 7: 492-502, 2015.
22. Grah JJ, Katalinic D, Juretic A, Santek F and Samarzija M: Clinical significance of immunohistochemical expression of cancer/testis tumor-associated antigens (MAGE-A1, MAGE-A3/4, NY-ESO-1) in patients with non-small cell lung cancer. *Tumori* 100: 60-68, 2014.
23. Sigalotti L, Coral S, Altomonte M, Natali L, Gaudino G, Cacciotti P, Libener R, Colizzi F, Vianale G, Martini F, *et al*: Cancer testis antigens expression in mesothelioma: Role of DNA methylation and bioimmunotherapeutic implications. *Br J Cancer* 86: 979-982, 2002.
24. Gaykalova DA, Vatapalli R, Wei Y, Tsai HL, Wang H, Zhang C, Hennessey PT, Guo T, Tan M, Li R, *et al*: Outlier analysis defines zinc finger gene family DNA methylation in tumors and saliva of head and neck cancer patients. *PLoS One* 10: e0142148, 2015.
25. Hentati A, Bejaoui K, Pericak-Vance MA, Hentati F, Speer MC, Hung WY, Figlewicz DA, Haines J, Rimmler J, Ben Hamida C, *et al*: Linkage of recessive familial amyotrophic lateral sclerosis to chromosome 2q33-q35. *Nat Genet* 7: 425-428, 1994.
26. Kiyokawa H and Ray D: In vivo roles of CDC25 phosphatases: Biological insight into the anti-cancer therapeutic targets. *Anticancer Agents Med Chem* 8: 832-836, 2008.
27. Galaktionov K, Lee AK, Eckstein J, Draetta G, Meckler J, Loda M and Beach D: CDC25 phosphatases as potential human oncogenes. *Science* 269: 1575-1577, 1995.
28. Kristjánssdóttir K and Rudolph J: Cdc25 phosphatases and cancer. *Chem Biol* 11: 1043-1051, 2004.
29. Kim J, Singh AK, Takata Y, Lin K, Shen J, Lu Y, Kerényi MA, Orkin SH and Chen T: LSD1 is essential for oocyte meiotic progression by regulating CDC25B expression in mice. *Nat Commun* 6: 10116, 2015.