

Review

Plasmonic Nanomaterial-Based Optical Biosensing Platforms for Virus Detection

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Received: 17 September 2017; Accepted: 11 October 2017; Published: 13 October 2017

Abstract: Plasmonic nanomaterials (P-NM) are receiving attention due to their excellent properties, which include surface-enhanced Raman scattering (SERS), localized surface plasmon resonance (LSPR) effects, plasmonic resonance energy transfer (PRET), and magneto optical (MO) effects. To obtain such plasmonic properties, many nanomaterials have been developed, including metal nanoparticles (MNP), bimetallic nanoparticles (bMNP), MNP-decorated carbon nanotubes (MNP-CNT), and MNP-modified graphene (MNP-GRP). These P-NMs may eventually be applied to optical biosensing systems due to their unique properties. Here, probe biomolecules, such as antibodies (Ab), probe DNA, and probe aptamers, were modified on the surface of plasmonic materials by chemical conjugation and thiol chemistry. The optical property change in the plasmonic nanomaterials was monitored based on the interaction between the probe biomolecules and target virus. After bioconjugation, several optical properties, including fluorescence, plasmonic absorbance, and diffraction angle, were changed to detect the target biomolecules. This review describes several P-NMs as potential candidates of optical sensing platforms and introduces various applications in the optical biosensing field.

Keywords: plasmonic nanomaterial; metal nanoparticle; nanoparticles; decorated carbon nanomaterial; optical biosensing system; virus detection

1. Introduction

Plasmon has an interesting property that occurs as an electron cloud on the surface of metallic nanomaterials [1–3]. When metallic nanoparticles (MNP) interact with incident light, the electron cloud in the MNP oscillates, and in this case, the collective excitation of electrons is regarded as plasmon [1,4]. Recently, the plasmonic effect induced by pi electrons has been discovered in carbon nanomaterials, such as graphene (GRP) and carbon nanotubes (CNT) [5–8]. In some cases, plasmonic properties can be tuned by interactions between plasmonic nanomaterials (P-NM) and shape control, thus controlling their optical properties to exhibit various colors [9–13].

Numerous P-NMs have been introduced, including MNP, such as gold, silver, platinum, CNT, and GRP. To achieve an enhanced plasmonic effect, hybrid plasmonic nanomaterials have been developed, such as bimetallic NP (bMNP), core/shell MNP, MNP-decorated CNT (MNP-CNT), and MNP-modified GRP (MNP-GRP) [14–17]. To prepare these P-NMs, various synthetic strategies have been suggested, such as chemical reduction processes, electrochemical deposition, microwave-assisted synthesis processes, photo-induced decoration processes, and environmentally friendly processes [18–21]. In addition, such P-NMs possess excellent optical properties, such as surface enhanced Raman scattering (SERS), plasmonic resonance energy transfer (PRET), enhanced

catalytic properties, a localized surface plasmon resonance (LSPR) effect, and the magneto optical (MO) effect [22–26].

Because P-NMs have such specialized optical properties, they are reported to have many potential applications including energy devices, nano optics, sensors, and nanobiomedicine [27–33]. Among these, many researchers are investigating development of highly sensitive and selective biosensing systems to address public concerns and health care issues [34–36]. In addition, P-NMs have improved the specificity of on-site detection and point-of-care diagnosis platforms. Attempts to monitor several infectious diseases have been made using P-NM-based sensing systems, including the influenza virus, norovirus, and tuberculosis (TB) [37–39]. Viral DNA and RNA have also been detected using P-NMs [30,40].

In this review, we introduce various P-NMs, such as MNPs, bMNPs, MNP-CNT, and MNP-GRP, as candidates for optical sensing platforms. We also describe several P-NM-based optical biosensing systems that are demonstrated via SERS, PRET, LSPR, and colorimetry, including viral DNA and RNA detection.

2. Various Functions of Plasmonic Nanomaterials (P-NMs)

P-NMs can be classified into in two categories: noble metal-based MNP and MNP-carbon nanomaterials including MNP-CNT and MNP-GRP. Both P-NMs have excellent plasmonic and optical properties. Among of these, noble metals, such as Au, Ag, and Pt-based MNPs, are well-established and widely applied in various fields [41,42] due to their stable plasmonic properties and particle stability [43]. In addition, MNP's plasmonic property can be easily tuned and controlled through particle structure or the distance between each MNP. On the other hands, tunable infra-red (IR) plasmonic property of GRP also has been interested due to excellent optical properties such as enhancement of light and matter interaction and integration of mid-IR photonics [44].

Oh et al. tuned the plasmonic property of AgNP by controlling its particle size [45]. In this case, the size was changed by the seed-mediated growth method, and as the Ag ion concentration increased, large AgNPs were produced. In addition, the plasmonic absorbance of AgNP was redshifted, so the solution color changed from yellow to red. Lee et al. tuned the plasmonic property of AuNP by controlling the AuNP assembly (Figure 1) [46]. Typically, the plasmonic absorbance of spherical AuNP is measured to be approximately 525 nm. However, in this study, the authors synthesized AuNP chains using *N*-(3-Dimethylaminopropyl)-*N'*-ethylcarbodiimide (EDC) and as the EDC concentration increased, the AuNP chain was elongated, changing its structure (Figure 1a–f). Thus, the absorbance tendency of the AuNP chain changed and was redshifted. This phenomenon occurs by the plasmonic coupling effect between AuNPs that are in close contact, and this effect may have induced the secondary plasmonic band that was observed at approximately 625 nm (Figure 1g).

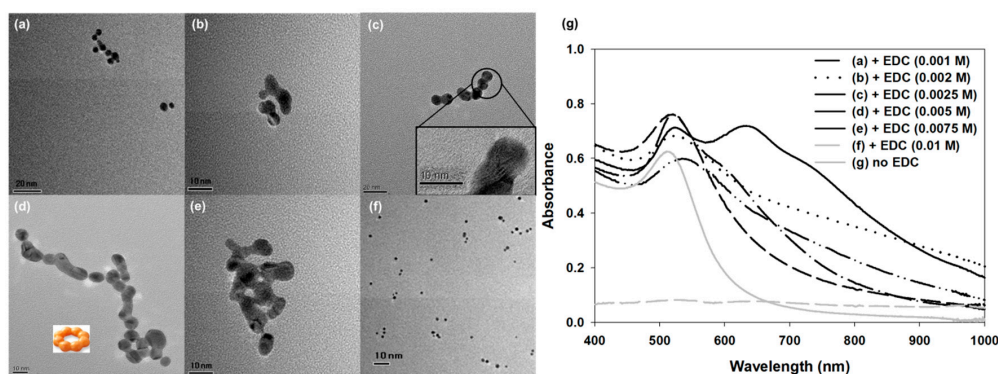


Figure 1. Various structure of Au-nanoparticles (NPs) and their plasmonic property. (a–f) Structural differences in AuNPs using TEM and (g) plasmonic absorbance measurement after adding varying concentrations of EDC. Reproduced from Ref. [46] with permission from the Royal Society of Chemistry.

To improve P-NM functionality and obtain a synergic effect, many core/shell structures have been developed. Zhou et al. reported an iron oxide AuNP core/shell particle structure that was urchin-shaped rather than spherical (Figure 2a–d) [47]. Thus, it could possess multi-functional properties including magnetic and plasmonic properties. In addition, this particle exhibited attractive optical properties due to the structurally induced plasmonic effect. This spiky structure could generate a plasmonic hotspot, and its electromagnetic field could be more enhanced than in the spherical case. Thus, this structure could possess improved SERS and catalytic activities [1]. In this case, the broadened absorbance spectrum of urchin-shaped structure was measured from 500 nm to 800 nm (Figure 2e). This urchin-shaped particle was prepared by the seed mediated synthesis method, and its plasmonic property was tuned depending on the concentration of seed material (Figure 2f). In another study, an Au-Ag core/shell structure was introduced to improve and tune the plasmonic property [48]. In this case, two plasmonic absorbance peaks were obtained, one approximately 424 nm and one with a broad peak between 538 nm and 560 nm. Using the plasmonic property of Au-Ag P-NM, they developed an optical sensing platform to detect the Zika virus. In other study, P-NM played a role as MO active substrate to detect a single molecule via light control [49,50]. In this case, single organic molecule has been detected under the strong external magnetic field with Au/Ag bimetallic hybrid structure that was used as MO active material [50].

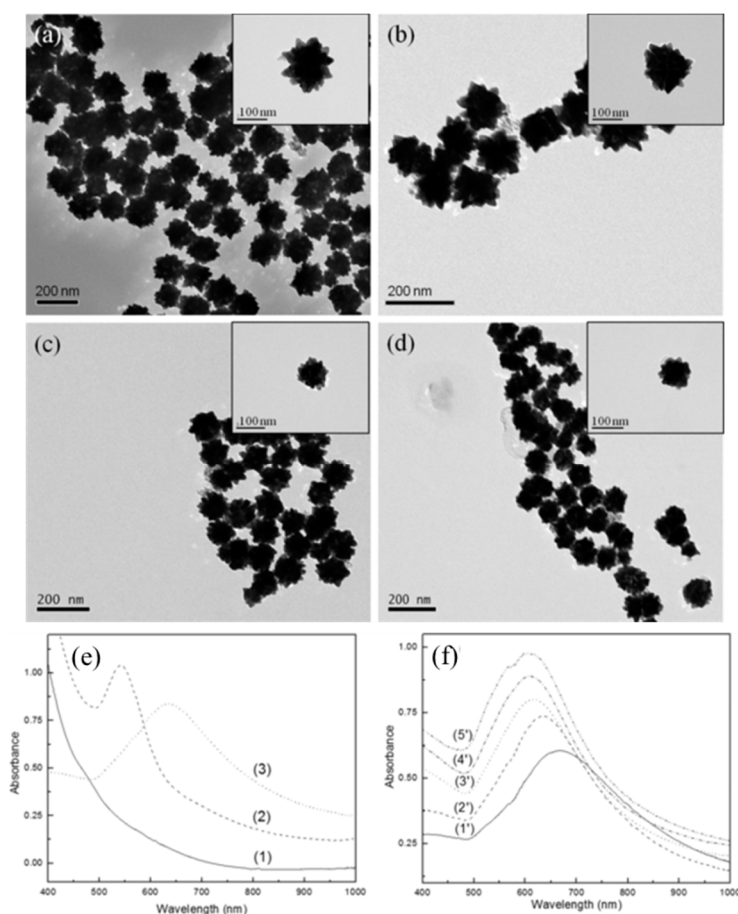


Figure 2. Urchin-shaped iron oxide-AuNP core/shell structure. (a–d) TEM images of iron oxide-AuNP core/shell structure; (e) UV/Vis absorbance spectra of (1) iron oxide, (2) spherical iron oxide@gold core/shell and (3) urchin type of iron oxide@gold core/shell and (f) plasmonic absorbance spectra of urchin-shaped structure depending on the concentration of seeds for assembly (1') 4.5 nM, (2') 8.9 nM, (3') 13.2 nM, (4') 17.5 nM and (5') 21.7 nM. Copyright Wiley and Sons. Reproduced with permission [47].

Recently, MNP decorated carbon nanomaterial (MNP-CNM) has been considered as a potential P-NM due to its excellent synergic properties, such as SERS, PRET and MO. Because CNM possesses pi electrons on its surface, plasmonic coupling between MNP and the CNM surface could occur. Thus, a strong and longitudinal plasmonic effect can be induced. For example, AuNP-CNMs were prepared as P-NM by Lee and colleagues, and in this study, AuNP-CNT and AuNP-GRP were synthesized through simple two steps without harsh conditions (Figure 3) [51,52]. In this case, Au ions and CNMs were sonicated in deionized water, then mild reducing agent was added to the mixture, and it was stirred for 3 h to obtain AuNP-CNT or AuNP-GRP. After preparation of hybrid material, its morphology was observed by TEM and in this case, a lot of Au NPs were well attached and distributed on the surface of CNT or GRP (Figure 3a,c). The plasmonic property of MNP-CNM was also characterized by UV/vis spectroscopy and as a result, plasmonic absorbance of MNP-CNM was clearly measured that was shown in Figure 3b,d. Using these hybrid nanomaterials, a PRET-based fluoro-immunoassay detected an infectious virus. In addition, AgNP-CNT was fabricated to enhance the property [53]. This hybrid structure served as a SERS substrate, and its sensitivity to the R6G molecule was enhanced by $\sim 10^{10}$. Other type of core/shell was also developed that was Au@cage (Au/Ag bimetallic cage) and this Au@cage structure was used for SERS active material to detect a malaria virus DNA [54]. It also showed highly sensitive detection performance and its LOD was about 100 attomole.

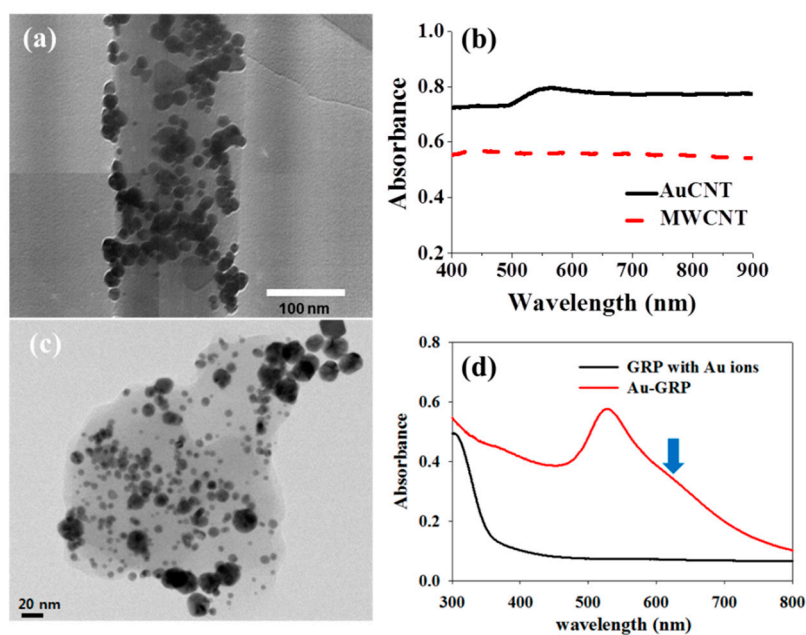


Figure 3. Structure of Au-CNT and Au-GRP. TEM images of (a) Au-CNT; (b) plasmonic absorbance of Au-CNT; (c) Au-GRP and (d) UV/Vis spectrum of Au-GRP as hybrid P-NM structures. (a,b) Copyright Elsevier B.V. Reproduced with permission [51] and (c,d) Reprinted with permission from [52], Copyright 2014, American Chemical Society.

Various P-NMs have been developed and investigated to use their specialized plasmonic and optical properties. They can be tuned, and synergic effects can be obtained through various strategies, such as assembly, core/shell, and hybrid structure with CNM.

3. Application of P-NM-Based Biosensing Systems

3.1. LSPR and PRET-Based Sensing Platform

P-NMs can be applied in various fields, such as nano optics, drug delivery systems (DDS), contrast agents, and sensing systems. Eventually, these plasmonic materials may play key roles in biosensing platforms. By using P-NMs, researchers have developed several types of biosensing

systems, including SERS and LSPR-based optical sensing platforms, PRET-based fluoro-immunoassays, and colorimetry-based detection systems. These P-NM-based biosensing platforms can detect viral antigens and viral DNA or RNA, thus this system could be applied universally.

Takemura and colleagues developed LSPR based-sensing systems to detect an influenza virus, and non-spherical AuNPs were used as plasmonic materials (Figure 4) [55].

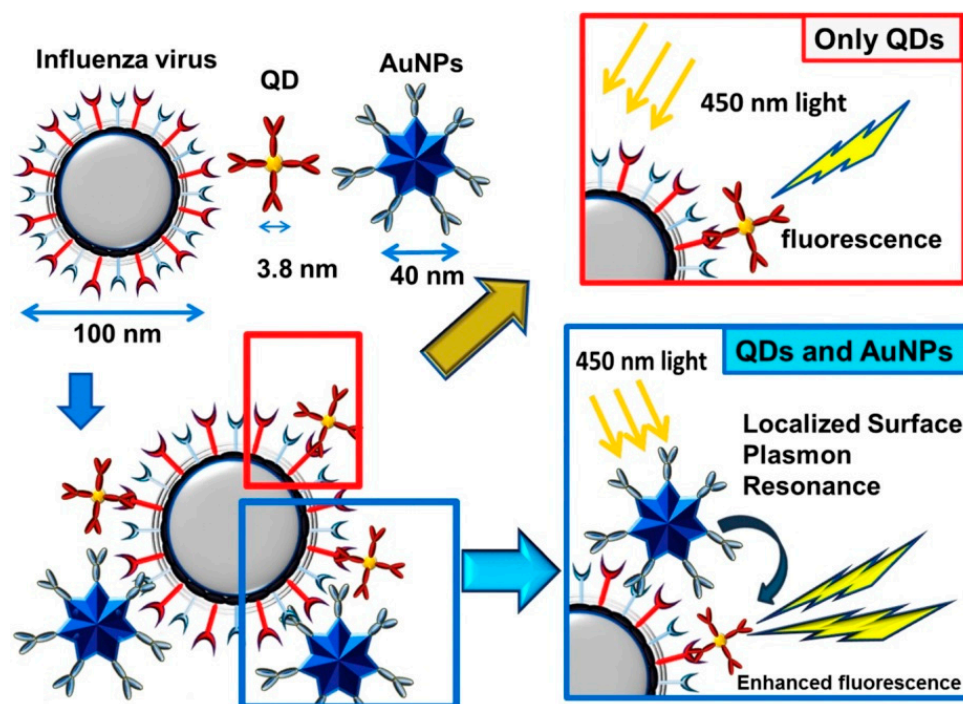


Figure 4. Schematic illustration of the localized surface plasmon resonance (LSPR)-based detection mechanism of biosensing for the influenza virus. Copyright Elsevier B.V. Reproduced with permission [55].

Fluorescent quantum dot (QD) was used to monitor the viral antigen. If QD is located near the P-NM, its fluorescence intensity can be enhanced by the LSPR-based PRET effect [56–58]. Therefore, if the target virus existed in the antibody (Ab) modified P-NM/QD system, key and lock interaction would occur between the target virus and P-NM/QD. As a result, the fluorescence intensity of QD could be enhanced by energy transfer in the virus/P-NM/QD structure. Depending on the concentration of viral antigen, the fluorescent enhancement factor was affected, and as the amount of the virus increased, the QD fluorescence gradually increased. In this study, the limit of detection (LOD) for the influenza virus was estimated at 0.03 pg/mL in deionized water and 0.4 pg/mL in a serum matrix. In addition, clinically isolated influenza virus was detected with 10 PFU/mL detection limitations. Moreover, this system was highly selective against other biomolecules. In a different study, Zika virus RNA was monitored using Au-Ag core/shell P-NM and QD [48]. The target RNA was detected by a similar approach to the influenza method. The molecular beacon was modified on the QD surface to capture the target RNA. When Zika virus RNA was placed in this system, hybridization occurred, resulting in enhanced fluorescence intensity. In this case, the LOD was 2.4 copies/mL, with extremely high selectivity.

MNP-GRP or MNP-CNT can also be used as LSPR substrate for PRET-based sensing systems to monitor the viral antigens. Lee et al. detected infectious TB antigen by using TB Ab-modified Au-GRP and QD through PRET-based fluoro-immunoassay (Figure 5a) [52]. In this study, a sandwich structure was formed with Au-GRP and QD by the TB antigen, i.e., CFP-10 and fluorescence enhancement of QD was measured by the antigen concentration. In Figure 5b, the sensing performance is depicted, and as

the target antigen concentration increased, the fluorescence intensity linearly increased. The LOD was estimated at 4.5 pg/mL. Moreover, other TB antigens, such as Antigen 85, were not monitored, thus it showed excellent selectivity. Au-CNT can also be a P-NM for fluoro-immunoassay, and the influenza virus was monitored by a similar strategy with an Au-GRP sensing mechanism [51]. Here, Au-CNT/QD was assembled as a sandwich structure by the influenza virus, resulting in enhanced fluorescence with each virus concentration. It was highly sensitive with 0.1 pg/mL, and a clinically isolated influenza virus was also detected with a 50 PFU/mL detection limitation.

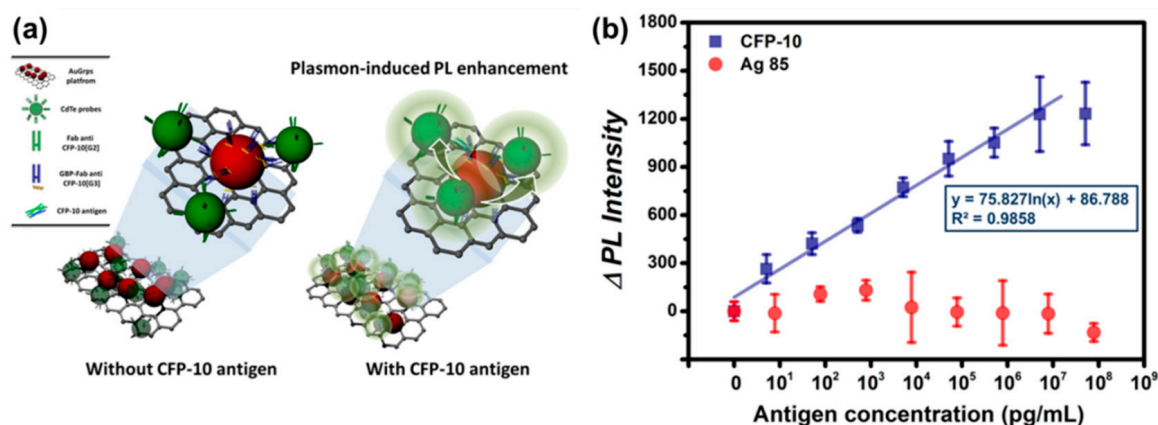


Figure 5. TB antigen detection using Au- Graphene (GRP)-assisted fluoro-immunoassay. (a) Au-GRP-assisted fluoro-immunoassay process and (b) detection behavior of the plasmonic resonance energy transfer (PRET)-based biosensing system for TB monitoring. Reprinted with permission from [52]. Copyright 2014, American Chemical Society.

Recently, other type of plasmon resonance mode also could be applied for biosensors such as Fano resonance. According to several literatures, the sensitivity of Fano resonance based detection system could be higher than that of conventional SPR based biosensing system [59–61].

3.2. SERS-Based Sensing Platform

The SERS effect on P-NMs has a remarkable optical property, and scientists have used it to develop a biosensing platform [62–65]. Zou et al. fabricated the SERS platform using an iron oxide-Au magnetoplasmonic NP (IO-Au) core/shell P-NM (Figure 6) [66]. Interestingly, IO-Au could be aligned and deposited on the substrate surface through an external magnetic field due to its magnetic property. Therefore, the SERS platform could be easily fabricated without expensive equipment or facilities. In their study, TB antigens were monitored using an IO-Au-based SERS platform and GRP QD. To detect a TB antigen, the TB Ab was modified on the IO-Au and GRP QD surface, and the Raman intensity of the GRP QD was monitored based on the TB antigen concentrations. Here, the linear calibration curve was obtained, and the Raman intensity of the GRP QD increased with an increase in TB antigen. The LOD was calculated at 0.0511 pg/mL. In another study, a reusable AgNP-GRP-based SERS platform was fabricated to detect methylated DNA [67]. The target DNA was captured by Ab-modified AuNPs, and this complex was attached to the surface of the AgNP-GRP-based sensing platform. The Raman intensity changed with the level of DNA methylation, and large increases in the Raman intensity were measured at higher methylation levels. In addition, as the DNA concentration increased, the Raman signal also increased. In this case, the LOD was estimated at 1.8 pg/mL, and the SERS platform was reused after washing it with deionized water and ethanol.

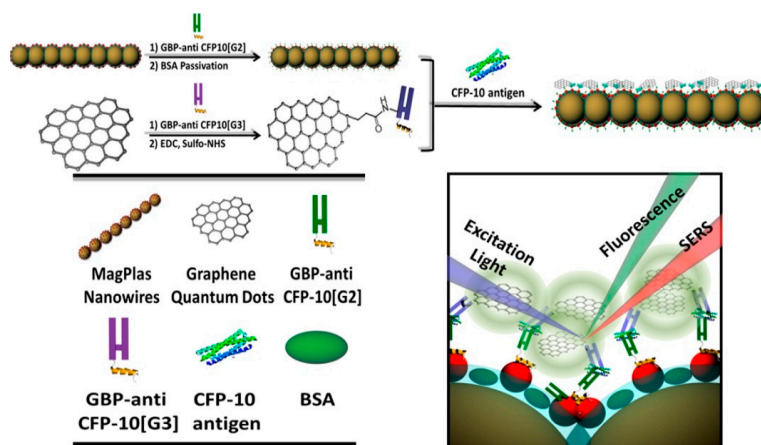


Figure 6. Illustration of the detection process for surface-enhanced Raman scattering (SERS)-based TB biosensing. Reprinted with permission from [66]. Copyright 2015, American Chemical Society.

3.3. Enzyme Like Activity-Based Colorimetry Sensing Platform

Such P-NM-based plasmonic structures could be applied to a colorimetry sensing platform due to the catalytic properties from free electrons [68]. This means that P-NM possesses enzyme activities, and this material-based sensing system can be an alternative to the enzyme-linked immunosorbent assay (ELISA) system [68]. For example, Rahin and colleagues detected Norovirus-like particles (No-VLPs) using Au-GRP as an enzyme-like catalyst (Figure 7) [69]. First, a specificity test was confirmed on various viruses and a color change was only observed in No-VLP (Figure 7a). Ab conjugation with Au-GRP was confirmed by FT-IR analysis (Figure 7b). In this case, Au-GRP induced the oxidation of the peroxidase substrate 3,3',5,5'-tetramethylbenzidine (TMB) with H_2O_2 , resulting in the appearance of a blue color in the aqueous solution (Figure 7c). Through an improved colorimetry sensing response, the LOD for the No-VLPs in this system was improved to 92.7 pg/mL, which is 112 times higher than that of conventional ELISA (Figure 7d).

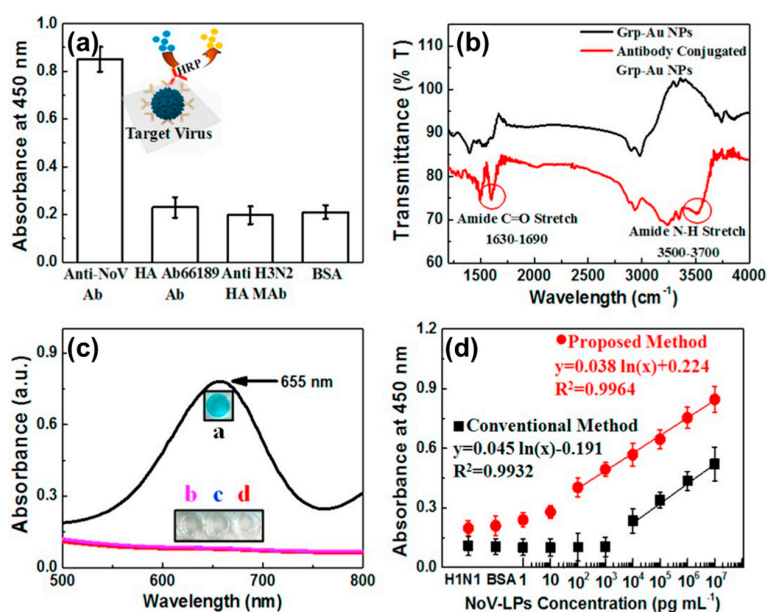


Figure 7. P-NM-based plasmonic application on colorimetry sensing platform. (a) Selectivity test with various viruses; (b) Confirmation of Ab conjugation with Au-GRP; (c) Catalytic activity test and (d) Biosensing demonstration with various concentrations of Norovirus-like particles (No-VLPs). Copyright Elsevier B.V. Reproduced with permission [69].

In a similar process, the influenza virus was monitored by Au-CNT-induced catalytic activity [70]. This case was highly sensitive at a 3.4 PFU/mL detection limitation, which is 385 times that of the ELISA detection limitation and 500 times that of commercial immunochromatography kits.

4. Conclusions

In recent years, many P-NMs have been developed for various applications, and their plasmonic properties have been improved and enhanced to achieve effective performance. Several P-NMs, such as MNPs, bMNPs, MNP-CNT, and MNP-GRP, were applied for novel biosensing platforms, such as the LSPR-based detection system, the SERS substrate, the PRET-induced sensing system, and colorimetry. Moreover, P-NM-based biosensing systems can detect antigens, DNA, and RNA in various diseases with excellent sensitivity and selectivity. Therefore, P-NM-based detection platforms can be widely used as highly accurate biosensing devices for public health.

Acknowledgments: This research was supported by the Japan Society for the Promotion of Science (JSPS) Postdoctoral Fellowship for Overseas Researchers (16F16361).

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviation

Antibodies (Ab); Bimetallic nanoparticles (bMNP); Carbon nanotubes (CNT); enzyme-linked immunosorbent assay (ELISA); Graphene (GRP); Infra-red (IR); Iron oxide-Au magnetoplasmonic NP (IO-Au); Limit of detection (LOD); Localized surface plasmon resonance (LSPR); Magneto optical (MO); Metal nanoparticles (MNP); MNP decorated carbon nanomaterial (MNP-CNM); MNP-decorated CNT (MNP-CNT); MNP-modified GRP (MNP-GRP); *N*-(3-Dimethylaminopropyl)-*N'*-ethylcarbodiimide (EDC); Norovirus-like particles (No-VLPs); Plasmonic nanomaterials (P-NM); Plasmonic resonance energy transfer (PRET); Quantum dot (QD); Surface-enhanced Raman scattering (SERS); Tuberculosis (TB); 3,3,5,5-tetramethylbenzidine (TMB).

References

1. Kelly, K.L.; Coronado, E.; Zhao, L.L.; Schatz, G.C. The optical properties of metal nanoparticles: The influence of size, shape, and dielectric environment. *J. Phys. Chem. B* **2003**, *107*, 668–677. [[CrossRef](#)]
2. Michaels, A.M.; Nirmal, M.; Brus, L.E. Surface enhanced raman spectroscopy of individual rhodamine 6G molecules on large Ag nanocrystals. *J. Am. Chem. Soc.* **1999**, *121*, 9932–9939. [[CrossRef](#)]
3. Tanabe, K. Field enhancement around metal nanoparticles and nanoshells: A systematic investigation. *J. Phys. Chem. C* **2008**, *112*, 15721–15728. [[CrossRef](#)]
4. Govorov, A.O.; Zhang, H.; Demir, H.V.; Gun'ko, Y.K. Photogeneration of hot plasmonic electrons with metal nanocrystals: Quantum description and potential applications. *Nano Today* **2014**, *9*, 85–101. [[CrossRef](#)]
5. Ni, G.X.; Wang, L.; Goldflam, M.D.; Wagner, M.; Fei, Z.; McLeod, A.S.; Liu, M.K.; Keilmann, F.; Özyilmaz, B.; Castro Neto, A.H.; et al. Ultrafast optical switching of infrared plasmon polaritons in high-mobility graphene. *Nat. Photonics* **2016**, *10*, 244–247. [[CrossRef](#)]
6. Chen, J.; Badioli, M.; Alonso-Gonzalez, P.; Thongrattanasiri, S.; Huth, F.; Osmond, J.; Spasenovic, M.; Centeno, A.; Pesquera, A.; Godignon, P.; et al. Optical nano-imaging of gate-tunable graphene plasmons. *Nature* **2012**, *487*, 77–81. [[CrossRef](#)] [[PubMed](#)]
7. García de Abajo, F.J. Graphene plasmonics: Challenges and opportunities. *ACS Photonics* **2014**, *1*, 135–152. [[CrossRef](#)]
8. Grigorenko, A.N.; Polini, M.; Novoselov, K.S. Graphene plasmonics. *Nat. Photonics* **2012**, *6*, 749–758. [[CrossRef](#)]
9. Tokarev, I.; Tokareva, I.; Minko, S. Gold-nanoparticle-enhanced plasmonic effects in a responsive polymer gel. *Adv. Mater.* **2008**, *20*, 2730–2734. [[CrossRef](#)] [[PubMed](#)]
10. Shipway, A.N.; Lahav, M.; Gabai, R.; Willner, I. Investigations into the electrostatically induced aggregation of Au nanoparticles. *Langmuir* **2000**, *16*, 8789–8795. [[CrossRef](#)]
11. Lin, S.; Li, M.; Dujardin, E.; Girard, C.; Mann, S. One-dimensional plasmon coupling by facile self-assembly of gold nanoparticles into branched chain networks. *Adv. Mater.* **2005**, *17*, 2553–2559. [[CrossRef](#)]

12. Issaad, D.; Moustauoui, H.; Medjahed, A.; Lalaoui, L.; Spadavecchia, J.; Bouafia, M.; de la Chapelle, M.L.; Djaker, N. Scattering correlation spectroscopy and raman spectroscopy of thiophenol on gold nanoparticles: comparative study between nanospheres and nanourchins. *J. Phys. Chem. C* **2017**, *121*, 18254–18262. [[CrossRef](#)]
13. Im, H.; Bantz, K.C.; Lindquist, N.C.; Haynes, C.L.; Oh, S.-H. Vertically oriented sub-10-nm plasmonic nanogap arrays. *Nano Lett.* **2010**, *10*, 2231–2236. [[CrossRef](#)] [[PubMed](#)]
14. Lee, J.; Lee, J. Magneto-optically active magnetoplasmonic graphene. *Chem. Commun.* **2017**, *53*, 5814–5817. [[CrossRef](#)] [[PubMed](#)]
15. Lee, J.; Lee, K.; Park, S. Environmentally friendly preparation of nanoparticle-decorated carbon nanotube or graphene hybrid structures and their potential applications. *J. Mater. Sci.* **2016**, *51*, 2761–2770. [[CrossRef](#)]
16. Jia, K.; Khaywah, M.Y.; Li, Y.; Bijeon, J.L.; Adam, P.M.; D eturche, R.; Guelorget, B.; Fran ois, M.; Louarn, G.; Ionescu, R.E. Strong improvements of localized surface plasmon resonance sensitivity by using Au/Ag bimetallic nanostructures modified with polydopamine films. *ACS Appl. Mater. Interfaces* **2014**, *6*, 219–227. [[CrossRef](#)] [[PubMed](#)]
17. Rao, V.K.; Ghildiyal, P.; Radhakrishnan, T.P. In situ fabricated Cu–Ag nanoparticle-embedded polymer thin film as an efficient broad spectrum SERS substrate. *J. Phys. Chem. C* **2017**, *121*, 1339–1348. [[CrossRef](#)]
18. Raghuv eer, M.S.; Agrawal, S.; Bishop, N.; Ramanath, G. Microwave-assisted single-step functionalization and in situ derivatization of carbon nanotubes with gold nanoparticles. *Chem. Mater.* **2006**, *18*, 1390–1393. [[CrossRef](#)]
19. Hussain, N.; Gogoi, A.; Sarma, R.K.; Sharma, P.; Barras, A.; Boukherroub, R.; Saikia, R.; Sengupta, P.; Das, M.R. Reduced graphene oxide nanosheets decorated with Au nanoparticles as an effective bactericide: Investigation of biocompatibility and leakage of sugars and proteins. *ChemPlusChem* **2014**, *79*, 1774–1784. [[CrossRef](#)]
20. Baro, M.; Nayak, P.; Baby, T.T.; Ramaprabhu, S. Green approach for the large-scale synthesis of metal/metal oxide nanoparticle decorated multiwalled carbon nanotubes. *J. Mater. Chem. A* **2013**, *1*, 482–486. [[CrossRef](#)]
21. Lee, J.; Park, E.Y.; Lee, J. Non-toxic nanoparticles from phytochemicals: Preparation and biomedical application. *Bioprocess Biosyst. Eng.* **2014**, *37*, 983–989. [[CrossRef](#)] [[PubMed](#)]
22. Sidorov, A.N.; S lawi nski, G.W.; Jayatissa, A.H.; Zamborini, F.P.; Sumanasekera, G.U. A surface-enhanced Raman spectroscopy study of thin graphene sheets functionalized with gold and silver nanostructures by seed-mediated growth. *Carbon* **2012**, *50*, 699–705. [[CrossRef](#)]
23. Lee, Y.H.; Polavarapu, L.; Gao, N.; Yuan, P.; Xu, Q.-H. Enhanced optical properties of graphene oxide–Au nanocrystal composites. *Langmuir* **2012**, *28*, 321–326. [[CrossRef](#)] [[PubMed](#)]
24. Seger, B.; Kamat, P.V. Electrocatalytically active graphene-platinum nanocomposites. Role of 2-D carbon support in PEM fuel cells. *J. Phys. Chem. C* **2009**, *113*, 7990–7995. [[CrossRef](#)]
25. Belotelov, V.I.; Akimov, I.A.; Pohl, M.; Kotov, V.A.; Kasture, S.; Vengurlekar, A.S.; Gopal, A.V.; Yakovlev, D.R.; Zvezdin, A.K.; Bayer, M. Enhanced magneto-optical effects in magnetoplasmonic crystals. *Nat. Nanotechnol.* **2011**, *6*, 370–376. [[CrossRef](#)] [[PubMed](#)]
26. Gonz alez-D iaz, J.B.; Garc a-Mart n, A.; Garc a-Mart n, J.M.; Cebollada, A.; Armelles, G.; Sep lveda, B.; Alaverdyan, Y.; K all, M. Plasmonic Au/Co/Au nanosandwiches with enhanced magneto-optical activity. *Small* **2008**, *4*, 202–205. [[CrossRef](#)] [[PubMed](#)]
27. Gong, J.; Zhou, T.; Song, D.; Zhang, L. Monodispersed Au nanoparticles decorated graphene as an enhanced sensing platform for ultrasensitive stripping voltammetric detection of mercury(II). *Sens. Actuators B Chem.* **2010**, *150*, 491–497. [[CrossRef](#)]
28. Lee, J.; Kim, J.; Go, J.; Lee, J.H.; Han, D.-W.; Hwang, D.; Lee, J. Transdermal treatment of the surgical and burned wound skin via phytochemical-capped gold nanoparticles. *Colloid. Surf. B-Biointerfaces* **2015**, *135*, 166–174. [[CrossRef](#)] [[PubMed](#)]
29. Lee, J.; Sun, F.; Lee, J. Fabrication of large area flexible and highly transparent film by a simple Ag nanowire alignment. *J. Exp. Nanosci.* **2013**, *8*, 130–137. [[CrossRef](#)]
30. Tran, V.T.; Zhou, H.; Kim, S.; Lee, J.; Kim, J.; Zou, F.; Kim, J.; Park, J.Y.; Lee, J. Self-assembled magnetoplasmonic nanochain for DNA sensing. *Sens. Actuators B Chem.* **2014**, *203*, 817–823. [[CrossRef](#)]

31. Guo, S.; Sun, S. FePt Nanoparticles assembled on graphene as enhanced catalyst for oxygen reduction reaction. *J. Am. Chem. Soc.* **2012**, *134*, 2492–2495. [[CrossRef](#)] [[PubMed](#)]
32. Jun, Y.; Bin, F.; Jin, L.; Bridgid, W.; Derrick, M.; Rameshowri, L.; Mei Shan, N.; Zheng, L.; Jian, H.; Whittingham, M.S.; et al. Nanoscale alloying effect of gold–platinum nanoparticles as cathode catalysts on the performance of a rechargeable lithium–oxygen battery. *Nanotechnology* **2012**, *23*, 305404.
33. Spackova, B.; Wrobel, P.; Bockova, M.; Homola, J. Optical biosensors based on plasmonic nanostructures: A review. *Proc. IEEE* **2016**, *104*, 2380–2408. [[CrossRef](#)]
34. Cheon, S.A.; Cho, H.H.; Kim, J.; Lee, J.; Kim, H.-J.; Park, T.J. Recent tuberculosis diagnosis toward the end TB strategy. *J. Microbiol. Methods* **2016**, *123*, 51–61. [[CrossRef](#)] [[PubMed](#)]
35. Ahmed, S.R.; Kim, J.; Tran, V.T.; Suzuki, T.; Neethirajan, S.; Lee, J.; Park, E.Y. In situ self-assembly of gold nanoparticles on hydrophilic and hydrophobic substrates for influenza virus-sensing platform. *Sci. Rep.* **2017**, *7*, 44495. [[CrossRef](#)] [[PubMed](#)]
36. Kim, J.; Jang, M.; Lee, K.G.; Lee, K.-S.; Lee, S.J.; Ro, K.-W.; Kang, I.S.; Jeong, B.D.; Park, T.J.; Kim, H.-J.; et al. Plastic-chip-based magnetophoretic immunoassay for point-of-care diagnosis of tuberculosis. *ACS Appl. Mater. Interfaces* **2016**, *8*, 23489–23497. [[CrossRef](#)] [[PubMed](#)]
37. Kim, J.; Lee, J.; Lee, K.-I.; Park, T.J.; Kim, H.-J.; Lee, J. Rapid monitoring of CFP-10 during culture of mycobacterium tuberculosis by using a magnetophoretic immunoassay. *Sens. Actuators B Chem.* **2013**, *177*, 327–333. [[CrossRef](#)]
38. Lee, J.; Takemura, K.; Kato, C.N.; Suzuki, T.; Park, E.Y. Binary nanoparticle-graphene hybrid structure-based highly sensitive biosensing platform for norovirus-like particle detection. *ACS Appl. Mater. Interfaces* **2017**, *9*, 27298–27304. [[CrossRef](#)] [[PubMed](#)]
39. Ahmed, S.R.; Hossain, M.A.; Park, J.Y.; Kim, S.-H.; Lee, D.; Suzuki, T.; Lee, J.; Park, E.Y. Metal enhanced fluorescence on nanoporous gold leaf-based assay platform for virus detection. *Biosens. Bioelectron.* **2014**, *58*, 33–39. [[CrossRef](#)] [[PubMed](#)]
40. Adegoke, O.; Park, E.Y. Gold nanoparticle-quantum dot fluorescent nanohybrid: Application for localized surface plasmon resonance-induced molecular beacon ultrasensitive DNA detection. *Nanoscale Res. Lett.* **2016**, *11*, 523. [[CrossRef](#)] [[PubMed](#)]
41. Hou, W.; Cronin, S.B. A review of surface plasmon resonance-enhanced photocatalysis. *Adv. Funct. Mater.* **2013**, *23*, 1612–1619. [[CrossRef](#)]
42. Wang, C.; van der Vliet, D.; More, K.L.; Zaluzec, N.J.; Peng, S.; Sun, S.; Daimon, H.; Wang, G.; Greeley, J.; Pearson, J.; et al. Multimetallic Au/FePt₃ nanoparticles as highly durable electrocatalyst. *Nano Lett.* **2011**, *11*, 919–926. [[CrossRef](#)] [[PubMed](#)]
43. Desireddy, A.; Conn, B.E.; Guo, J.; Yoon, B.; Barnett, R.N.; Monahan, B.M.; Kirschbaum, K.; Griffith, W.P.; Whetten, R.L.; Landman, U.; et al. Ultrastable silver nanoparticles. *Nature* **2013**, *501*, 399–402. [[CrossRef](#)] [[PubMed](#)]
44. Rodrigo, D.; Limaj, O.; Janner, D.; Etezadi, D.; García de Abajo, F.J.; Pruneri, V.; Altug, H. Mid-infrared plasmonic biosensing with graphene. *Science* **2015**, *349*, 165–168. [[CrossRef](#)] [[PubMed](#)]
45. Oh, S.; Jang, M.; Kim, J.; Lee, J.; Zhou, H.; Lee, J. Synthesis of silver nanoparticles using analogous reducibility of phytochemicals. *Curr. Appl. Phys.* **2016**, *16*, 738–747. [[CrossRef](#)]
46. Lee, J.; Zhou, H.; Lee, J. Small molecule induced self-assembly of Au nanoparticles. *J. Mater. Chem.* **2011**, *21*, 16935. [[CrossRef](#)]
47. Zhou, H.; Kim, J.-P.; Bahng, J.H.; Kotov, N.A.; Lee, J. Self-assembly mechanism of spiky magnetoplasmonic supraparticles. *Adv. Funct. Mater.* **2014**, *24*, 1439–1448. [[CrossRef](#)]
48. Adegoke, O.; Morita, M.; Kato, T.; Ito, M.; Suzuki, T.; Park, E.Y. Localized surface plasmon resonance-mediated fluorescence signals in plasmonic nanoparticle-quantum dot hybrids for ultrasensitive Zika virus RNA detection via hairpin hybridization assays. *Biosens. Bioelectron.* **2017**, *94*, 513–522. [[CrossRef](#)] [[PubMed](#)]
49. Maccaferri, N.; Gregorczyk, K.E.; de Oliveira, T.; Kataja, M.; van Dijken, S.; Pirzadeh, Z.; Dmitriev, A.; Akerman, J.; Knez, M.; Vavassori, P. Ultrasensitive and label-free molecular-level detection enabled by light phase control in magnetoplasmonic nanoantennas. *Nat. Commun.* **2015**, *6*, 6150. [[CrossRef](#)] [[PubMed](#)]

50. Melnikau, D.; Govyadinov, A.A.; Sánchez-Iglesias, A.; Grzelczak, M.; Liz-Marzán, L.M.; Rakovich, Y.P. Strong magneto-optical response of nonmagnetic organic materials coupled to plasmonic nanostructures. *Nano Lett.* **2017**, *17*, 1808–1813. [[CrossRef](#)] [[PubMed](#)]
51. Lee, J.; Ahmed, S.R.; Oh, S.; Kim, J.; Suzuki, T.; Parmar, K.; Park, S.S.; Lee, J.; Park, E.Y. A plasmon-assisted fluoro-immunoassay using gold nanoparticle-decorated carbon Nanotubes for monitoring the influenza virus. *Biosens. Bioelectron.* **2015**, *64*, 311–317. [[CrossRef](#)] [[PubMed](#)]
52. Lee, J.; Kim, J.; Ahmed, S.R.; Zhou, H.; Kim, J.-M.; Lee, J. Plasmon-induced photoluminescence immunoassay for tuberculosis monitoring using gold nanoparticles decorated graphene. *ACS Appl. Mater. Interfaces* **2014**, *6*, 21380–21388. [[CrossRef](#)] [[PubMed](#)]
53. Zhang, J.; Zhang, X.; Lai, C.; Zhou, H.; Zhu, Y. Silver-decorated aligned CNT arrays as SERS substrates by high temperature annealing. *Optics Express* **2014**, *22*, 21157–21166. [[CrossRef](#)] [[PubMed](#)]
54. Ngo, H.T.; Gandra, N.; Fales, A.M.; Taylor, S.M.; Vo-Dinh, T. Sensitive DNA detection and SNP discrimination using ultrabright SERS nanorattles and magnetic beads for malaria diagnostics. *Biosens. Bioelectron.* **2016**, *81* (Suppl. C), 8–14. [[CrossRef](#)] [[PubMed](#)]
55. Takemura, K.; Adegoke, O.; Takahashi, N.; Kato, T.; Li, T.-C.; Kitamoto, N.; Tanaka, T.; Suzuki, T.; Park, E.Y. Versatility of a localized surface plasmon resonance-based gold nanoparticle-alloyed quantum dot nanobiosensor for immunofluorescence detection of viruses. *Biosens. Bioelectron.* **2017**, *89*, 998–1005. [[CrossRef](#)] [[PubMed](#)]
56. Lilly, G.D.; Lee, J.; Kotov, N.A. “Cloud” assemblies: Quantum dots form electrostatically bound dynamic nebulae around large gold nanoparticles. *Phys. Chem. Chem. Phys.* **2010**, *12*, 11878–11884. [[CrossRef](#)] [[PubMed](#)]
57. Lee, J.; Hernandez, P.; Lee, J.; Govorov, A.O.; Kotov, N.A. Exciton-plasmon interactions in molecular spring assemblies of nanowires and wavelength-based protein detection. *Nat. Mater.* **2007**, *6*, 291–295. [[CrossRef](#)] [[PubMed](#)]
58. Lee, J.; Govorov, A.O.; Kotov, N.A. Nanoparticle assemblies with molecular springs: A nanoscale thermometer. *Angew. Chem. Int. Ed.* **2005**, *44*, 7439–7442. [[CrossRef](#)] [[PubMed](#)]
59. Lee, K.-L.; Huang, J.-B.; Chang, J.-W.; Wu, S.-H.; Wei, P.-K. Ultrasensitive biosensors using enhanced fano resonances in capped gold nanoslit arrays. *Sci. Rep.* **2015**, *5*, 8547. [[CrossRef](#)] [[PubMed](#)]
60. Wu, C.; Khanikae, A.B.; Adato, R.; Arju, N.; Yanik, A.A.; Altug, H.; Shvets, G. Fano-resonant asymmetric metamaterials for ultrasensitive spectroscopy and identification of molecular monolayers. *Nat. Mater.* **2012**, *11*, 69–75. [[CrossRef](#)] [[PubMed](#)]
61. Ruan, B.; Guo, J.; Wu, L.; Zhu, J.; You, Q.; Dai, X.; Xiang, Y. Ultrasensitive terahertz biosensors based on fano resonance of a graphene/waveguide hybrid structure. *Sensors* **2017**, *17*, 1924. [[CrossRef](#)] [[PubMed](#)]
62. Laing, S.; Gracie, K.; Faulds, K. Multiplex in vitro detection using SERS. *Chem. Soc. Rev.* **2016**, *45*, 1901–1918. [[CrossRef](#)] [[PubMed](#)]
63. Marks, H.; Schechinger, M.; Garza, J.; Locke, A.; Côté, G. Surface enhanced raman spectroscopy (SERS) for in vitro diagnostic testing at the point of care. *Nanophotonics* **2017**, *6*, 681. [[CrossRef](#)]
64. Ngo, H.T.; Wang, H.-N.; Fales, A.M.; Vo-Dinh, T. Plasmonic SERS biosensing nanochips for DNA detection. *Anal. Bioanal. Chem.* **2016**, *408*, 1773–1781. [[CrossRef](#)] [[PubMed](#)]
65. Kahraman, M.; Mullen Emma, R.; Korkmaz, A.; Wachsmann-Hogiu, S. Fundamentals and applications of SERS-based bioanalytical sensing. *Nanophotonics* **2017**, *6*, 831. [[CrossRef](#)]
66. Zou, F.; Zhou, H.; Tan, T.V.; Kim, J.; Koh, K.; Lee, J. Dual-mode SERS-fluorescence immunoassay using graphene quantum dot labeling on one-dimensional aligned magnetoplasmonic nanoparticles. *ACS Appl. Mater. Interfaces* **2015**, *7*, 12168–12175. [[CrossRef](#)] [[PubMed](#)]
67. Ouyang, L.; Hu, Y.; Zhu, L.; Cheng, G.J.; Irudayaraj, J. A reusable laser wrapped graphene-Ag array based SERS sensor for trace detection of genomic DNA methylation. *Biosens. Bioelectron.* **2017**, *92*, 755–762. [[CrossRef](#)] [[PubMed](#)]
68. Ahmed, S.R.; Kim, J.; Suzuki, T.; Lee, J.; Park, E.Y. Detection of influenza virus using peroxidase-mimic of gold nanoparticles. *Biosens. Bioelectron.* **2016**, *113*, 2298–2303. [[CrossRef](#)] [[PubMed](#)]

69. Ahmed, S.R.; Takemeura, K.; Li, T.-C.; Kitamoto, N.; Tanaka, T.; Suzuki, T.; Park, E.Y. Size-controlled preparation of peroxidase-like graphene-gold nanoparticle hybrids for the visible detection of norovirus-like particles. *Biosens. Bioelectron.* **2017**, *87*, 558–565. [[CrossRef](#)] [[PubMed](#)]
70. Ahmed, S.R.; Kim, J.; Suzuki, T.; Lee, J.; Park, E.Y. Enhanced catalytic activity of gold nanoparticle-carbon nanotube hybrids for influenza virus detection. *Biosens. Bioelectron.* **2016**, *85*, 503–508. [[CrossRef](#)] [[PubMed](#)]



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