Do Genetic Markers of Inflammation Modify the Relationship between Periodontitis and Nonalcoholic Fatty Liver Disease? Findings from the SHIP Study

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Abstract

An association between periodontitis and nonalcoholic fatty liver disease (NAFLD) has been reported by experimental animal and epidemiologic studies. This study investigated whether circulating levels of serum C-reactive protein (CRP) and a weighted genetic CRP score representing markers of inflammatory burden modify the association between periodontitis and NAFLD. Data came from 2,481 participants of the Study of Health in Pomerania who attended baseline examination that occurred between 1997 and 2001. Periodontitis was defined as the percentage of sites (0%, <30%, \geq 30%) with probing pocket depth (PD) \geq 4 mm, and NAFLD status was determined using liver ultrasound assessment. Serum CRP levels were assayed at a central laboratory, and single-nucleotide polymorphisms previously identified through genome-wide association studies as robustly associated with serum CRP were combined into a weighted genetic CRP score (wGS_{CRP}). Logistic regression models estimated the association between periodontitis and NAFLD within strata of serum CRP and separately within strata of the wGS_{CRP}. The prevalence of NAFLD was 26.4% (95% confidence interval [CI], 24.6, 28.1) while 17.8% (95% CI, 16.0–19.6) had \geq 30% of sites with PD \geq 4 mm. Whereas the wGS_{CRP} was not a modifier ($P_{interaction} = 0.8$) on the multiplicative scale, serum CRP modified the relationship between periodontitis and NAFLD ($P_{interaction} = 0.01$). The covariate-adjusted prevalence odds ratio of NAFLD comparing participants with \geq 30% of sites with PD \geq 4 mm to those with no site affected was 2.39 (95% CI, 1.32–4.31) among participants with serum CRP <1 mg/L. The corresponding estimate was 0.97 (95% CI, 0.57–1.66) for participants with serum CRP levels of 1 to 3 mg/L and 1.12 (95% CI, 0.65–1.93) for participants with serum CRP levels.

Keywords: periodontal disease, epidemiology, genetic score, C-reactive protein, inflammatory mediator, hepatic steatosis

A supplemental appendix to this article is available online.

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Introduction

In response to an injury or infection, inflammation occurs as a complex series of short-term adaptive responses accompanied by local tissue damage with manifestations that gradually resolves as inflammation abates, leaving little to no permanent damage (Kumar et al. 2014). Inflammation is regulated primarily by the innate immune system (Takashiba and Naruishi 2006; Kumar et al. 2014), and it involves a coordinated cascade of biological events regulated by specific cells and molecular signals (Naitza et al. 2012).

An excessive inflammatory response that occurs upon stimulation of the innate immune system has been described as a hyperresponsive trait (Shaddox et al. 2010) that presents systemically as a heightened expression of systemic markers of inflammation (Southerland et al. 2006). Population-based genetic studies suggest that natural selection has shaped the evolution of the innate immunity with specific focus on inflammatory genes that are pivotal in host-pathogen interactions (Barreiro and Quintana-Murci 2010). Inflammatory biomarkers are reported to be highly heritable, with studies among ethnically homogeneous groups and twins indicating that about half of interindividual variability in markers of inflammation is genetically determined (Pankow et al. 2001; Dupuis et al. 2005). For instance, the Framingham Heart Study reported ageand sex-adjusted heritability of 25.3%, 25.4%, and 45.2% for C-reactive protein (CRP), interleukin 6 (IL-6), and monocyte chemoattractant protein 1 (MCP-1), respectively (Dupuis et al. 2005), while the National Heart Lung and Blood Institute (NHLBI) Family Heart Study reported heritability ranging from 35% to 40% for CRP, white blood cells, and albumin (Pankow et al. 2001; Dupuis et al. 2005).

Findings among humans and from mice models (Yoneda et al. 2012) suggest a relationship between periodontitis and nonalcoholic fatty liver disease (NAFLD), both of which are chronic health conditions characterized by a heightened inflammatory burden (Day and James 1998; Haukeland et al. 2006; Targher 2006; Tilg and Moschen 2010; Schenkein and Loos 2013; Gocke et al. 2014). Indeed, individuals with periodontitis present with frequent bacteremia (Schenkein and Loos 2013) that promotes a proinflammatory state, while obesity, a precursor for NAFLD, is characterized by a state of chronic low-grade systemic inflammation (Shoelson et al. 2006; Ouchi et al. 2011).

Genome-wide association studies (GWAS) have identified 19 independent loci that are robustly associated with CRP levels (Dehghan et al. 2011; Naitza et al. 2012), an acute phase reactant, and a marker of systemic inflammation (Pearson et al. 2003; Raman et al. 2013). Because genetic determinants of inflammatory biomarkers can more accurately indicate lifelong inflammatory status (Raman et al. 2013) compared to biomarker concentrations obtained at a given point in time, polymorphisms in genes regulating inflammatory processes may influence the expression of periodontitis and NAFLD, as well as modify the relationship between these 2 conditions.

The purpose of this investigation was to determine whether CRP-associated genetic loci and serum CRP levels modify the association between periodontitis and NAFLD.

Methods

Data Source

The Study of Health in Pomerania (SHIP) is a populationbased cohort sampled from the Western Pomeranian region of northeastern Germany (John et al. 2001; Volzke et al. 2011). The SHIP was designed to provide prevalence estimates for various diseases and disease risk factors, incidence of common risk factors, subclinical disorders, and clinical diseases and to evaluate associations among these factors. From eligible inhabitants of West Pomerania in 1996, 6,265 adults aged 20 to 79 y were invited to participate. A total of 4,308 participated in the baseline examination conducted between 1997 and 2001. Study participants underwent rigorous examinations, and interviewer-administered questionnaires were used to collect information on relevant covariates. Ethics approval for this study was obtained from the institutional review board of the University of North Carolina at Chapel Hill, and this study conformed to the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) guidelines for reporting observational studies.

Exposure Assessment and Definition

Dental examiners performed periodontal examination on study participants with no medical contraindication. Measurements of probing pocket depth (PD) and clinical attachment level (CAL) were obtained on 4 sites per tooth: distobuccal, mesiobuccal, midbuccal, and midlingual or midpalatal (except the third molars) on 2 quadrants (quadrants 1 and 4 or quadrants 2 and 3). For this investigation, periodontitis was defined as the proportion of periodontal sites with PD \geq 4 mm categorized as none (0%), moderate (<30%), and extensive ($\geq30\%$). In a secondary analysis, periodontitis was investigated as the proportion of sites with CAL \geq 3 mm (none, moderate, and extensive) and according to the Centers for Disease Control and Prevention-American Association for Periodontology (CDC-AAP) criteria that define severe periodontitis as ≥ 2 interproximal sites with CAL of ≥ 6 mm (not on the same tooth) and ≥ 1 interproximal sites with PD of ≥5 mm, and moderate periodontitis as ≥ 2 interproximal sites (not on the same tooth) with CAL of ≥ 4 mm or ≥ 2 interproximal sites (not on the same tooth) with PD of ≥5 mm (Page and Eke 2007). Individuals with moderate or severe periodontitis were categorized as having periodontitis, while participants not meeting these criteria were categorized as healthy/mild.

Outcome Assessment and Definition

Trained physicians performed liver ultrasound on study participants using a 7.5-MHz transducer (John et al. 2001; Volzke et al. 2011). A positive finding on ultrasound was defined as a significant increase in liver echogenicity (brightness) relative to the kidneys, with the diaphragm indistinct or the echogenic walls of the portal veins invisible (Baumeister et al. 2008; Williams et al. 2011).

Covariates

Covariates identified as confounders were determined after analyzing a directed acyclic graph (Greenland et al. 1999) and included age modeled with a quadratic term, sex, alcohol consumption in the past 30 days, and waist circumference, dichotomized at \geq 88 cm for women and \geq 102 cm for men and is indicative of abdominal obesity (Grundy et al. 2005). Diabetes was based on self-reported physician's diagnosis or study's measurement of hemoglobin A1c (HbA1c) \geq 6.5%. Selfreported smoking status was categorized as never, former, and current. Physical activity was based on self-report of the number of hours per week of moderate physical activity.

Laboratory Measurements

Nonfasting blood samples were drawn from the cubital vein in the supine position. HbA1c was measured by high-performance liquid chromatography (ClinRep HbA1C, Recipe Chemicals Instruments GmbH), while serum levels of high-sensitivity CRP (hs-CRP) were estimated with the Behring Nephelometer II (Dade Behring) (Gocke et al. 2014).

Genotyping

Genomic DNA from blood samples was collected using standardized procedures. Blood aliquots were immediately placed on ice after collection and stored at -80° C in a biobank (John et al. 2001). A total of 4,096 samples were genotyped using the Human SNP 6.0 Array (Affymetrix) with overall genotyping efficiency of 98.6% and imputed to the 1000 Genomes v3 reference panel released March 2012 (ALL ancestries panel, build 37) (Volzke et al. 2011; Teumer et al. 2013).

A total of 19 CRP single-nucleotide polymorphisms (SNPs) from 19 loci, 1 IL-6 SNP, 2 MCP-1 SNPs, and 2 erythrocyte sedimentation rate (ESR) SNPs were identified as genetic markers of inflammation for this investigation. After quality control analysis, genotype data were available for 4,070 participants. None of the SNPs deviated significantly from Hardy-Weinberg equilibrium (P > 0.003), and call rates were >95% for all SNPs.

CRP-Specific Weighted Genetic Score

To study the cumulative effect of multiple gene loci, a weighted genetic score for CRP was computed for each study participant. A risk allele was defined as the allele associated with a unit increase in log-transformed serum CRP level. The corresponding effect sizes from previous GWAS of inflammatory mediators (Naitza et al. 2012) and a meta-analysis of GWAS of CRP levels (Dehghan et al. 2011) were used to weigh the contribution of each risk allele within the weighted genetic CRP score (wGS_{CRP}) as previously described (Pharoah et al. 2008) and implemented (Thanassoulis et al. 2012; Xiao et al. 2015). The lead SNP in each identified locus were used in creating the weighted genetic score. The list of SNPs and effect sizes

(β estimates) are presented in Appendix Table 1. The weighted genetic score for CRP was calculated as follows:

$$wGS_{CRP} = \alpha_1 x_1 + \alpha_2 x_2 + \dots \alpha_k x_k + \alpha_n x_n,$$

where α_k is the per-allele β estimate associated with the risk allele for CRP SNP *k*, and X_k is the number of risk alleles for the same SNP, and *n* is total number of SNPs used in creating the genetic score. In addition to CRP, weighted genetic scores were also created for IL-6, MCP-1, and ESR.

Exclusions

From 4,308 eligible participants at baseline, participants with no genotype data (n = 238) were excluded. Also excluded were participants who reported excessive alcohol consumption, defined as 70 g of ethanol/wk (equivalent to 1 standard drink/ day) for women and 140 g of ethanol/wk (equivalent to 2 standard drinks/day) for men (n = 970). Participants with no liver ultrasound reading (n = 48), as well as those with no periodontal examination (n = 14) or edentulous (n = 563), were also excluded. Also excluded were participants with chronic or autoimmune viral hepatitis as well as those who self-reported use of the following steatosis-promoting medications: tamoxifene, methotrexate, and amiodarone. The resulting analytic sample size was 2,481, noting that some participants were ineligible for multiple reasons.

Statistical Analysis

The respective genetic scores, including the wGS_{CRP} and logtransformed serum CRP levels, were modeled as continuous traits in separate linear regression models investigating whether these traits were associated with periodontitis and NAFLD.

Logistic regression models stratified according to the median value for the wGS_{CRP} (<1.98 vs. \geq 1.98) assessed the relationship between periodontitis and NAFLD. In separate stratified analyses, a similar association was investigated within strata of low (<1 mg/L), intermediate (1 to 3 mg/L), and high (>3 mg/L) (Pearson et al. 2003) serum CRP levels. Statistical tests were 2-sided, and the test for statistical significant interaction was set a priori at P < 0.1. Data analysis was conducted in SAS version 9.4 (SAS Institute).

Results

The prevalence of NAFLD was 26.4% (95% confidence interval [CI], 24.6%–28.1%), and 52.7% (95% CI, 50.5%–54.8%) had <30% of sites with PD ≥4 mm while 17.8% (95% CI, 16.0%–19.6%) of the SHIP examinees had ≥30% of sites with PD ≥4 mm. Participants with periodontitis (PD ≥4 mm) and those with NAFLD had higher serum CRP levels compared to their counterparts with a healthy periodontium and without NAFLD. The median serum CRPs were 1.48 mg/L (interquartile range [IQR], 0.68–3.01) and 1.82 mg/L (IQR, 0.82–3.90) for participants with <30% and ≥30% of sites with PD ≥4 mm,

		Proportion of Sites with PD \ge 4 mm			NAFLD	
	Total (n = 2,481)	None (0%) (n = 733)	Moderate (<30%) (n = 1,307)	Extensive (≥30%) (n = 441)	No (n = 1,827)	Yes (n = 654)
CRP, mg/L	1.42 (0.64, 3.01)	1.11 (0.51, 2.95)	1.48 (0.68, 3.01)	1.82 (0.82, 3.90)	1.20 (0.55, 2.95)	2.16 (0.99, 4.41)
Age, y	47.0 (34.0, 60.0)	36.0 (27.0, 56.0)	47.0 (35.0, 60.0)	55.0 (46.0, 65.0)	42.0 (31.0, 57.0)	56.0 (46.0, 65.0)
Waist circumference, cm	88.0 (77.2, 98.1)	83.0 (73.0, 94.0)	89.0 (78.0, 98.0)	94.0 (84.5, 103)	84.0 (74.3, 93.8)	98.5 (90.8, 106)
Alcohol ^a	4.10 (1.31, 6.86)	4.56 (1.39, 6.71)	4.09 (1.31, 7.17)	3.92 (0.65, 6.41)	4.08 (1.31, 6.86)	4.28 (0.83, 6.86)
Abdominal obesity ^b	747 (30.1)	170 (23.2)	389 (29.8)	188 (42.6)	383 (21.0)	364 (55.7)
Physical activity	1,099 (44.5)	388 (53.0)	586 (45.0)	125 (28.5)	881 (48.5)	218 (33.3)
Men	1,116 (45.0)	281 (38.3)	592 (45.3)	243 (55.1)	740 (40.5)	376 (57.5)
Smoking		. ,	. ,			. ,
Never	966 (39.1)	323 (44.1)	515 (39.6)	128 (29.2)	717 (39.4)	249 (38.1)
Former	780 (31.6)	203 (27.7)	434 (33.4)	143 (32.7)	527 (29.0)	253 (38.7)
Current	726 (29.4)	207 (28.2)	352 (27.1)	167 (38.I)	574 (31.6)	152 (23.2)
Diabetes mellitus	465 (18.7)	98 (13.4)	248 (19.0)	119 (27.0)	226 (12.4)	239 (36.5)
NAFLD	654 (26.4)	133 (18.1)	348 (26.6)	173 (39.2)		

Table 1. Distribution of Baseline Factors According to the Proportion of Sites with PD \geq 4 mm and Nonalcoholic Fatty Liver Disease Status amongParticipants of the Study of Health in Pomerania, 1997 to 2001.

Data are presented as No. (%) or median (lower quartile, upper quartile).

CDC-AAP, Centers for Disease Control and Prevention–American Academy of Periodontology; CRP, C-reactive protein; NAFLD, nonalcoholic fatty liver disease; PD, pocket depth.

^aAverage number of standard drinks in the past 30 days.

^bSex-specific weight circumference defined as ≥88 cm for women and ≥102 cm for men (Grundy et al. 2005).

respectively, and 1.11 mg/L (IQR, 0.51–2.95) for participants with no site with PD \geq 4 mm (Table 1). Participants with NAFLD were less likely to report physical activity (33% vs. 49%), equally as likely to consume alcohol, and more likely to be men (58% vs. 41%) than participants without NAFLD (Table 1).

Appendix Table 1 shows the full list of SNPs with the corresponding effect sizes used in creating the respective weighted genetic scores. Most of the CRP SNPs were positively associated with serum CRP levels in this study population (Appendix Fig. 1), and the wGS_{CRP} aligns well with serum CRP levels such that participants with a low genetic CRP score had lower mean serum CRP levels compared to participants with a high genetic CRP score (Fig. 1). The median value for the wGS_{CRP} was 1.98 (IQR, 1.82-2.14). Participants with wGS_{CRP} above the median had a mean (standard deviation [SD]) serum CRP of 3.24 (4.72) mg/L, while participants with wGS_{CRP} at or below

the median had a mean (SD) serum CRP of 2.65 (6.62) mg/L.

As expected, serum CRP was associated with periodontitis. Specifically, each unit increase in log-transformed serum CRP was associated with a 23% increase in adjusted prevalence odds of having \geq 30% sites with PD \geq 4 mm, prevalence odds ratio [POR] = 1.23 (95% CI, 1.09–1.39), while the corresponding estimate for having <30% of sites with PD \geq 4 mm was POR = 1.16 (95% CI, 1.06–1.26) (Table 2). There was no meaningful association between individual CRP SNPs and periodontitis (Appendix



Figure 1. Mean serum C-reactive protein (CPR) levels (right vertical axis) shown as solid black line for categories of the weighted genetic CRP score. The shaded bars show the distribution of the weighted CRP score in the study population (left vertical axis). hs, high sensitivity.

Figs. 2–4). Even after combining into a score, the wGS_{CRP} was not associated with the prevalence odds of having <30% sites or \geq 30% sites with PD \geq 4 mm, POR = 1.02 (95% CI, 0.69–1.51) and 1.01 (95% CI, 0.61–1.69), respectively (Table 2).

As was observed for periodontitis, each unit increase in logtransformed serum CRP was associated with a higher prevalence odds of NAFLD, with adjusted POR = 1.26 (95% CI, 1.14-1.39) (Table 2). Likewise, most of the individual CRP SNPs had no meaningful effect on the prevalence odds of

PD ≥4 mm	Prevalence Odds Ratio (95% Sites with PD ≥4 mr	6 Cl) for the Proportion of n (Reference = 0%)	Prevalence Odds Ratio (95% CI) for NAFLD		
	Crude	Adjusted ^a	Crude	Adjusted ^a	
Serum CRP ^b					
<30%	1.19 (1.10–1.29)	1.16 (1.06–1.26)	1.49 (1.37–1.61)	1.26 (1.14–1.39)	
≥30%	1.40 (1.26–1.56)	1.23 (1.09–1.39)			
Genetic scores ^b	× ,	, , , , , , , , , , , , , , , , , , ,			
wGS _{CRP}					
<30%	1.02 (0.69–1.51)		1.21 (0.82–1.78)		
≥30%	1.01 (0.61–1.69)				
wGS _{IL-6}					
<30%	0.83 (0.49–1.40)		0.54 (0.32-0.90)		
≥30%	0.60 (0.31-1.19)				
wGS _{ESR}					
<30%	1.24 (0.53–2.91)		1.61 (0.69–3.76)		
≥30%	0.99 (0.33-3.04)				
wGS _{MCP}					
<30%	0.88 (0.60 1.29)		1.07 (0.73–1.57)		
≥30%	1.09 (0.66–1.80)				

Table 2. Association between Serum CRP and Genetic Determinants of Inflammatory Mediators with Proportion of Sites with PD \geq 4 mm and NAFLD in the Study of Health in Pomerania, 1997 to 2001.

CI, confidence interval; CRP, C-reactive protein; NAFLD, nonalcoholic fatty liver disease; PD, pocket depth; wGS_{CRP} , weighted genetic CRP score; wGS_{ESR} , weighted genetic erythrocyte sedimentation rate score; wGS_{IL-6} , weighted genetic interleukin 6 score; wGS_{MCP} , weighted genetic monocyte chemoattractant protein score.

^aAdjusted for age, sex, waist circumference, smoking, physical activity, alcohol consumption, and diabetes. ^bEstimate is for each unit increase in the respective genetic scores or log-transformed CRP.



Figure 2. Predicted probability of nonalcoholic fatty liver disease (NAFLD) according to tertile of the weighted genetic C-reactive protein (CRP) score (A) and categories of serum CRP (B) for study participants with no site (square), <30% of sites (circle), and \geq 30% of sites (diamond) with pocket depth \geq 4 mm.

NAFLD (Appendix Fig. 5), while each unit increase in the wGS_{CRP} was associated with a 21% increase in the prevalence odds of NAFLD; POR = 1.21 (95% CI, 0.82-1.78) (Table 2).

Although participants with periodontitis had a higher predicted probability of NAFLD compared to participants with a healthy periodontium, there was no significant statistical interaction (i.e., effect measure modification on the multiplicative scale) according to tertile of the wGS_{CRP} ($P_{\text{interaction}} = 0.8$) (Fig. 2A). Despite this, estimates stratified at the median value for wGS_{CRP} are presented in Table 3. Participants in the stratum of wGS_{CRP} at or below the median had slightly lower unadjusted and covariate-adjusted prevalence odd ratios for the relationship between periodontitis (PD ≥4 mm) and NAFLD compared to participants in the wGS_{CRP} stratum above the median (Table 3).

In contrast, the higher predicted probability of NAFLD for participants with periodontitis (PD \geq 4 mm) differed according to serum CRP levels $(P_{\text{interaction}} = 0.01)$ (Fig. 2B). And in contrast to the wGS_{CRP}, stratified analyses show a magnitude of unadjusted and covariate-adjusted prevalence odds of NAFLD comparing participants with periodontitis to those without to be highest in the low serum CRP (CRP <1 mg/L) stratum. For instance, the unadjusted and covariate-adjusted NAFLD prevalence odds ratio comparing participants with $\geq 30\%$ sites with PD ≥ 4 mm to those with no sites affected in

the low-serum CRP (<1 mg/L) stratum were 5.34 (95% CI, 3.18-8.97) and 2.39 (95% CI, 1.32-4.31), respectively, while the corresponding estimates for participants in the high-serum CRP (>3 mg/L) stratum were 1.66 (95% CI, 1.04-2.65) and 1.12 (95% CI, 0.65-1.93), respectively (Table 3).

Contrary to the finding of a significant statistical interaction between serum CRP and PD \geq 4 mm, there was no significant interaction with CAL \geq 3 mm ($P_{\text{interaction}} = 0.2$). This suggests that the significant interaction observed with the CDC-AAP periodontitis classification ($P_{\text{interaction}} = 0.014$) appeared to have been driven mostly by PD instead of CAL. Stratified estimates for periodontitis defined as the proportion of sites with CAL \geq 3 mm and based on the CDC-AAP criteria are presented in Appendix Tables 2 and 3, respectively.

	NAFLD, n		Prevalence Odds Ratio (95% CI)	
-	Cases	Noncases	Crude	Adjusted ^a
wGS _{CRP} ≤1.98 (<i>n</i> = 1,222)				
No site (0%)	65	292	Reference	Reference
Moderate (<30%)	166	479	1.56 (1.13–2.15)	1.08 (0.75–1.57)
Extensive (≥30%)	81	139	2.62 (1.78-3.84)	1.14 (0.72–1.80)
$wGS_{CRP} > 1.98 (n = 1,259)$				
No site (0%)	68	308	Reference	Reference
Moderate (<30%)	182	480	1.72 (1.26–2.35)	1.33 (0.94–1.89)
Extensive (≥30%)	92	129	3.23 (2.22-4.70)	1.65 (1.07-2.55)
Serum CRP $<$ I mg/L ($n = 972$)				
No site (0%)	29	313	Reference	Reference
Moderate (<30%)	89	402	2.40 (1.53–3.73)	1.62 (1.00-2.61)
Extensive (≥30%)	46	93	5.34 (3.18-8.97)	2.39 (1.32-4.31)
Serum CRP I to 3 mg/L ($n = 887$)				
No site (0%)	48	190	Reference	Reference
Moderate (<30%)	151	337	1.77 (1.23–2.57)	1.37 (0.90-2.08)
Extensive (≥30%)	58	103	2.23 (1.42-3.50)	0.97 (0.57–1.66)
Serum CRP >3 mg/L ($n = 622$)				
No site (0%)	56	97	Reference	Reference
Moderate (<30%)	108	220	0.85 (0.57–1.27)	0.70 (0.45-1.10)
Extensive (≥30%)	69	72	1.66 (1.04–2.65)	1.12 (0.65–1.93)

Table 3. Stratified Analysis for the Association between the Proportion of Periodontal Sites with Pocket Depth ≥ 4 mm (0%, <30%, $\geq 30\%$) andNAFLD According to Categories of the Weighted Genetic CRP Score and Serum CRP Levels, in the Study of Health in Pomerania, 1997 to 2001.

Interaction P value for the genetic CRP score = 0.6. Interaction P value for serum CRP levels = 0.01. CI, confidence interval; CRP, C-reactive protein; NAFLD, nonalcoholic fatty liver disease; wGS_{CRP}, weighted genetic score for CRP.

^aAdjusted for age, sex, waist circumference, smoking, physical activity, alcohol consumption, and diabetes.

Discussion

Findings from this investigation aimed at assessing whether inflammatory burden of which CRP was used as a marker modified the relationship between periodontitis and NAFLD, were consistent with a positive association between serum CRP and NAFLD as well as serum CRP and periodontitis. Under the premise that genetic determinants of inflammatory markers are better able to indicate lifelong inflammatory status, genetic variants robustly associated with CRP levels were combined into a genetic score that was substituted for serum CRP in this study. Contrary to the findings for serum CRP, the wGS_{CRP} predicted NAFLD to a greater extent than it did for periodontitis. And while serum CRP was a significant modifier, the wGS_{CRP} was not a modifier of the association between periodontitis and NAFLD.

NAFLD has a multifactorial etiology with conditions like insulin resistance and obesity identified as risk factors (Farrell and Larter 2006; Angulo 2007). Increased levels of inflammatory mediators have also been reported in individuals with NAFLD (Haukeland et al. 2006; Targher 2006). While a formal mediation analysis was beyond the scope of this study because of its cross-sectional nature, investigating the periodontitis-NAFLD association conditioning on or stratifying by markers of inflammatory burden while adjusting for confounders and NAFLD risk factors offers insights into the potential role of other factors besides inflammation in NAFLD etiology.

Despite the high CRP heritability (Pankow et al. 2001; Dupuis et al. 2005), currently identified genetic loci explain ~5% of the variation in serum CRP levels (Dehghan et al. 2011). Thus, currently identified loci may not be sufficiently robust to characterize associations or detect gene-environment interactions, even after combining the lead SNPs into a score as was done in this study. This may in part explain why no significant effect measure modification within strata of the genetic CRP score was detected.

Serum CRP represents a systemic marker of an inflammatory response and can be instrumental in detecting effect measure modification. The associations between periodontitis and NAFLD within levels of serum CRP were contrary to expectation. Indeed, the greater magnitude of association in the stratum of serum CRP <1 mg/L suggests a contribution of periodontitis to NAFLD burden independent of chronic lowgrade systemic inflammation. This association may have been induced via pathways involving an alteration in the gut microbial composition by swallowed periodontal pathogens (Arimatsu et al. 2014). Furthermore, participants with serum CRP <1 mg/L were less likely to have established NAFLD risk factors like abdominal obesity and diabetes (Appendix Table 4), consistent with an association of periodontitis with NAFLD independent of these factors.

While periodontitis may also contribute to NAFLD burden in the intermediate- and high-serum CRP strata, these associations in opposite of expectation suggest that the effects of periodontitis on NAFLD may be conditioned by a systemic inflammatory response of which CRP is a marker. Alternatively, the higher levels of serum CRP may indicate the presence to a larger extent of competing risk factors for NAFLD (Appendix Table 4), whose effects likely overshadowed those of periodontitis. Last, it is also possible that the high CRP levels may have been generated in response to hepatic injury as a result of NAFLD.

In theory, polymorphisms robustly associated with serum CRP levels are expected to predict an increase in the risk of coronary heart disease (CHD) and other cardiovascular disease-related events. However, these polymorphisms are not associated with an increased risk of CHD (Zacho et al. 2008; Elliott et al. 2009; Dehghan et al. 2011; Wensley et al. 2011), while serum CRP levels have been reported in several longitudinal studies to be associated with increased risk of CHD and myocardial infarction (Danesh et al. 2004; Lange et al. 2006). The SNPs used in creating the genetic CRP score in this study were robustly associated with CRP levels (Appendix Fig. 1), but these SNPs did not independently predict periodontitis or NAFLD risk (Appendix Figs. 2-5). While the findings for the genetic CRP score are not entirely surprising, it is also likely that the inability to detect effect measure modification by the genetic CRP score was also due to the relatively small effect of the individual SNPs even after combining them into a score.

Strengths and Limitations

Given the modest size, this study may be insufficiently powered to detect effect measure modification, especially for the genetic CRP score. In addition, the cross-sectional design makes it difficult to infer how the modification by serum CRP levels might affect the periodontitis-NAFLD association over time. Last, due to the mostly homogeneous study population, findings may not generalize to other racial/ethnic groups especially given that the genetic architecture of CRP differs by ethnicity (Carlson et al. 2005). Study strengths include a good characterization of the cohort that enabled the implementation of relevant exclusions of factors like alcohol consumption that might bias findings if not accounted for. Also, the availability of genotype and phenotype data for CRP allowed an investigation of this factor as a potential effect measure modifier.

Conclusion

Serum CRP was a significant modifier of the relationship between periodontitis and NAFLD, and there was a discordance of effect measure modification of this association by serum CRP and the weighted CRP genetic score. Given that only a fraction of the variability in CRP is explained by currently identified genetic loci, more research may be needed to identify missing CRP heritability that could provide a more robust picture of genetic loci predictive of CRP levels and inflammatory markers in general.

Author Contributions

A.A. Akinkugbe, contributed to conception, design, data acquisition, analysis, and interpretation, drafted and critically revised the manuscript; C.L. Avery, A.S. Barritt, G.D. Slade, T. Kocher, B. Holtfreter, contributed to data acquisition, analysis, and interpretation, critically revised the manuscript; S.R. Cole, contributed to data analysis, critically revised the manuscript; M. Lerch, J. Mayerle, S. Offenbacher, A. Petersmann, M. Nauck, H. Völzke, contributed to data acquisition, critically revised the manuscript; G. Heiss, contributed to conception, design, data acquisition, analysis, and interpretation, critically revised the manuscript. All authors gave final approval and agree to be accountable for all aspects of the work.

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