

Use of Mac-2 Binding Protein as a Biomarker for Nonalcoholic Fatty Liver Disease Diagnosis

Yoshihiro Kamada,^{1,2} Masafumi Ono,³ Hideyuki Hyogo,⁴ Hideki Fujii,⁵ Yoshio Sumida,⁶ Makoto Yamada,⁷ Kojiroh Mori,⁸ Saiyu Tanaka,⁸ Tomohiro Maekawa,¹ Yusuke Ebisutani,¹ Akiko Yamamoto,¹ Shinji Takamatsu,¹ Masashi Yoneda,⁶ Norifumi Kawada,⁵ Kazuaki Chayama,⁹ Toshiji Saibara,³ Tetsuo Takehara,² and Eiji Miyoshi¹,
Japan Study Group of Nonalcoholic Fatty Liver Disease (JSG-NAFLD)

In contrast to patients with viral hepatitis, patients with nonalcoholic fatty liver disease (NAFLD) can progress to hepatocellular carcinoma during the initial stages of liver fibrosis. Development and implementation of noninvasive methods for diagnosis and progression prediction are important for effective NAFLD surveillance. Mac-2 binding protein (Mac-2bp) is a useful nonalcoholic steatohepatitis (NASH) diagnosis biomarker and a powerful prediction biomarker for NAFLD fibrosis stage. *Wisteria floribunda* agglutinin (WFA)-positive Mac-2bp (WFA⁺-M2BP) is a novel serum fibrosis biomarker for chronic hepatitis C that has clinical validity. Mac-2bp and WFA⁺-M2BP are also clinical NAFLD biomarker candidates. We examined the efficacy of Mac-2bp and WFA⁺-M2BP for NAFLD assessment using patients with biopsy-proven NAFLD (n = 510; NAFLD cohort) and subjects who received a health check-up (n = 2,122; check-up cohort). In the NAFLD cohort, we set the fibrosis predicting cutoff values as 1.80 (F1), 2.21 (F2), and 2.24 $\mu\text{g/mL}$ (F3). In the subjects with fatty liver from the check-up cohort (n = 1,291), the serum Mac-2bp levels were $>1.80 \mu\text{g/mL}$ in 38.6% of the subjects (n = 498), and $>2.24 \mu\text{g/mL}$ in 24.6% of the subjects (n = 318). The NAFLD cohort results indicated that Mac-2bp and WFA⁺-M2BP were equally useful for NASH diagnosis. During the early stages of fibrosis (F1, F2), the increase in Mac-2bp was statistically significant but WFA⁺-M2BP did not increase. Logistic regression analysis revealed that Mac-2bp was an independent determinant for the prediction of advanced fibrosis stage ($\geq\text{F2}$), even when adjusted for WFA⁺-M2BP. Immunohistochemical staining of Mac-2bp revealed that hepatocytes strongly expressed Mac-2bp in patients with NAFLD. **Conclusion:** Our results indicated that hepatocyte-derived Mac-2bp would be a useful single biomarker for NASH diagnosis and fibrosis stage prediction in patients with NAFLD. (*Hepatology Communications* 2017;1:780-791)

Introduction

Nonalcoholic fatty liver disease (NAFLD) is among the most common causes of chronic liver disease worldwide and is a growing medical problem in industrialized countries.⁽¹⁾ A wide spectrum of hepatic histologic changes occurs in

patients with NAFLD. These changes range from the generally nonprogressive nonalcoholic fatty liver (NAFL) to nonalcoholic steatohepatitis (NASH). During the clinical progression of NAFLD, assessment of liver fibrosis degree is important to predict further progression and to make therapeutic management decisions.^(2,3) Hepatocellular carcinoma (HCC)

Abbreviations: Alb, albumin; ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; AUROC, area under the receiver operating characteristic curve; BMI, body mass index; CHC, chronic hepatitis type C; COI, cutoff index; FBS, fasting blood glucose; FIB4, fibrosis-4; HCC, hepatocellular carcinoma; HSC, hepatic stellate cell; IHC, immunohistochemical; IRL, immunoreactive insulin; Mac-2bp, Mac-2 binding protein; NAFL, nonalcoholic fatty liver; NAFLD, nonalcoholic fatty liver disease; NAS, NAFLD activity scoring; NASH, nonalcoholic steatohepatitis; NC, negative control; NFS, NAFLD fibrosis score; PC, positive control; PNPLA3, patatin-like phospholipase domain containing 3; WFA, *Wisteria floribunda* agglutinin; WFA⁺-M2BP, WFA-positive Mac-2bp.

Received May 24, 2017; accepted July 14, 2017.

Additional Supporting Information may be found at onlinelibrary.wiley.com/doi/10.1002/hep4.1080/full.

Supported by a Japan Society for the Promotion of Science KAKENHI grant (grant number 15H04810, 16H05226, and 16K15428), the Program for Basic and Clinical Research on Hepatitis from the Japan Agency for Medical Research and Development, and research funding from Sysmex Co. (Kobe, Hyogo, Japan) (number J770701633).

incidence increases with the progression of liver fibrosis in chronic hepatitis type C (CHC) patients.⁽⁴⁻⁶⁾ In contrast with viral hepatitis, patients with NAFLD can progress to HCC during the early stages of liver fibrosis.⁽⁷⁻¹⁰⁾ Therefore, diagnosis of NASH during the early stages is important during clinical follow-up of the patient. Liver biopsy remains the gold standard for liver fibrosis assessment.^(11,12) However, liver biopsy has significant limitations, including pain, risk of severe complications, sampling error,⁽¹³⁾ cost,⁽¹⁴⁾ and patient unwillingness to undergo invasive testing. Recently, Angulo et al.⁽¹⁵⁾ reported that liver fibrosis is independently associated with long-term outcome in patients with NAFLD. Therefore, there is an urgent need for a reliable and noninvasive test that can be used to accurately assess the degree of liver fibrosis.

Novel noninvasive approaches, e.g., transient elastography (FibroScan, acoustic radiation force impulse, and magnetic resonance elastography), and various scoring systems can be used to measure the severity of fibrosis in patients with chronic liver disease.⁽¹⁶⁻²¹⁾ However, transient elastography has reduced interobserver agreement in patients with early stage fibrosis, moderate to severe steatosis, or increased body mass index (BMI). Distinguishing early liver fibrosis (F1-F2) from the nonfibrotic liver is difficult when transient elastography is used. Various scoring systems for measurement of liver fibrosis severity are available as

noninvasive methods,⁽¹⁹⁻²⁵⁾ but few individual biomarkers have clinical validity.

Recently, we found that serum Mac-2 binding protein (Mac-2bp) levels can be used to predict histologic severity of hepatic fibrosis in patients with NAFLD.^(26,27) Mac-2bp is a glycoprotein with seven potential *N*-glycosylation sites.^(28,29) Serum Mac-2bp concentrations increase in patients with pancreatic, breast, and lung cancers, viral hepatitis, and autoimmune disease.⁽²⁸⁾ Mac-2bp is almost undetectable in normal liver but is easily detected in hepatocytes from CHC patients as liver fibrosis progresses.⁽³⁰⁾ Cheung et al.⁽³¹⁾ used proteome analysis and found that serum Mac-2bp is a potential marker of fibrosis progression in CHC patients. *Wisteria floribunda* agglutinin (WFA)-positive Mac-2bp (WFA⁺-M2BP) is a novel serum fibrosis biomarker for CHC.⁽³²⁾ This biomarker can distinguish the glycan structure of WFA-detectable Mac-2bp. It was developed using a glycan-based immunoassay for the assessment of liver fibrosis severity in CHC patients.⁽³²⁾ WFA⁺-M2BP can also be used for the assessment of liver fibrosis stage in patients with NAFLD.⁽³³⁾ Taken together, these findings indicate that serum Mac-2bp and WFA⁺-M2BP levels are useful liver fibrosis biomarkers. However, the validity of the use of these two biomarkers for NASH diagnosis and fibrosis stage prediction in patients with NAFLD is not fully understood.

Copyright © 2017 The Authors. *Hepatology Communications* published by Wiley Periodicals, Inc., on behalf of the American Association for the Study of Liver Diseases. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

View this article online at wileyonlinelibrary.com.

DOI 10.1002/hep4.1080

Potential conflict of interest: Tetsuo Takehara and Eiji Miyoshi received research funding from Sysmex Co. (Kobe, Hyogo, Japan). The other authors have no conflicts of interests to disclose.

ARTICLE INFORMATION:

From the Departments of ¹Molecular Biochemistry and Clinical Investigation and ²Gastroenterology and Hepatology, Osaka University, Graduate School of Medicine, Osaka, Japan; ³Department of Gastroenterology and Hepatology, Kochi Medical School, Kochi, Japan; ⁴Department of Gastroenterology and Hepatology, JA Hiroshima General Hospital, Hiroshima, Japan; ⁵Department of Hepatology, Osaka City University Graduate School of Medicine, Osaka, Japan; ⁶Division of Hepatology and Pancreatology, Department of Internal Medicine, Aichi Medical University School of Medicine, Aichi, Japan; ⁷aMs New Otani Clinic, Osaka, Japan; ⁸Center for Digestive and Liver Diseases, Nara City Hospital, Nara, Japan; ⁹Department of Gastroenterology and Metabolism, Institute of Biomedical and Health Sciences, Hiroshima University, Hiroshima, Japan.

ADDRESS CORRESPONDENCE AND REPRINT REQUESTS TO:

Eiji Miyoshi, M.D., Ph.D.
Department of Molecular Biochemistry & Clinical Investigation
Osaka University Graduate School of Medicine
1-7, Yamada-oka, Suita

Osaka 565-0871, Japan
E-mail: emiyoshi@sahs.med.osaka-u.ac.jp
Tel: +81-6-6879-2590

The objective of this study was to examine the efficacy of Mac-2bp and WFA⁺-M2BP for the assessment of NAFLD. We investigated these biomarkers in patients with biopsy-proven NAFLD (n = 510) and subjects who received a health check-up (n = 2,122).

Materials and Methods

ETHICAL COMMITTEE APPROVAL

The research and informed consent protocols were approved for use in a multicenter study by the institutional review boards at Osaka University Hospital, Kochi Medical School Hospital, Hiroshima University Hospital, Osaka City University Hospital, Nara City Hospital, and aMs New Otani Clinic. Written informed consent was obtained from each subject at the time of liver biopsy or at the time of enrollment at each institute. The study was conducted in accordance with the Helsinki Declaration.

PATIENTS WITH BIOPSY-PROVEN NAFLD

From 2002 to 2013, 510 patients (200 with NAFL and 310 with NASH) with biopsy-confirmed NAFLD who were treated at one of five hepatology centers in Japan (Osaka University Hospital, Kochi Medical School Hospital, Hiroshima University Hospital, Osaka City University Hospital, and Nara City Hospital) were enrolled in the study.

Each patient with biopsy-proven NAFLD had received a percutaneous liver needle biopsy. The liver samples were embedded in paraffin blocks following standard procedures and were stained with hematoxylin and eosin and Masson's trichrome stains. All biopsy specimens were centrally evaluated by two hepatic pathologists (Y.K. and H.F.) who were blinded to the clinical data. An adequate liver sample was defined as >1.5 cm in length or containing more than six portal tracts or both. Matteoni's classification was used to confirm the presence of NASH.⁽³⁴⁾ Patients with NAFLD with ballooning hepatocytes (Matteoni type 3) and patients with NAFLD with liver fibrosis (Matteoni type 4) were assigned to the NASH cohort. Patients whose liver biopsy analysis revealed simple steatosis or steatosis with nonspecific inflammation were assigned to the NAFL cohort. NAFLD activity scoring (NAS) was used to assess and quantify each

sample.⁽³⁵⁾ The stages of steatosis (0-3), lobular inflammation (0-2), and hepatocellular ballooning (0-2) were quantified. Individual fibrosis parameters were scored independently using the NASH Clinical Research Network scoring system.⁽³⁵⁾ Advanced fibrosis was classified as a stage 2-4 (F2-F4) disease. The exclusion criteria included a history of other hepatic disease, a substance abuse-induced hepatic disorder, and a history of alcohol abuse (>20 g of alcohol daily). In this study, we could measure both Mac-2bp and WFA⁺-M2BP in 116 of the 510 patients with NAFLD.

Mac-2bp IMMUNOSTAINING

Normal liver tissue was obtained from the livers of patients with hepatic colorectal carcinoma metastasis who had surgery at Osaka University Hospital. The liver biopsy specimens from the patients with NAFLD and the normal liver tissues were prepared using the immunohistochemical (IHC) staining VECTASTAIN ABC kit (Funakoshi, Co., Ltd. Tokyo, Japan). Mouse anti-human Mac-2bp antibody (1:200; ab67353; Abcam, Cambridge, MA) was used for immunodetection. We classified the IHC liver histology into three classes according to IHC intensity (negative/weak/strong; 0/1/2) (Supporting Figs. S1 and S2).

SUBJECTS IN MEDICAL HEALTH CHECK-UPS

A total of 2,122 (1,495 male; 627 female) of the 2,215 Japanese adult subjects (1,538 male; 677 female) who underwent a health check-up at the aMs New Otani Clinic (Osaka, Japan) between 2008 to 2011 were initially recruited into the study. The exclusion criteria included a history of hepatic disease, such as CHC or concurrent active hepatitis B (seropositive for hepatitis B surface antigen), autoimmune hepatitis, primary biliary cirrhosis, sclerosing cholangitis, hemochromatosis, α 1-antitrypsin deficiency, Wilson's disease, hepatic injury caused by substance abuse, or a current or previous history of daily consumption of >20 g alcohol. The diagnosis of fatty liver was based on the results of an abdominal ultrasound examination performed by trained technicians after exclusion of other etiologies of steatogenic liver disease. A fatty liver was defined as a liver parenchyma with an echogenicity greater than that of the kidney cortex, the presence of vascular blurring, and deep attenuation of the

ultrasound signal.^(36,37) Among the 2,122 subjects, 1,291 (1,022 male, 269 female) were diagnosed as having a fatty liver and 831 (473 male, 358 female) were diagnosed as not having a fatty liver, using abdominal ultrasound. Blood serum was collected from each subject during the health check-up and was kept frozen at -80°C until analysis.

ANTHROPOMETRIC AND LABORATORY EVALUATION

Anthropometric variables (height and weight) were measured while each subject was in the standing position. BMI was calculated as weight (kg) divided by the square of height in meters (m^2). Serum biochemical variables (aspartate aminotransferase [AST], alanine aminotransferase [ALT], γ -glutamyltransferase, alkaline phosphatase [ALP], choline esterase, total cholesterol, triglyceride, high-density lipoprotein cholesterol, fasting blood glucose [FBS], immunoreactive insulin [IRI], albumin [Alb], ferritin, hyaluronic acid, and platelet count) were measured using a conventional automated blood analyzer. The fibrosis-4 (FIB4) index (based on the age, serum AST and ALT levels, and platelet counts) was calculated for each of the subjects as reported ($\text{age} \times \text{AST} [\text{U/L}] / \text{platelet count} [\times 10^9/\text{L}] / \sqrt{\text{ALT} [\text{U/L}]}$).^(24,38) The NAFLD fibrosis score (NFS) was calculated for each of the subjects as reported ($-1.675 + 0.037 \times \text{age} [\text{years}] + 0.094 \times \text{BMI} [\text{kg}/\text{m}^2] + 1.13 \times \text{impaired fasting glycemia/diabetes mellitus} [\text{yes} = 1, \text{no} = 0] + 0.99 \times \text{AST}/\text{ALT} - 0.013 \times \text{platelet count} [\times 10^9/\text{L}] - 0.66 \times \text{Alb} [\text{g}/\text{dL}]$).⁽²⁰⁾ Impaired fasting glucose (IFG) was defined as FBS of 110–125 mg/dL. The presence of diabetes mellitus (DM) was defined as FBS ≥ 126 mg/dL, HbA1c (NGSP) $\geq 6.5\%$, or treatment with anti-diabetic drugs.

Mac-2bp AND WFA⁺-M2BP MEASUREMENT

We used an enzyme-linked immunosorbent assay kit (code #27362; Immuno-Biological Laboratory Co., Ltd., Fujioka, Japan) to measure serum Mac-2bp.⁽²⁶⁾ WFA⁺-M2BP quantification was based on a lectin-antibody sandwich immunoassay performed using a fully automatic immunoanalyzer (HISCL-2000i; Sysmex Co., Hyogo, Japan).⁽³²⁾ The measured values of WFA⁺-M2BP conjugated to WFA were indexed with the obtained values using the following equation:

$$\text{Cutoff index(COI)} = \frac{([\text{WFA}^+ - \text{M2BP}]_{\text{sample}} - [\text{WFA}^+ - \text{M2BP}]_{\text{NC}})}{([\text{WFA}^+ - \text{M2BP}]_{\text{PC}} - [\text{WFA}^+ - \text{M2BP}]_{\text{NC}})},$$

where $[\text{WFA}^+ - \text{M2BP}]_{\text{sample}}$ was the WFA⁺-M2BP count for the serum sample, PC was the positive control, and NC was the negative control. The PC was supplied as a calibration solution preliminarily standardized to yield a COI value of 1.0.⁽³⁹⁾ Measurement of WFA⁺-M2BP was supported by the Sysmex Co.

STATISTICAL ANALYSIS

Statistical analysis was performed using JMP Pro 12.0 software (SAS Institute Inc., Cary, NC). Results were expressed as mean \pm SD values. Statistical analysis included descriptive statistics, analysis of variance, the Wilcoxon and Kruskal–Wallis tests, the Pearson test, and Spearman R correlation analysis. The diagnostic performance of each marker was assessed using receiver operating characteristic (ROC) curves. The probabilities of true positive (sensitivity) and true negative (specificity) results were determined for selected cutoff values, and the area under the ROC curve (AUROC) was calculated for each index. The Youden index was used to identify the optimal cutoff points. Multivariate logistic regression analyses were performed to identify parameters that significantly contributed to the estimation of liver fibrosis severity. Differences were considered to be statistically significant at $P < 0.05$.

Results

CHARACTERISTICS OF THE SUBJECTS

The results for the clinical and biochemical characteristics of the individuals included in the study population (i.e., patients with biopsy-proven NAFLD [NAFLD cohort], subjects with health check-ups [check-up cohort]) are presented in Table 1. In the NAFLD cohort, the ratio of males was increased in the NASH patient group compared with the NAFL patient group. In the check-up cohort, the ratio of males was greater in the fatty liver subject group [FL(+)] compared with the group of subjects without fatty liver [FL(-)]. Age and BMI were greater in the patients with NASH and in the FL(+) subjects in the NAFLD cohort and check-up cohort, respectively. In the NAFLD cohort, the serum levels of AST, ALT,

TABLE 1. CLINICAL AND SEROLOGICAL CHARACTERISTICS OF THE SUBJECTS

Variables	Biopsy-Proven NAFLD Patients			Health Check-Up Subjects		
	NAFL	NASH	<i>P</i> Value*	Fatty Liver (-)	Fatty Liver (+)	<i>P</i> Value*
Number	200	310		831	1291	
Age (y)	50.7 ± 10.3	52.8 ± 14.2	<0.005	54.1 ± 8.7	54.6 ± 7.0	<0.05
Sex (M/F)	69/131	141/169	<0.0001	473/358	1022/269	<0.0001
BMI (kg/m ²)	26.0 ± 3.9	28.1 ± 5.1	<0.0005	21.9 ± 2.7	26.0 ± 3.6	<0.0001
AST (U/L)	42.0 ± 26.3	64.2 ± 44.1	<0.0001	24.3 ± 14.2	31.1 ± 16.8	<0.0001
ALT (U/L)	75.4 ± 49.8	98.0 ± 73.8	<0.01	23.4 ± 23.0	41.1 ± 24.0	<0.0001
GGT (U/L)	111.0 ± 143.1	88.5 ± 79.7	N.S.	50.5 ± 94.4	74.5 ± 94.2	<0.0001
ALP (U/L)	238.3 ± 85.4	274.9 ± 125.8	<0.05	200.9 ± 74.8	210.9 ± 58.0	<0.0001
CHE (U/L)	368.4 ± 75.3	363.4 ± 89.1	N.S.	316.1 ± 71.7	376.9 ± 67.2	<0.0001
T-Chol (mg/dL)	193.2 ± 36.7	206.1 ± 36.0	N.S.	194.9 ± 30.8	208.7 ± 35.3	<0.0001
TG (mg/dL)	136.2 ± 93.6	143.2 ± 64.4	N.S.	92.3 ± 80.3	155.4 ± 106.9	<0.0001
HDL-C (mg/dL)	49.2 ± 12.0	49.2 ± 15.2	N.S.	64.7 ± 14.3	54.3 ± 11.7	<0.0001
FBS (mg/dL)	98.6 ± 15.1	106.1 ± 25.0	N.S.	108.3 ± 28.1	120.2 ± 32.1	<0.0001
IRI (mU/mL)	10.1 ± 5.6	14.7 ± 9.1	<0.05	N.D.	N.D.	
Albumin (g/dL)	4.57 ± 0.38	4.42 ± 0.45	<0.05	4.24 ± 0.22	4.39 ± 0.22	<0.0001
Ferritin (ng/mL)	179.2 ± 127.3	287.0 ± 285.7	<0.005	N.D.	N.D.	
Hyaluronic acid (ng/mL)	37.4 ± 37.4	106.5 ± 206.6	<0.05	N.D.	N.D.	
Platelet count (× 10 ⁴ /μL)	23.8 ± 5.6	22.1 ± 6.9	N.S.	21.7 ± 5.4	22.0 ± 5.1	0.07
FIB4 index	1.00 ± 0.53	1.97 ± 1.47	<0.0001	1.39 ± 0.65	1.33 ± 0.70	<0.005
NFS	-2.40 ± 1.32	-1.56 ± 1.56	<0.0001	-1.71 ± 1.11	-1.53 ± 1.13	<0.0005
Mac-2bp (μg/mL)	1.46 ± 0.77	2.74 ± 1.52	<0.0001	1.11 ± 0.69	1.81 ± 1.09	<0.0001
WFA ⁺ -M2BP (COI)	N.D.	N.D.		0.51 ± 0.36	0.60 ± 0.31	<0.0001

Data are presented as the mean ± SD. **P* values correspond to the comparison between NAFL and NASH groups. Wilcoxon test for continuous factors and Pearson's chi-square test for categorical factors were used.

Abbreviations: CHE, choline esterase; GGT, gamma glutamyltranspeptidase; HDL-C, high-density lipoprotein cholesterol; N.D., not determined; N.S., not significant; T-Chol, total cholesterol; TG, triglyceride.

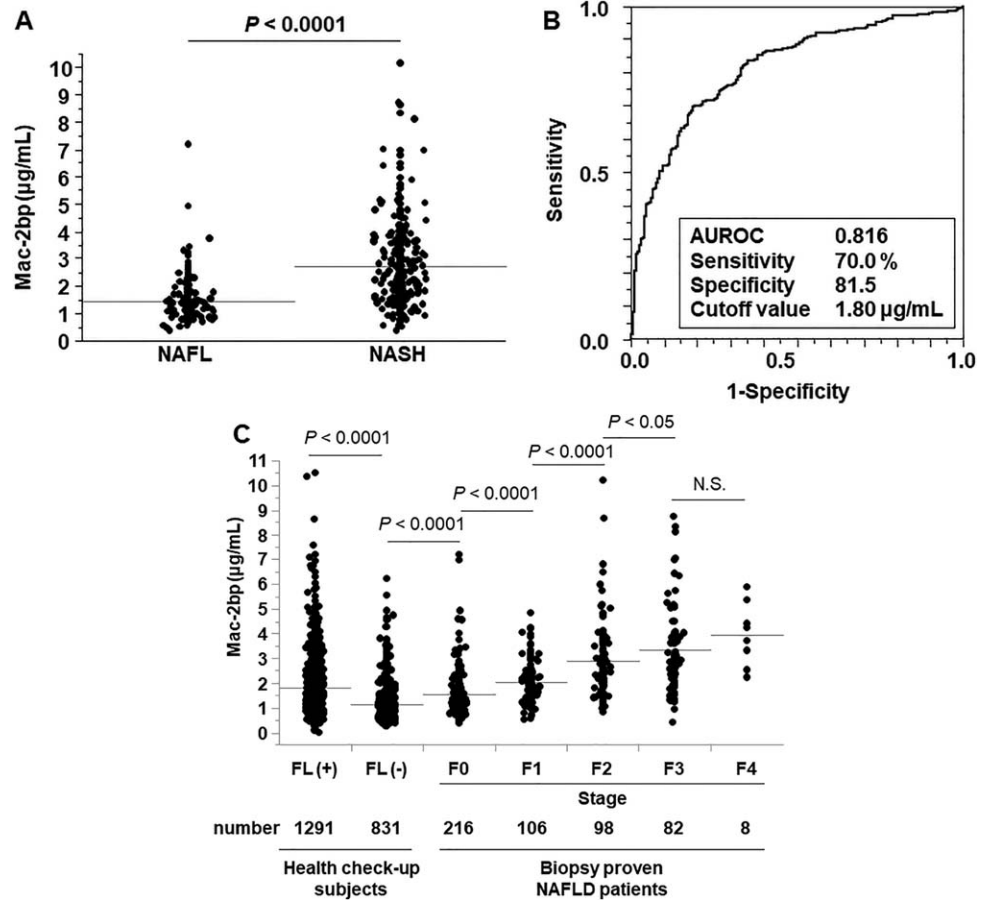
ALP, IRI, and ferritin were also higher in the patients with NASH compared with the patients with NAFL. The values of the FIB4 index and NFS also increased in patients with NASH. The values for Alb and platelet count were lower in the patients with NASH. In the check-up cohort, the serum levels of AST, ALT, γ -glutamyltransferase, ALP, choline esterase, total cholesterol, triglyceride, FBS, and Alb were higher in the FL(+) subjects than in the FL(-) subjects. The NFS was higher but the FIB4 index was lower in the FL(+) subjects than in the FL(-) subjects. Serum high-density lipoprotein cholesterol levels were lower in the FL(+) subjects. In the NAFLD cohort, serum Mac-2bp levels were significantly increased in the patients with NASH compared with the patients with NAFL (Table 1; Fig. 1A). In the check-up cohort, both serum Mac-2bp and WFA⁺-M2BP levels were higher in the FL(+) subjects.

Mac-2bp IS A USEFUL BIOMARKER FOR NASH DIAGNOSIS AND LIVER FIBROSIS PREDICTION

Next, we used ROC analysis to evaluate the utility of Mac-2bp as a NASH diagnosis biomarker (510

patients with NAFLD; Fig. 1B). When the cutoff value was 1.80 μ g/mL, the values for the AUROC and for sensitivity and specificity for Mac-2bp for NASH diagnosis were 0.816, 70.0%, and 81.5%, respectively. In contrast, the values for the AUROC and for sensitivity and specificity were 0.733, 58.6%, and 91.4% (FIB4 index) and 0.654, 50.9%, and 74.9% (NFS), respectively. Mac-2bp was superior to these scoring systems for NASH diagnosis, although the FIB4 index showed high specificity.

Our previous study revealed that Mac-2bp is a useful biomarker for the prediction of liver fibrosis severity in patients with NAFLD.⁽²⁶⁾ In the present study, we measured serum Mac-2bp levels in the NAFLD and check-up cohorts. In patients with biopsy-proven NAFLD, serum Mac-2bp levels increased stepwise as the liver fibrosis stage increased (F0/F1/F2/F3/F4, 1.55 ± 0.91, 2.05 ± 0.88, 2.89 ± 1.59, 3.34 ± 1.72, 3.96 ± 1.28 μ g/mL, respectively) (Fig. 1C). In the check-up cohort, the mean serum Mac-2bp value was higher in the FL(+) group than in the FL(-) group (1.81 ± 1.09, 1.11 ± 0.69 μ g/mL, respectively). The serum Mac-2bp levels of the FL(+) group subjects were between the levels of F0 and F1 in the patients with biopsy-proven NAFLD. Using ROC analysis, we



set Mac-2bp cutoff values to predict each fibrosis stage in patients with biopsy-proven NAFLD (Supporting Table S1). The cutoff values were 1.80 $\mu\text{g}/\text{mL}$ (\leq stage 1), 2.21 $\mu\text{g}/\text{mL}$ (\leq stage 2), and 2.24 $\mu\text{g}/\text{mL}$ (\leq stage 3). In 1,291 FL(+) subjects, the serum Mac-2bp levels were $>1.80 \mu\text{g}/\text{mL}$ in 498 subjects (38.6%) and $>2.24 \mu\text{g}/\text{mL}$ in 318 subjects (24.6%).

SERUM Mac-2bp LEVELS AND WFA⁺-M2BP LEVELS WERE WELL CORRELATED IN THE HEALTH CHECK-UP GROUP

We measured serum levels of Mac-2bp and WFA⁺-M2BP in 1,830 subjects receiving health check-ups (1,337 male subjects, 493 female subjects). The results indicated that serum Mac-2bp and WFA⁺-M2BP levels were well correlated ($R = 0.44$, $P < 0.0001$) (Fig. 2).

SERUM Mac-2bp LEVELS AND WFA⁺-M2BP LEVELS WERE EQUALLY USEFUL BIOMARKERS FOR NASH DIAGNOSIS

We measured both Mac-2bp and WFA⁺-M2BP in 116 (48 NAFL, 68 NASH) of 510 patients in the NAFLD cohort. The results for the clinical and biochemical characteristics of the 116 patients with NAFLD are presented in Table 2. Age, AST, ALT, ALP, IRI, and ferritin levels were significantly higher and FBS, Alb, and platelet count were significantly lower in the patients with NASH. The ratio of female to male patients was significantly higher in the NASH patient group. Serum Mac-2bp and WFA⁺-M2BP levels were significantly higher in the patients with NASH compared with the patients with NAFL.

We compared the significance of NASH diagnostic ability between Mac-2bp and WFA⁺-M2BP in the NASH patient group. The serum levels of Mac-2bp

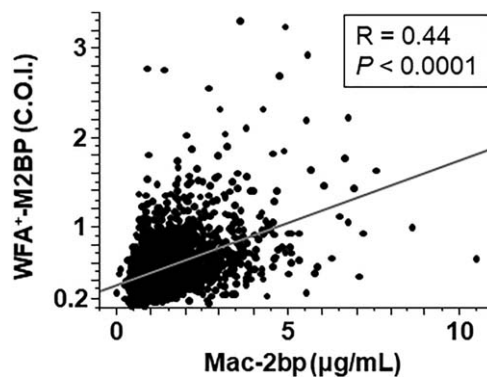


FIG. 2. Serum Mac-2bp and WFA⁺-M2BP levels were well correlated in subjects with health check-ups.

and WFA⁺-M2BP were well correlated (Fig. 3A). The ROC analysis revealed that the utility of Mac-2bp and WFA⁺-M2BP as NASH biomarkers was similar for each marker (Fig. 3B).

COMPARISON OF Mac-2bp AND WFA⁺-M2BP AS LIVER FIBROSIS PREDICTION BIOMARKERS

The serum Mac-2bp and WFA⁺-M2BP levels increased as liver fibrosis severity increased (116 patients with NAFLD; Fig. 4A,B). Serum Mac-2bp levels were significantly increased between F0 and F1 and between F1 and F2. We used ROC analysis to compare the utility of these two biomarkers for the assessment of advanced liver fibrosis (\geq F2) in patients with NAFLD (Fig. 4C). The AUROC for Mac-2bp was greater than that of WFA⁺-M2BP (0.809 versus 0.758, respectively). Logistic regression analysis revealed that Mac-2bp was an independent determinant for the prediction of advanced liver fibrosis, even after adjustment for WFA⁺-M2BP results (Table 3). The AUROC values for Mac-2bp and WFA⁺-M2BP were 0.750 and 0.731, respectively, for the prediction of early liver fibrosis (\geq F1); the values were 0.738 and 0.701, respectively, for the prediction of severe advanced fibrosis (\geq F3) (Supporting Fig. S3). In contrast, the values for the AUROC for the assessment of advanced liver fibrosis (\geq F2) in patients with NAFLD were 0.791 (FIB4 index) and 0.719 (NFS). Mac-2bp was superior to these scoring systems for the prediction of advanced liver fibrosis in NAFLD.

Mac-2bp PROTEIN IS STRONGLY EXPRESSED IN NAFLD PATIENT HEPATOCYTES

We used IHC analysis of Mac-2bp to determine the kinds of cells that were the source of Mac-2bp protein production in livers of patients with NAFLD (Fig. 5). The Mac-2bp protein was barely expressed in normal liver; strong cytoplasmic staining was apparent in the hepatocytes from patients with NAFLD. Hepatocytes of patients with NAFLD had whole cytoplasmic staining or peripheral nuclear lesion staining of Mac-2bp. Surprisingly, there were significant negative correlations between serum Mac-2bp levels and degree of hepatic Mac-2bp staining (Supporting Fig. S2).

Discussion

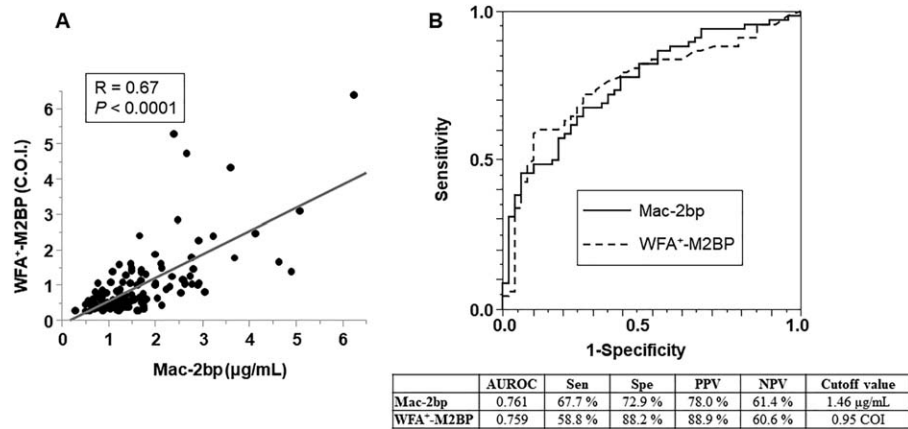
We investigated serum Mac-2bp and WFA⁺-M2BP in a large sample size of NAFLD subjects. Serum Mac-2bp levels increased in a stepwise pattern in patients with biopsy-proven NAFLD. The diagnostic ability for NASH was not different between

TABLE 2. CLINICAL AND SEROLOGICAL CHARACTERISTICS OF THE PATIENTS WITH BIOPSY-PROVEN NAFLD WHO WERE MEASURED FOR BOTH MAC-2BP AND WFA⁺-M2BP

	NAFL	NASH	P Value
Number	48	68	
Age (y)	51.8 \pm 9.8	55.6 \pm 12.4	<0.05
Sex (M/F)	35/13	28/40	<0.01
BMI (kg/m ²)	26.7 \pm 3.9	26.9 \pm 4.8	N.S.
AST (U/L)	37.9 \pm 26.8	64.3 \pm 37.0	<0.0001
ALT (U/L)	56.2 \pm 39.6	93.9 \pm 60.3	<0.0005
GGT (U/L)	104.9 \pm 133.4	100.7 \pm 121.4	N.S.
ALP (U/L)	244.4 \pm 101.8	275.3 \pm 100.2	<0.05
CHE (U/L)	349.7 \pm 54.8	352.3 \pm 71.5	N.S.
T-Chol (mg/dL)	208.8 \pm 33.2	206.4 \pm 39.9	N.S.
TG (mg/dL)	139.0 \pm 59.3	161.4 \pm 92.8	N.S.
HDL-C (mg/dL)	51.6 \pm 11.5	48.9 \pm 15.6	N.S.
FBS (mg/dL)	111.2 \pm 22.1	103.9 \pm 28.2	<0.05
IRI (mU/mL)	8.16 \pm 3.83	14.7 \pm 11.2	<0.05
Albumin (g/dL)	4.36 \pm 0.29	4.12 \pm 0.35	<0.0001
Ferritin (ng/mL)	185.8 \pm 148.2	375.2 \pm 419.8	0.07
Platelet count ($\times 10^4/\mu$ L)	22.6 \pm 5.7	19.2 \pm 5.4	<0.01
Hyaluronic acid (ng/mL)	39.6 \pm 60.6	79.1 \pm 109.5	<0.05
Mac-2bp (μ g/mL)	1.23 \pm 0.56	2.05 \pm 1.12	<0.0001
WFA ⁺ -M2BP (COI)	0.67 \pm 0.67	1.26 \pm 1.10	<0.0001

Abbreviations: CHE, choline esterase; GGT, gamma glutamyl-transpeptidase; HDL-C, high-density lipoprotein cholesterol; N.D., not determined; N.S., not significant; T-Chol, total cholesterol; TG, triglyceride.

FIG. 3. Serum Mac-2bp levels and WFA⁺-M2BP levels were equally useful biomarkers for NASH diagnosis. (A) Correlation between serum Mac-2bp and WFA⁺-M2BP in patients with biopsy-proven NAFLD. (B) ROC curves of Mac-2bp and WFA⁺-M2BP for the discrimination of NASH in patients with biopsy-proven NAFLD. The table presents the results for the efficacy of each biomarker for NASH diagnosis. Abbreviations: NPV, negative predictive value; PPV, positive predictive value; Sen, sensitivity; Spe, specificity.



Mac-2bp and WFA⁺-M2BP; however, the utility of Mac-2bp for the prediction of advanced liver fibrosis was slightly superior to that of WFA⁺-M2BP. IHC staining of Mac-2bp revealed that hepatocytes strongly expressed Mac-2bp in the livers of some patients with NAFLD; Mac-2bp was barely expressed in the non-NAFLD patients' livers. Our

results indicated that hepatocytes were a main source of circulating Mac-2bp in the patients with NAFLD; however, our study demonstrated that serum Mac-2bp levels negatively correlated with the degree of Mac-2bp staining in liver. Not only production but also secretion of Mac-2bp should be involved in these phenomena.

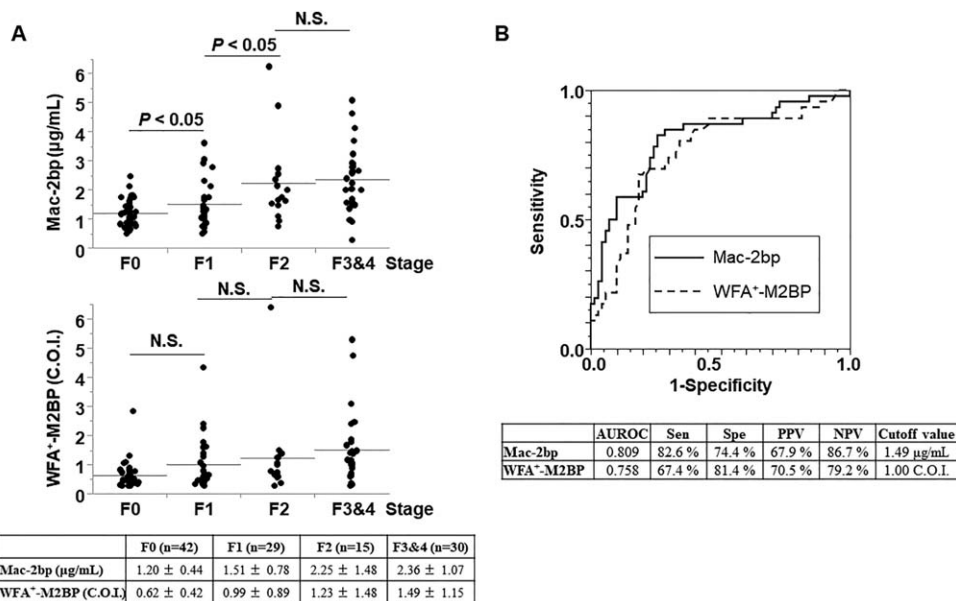


FIG. 4. Comparison of Mac-2bp and WFA⁺-M2BP as liver fibrosis prediction biomarkers. (A) Correlation between serum Mac-2bp levels and liver fibrosis stage (upper panel). Correlation between serum WFA⁺-M2BP levels and liver fibrosis stage (middle panel). The table presents the results for the mean values of Mac-2bp and WFA⁺-M2BP at each liver fibrosis stage. (B) ROC curves of Mac-2bp and WFA⁺-M2BP for the prediction of advanced liver fibrosis (≥F2) in patients with biopsy-proven NAFLD. The table presents the results for the efficacy of each biomarker. NPV, negative predictive value; N.S., not significant; PPV, positive predictive value; Sen, sensitivity; Spe, specificity.

TABLE 3. LOGISTIC REGRESSION ANALYSIS FOR THE PREDICTION OF EARLY LIVER FIBROSIS (\geq F1)

Parameters	Odds Ratio	95% CI	P Value
Mac-2bp (μ g/mL)	4.43	2.12-10.40	<0.0001
WFA ⁺ -M2BP (COI)	1.31	0.68-3.05	N.S.

Abbreviations: CI, confidence interval; N.S., not significant.

Kuno et al.⁽³²⁾ studied a group of 125 CHC patients and found that serum Mac-2bp levels are greater and that WFA has the best correlation with liver fibrosis progression in these patients. They developed an automatic measurement system that can detect WFA⁺-M2BP within 20 minutes. WFA⁺-M2BP has a characteristic glycan structure that is recognized by WFA. WFA recognizes terminal *N*-acetylgalactosaminides and specifically binds with the disaccharide LacdiNAc (β -D-GalNAc-[1 \rightarrow 4]-D-GlcNAc; GalNAc *N*-acetylgalactosamine, GlcNAc *N*-acetylglucosamine).⁽⁴⁰⁾ WFA⁺-M2BP is very useful as a liver fibrosis biomarker for CHC, but the liver fibrosis prediction abilities for the other chronic liver diseases are relatively lower than that for CHC.^(32,33,41-43) Our study revealed that compared with WFA⁺-M2BP, Mac-2bp had greater ability to predict NAFLD fibrosis stage. Mac-2bp is a glycoprotein with seven *N*-glycosylation sites⁽²⁹⁾; the Mac-2bp measured in this study included whole types of Mac-2bp, including WFA⁺-M2BP.

Investigation of the NAFLD-specific glycan modification of Mac-2bp is necessary to find more useful NAFLD glycol-biomarkers.

Bekki et al.⁽⁴⁴⁾ found that hepatic stellate cells (HSCs) produce WFA⁺-M2BP. WFA⁺-M2BP produced from HSCs stimulate Mac-2 (galectin 3) production from Kupffer cells and accelerate further HSC activation, which results in liver fibrosis progression. In our study, the hepatocytes in the patients with NAFLD mainly produced Mac-2bp. Secreted glycoproteins have different types of glycosylation patterns due to the different origins of cell types and stimuli.^(45,46) The differences in the Mac-2bp-producing cells or in the type of stimulation or both should account for the variant glycosylation patterns of Mac-2bp. With comprehensive analysis of glycosylation patterns on Mac-2bp, we would like to establish, in a future study, novel NAFLD biomarkers that can predict the severity of the NAFLD clinical course more precisely.

We found that Mac-2bp was superior to WFA⁺-M2BP for the prediction of NAFLD fibrosis stage, especially for early stage fibrosis. More than 90% of virus-associated hepatitis patients with liver cirrhosis will develop HCC.⁽⁷⁾ In contrast, more than 50% of male patients with NAFLD with NAFLD-associated HCC do not have liver cirrhosis.⁽⁷⁻⁹⁾ Therefore, it is

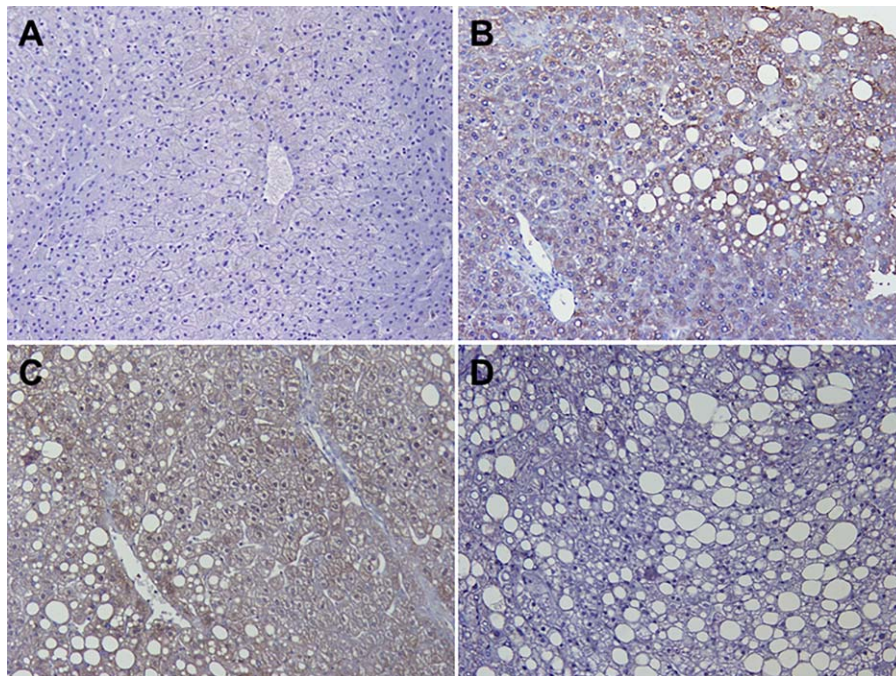


FIG. 5. Mac-2bp protein was strongly expressed in hepatocytes from patients with NAFLD. Photomicrographs of representative samples from human livers stained with anti-human Mac-2bp antibody (original magnification \times 200). (A) Normal liver tissues obtained from livers of patients who underwent surgery for hepatic colorectal carcinoma metastasis. (B-D) Livers from patients with NAFLD. Serum Mac-2bp levels were (B) 0.932 μ g/mL, (C) 1.26 μ g/mL, and (D) 3.65 μ g/mL.

important to diagnosis NAFLD at an early stage. These findings indicated that compared with WFA⁺-M2BP, Mac-2bp would be superior as a NAFLD biomarker for prediction of the extent of early liver fibrosis. In the subjects who received health check-ups, the serum Mac-2bp levels of the FL(+) subjects were between the levels of F0 and F1 found in the patients with biopsy-proven NAFLD (Fig. 1C). We set the fibrosis-predicting cutoff value as 1.80 (F1), 2.21 (F2), and 2.24 $\mu\text{g/mL}$ (F3) for the group of 510 patients with biopsy-proven NAFLD. Serum Mac-2bp levels were $>1.80 \mu\text{g/mL}$ in 38.6% and $>2.24 \mu\text{g/mL}$ in 24.6% of the FL(+) subjects. These findings indicated that approximately 25% of the NAFLD subjects would have severe advanced fibrosis (\geq F3).

Recent studies demonstrated that there is a significant association between the patatin-like phospholipase domain containing 3 (PNPLA3) genotype and fibrosis progression in patients with NAFLD.^(47,48) Although we did not analyze the PNPLA3 genotype in our study subjects, the information about the PNPLA3 genotype should be useful for the clinical follow-up of patients with NAFLD.

Our previous study revealed that serum Mac-2bp levels are higher in patients with NAFL compared with healthy volunteers.⁽²⁶⁾ Serum Mac-2bp was increased in a mouse simple fatty liver model and was further elevated in mouse NASH models by use of our enzyme-linked immunosorbent assay system.⁽⁴⁹⁾ However, in human patients with NAFLD, serum Mac-2bp levels do not change with the degree of hepatic steatosis but do increase with hepatic inflammation and liver fibrosis progression.^(26,27) In the present study, serum Mac-2bp levels did not increase with steatosis degree in patients with NAFLD (Supporting Fig. S4). Taken together, these results indicate that serum Mac-2bp increases in patients with fatty liver disease regardless of steatosis degree; Mac-2bp is further elevated by inflammation and fibrosis. We found that there was significant negative correlation between serum Mac-2bp levels and hepatic Mac-2bp staining degree in patients with NAFLD (Supporting Figs. S1 and S2). We hypothesized that the mechanism of secretion of Mac-2bp from the hepatocytes would be different in each NAFLD subject and that the balance between secretion and production of Mac-2bp would contribute to the different accumulation patterns that occur in NAFLD hepatocytes. There are many etiologies for NAFLD (e.g., free fatty acids, inflammatory cytokines, and lipopolysaccharide).⁽⁵⁰⁻⁵³⁾ These factors would have significant effects on the secretion of Mac-2bp from hepatocytes in

patients with NAFLD. The molecular mechanism underlying the induction of Mac-2bp during fatty liver changes should be investigated.

In conclusion, both Mac-2bp and WFA⁺-M2BP were useful biomarkers for NASH diagnosis. Mac-2bp would be superior to WFA⁺-M2BP for the prediction of liver fibrosis stage in patients with NAFLD. Our study revealed that hepatocytes are the main producer of Mac-2bp in patients with NAFLD. The balance between production and secretion of Mac-2bp contributes to the determination of Mac-2bp serum levels. The mechanisms that contribute to serum Mac-2bp levels in patients with NAFLD should be investigated.

Acknowledgment: We thank Atsuko Sawanobori, Tomomi Maeda, and Haruka Maeda for their excellent technical assistance.

REFERENCES

- 1) Ford ES, Giles WH, Dietz WH. Prevalence of the metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. *JAMA* 2002;287:356-359.
- 2) Shirabe K, Takeishi K, Taketomi A, Uchiyama H, Kayashima H, Maehara Y. Improvement of long-term outcomes in hepatitis C virus antibody-positive patients with hepatocellular carcinoma after hepatectomy in the modern era. *World J Surg* 2011;35:1072-1084.
- 3) Mazzaferro V, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, et al. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996;334:693-699.
- 4) Liang TJ, Heller T. Pathogenesis of hepatitis C-associated hepatocellular carcinoma. *Gastroenterology* 2004;127:S62-S71.
- 5) Imai Y, Kawata S, Tamura S, Yabuuchi I, Noda S, Inada M, et al. Relation of interferon therapy and hepatocellular carcinoma in patients with chronic hepatitis C. Osaka Hepatocellular Carcinoma Prevention Study Group. *Ann Intern Med* 1998;129:94-99.
- 6) Yoshida H, Shiratori Y, Moriyama M, Arakawa Y, Ide T, Sata M, et al. Interferon therapy reduces the risk for hepatocellular carcinoma: national surveillance program of cirrhotic and noncirrhotic patients with chronic hepatitis C in Japan. IHIT Study Group. Inhibition of hepatocarcinogenesis by interferon therapy. *Ann Intern Med* 1999;131:174-181.
- 7) Ertle J, Dechene A, Sowa JP, Penndorf V, Herzer K, Kaiser G, et al. Non-alcoholic fatty liver disease progresses to hepatocellular carcinoma in the absence of apparent cirrhosis. *Int J Cancer* 2011;128:2436-2443.
- 8) Yasui K, Hashimoto E, Komorizono Y, Koike K, Arai S, Imai Y, et al.; Japan NASH Study Group, Ministry of Health, Labour, and Welfare of Japan. Characteristics of patients with nonalcoholic steatohepatitis who develop hepatocellular carcinoma. *Clin Gastroenterol Hepatol* 2011;9:428-433.

- 9) Tokushige K, Hyogo H, Nakajima T, Ono M, Kawaguchi T, Honda K, et al. Hepatocellular carcinoma in Japanese patients with nonalcoholic fatty liver disease and alcoholic liver disease: multicenter survey. *J Gastroenterol* 2016;51:586-596.
- 10) Piscaglia F, Svegliati-Baroni G, Barchetti A, Pecorelli A, Marinelli S, Tiribelli C, et al.; HCC-NAFLD Italian Study Group. Clinical patterns of hepatocellular carcinoma in nonalcoholic fatty liver disease: a multicenter prospective study. *Hepatology* 2016;63:827-838.
- 11) Bravo AA, Sheth SG, Chopra S. Liver biopsy. *N Engl J Med* 2001;344:495-500.
- 12) Gebo KA, Herlong HF, Torbenson MS, Jenckes MW, Chander G, Ghanem KG, et al. Role of liver biopsy in management of chronic hepatitis C: a systematic review. *Hepatology* 2002;36(5 Suppl. 1):S161-S172.
- 13) Piccinino F, Sagnelli E, Pasquale G, Giusti G. Complications following percutaneous liver biopsy. A multicentre retrospective study on 68,276 biopsies. *J Hepatol* 1986;2:165-173.
- 14) Ratziu V, Charlotte F, Heurtier A, Gombert S, Giral P, Bruckert E, et al.; LIDO Study Group. Sampling variability of liver biopsy in nonalcoholic fatty liver disease. *Gastroenterology* 2005;128:1898-1906.
- 15) Angulo P, Kleiner DE, Dam-Larsen S, Adams LA, Bjornsson ES, Charatcharoenwitthaya P, et al. Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* 2015;149:389-397.e310.
- 16) Yoneda M, Yoneda M, Fujita K, Inamori M, Tamano M, Hiriishi H, et al. Transient elastography in patients with nonalcoholic fatty liver disease (NAFLD). *Gut* 2007;56:1330-1331.
- 17) Yoneda M, Suzuki K, Kato S, Fujita K, Nozaki Y, Hosono K, et al. Nonalcoholic fatty liver disease: US-based acoustic radiation force impulse elastography. *Radiology* 2010;256:640-647.
- 18) Castera L, Forns X, Alberti A. Non-invasive evaluation of liver fibrosis using transient elastography. *J Hepatol* 2008;48:835-847.
- 19) Harrison SA, Oliver D, Arnold HL, Gogia S, Neuschwander-Tetri BA. Development and validation of a simple NAFLD clinical scoring system for identifying patients without advanced disease. *Gut* 2008;57:1441-1447.
- 20) Angulo P, Hui JM, Marchesini G, Bugianesi E, George J, Farrell GC, et al. The NAFLD fibrosis score: a noninvasive system that identifies liver fibrosis in patients with NAFLD. *Hepatology* 2007;45:846-854.
- 21) Sumida Y, Yoneda M, Hyogo H, Yamaguchi K, Ono M, Fujii H, et al.; Japan Study Group of Nonalcoholic Fatty Liver Disease (JSG-NAFLD). A simple clinical scoring system using ferritin, fasting insulin, and type IV collagen 7S for predicting steatohepatitis in nonalcoholic fatty liver disease. *J Gastroenterol* 2011;46:257-268.
- 22) Dixon JB, Bhathal PS, O'Brien PE. Nonalcoholic fatty liver disease: predictors of nonalcoholic steatohepatitis and liver fibrosis in the severely obese. *Gastroenterology* 2001;121:91-100.
- 23) Ratziu V, Giral P, Charlotte F, Bruckert E, Thibault V, Theodorou I, et al. Liver fibrosis in overweight patients. *Gastroenterology* 2000;118:1117-1123.
- 24) Sterling RK, Lissen E, Clumeck N, Sola R, Correa MC, Montaner J, et al.; APRICOT Clinical Investigators. Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. *Hepatology* 2006;43:1317-1325.
- 25) Vallet-Pichard A, Mallet V, Nalpas B, Verkarre V, Nalpas A, Dhalluin-Venier V, et al. FIB-4: an inexpensive and accurate marker of fibrosis in HCV infection. Comparison with liver biopsy and fibrotest. *Hepatology* 2007;46:32-36.
- 26) Kamada Y, Fujii H, Fujii H, Sawai Y, Doi Y, Uozumi N, et al. Serum Mac-2 binding protein levels as a novel diagnostic biomarker for prediction of disease severity and nonalcoholic steatohepatitis. *Proteomics Clin Appl* 2013;7:648-656.
- 27) Kamada Y, Ono M, Hyogo H, Fujii H, Sumida Y, Mori K, et al. A novel noninvasive diagnostic method for nonalcoholic steatohepatitis using two glycobiomarkers. *Hepatology* 2015;62:1433-1443.
- 28) Grassadonia A, Tinari N, Iurisci I, Piccolo E, Cumashi A, Innominato P, et al. 90K (Mac-2 BP) and galectins in tumor progression and metastasis. *Glycoconj J* 2002;19:551-556.
- 29) Przybylo M, Martuszczyńska D, Pocheć E, Hoja-Lukowicz D, Lityńska A. Identification of proteins bearing beta1-6 branched N-glycans in human melanoma cell lines from different progression stages by tandem mass spectrometry analysis. *Biochim Biophys Acta* 2007;1770:1427-1435.
- 30) Artini M, Natoli C, Tinari N, Costanzo A, Marinelli R, Balsano C, et al. Elevated serum levels of 90K/MAC-2 BP predict unresponsiveness to alpha-interferon therapy in chronic HCV hepatitis patients. *J Hepatol* 1996;25:212-217.
- 31) Cheung KJ, Tilleman K, Deforce D, Colle I, Van Vlierberghe H. The HCV serum proteome: a search for fibrosis protein markers. *J Viral Hepat* 2009;16:418-429.
- 32) Kuno A, Ikehara Y, Tanaka Y, Ito K, Matsuda A, Sekiya S, et al. A serum "sweet-doughnut" protein facilitates fibrosis evaluation and therapy assessment in patients with viral hepatitis. *Sci Rep* 2013;3:1065.
- 33) Abe M, Miyake T, Kuno A, Imai Y, Sawai Y, Hino K, et al. Association between Wisteria floribunda agglutinin-positive Mac-2 binding protein and the fibrosis stage of non-alcoholic fatty liver disease. *J Gastroenterol* 2015;50:776-784.
- 34) Matteoni CA, Younossi ZM, Gramlich T, Boparai N, Liu YC, McCullough AJ. Nonalcoholic fatty liver disease: a spectrum of clinical and pathological severity. *Gastroenterology* 1999;116:1413-1419.
- 35) Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, et al.; Nonalcoholic Steatohepatitis Clinical Research Network. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005;41:1313-1321.
- 36) Saadeh S, Younossi ZM, Remer EM, Gramlich T, Ong JP, Hurley M, et al. The utility of radiological imaging in nonalcoholic fatty liver disease. *Gastroenterology* 2002;123:745-750.
- 37) Hamano M, Kamada Y, Kiso S, Furuta K, Kizu T, Chatani N, et al. Adiponectin negatively correlates with alcoholic and nonalcoholic liver dysfunction: health check-up study of Japanese men. *Hepatol Res* 2013;43:238-248.
- 38) Shah AG, Lydecker A, Murray K, Tetri BN, Contos MJ, Sanyal AJ; Nash Clinical Research Network. Comparison of noninvasive markers of fibrosis in patients with nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 2009;7:1104-1112.
- 39) Kuno A, Sato T, Shimazaki H, Unno S, Saitou K, Kiyohara K, et al. Reconstruction of a robust glycodiagnostic agent supported by multiple lectin-assisted glycan profiling. *Proteomics Clin Appl* 2013;7:642-647.
- 40) Haji-Ghassemi O, Gilbert M, Spence J, Schur MJ, Parker MJ, Jenkins ML, et al. Molecular basis for recognition of the cancer glycobiomarker, LacdiNAc (GalNAc[beta1-->4]GlcNAc), by Wisteria floribunda agglutinin. *J Biol Chem* 2016;291:24085-24095.
- 41) Zou X, Zhu MY, Yu DM, Li W, Zhang DH, Lu FJ, et al. Serum WFA+ -M2BP levels for evaluation of early stages of liver fibrosis in patients with chronic hepatitis B virus infection. *Liver Int* 2017;37:35-44.
- 42) Nishikawa H, Enomoto H, Iwata Y, Hasegawa K, Nakano C, Takata R, et al. Impact of serum Wisteria floribunda agglutinin

- positive Mac-2-binding protein and serum interferon-gamma-inducible protein-10 in primary biliary cirrhosis. *Hepatol Res* 2016;46:575-583.
- 43) Nishikawa H, Enomoto H, Iwata Y, Hasegawa K, Nakano C, Takata R, et al. Clinical significance of serum *Wisteria floribunda* agglutinin positive Mac-2-binding protein level and high-sensitivity C-reactive protein concentration in autoimmune hepatitis. *Hepatol Res* 2016;46:613-621.
- 44) Bekki Y, Yoshizumi T, Shimoda S, Itoh S, Harimoto N, Ikegami T, et al. Hepatic stellate cells secrete WFA+ -M2BP: its role in biological interactions with Kupffer cells. *J Gastroenterol Hepatol* 2017;32:1387-1393.
- 45) Hart GW, Copeland RJ. Glycomics hits the big time. *Cell* 2010;143:672-676.
- 46) Connelly MA, Gruppen EG, Otvos JD, Dullaart RP. Inflammatory glycoproteins in cardiometabolic disorders, autoimmune diseases and cancer. *Clin Chim Acta* 2016;459:177-186.
- 47) Seko Y, Sumida Y, Tanaka S, Mori K, Taketani H, Ishiba H, et al. Development of hepatocellular carcinoma in Japanese patients with biopsy-proven non-alcoholic fatty liver disease: association between PNPLA3 genotype and hepatocarcinogenesis/fibrosis progression. *Hepatol Res* 2016, doi: 10.1111/hepr.12840. [Epub ahead of print]
- 48) Bugianesi E, Bizzarri C, Rosso C, Mosca A, Panera N, Veraldi S, et al. Low birthweight increases the likelihood of severe steatosis in pediatric non-alcoholic fatty liver disease. *Am J Gastroenterol* 2017, doi: 10.1038/ajg.2017.140. [Epub ahead of print]
- 49) Iwata A, Kamada Y, Ebisutani Y, Yamamoto A, Ueda Y, Arai H, et al. Establishment of mouse Mac-2 binding protein enzyme-linked immunosorbent assay and its application for mouse chronic liver disease models. *Hepatol Res* 2016, doi: 10.1111/hepr.12819. [Epub ahead of print]
- 50) Miura K, Yang L, van Rooijen N, Brenner DA, Ohnishi H, Seki E. Toll-like receptor 2 and palmitic acid cooperatively contribute to the development of nonalcoholic steatohepatitis through inflammasome activation in mice. *Hepatology* 2013;57:577-589.
- 51) Puri P, Wiest MM, Cheung O, Mirshahi F, Sargeant C, Min HK, et al. The plasma lipidomic signature of nonalcoholic steatohepatitis. *Hepatology* 2009;50:1827-1838.
- 52) Tilg H, Diehl AM. Cytokines in alcoholic and nonalcoholic steatohepatitis. *N Engl J Med* 2000;343:1467-1476.
- 53) Imajo K, Fujita K, Yoneda M, Nozaki Y, Ogawa Y, Shinohara Y, et al. Hyperresponsivity to low-dose endotoxin during progression to nonalcoholic steatohepatitis is regulated by leptin-mediated signaling. *Cell Metab* 2012;16:44-54.

Supporting Information

Additional Supporting Information may be found at onlinelibrary.wiley.com/doi/10.1002/hep4.1080/full.