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# Murine In vitro Memory T Cell Differentiation

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### Abstract

Upon pathogen encounter, naïve CD8<sup>+</sup> T cells are primed and undergo massive clonal expansion. A fraction of effector CD8<sup>+</sup> T cells remains during the contraction phase and differentiate into memory T cells critical for mounting robust recall responses in response to secondary infection. Low frequency of memory T cells *in vivo* is a major obstacle to investigate their functional aspects including migration capacity and genetic regulation. Here, we describe detailed protocol for memory T cell differentiation developed by von Andrian's group to generate large number of CD44<sup>hi</sup>CD62L<sup>hi</sup> antigen-specific memory T cells *in vitro*.

## Materials and Reagents

- 1. Recombinant mouse IL-15 (rmIL15) (BioLegend, catalog number: 566302)
- 2. RPMI-1640 medium (Life Technologies, Gibco<sup>®</sup>, catalog number: 11875-119)
- 3. Fetal bovine serum (Atlanta Biologicals, catalog number: S11055H)
- 4. Penicillin/streptomycin (Gemini Bio-Products, catalog number: F52M00E)
- 5. L-Glutamine (Life Technologies, Gibco<sup>®</sup>, catalog number: 25030-081)
- 6. 100x 1 M Hepes (Life Technologies, Gibco<sup>®</sup>, catalog number: 15630-080)
- 100x MEM Non-essential amino acids (Life Technologies, Gibco<sup>®</sup>, catalog number: 11140-050)
- 8. 100x sodium pyruvate (100 mM) (Life Technologies, Gibco<sup>®</sup>, catalog number: 11360-070)
- **9.** 100x 2-mercaptoethanol (Life Technologies, Gibco<sup>®</sup>, catalog number: 21985-023)
- **10.** OVA<sub>257-264</sub> synthetic peptide (Sigma-Aldrich, catalog number: S7951)

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<b>11.</b> Ficoll-Paque <sup>™</sup> Premium 1.084 (GE Healthcare, catalog number: 17-5446-0)	11.	Ficoll-Paque <sup>TM</sup>	<sup>1</sup> Premium 1.084	(GE Healthcare,	catalog number:	: 17-5446-02
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- 12. Antibodies:
  - **a.** Anti-CD44 PerCpCy5.5 (clone: IM7) (eBioscience, catalog number: 45-0441)
  - **b.** Anti-CD62L APC (clone: MEL-14) (eBioscience, catalog number: 17-0621)
- **13.** RBC lysis buffer (eBioscience, catalog number: 00-4333-57)
- 14. Bovine serum albumin (Thermo Fisher Scientific, catalog number: BP1605-100)
- **15.** NaN<sub>3</sub> (Sigma-Aldrich, catalog number: S8032)
- **16.** T cell media (see Recipes)
- 17. Staining buffer (in PBS) (see Recipes)

### Equipment

- 1. Centrifuge (Thermo Fischer Scientific, Sorvall<sup>™</sup> Legend RT)
- 2. 70 µm cell strainer (BD Biosciences, Falcon<sup>®</sup>, catalog number: 352350)
- **3.** 15 ml and 50ml Falcon tubes
- 4. 24 well plates (BD Biosciences, Falcon<sup>®</sup>, catalog number: 353226)
- 5. T75 culture flask (Corning, catalog number: 430641)
- 6. 37 °C 5% CO<sub>2</sub> Cell Culture incubator

### Procedure

# A. CD44<sup>hi</sup>CD62L<sup>lo</sup> Memory T cell differentiation proceeds under sterile tissue culture conditions

- 1 Euthanize a OT-1 CD8 TCR transgenic mouse and take spleen, and (optional) lymph nodes.
- 2 Splenocytes are RBC lysed followed by washing with PBS twice.

#### B. OT-1 TCR stimulation with cognate peptide antigen

- 3 Resuspend cells in 1 ml of T cell media and add  $OVA_{257-264}$  synthetic peptide to 1  $\mu$ M.
- 4 Incubate in the 5%  $CO_2$  at 37 °C for 1 h.
- 5 Spin down cells at 1,500 rpm for 3 min at 4 °C and wash once with T cell media.
- 6 Resuspend cells in 12 ml of T cell media and plate 1ml/well of a 24 well plate.
- 7 Incubate in the 5%  $CO_2$  at 37 °C for 2 days.
- 8 Harvest the cells by pipetting up and down, and pellet cells.

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- 9 Resuspend cells in 5 ml of T cell media, and load on to 2.5 ml of Ficoll.
- 10 Spin down at 400 x g for 15 min at 4 °C.
- 11 Transfer live cells on the interphase to a new 15 ml tube and fill up the tube with T cell media.
- 12 Spin down cells at 1,500 rpm for 3 min at 4 °C.

#### C. Memory T cell culture in the presence of IL-15

- 13 Resuspend cells in 24 ml of T cell media containing rmIL15 (20 ng/ml). Culture cells in T75 flask for four days.
- 14 Harvest and pellet cells for Ficoll gradient (repeat steps 9–12).
- 15 Resuspend cells in 40 ml of T cell media containing rmIL15 (20 ng/ml). Culture in T75 flask for two days.
- 16 Staining cells with anti-CD44 and CD62L antibodies in staining buffer for 15 min on ice.
- 17 Wash with staining buffer twice, then proceeds flow cytometry analysis.

### Recipes

1. T cell media

RPMI-1640

10% fetal bovine serum

1% penicillin/streptomycin

1% L-Glutamine

1x 1 M Hepes

1x MEM non-essential amino acids

1x sodium pyruvate 100 mM

- 1x 2-mercaptoethanol
- 2. Staining buffer (in PBS)

1% BSA

0.02% NaN3

### Acknowledgments

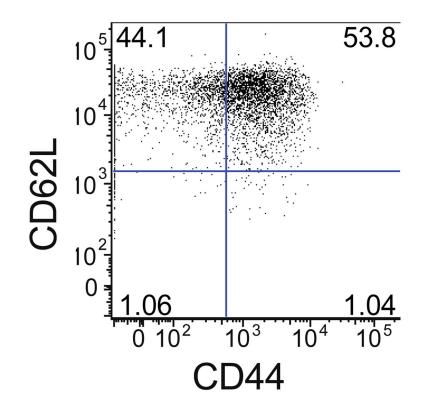
The protocol was adapted from a previously described study (Manjunath *et al.*, 2001). This work was supported by the Starr Cancer Consortium (13-A123 to M.O.L. and M.Q.Z.), the Rita Allen Foundation (M.O.L.), the NBRPC (2012CB316503 to M.Q.Z), and the NIH (HG001696 to M.Q.Z.).

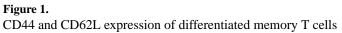
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