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Exposure to di-2-ethylhexyl terephthalate in a convenience sample of U.S. adults from 2000 to 2016

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Abstract

Di-2-ethylhexyl terephthalate (DEHTP), a structural isomer of di-2-ethylhexyl phthalate (DEHP), is a plasticizer used in a variety of commercial applications, but data on Americans' exposure to DEHTP do not exist. We investigated the exposure to DEHTP in a convenience group of U.S. adults by analyzing urine collected anonymously in 2000 ($N=44$), 2009 ($N=61$), 2011 ($N=81$), 2013 ($N=92$), and 2016 ($N=149$) for two major DEHTP oxidative metabolites: mono-2-ethyl-5-carboxypentyl terephthalate (MECPTP) and mono-2-ethyl-5-hydroxyhexyl terephthalate (MEHHTP). For comparison, we also quantified the analogous DEHP metabolites mono-2-ethyl-5-hydroxyhexyl phthalate (MEHHP) and mono-2-ethyl-5-carboxypentyl phthalate (MECPP). We detected MECPTP, MEHHP, and MECPP in all samples collected in 2016 with geometric means of 13.1, 4.1, and 6.7 ng/mL, respectively; we detected MEHHTP in 91% of the samples (geometric mean = 3.1 ng/mL). Concentrations of MECPTP correlated well with those of MEHHTP ($R^2=0.8$, $p<0.001$), but did not significantly correlate with those of MEHHP ($p>0.05$) suggesting different sources of exposure to DEHP and DEHTP. We also evaluated the fraction of the metabolites eliminated in their free (i.e., unconjugated) form. The median percent of unconjugated species was lower for the DEHP metabolites (MECPP [45.5%], MEHHP [1.9%]) compared to the DEHTP metabolites (MECPTP [98.8%], MEHHTP [21.2%]). Contrary to the downward trend from 2000 to 2016 in urinary concentrations of MEHHP and MECPP, we observed an upward trend for MEHHTP and MECPTP. These preliminary data suggest that exposure to DEHTP may be on the rise. Nevertheless, general population exposure data using MEHHTP and MECPTP as exposure biomarkers would increase our understanding of exposure to DEHTP, one of the known DEHP alternatives.

Keywords

Di-2-ethylhexyl terephthalate; Di-2-ethylhexyl phthalate; DEHTP; DEHP; Plasticizers; Exposure; Oxidative metabolites

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Introduction

Di-2-ethylhexyl terephthalate (DEHTP) is used as a plasticizer in food and drink contact materials, medical devices, children's toys, and childcare articles, among other applications (Beeler 1976). DEHTP is a structural isomer of another plasticizer: di-2-ethylhexyl phthalate (DEHP). In the past decade, use of DEHP has been restricted in the United States and some European countries because of concerns regarding DEHP potential human toxicity (CPSC 2014; European Union 2005). In contrast, DEHTP has Food Contact Notification clearance from the US Food & Drug Administration (Eastman chemical company 2014); DEHTP also complies with the European Commission regulation for use in food contact applications (EFSA 2008). Therefore, DEHTP has been used as an alternative to DEHP, and both production volume and application of DEHTP are expected to increase (Eastman chemical company 2014). As a result, human exposure to DEHTP might also increase.

DEHTP is used as an alternative to DEHP (Eastman chemical company 2014), but some animal and human studies suggest potential adverse effects upon exposure to high doses of DEHTP. For example, chronic dietary exposure caused general toxicity in rats (Ball et al. 2012). Female Sprague–Dawley rats administered DEHTP in the diet for 90 days showed no major organ or systemic toxicity at 0.5% (v/v) dose levels, but some effects on hematology parameters and increase in relative liver weights were observed at the 1.0% dose (Barber and Topping 1995). In a study conducted with 203 human volunteers for evidence of sensitization to DEHTP (0.5%, v/v) following 3 weeks of dermal application three times a week, two subjects had slight erythema to DEHTP (David et al. 2003).

In order to better understand the extent of human exposure to DEHTP and its potential impact on health using biomonitoring, identifying and quantifying exposure biomarkers is a necessary step. Oxidative metabolites of DEHTP, mono-2-ethyl-5-hydroxyhexyl terephthalate (MEHHTP), mono-2-ethyl-5-oxohexyl terephthalate (MEOHTP), and mono-2-ethyl-5-carboxypentyl terephthalate (MECPTP), have been identified in vitro (Silva et al. 2015) and in vivo (Lessmann et al. 2016a, b). In humans, MECPTP was identified as the major metabolite of DEHTP, followed by MEHHTP (Lessmann et al. 2016b). Biomonitoring data are limited to one study in which both compounds were detected in the urine of 34 German adults (Lessmann et al. 2016b), but information about the extent of Americans' exposure to DEHTP does not exist. To fill in this gap, we quantified MECPTP and MEHHTP in urine samples collected in a period of 16 years [2000 ($N= 44$), 2009 ($N= 61$), 2011 ($N= 81$), 2013 ($N= 92$), and 2016 ($N= 149$)] from U.S. adults with no documented DEHTP exposure. We also compared the concentrations of these biomarkers to mono-2-ethyl-5-hydroxyhexyl phthalate (MEHHP) and mono-2-ethyl-5-carboxypentyl phthalate (MECPP), two metabolites of DEHP.

Materials and methods

Chemicals

We purchased MEHHTP, MECPTP, d_4 -MEHHTP, d_4 -MECPTP, and $^{13}C_6$ -MECPP from CanSyn (Ontario, Canada), and MEHHP, MECPP, and d_4 -MEHHP from ADM (Germany).

Acetonitrile (HPLC grade), water (HPLC grade), and methanol (99.8%, HPLC grade) were purchased from Honeywell Burdick & Jackson (Muskegon, MI). β -glucuronidase (*Escherichia coli*-K12) was purchased from Roche Biomedical (Mannheim, Germany). All chemicals and reagents were used without further purification.

Analytical method

The analytical method for measuring phthalate oxidative metabolites in urine is adapted from previously published methods (Silva et al. 2007). Briefly, urine (0.1 mL) and calibration standards were spiked with an internal standard solution containing deuterium- or ^{13}C -labeled analogs of the target metabolites and a buffered solution of β -glucuronidase (*Escherichia coli*-K12; 25 μL , pH 6.5/1 M ammonium acetate), and incubated at 37 °C for a minimum of 120 min. The target analytes in the spiked urine were extracted using on-line solid phase extraction, chromatographically resolved by high-performance liquid chromatography (Figure S1) and quantified using negative ion electrospray-ionization tandem mass spectrometry (Table 1). Analysis of urine samples collected in 2016 was repeated with addition of 1M ammonium acetate buffer solution without β -glucuronidase to quantify unconjugated metabolites (Silva et al. 2007); we did not investigate for other potential conjugates (e.g., sulfates). The limits of detection (LODs), calculated as $3S_0$, where S_0 is the standard deviation as the concentration approaches zero (Taylor 1987), were 0.4 ng/mL for MEHHTP and MECPTP and 0.2 ng/mL for MECPP and MEHHP. For concentrations <LOD, we imputed a value equal to the LOD divided by the 2. Statistical significance was as set to $p < 0.05$. Trend analysis for MECPP and MECPTP using regression analysis was performed by SAS software.

Subjects

We collected urine anonymously in 2000, 2009, 2011, 2013, and 2016 from demographically diverse groups of U.S. male and female adults. No personal information from the subjects was available. Samples were collected between 8:00 AM and 5:00 PM and were not necessarily first-morning voids. We cannot rule out the possibility that some donors contributed urine in different collection years and/or multiple urine specimens, albeit collected during different days or time of day, during any given collection year. After collection, samples were stored at $-70\text{ }^\circ\text{C}$ until analysis. The Centers for Disease Control and Prevention (CDC) Internal Review Board approved the urine collection and analysis. A waiver of informed consent was requested under 45 CFR 46.116(d).

Results and discussion

In humans, DEHTP metabolizes extensively before elimination in urine (Lessmann et al. 2016b) and its oxidative metabolites (e.g., MEHHTP, MECPTP) can be used as biomarkers of exposure to DEHTP (Lessmann et al. 2016a, b). Interestingly, MEHHTP and MECPTP are structural isomers of MEHHP and MECPP, respectively, which have been used extensively as DEHP exposure biomarkers (Koch et al. 2004, 2005, 2006; Silva et al. 2006).

In this study, we quantified the concentrations of MEHHTP, MECPTP, MEHHP, and MECPP in urine collected in 2000, 2009, 2011, 2013, and 2016 from convenience samples

of U.S. adults. The geometric mean and select percentile concentrations, and detection frequencies are listed in Table 2. We observed an increase in detection frequencies of MEHHTP (from 7 to 91%) and MECPTP (18–100%) from 2000 to 2016 (Table 2). Similarly, we observed an upward trend in the geometric mean concentrations of MEHHTP (<LOD to 3.1 ng/mL from 2011 to 2016) and MECPTP (<LOD to 13.1 ng/mL from 2009 to 2016, $p < 0.001$ -trend analysis 2011–2016) (Table 2). By contrast, during the same time period, geometric means of the two DEHP metabolites decreased from 8.7 to 4.1 ng/mL (MEHHP) and 12.1 to 6.7 ng/mL (MECPP, $p = 0.005$ -trend analysis 2000–2016) (Table 2). Together, these findings suggest that DEHTP exposures may be on the rise perhaps because DEHTP can be used as a replacement of DEHP in consumer products (Eastman chemical company 2014).

We observed higher median concentrations of MEHHTP (3.0 ng/mL) and MECPTP (15.7 ng/mL) in 2016 samples than concentrations reported in a German pilot study (<0.3 and 0.9 ng/mL for MEHHTP and MECPTP, respectively) conducted around the same time period (Lessmann et al. 2016b). These differences may reflect potentially higher exposure to DEHTP in the United States than in Germany. However, because of the pilot nature of the two studies and non-representative nature of the samples analyzed, these results must be interpreted with caution until representative data from both countries become available.

Both DEHP and DEHTP produced a carboxylic acid (MECPP and MECPTP) as their major metabolite (Table 2), but we observed some differences in their concentration patterns. In the 2016 samples tested, the geometric mean concentration of MECPTP was higher than MECPP (13.1 vs. 6.7 ng/mL); however, the geometric mean concentration of MEHHTP was lower than of MEHHP (3.1 vs. 4.1 ng/mL) (Table 2). These results may suggest potential differences in toxicokinetics (i.e., adsorption, distribution, metabolism, elimination) between DEHP and DEHTP.

Glucuronidation facilitates urinary elimination of phthalate metabolites (Silva et al. 2003). Despite structural similarities between DEHP and DEHTP, interestingly, we observed that glucuronidation of their analogous metabolites differed. Specifically, we found that 98.8% of MECPTP but only 45.5% of MECPP eliminated in their free (i.e., unconjugated) form (Table 2). In contrast, MEHHP and MEHHTP eliminated mostly conjugated (median free metabolites were 1.9 and 21.2%, respectively). Our MECPTP results are in agreement with previous findings where humans administered with DEHTP eliminated 91.2% of MECPTP in its free form (Lessmann et al. 2016b). However, in our study population, only 21.2% of MEHHTP eliminated as the free metabolite, compared to >70% of unconjugated MEHHTP in males administered DEHTP (Lessmann et al. 2016b). The reasons for the differences in the percentage of MEHHTP eliminated as free species in these two studies remain, at present, unknown, but may relate to inter-individual differences because the human metabolism study relied on data from three male adults (Lessmann et al. 2016b).

As expected, because MEHHTP and MECPTP share a common precursor, their concentrations correlated well ($R^2 = 0.8$, Fig. 1). However, urinary MECPTP did not correlate well with MEHHP ($p > 0.05$, Fig. 1) suggesting that the sources of DEHTP and

DEHP exposure likely differ as expected if DEHTP is indeed replacing DEHP in certain commercial applications.

In conclusion, the frequent detection of DEHTP metabolites among a diverse group of U.S. male and female adults suggests widespread exposure to DEHTP. Further, the apparent increase in detection frequency and magnitude of the concentrations with time suggests that DEHTP exposure in the United States may be on the rise. Last, these pilot data suggest that the DEHTP metabolites, MEHHTP and MECPTP can serve as biomarkers of exposure to DEHTP in large-scale biomonitoring studies such as the National Health and Nutrition Examination Survey.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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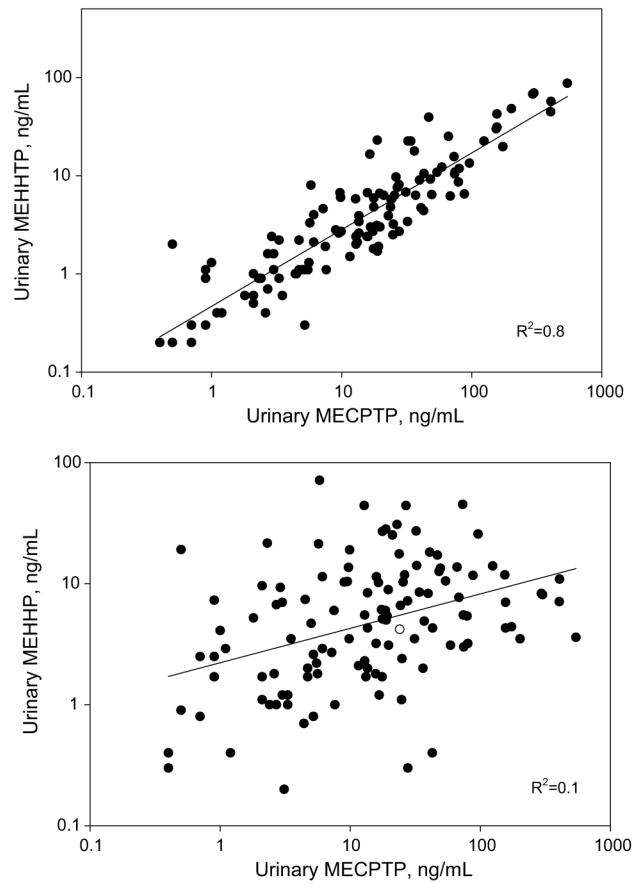


Fig. 1. Correlation analyses of urinary concentrations of MECPTP and MEHHTP (*top*); and MECPTP and MEHHP (*bottom*)

Table 1

Analytical parameters for the quantification of DEHP and DEHTP metabolites

Parent chemical	Urinary metabolite	Internal standard	MS/MS scan, native (ISD ^a)	Collision energy (eV) ^b
DEHP	MEHHP	d ₄ -MEHHP	293/145 (297/145)	17
	MECPP	¹³ C ₆ -MECPP	307/159 (313/159)	17
DEHTP	MEHHTP	d ₄ -MEHHTP	293/121 (297/125)	22
	MECPTP	d ₄ -MECPTP	307/165 (311/169)	20

^aISD-internal standard^bCollision energy applied in ThermoScientific Vantage mass spectrometer

Table 2
Urinary concentrations of DEHP and DEHTP metabolites in a convenience sample of U.S. adults

Parent chemical	Urinary metabolite	Collection year	N	Select percentiles ^b (ng/mL)				Frequency of detection (%)	Geometric Mean ^b (ng/mL)	Median unconjugated metabolite ^{c,d} (%)
				25th	Median	75th	90th			
DEHP	MEHHP	2016	149	1.8	4.3	10.2	19.0	100	4.1	1.9
		2013	92	2.2	4.9	9.4	16.2	100	4.3	
		2011	81	17	4.2	10.2	22.0	100	4.1	
		2009	61	2.7	5.1	13.5	21.5	100	5.9	
		2000	44	4.8	11.2	22.5	30.0	100	8.7	
		2016	149	2.6	6.6	15.3	30.9	100	6.7	45.5
		2013	92	4.6	7.9	15.9	29.6	100	7.8	
		2011	81	4.0	7.3	17.9	41.5	100	9.3	
		2009	61	5.9	10.9	11.6	32.6	100	10.1	
		2000	44	6.7	16.6	28.0	37.9	100	12.1	
DEHTP	MEHHTP	2016	149	1.1	3.0	7.6	22.5	91	3.1	21.2
		2013	92	0.6	1.1	3.0	7.1	77	1.3	
		2011	81	<LOD ^a	<LOD	1.1	4.5	44	<LOD	
		2009	61	<LOD	<LOD	0.6	1.8	34	<LOD	
		2000	44	<LOD	<LOD	<LOD	1.2	7	<LOD	
		2016	149	4.5	15.7	37.1	92.2	100	13.1	98.8
		2013	92	2.1	4.8	10.8	44.6	95	4.6	
		2011	81	1.5	2.0	4.8	14.1	86	2.0	
		2009	61	<LOD	<LOD	0.6	1.1	38	<LOD	
		2000	44	<LOD	<LOD	<LOD	1.3	18	<LOD	

^a LOD-limit of detection; LOD-0.4 ng/mL for both MEHHTP and MECPTP

^b <LOD was treated as LOD 2

^c Only total concentrations >LOD were used in the calculation

^d Only urine samples collected in 2016 were analyzed without β -glucuronidase for unconjugated metabolites