

RESEARCH ARTICLE

# In-depth phenotypic characterization of multicellular tumor spheroids: Effects of 5-Fluorouracil

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## Abstract

MultiCellular Tumor Spheroids (MCTS), which mimic the 3-Dimensional (3D) organization of a tumor, are considered as better models than conventional cultures in 2-Dimensions (2D) to study cancer cell biology and to evaluate the response to chemotherapeutic drugs. A real time and quantitative follow-up of MCTS with simple and robust readouts to evaluate drug efficacy is still missing. Here, we evaluate the chemotherapeutic drug 5-Fluorouracil (5-FU) response on the growth and integrity of MCTS two days after treatment of MCTS and for three colorectal carcinoma cell lines with different cohesive properties (HT29, HCT116 and SW480). We found different sensitivity to 5-FU for the three CRC cell lines, ranging from high (SW480), intermediate (HCT116) and low (HT29) and the same hierarchy of CRC cell lines sensitivity is conserved in 2D. We also evidence that 5-FU has a strong impact on spheroid cohesion, with the apparition of a number of single detaching cells from the spheroid in a 5-FU dose- and cell line-dependent manner. We propose an innovative methodology for the chemosensitivity evaluation in 3D MCTS that recapitulates and regionalizes the 5-FU-induced changes within MCTS over time. These robust phenotypic read-outs could be easily scalable for high-throughput drug screening that may include different types of cancer cells to take into account tumor heterogeneity and resistance to treatment.

## Introduction

Significant improvements have been made in cancer therapy but there is still a need for real time quantification of the progression of various biological processes (differentiation, proliferation, invasion, death. . .) on fresh living samples and for innovative drug screening methodologies. Functional analysis of cancer cells survival in response to chemotherapeutic agents could be used to adjust the treatment strategy and to predict the therapeutic outcome. Traditional two-dimensional (2D) cell-based assays are commonly employed to evaluate drug sensitivity patterns [1]. However, results from such 2D platforms are often very different from the

*in vivo* as cell interactions are restrained to neighbouring flat cells and underlying extracellular matrix [2,3]. Three dimensional (3D) cells aggregates, called Multicellular Tumor Spheroids (MCTS), recapitulate with better fidelity the organization of cells found *in vivo* and represent a recognized non-vascularized tumor model [4]. It is now well acknowledged that MCTS are apt *in vitro* models for drug screening in the field of oncology and especially for the translation of anti-cancer therapeutics to the clinic, as it mimics not only 3D cell-cell and cell-matrix interactions, but also the biochemical environment of the *in vivo* tumour mass [4]. However, even though biologists have been using MCTS since more than 40 years in laboratories [5–7], MCTS are just beginning to be routinely employed for drug screening [8]. Recent studies showing that chemotherapeutic molecules identified in 3D models are distinct from those found in 2D [9] have renewed the interest of MCTS in drug screening platforms to better predict *in vivo* efficacy of drug candidates [10]. The slow emergence of MCTS model, despite its non-ambiguous relevance, arises from the increased costs and complex preparation compared to its 2D counterparts, and more importantly, from the lack of standard protocol for the quantification of drug potency.

Many complex methodologies have been developed to assess treatment efficacy in 3D by following the number of proliferating cells using the analysis of frozen or paraffin-embedded slices [11] or tricky selective dissociation protocols [12] and FACS sorting [13,14]. The dynamic and non-destructive follow-up of MCTS over time include daily analysis of spheroid volume growth extracted from standard phase contrast microscopy [15] or more global analysis of metabolic activity within spheroids such as the Acid Phosphatase Assay (APH assay [8]) and Tetrazolium-based assay (MTT assay, [16]). Many studies are also focusing on genes and proteins expression modifications in MCTS and its implication on cell resistance to drug treatment [17,18] with few correlating these modifications to phenotyping changes within spheroids [16,19]. Overall, there is still a crucial need for innovative methodologies that would allow the quantification of drug efficacy on 3D models with simple and easy readouts to accelerate the translation of novel therapies from the lab to the clinic [20].

The aim of this study is to systematically evaluate the dose response to 5-Fluorouracil (5-FU) during the first 48h of treatment in 3D colorectal carcinoma (CRC) models and to establish robust phenotypic read-outs for MCTS response to treatment using microscopies widely available in research labs. 5-FU is one of the most commonly used drug in chemotherapy against CRC. It is an anti-metabolite inducing DNA damage, and is widely given in first-line of treatment in many types of solid cancers [21]. As it is well known that cells at different tumour stages are exhibiting different sensitivity to drugs, we have chosen to compare three classical CRC cell lines: HT29, HCT116 and SW480.

We evidence that 5-FU has a strong impact on spheroid cohesion, with the apparition of a number of single cells detaching from the spheroid in a 5-FU dose- and cell line-dependent manner. We describe a methodology based on a new 3D-IC<sub>50</sub> calculated from spheroid residual and cohesive core size after transfer in fresh medium to new wells. We show that the disintegration degree of peripheral cells and the nature of detached cells after drug is a valuable independent measure of drug efficacy. Using this simple methodology, we can quantify indirectly the MCTS cohesion that is an indicator of cell sensitivity to 5-FU. We propose it could be easily scalable for high-throughput drug screening that may include different types of cancer cells to take into account tumor heterogeneity and resistance to treatments.

## Materials and methods

### Colorectal cancer lines

HT29 colorectal adenocarcinoma (HTB-38) cell line, HCT116 colorectal carcinoma (CCL-247) cell line, and SW480 (CCL-228) adenocarcinoma cell line were purchased from the

American Type Culture Collection (ATCC, Virginia, USA). All cell lines were cultured in Dulbecco's Modified Eagle's medium (DMEM-Glutamax), supplemented with 10% of heat-inactivated fetal bovine serum (FBS; Sigma, St. Louis, Missouri, US), 100 units/100  $\mu$ g of penicillin/streptomycin.

### Cell culture in two dimensions (2D)

The three cell lines are grown in petri dishes and placed in an incubator at 37°C and 5% CO<sub>2</sub> in an atmosphere saturated with H<sub>2</sub>O. The culture medium is changed regularly, and the cell passage is carried out at 70% confluency. Trypsin (0,25%, Gibco) is used to detach the cells from the substrate.

### 3D cell model: MCTS formation

MCTS are formed in Ultra Low Attachment (ULA) 96 wells Round-Bottom plate treated to be non-adhesive (Greiner bio-one), thereby avoiding cell-substrate attachment. The cells are trypsinized, a coloration of dead cells with trypan blue (0,4%, Sigma) is assessed and cells are counted using a Malassez grid in order to obtain 2400 cells per milliliter. This concentration of cells (*i.e.*, 480 cells per well in a volume of 200  $\mu$ L) is chosen in order to obtain a single spheroid per well, with a spheroid diameter at the end of the experimentation not exceeding 500  $\mu$ m. The plate is centrifuged for 5 minutes at 1200 g at room temperature to initiate the formation of spheroids. The plate is placed in the incubator under agitation at 37°C and 5% CO<sub>2</sub> during all the experiment. At the end of the first day after seeding, 100  $\mu$ L of culture is added to ensure proper 3D growth.

### Spreading assay

Multi-wells plates ( $\mu$ -Slide 8 Wells ibiTreat, ibidi, Germany) are covered with 100  $\mu$ L of a 0,5 mg/ml collagen type I (rat tail, Gibco) solution at 4°C and then kept at 37°C for 1 hour. MCTS are transferred on these plates, placed in a stage incubator at 37°C, 5% CO<sub>2</sub>, 100% humidity (INUBG2E-GSI-BP-H2-SL, Tokai Hit) and recorded during 50 hours with a confocal microscope (Leica SP5) using XYZt scans.

### 5-FU treatment

5-Fluorouracil (5-FU) was kindly provided by Centre Léon Bérard (Lyon, France) purchased at Sanofi-Aventis, stored at room temperature at 384 mM in sterile water and diluted in cell culture medium. 5-FU acts during the S phase of the cell cycle inhibiting DNA synthesis by restricting availability of thymidylate and can also inhibit RNA synthesis [14]. MCTS are formed following the usual protocol of 3D cell culture described above. After two days, they are treated with 5-FU at various concentrations: 1, 2, 5, 10, 25, 50, 100 and 300  $\mu$ M. For control spheroids, only the culture medium is renewed. MCTS are followed during 48h post 5-FU. Twelve samples (N = 12) are made for each concentration after checking the homogeneity of spheroid size in each wells.

### Spheroids transfer protocol

A ring of detaching cells appears spontaneously after one-day of 5-FU treatment. By transferring spheroids into new well plates, we eliminate mechanically this uncohesive peripheral cell layer. We thus obtain an unbiased area of the spheroid cohesive core. For each transfer, the drug and culture medium are renewed.

## Live and dead assay

Spheroids are labeled directly in 96-well plates after removing the maximum of culture medium using calcein (ThermoFisher, 1  $\mu\text{M}$ ) and Propidium Iodide (PI; Sigma, 2mg/ml) diluted in sterile PBS 1X (Gibco). Calcein labels viable cells cytoplasm in green, PI labels nuclei of dead cells in red showing red dots intercalating DNA (PI+ cells). In order to avoid cell dispersion especially in the loose peripheral layer all around spheroids for the labeling, we add 50  $\mu\text{L}$  of a 2% low temperature gelling agarose solution prior to the live and dead labeling. The double staining was performed during 20 minutes at 37°C before epifluorescence microscope analysis.

## Phase-contrast follow-up of MCTS core diameter

3D MCTS photographs are taken with an inverted microscope (Leica DMIRB) in phase contrast inside the 96-well plates at 24h, 32h and 48h after 5-FU exposure. MCTS are transferred to new 96-well plates at 24h and 48h and photographs are taken before and after transfers. A segmentation based on a sobel threshold is made for each spheroid to isolate properly the core using ImageJ free software. From these binary images, the spheroids are fitted with an ellipse and major ( $L_M$ ) and minor ( $L_m$ ) axes are calculated using ImageJ "Analyze Particles" plugins. From this, a mean diameter is calculated ( $D = L_m + L_M / 2$ ). The volume  $V$  is then extrapolated making the assumption that the spheroids are spherical ( $V = \frac{4}{3} \pi R^3$ ).

## 2D-cellular impedance assay by real-time cell analysis (RTCA)

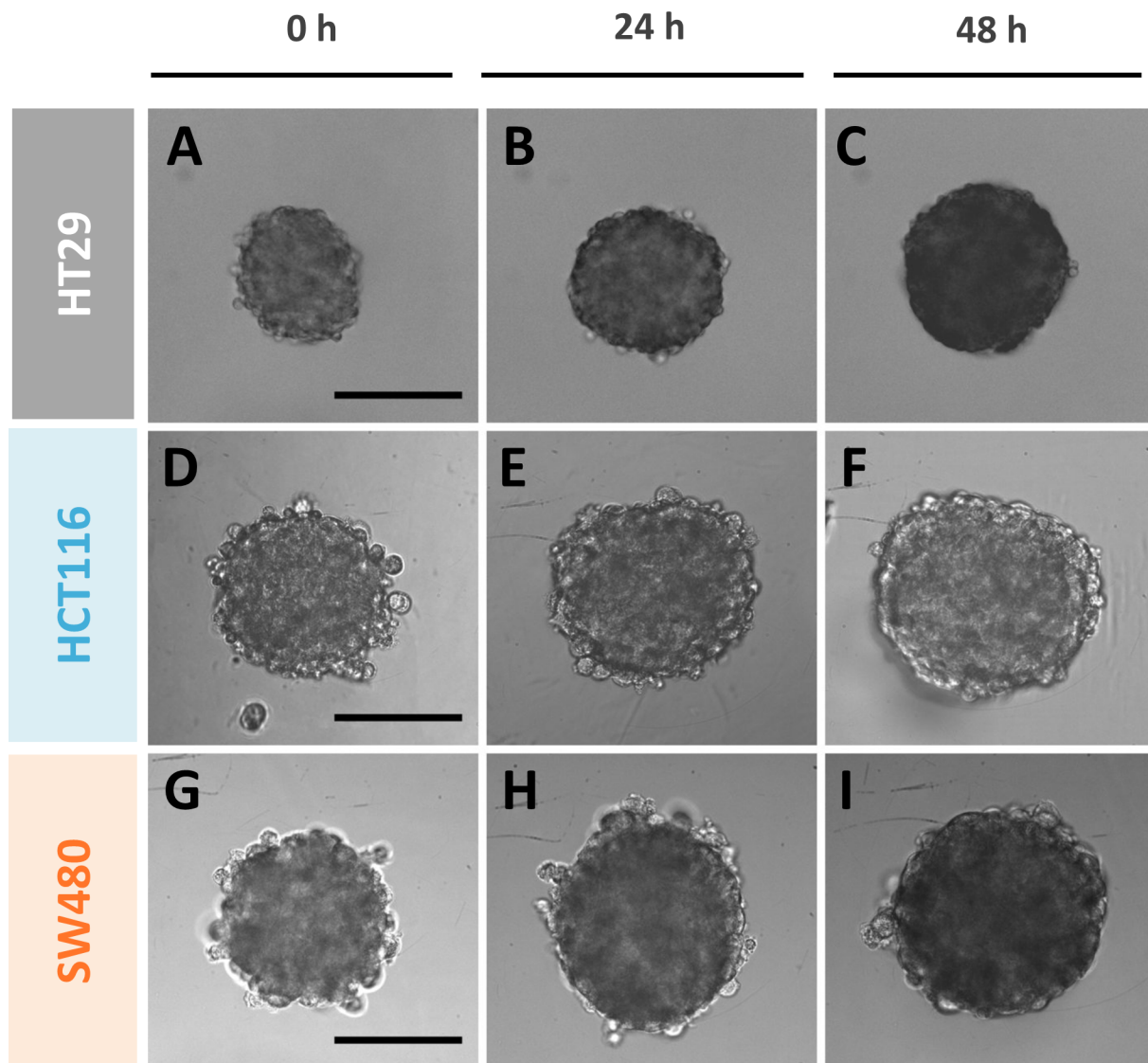
The RTCA impedance assay (xCELLigence RTCA, ACEA Biosciences) is a noninvasive real-time monitoring of cell proliferation, cell size and morphology, continuously providing data over days. Three separate 16-well plates with electrodes embedded are controlled and monitored in parallel. The instrument is placed in a standard CO<sub>2</sub> cell culture incubator and is powered and controlled via a cable connected to the control unit housed outside the incubator. The functional unit of a cellular impedance assay is a set of gold microelectrodes fused to the bottom surface of a microtiter plate well (E-Plates®). The impedance magnitude is dependent on the number of cells, the size and shape of the cells, and the cell-substrate attachment quality and is reported as a Cell Index parameter. Cells are seeded at a concentration of 5000 cells per well in culture medium. 5-FU is added 48 hours after cells seeding, by removal of culture medium and replacing by 5-FU diluted in culture medium at concentrations of 1, 2, 5, 10, 25, 50 and 100  $\mu\text{M}$  (200  $\mu\text{L}$  per well). Cell proliferation was monitored during 50 hours.

## 2D videomicroscopy analyses

2D cell proliferation is followed with an inverted microscope (Leica DMIRB) using time lapse phase contrast imaging in controlled CO<sub>2</sub>, temperature and humidity parameters during 120 hours in 24-wells plate (Corning). The medium volume per well is 1 mL. 5-FU is added 48h post incubation at concentrations of 1, 2, 5, 10, 25, 50 and 100  $\mu\text{M}$ . Control spheroids are supplemented with new culture medium at the same time and the kinetics of 2D cell proliferation is assessed with a homemade Matlab program (The MathWorks, Inc.) by measuring the whole occupied cell area. This area is normalized by the initial area.

## Statistics

Values were compared using a non-parametric two-tailed Mann–Whitney test. The level of statistical significance was set at  $p < 0.05$ . In figure captions,  $n$  represents the number of MCTS investigated for each particular condition or cell line.



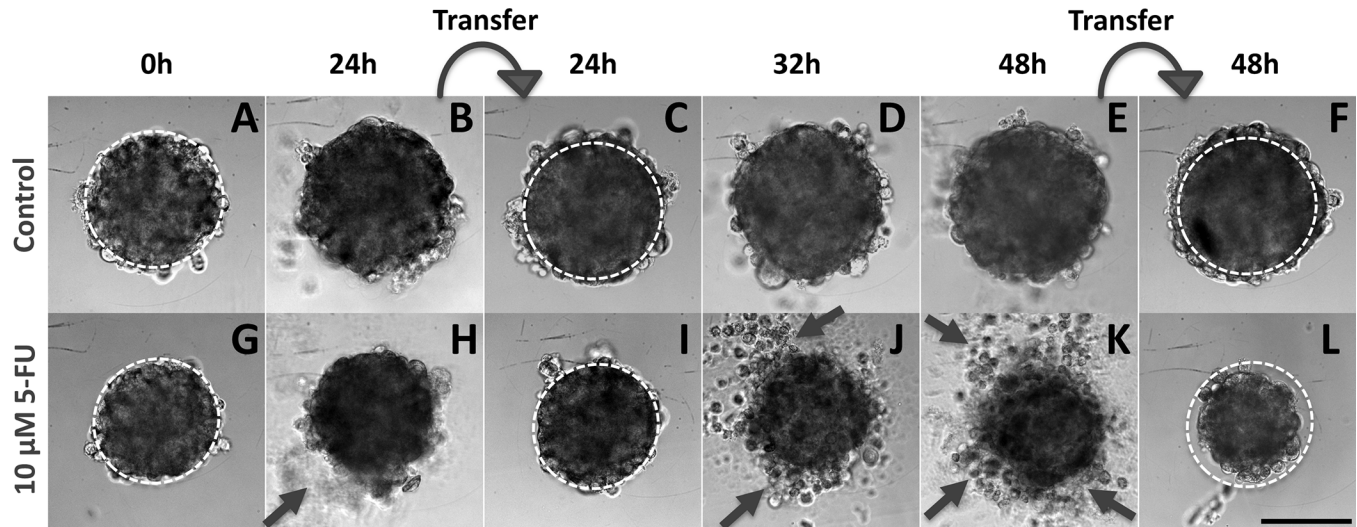
**Fig 1. Typical phase contrast images of the MCTS growth for the three CRC lines taken at 0h, 24h and 48h in the absence of drug. (A-C) HT29, (D-F) HCT116 and (G-I) SW480. Scale bars: 200  $\mu$ m.**

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## Results

### MCTS growth in 3D

Spheroid growth is assessed over time using classical bright field microscope. In the absence of drug, the MCTS mean diameter continuously increases with time from  $219 \pm 2 \mu\text{m}$  at day 0 to  $277 \pm 3 \mu\text{m}$  at day 2 (48h) for HT29 MCTS,  $303 \pm 3 \mu\text{m}$  to  $358 \pm 5 \mu\text{m}$  for HCT116 and  $315 \pm 3 \mu\text{m}$  to  $364 \pm 4 \mu\text{m}$  for SW480 (Fig 1). Assuming a spherical shape, the spheroid diameter or projected area extracted from optical microscopy images enables the calculation of the spheroid volume, which is a straightforward read-out of growth. The MCTS volume increases by a factor of two for HT29 cell line and slightly less for the two other cell lines.



**Fig 2. Typical images of the follow-up of a SW480 MCTS.** Images taken at 0h, 24h, 32h and 48h in the absence of drug (A-F) or in presence of 10  $\mu$ M of 5-FU dispensed at initial time 0h (G-L). The dotted line representing the initial spheroid size in A, G is reported on other images as a guide for the eye. Curved arrows indicate the spheroids transfer into a new well. Straight short arrows show the diffuse cell outer layer. Scale bar: 200  $\mu$ m.

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## MCTS spreading on collagen to rank the cohesion degree of MCTS for the three CRC cell lines

Following the kinetics of spreading of MCTS placed on a surface coated with extracellular cell matrix (ECM) molecules is a simple assay to qualitatively characterize cell-cell cohesion [22], the slower the spreading the higher the cell-cell cohesion. Because, the difference between SW480 and HCT116 was not clearly reported in the literature, we performed this simple assay with our three CRC MCTS placed on collagen film. It appears that the three CRC cell lines tested do not invade inside the collagen (data not shown). By quantifying the spreading area, we show that HT29 are much more cohesive than the two other CRC cell lines. Among them, the HCT116 cell line is slightly but significantly more cohesive than the SW480 cell line (See supplementary S1 Fig).

## An outer loose layer of cells breaks away from the MCTS under 5-FU treatment

When exposed to 5-FU, the measurement of the MCTS volume is not so straightforward as exemplified in Fig 2 with the case of the SW480 cell line (10  $\mu$ M 5-FU condition). A diffuse outer layer of cells appears after 24h (Fig 2H). This layer is loosely attached to the core of the spheroid as a simple transfer by pipetting from one to another well with a fresh renewed medium with drug removes it (Fig 2I). Within eight supplementary hours (*i.e.*, up to 32 h), loose cells again leave the MCTS core (Fig 2J) and at 48h we again performed a transfer to clean up the MCTS (Fig 2K–2L). We checked that control spheroids do not present this loose diffuse layer and that transferring them from one to another well does not change their shape and size (Fig 2A–2F). Using calcein and propidium iodide (PI) dyes (green and red respectively), we assessed at 48h the number of living and dead cells respectively as a function of the 5-FU concentration for SW480 cell line at day 2 without transfer (Fig 3). In order to keep most of the cells of the diffuse layer, we embedded gently the MCTS in agarose before staining cells (see materials and methods). Several important conclusions can be drawn from these observations. First, dead cells (PI+) are present both in the core and in the outer layer of the MCTS even if they are found at higher density (proportionally to the cellular volume) in the diffuse layer. Getting a closer look to this outer layer, we found that the

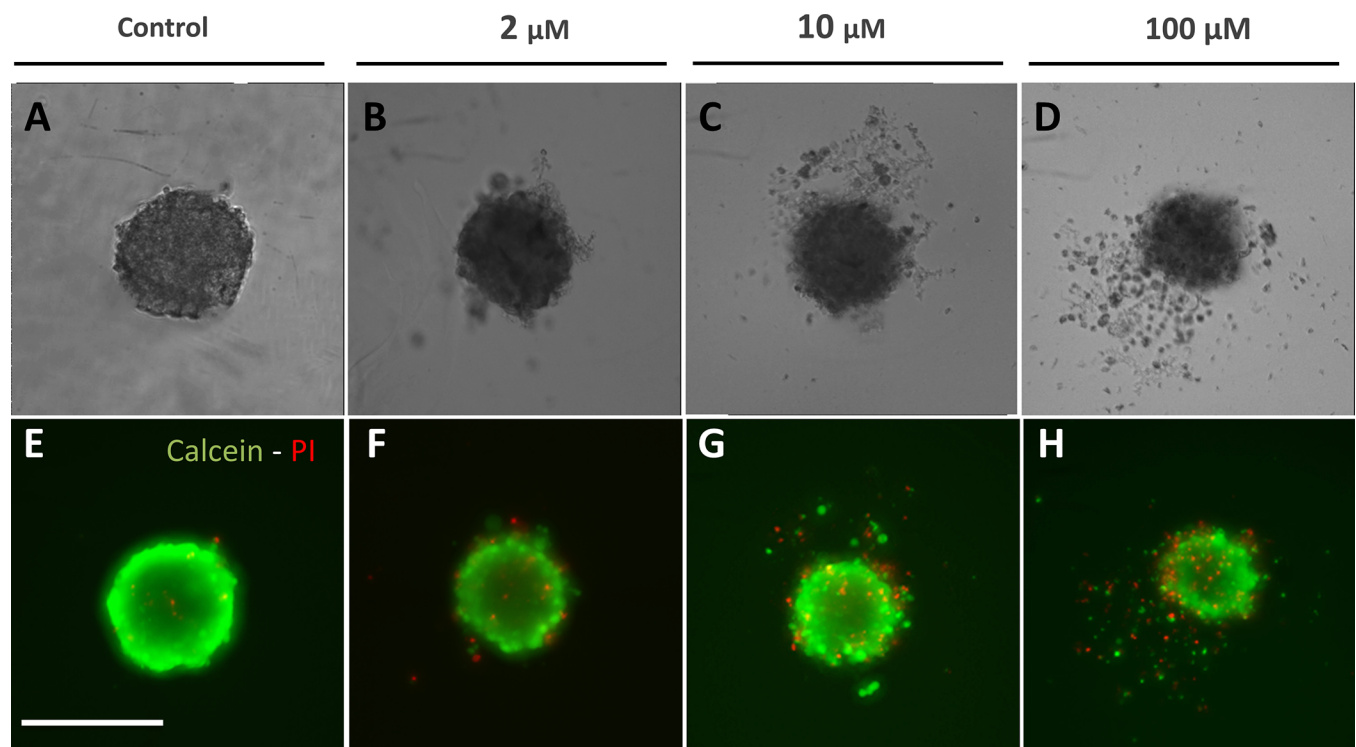
number of dead cell is highly 5-FU concentration dependent (See supplementary S2 Fig). Finally and more surprisingly, the diffuse outer layer is composed not only of dead cells, but also of living cells, with a proportion of living or dead cell roughly equivalent in the outer layer whatever the 5-FU concentration (50%/50%, see supplementary S2 Fig). This last observation indicates that this outer layer is partially composed of live 5-FU resistant cells that detached from the core of the MCTS. For HCT116, we also observe the formation of an outer layer (Fig 4D–4F) but with a slight timing difference as it takes more than eight hours after the first transfer to rebuild it from scratch (Fig 4E versus Fig 4H for HCT116 and SW480 respectively, and quantification in supplementary S3 Fig). For HT29 MCTS, we do not observe any loose outer layer (Fig 4A–4C), the situation is similar to control MCTS.

### Death in the spheroid center

We computed the number of PI+ cells within the core of the MCTS at 48h after transferring the MCTS to a new plate in order to remove the diffuse layer. The number of PI+ particles increases proportionally to the concentration of 5-FU for all cell lines (Fig 5). A higher number of dead cells per unit volume is observed for SW480 spheroids at low 5-FU concentrations ( $[5\text{-FU}] = 5\text{--}10\ \mu\text{M}$ ) as compared to the other metastatic cell line HCT116 spheroids while HT29 spheroids present the lower number of PI+.

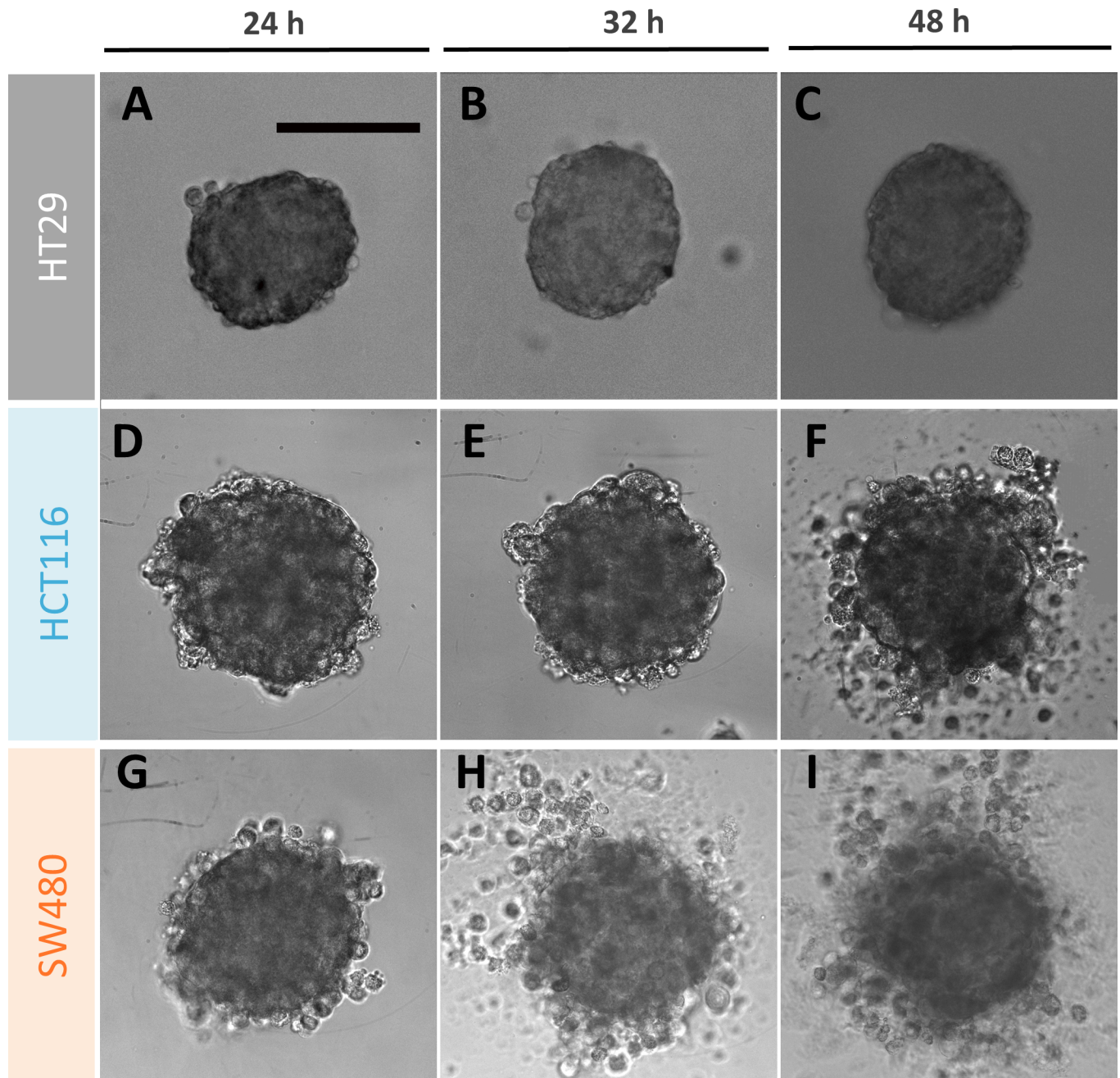
### Drug effect on the cohesive core growth

The outer diffuse layer, if accounted in the area measurement, induces an overestimation of the growth. For that reason, we monitor every day MCTS growth after transferring the MCTS



**Fig 3. Nature of cells in the outer layer of the SW480 spheroids at 48h.** (A–D) Phase contrast and (E–H) corresponding fluorescent images of living cells in green (calcein+) and dead cells in red (PI+) treated with 0, 2, 10 and 100  $\mu\text{M}$  of 5-FU. Compared to control (A,E), a gradual disaggregation of peripheral cells appears clearly since 2  $\mu\text{M}$  of 5-FU (B,F) to 100  $\mu\text{M}$  (D,H). Cells in the outer layer are equally dead and living cells. Scale bar: 200  $\mu\text{m}$ .

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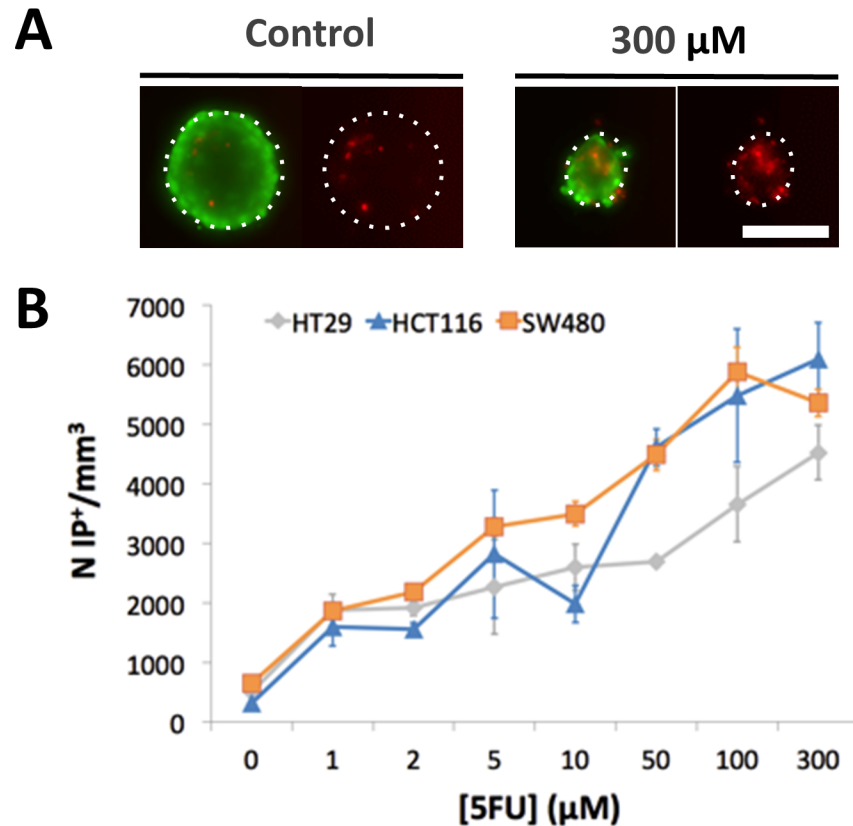


**Fig 4. An outer loose layer of cells breaks away from the MCTS under 10  $\mu$ M of 5-FU treatment.** Typical images of MCTS of the three different CRC cell lines, HT29 (A-C), HCT116 (D-F) and SW480 (G-I) at three different times after drug administration ([5-FU] = 10 $\mu$ M): 24h (A, D, G), 32h (B, E, H) and 48h (C, F, I). In the absence of transfer, numerous dead cells appear on the diffuse outer cell layer for the HCT116 and SW480 MCTS. Scale bar: 200  $\mu$ m.

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in a new well with fresh medium (Fig 2), leaving only the intact cohesive MCTS core but eliminating the peripheral loose cells (Fig 6). Quantitative measurements of the core volume are performed at 0h, 24h and 48h after transfer (Fig 6D, 6H and 6L). With respect to the control, the core volume gradually decreases as a function of the 5-FU concentration. 3D IC<sub>50</sub> are estimated from the 50% value in 3D cell survival curves (Fig 7A at 48h). Here, 3D cell survival is





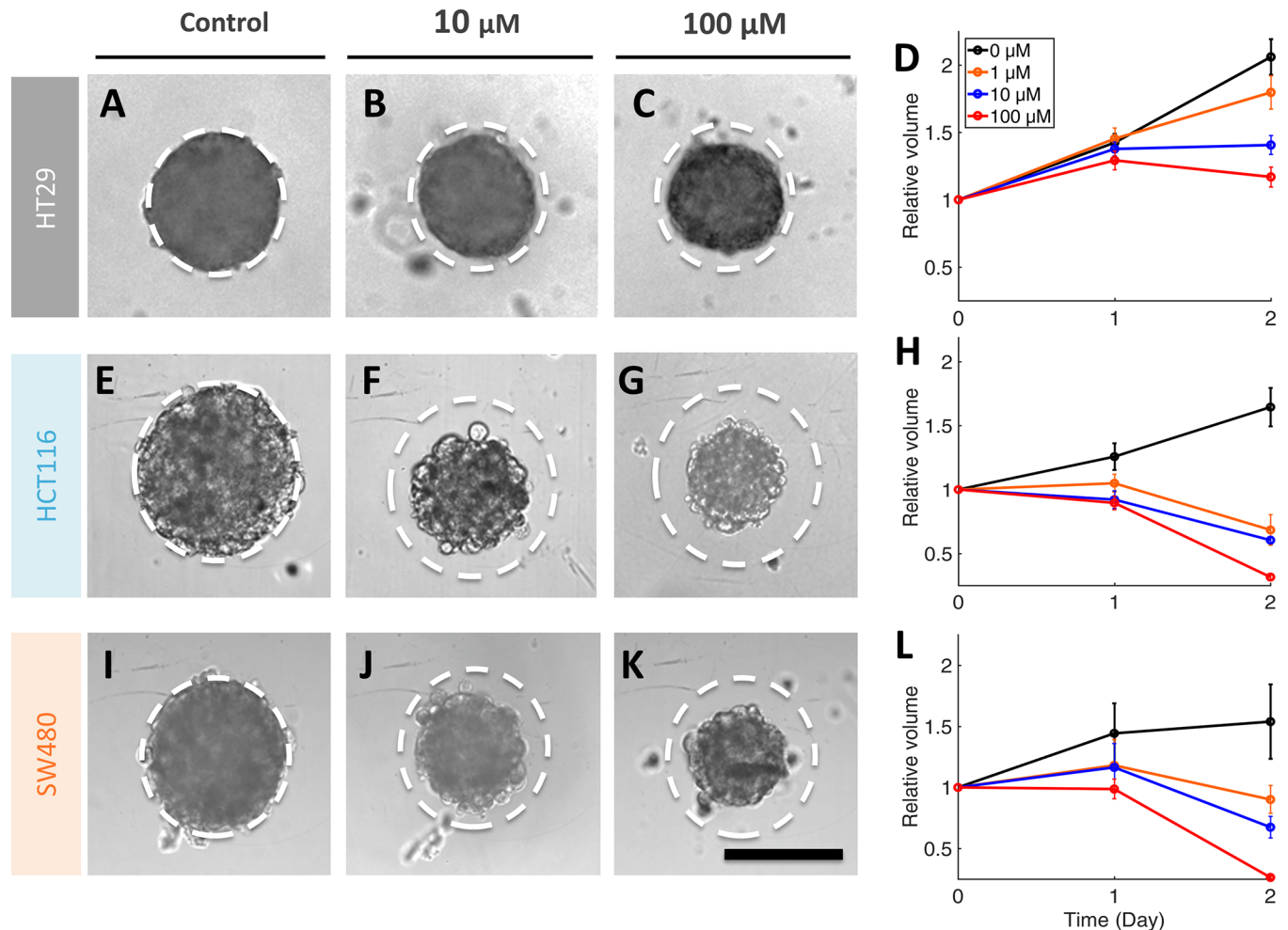
**Fig 5. Number of dead cells within the cohesive core of MCTS under drug treatment after transfer to new wells.** (A) Typical fluorescence images at 48h of living cells in green (calcein +) and dead cells in red (PI+) in control SW480 MCTS or in presence of 5-FU (300 μM). (B) Quantification of PI+ dead cells per unit volume for each cell line as a function of the 5-FU concentration. Error bar = SEM (standard error of the mean), (n = 12 for each cell line). Scale bar = 200 μm.

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defined as the ratio between the core volume for a given condition to the control volume at the same time. This quantity decreases continuously from 1 to 300 μM 5-FU. IC<sub>50</sub> values at 24h and 48h are reported in Table 1: they are [1–5 μM] for both SW480 and HCT116 and larger than 300 μM for HT29. We clearly observe a higher resistance of HT29 spheroids against 5-FU in comparison with the higher metastatic ones HCT116 and SW480. In addition, as opposed to HCT116 and SW480, HT29 spheroids keep growing for all concentrations below 300 μM (Fig 6D) and are very cohesive even after two days' drug exposure as judged by the absence of the diffuse cell layer (Fig 4).

### Drug effect on 2D cell growth

In order to compare the drug effect obtained in 3D to the most classical one obtained in 2D, we also quantify the effect of 5-FU on 2-D cell growth of our three CRC cell lines using two complementary real time cell analysis methods, namely impedimetric and optical methods (Fig 8). The electrical signal collected via the impedimetric method (Cell Index) is correlated with the whole 2D cell area (Fig 8). This Cell Index is monotonously increasing over time for control conditions (no 5-FU added) for the three CRC cell lines. But it follows a non-monotonous cell line dependent variation in presence of the anti-cancer drug 5-FU. The Cell Index of the HT29 cell population is increasing faster than the control for all 5-FU concentrations. It

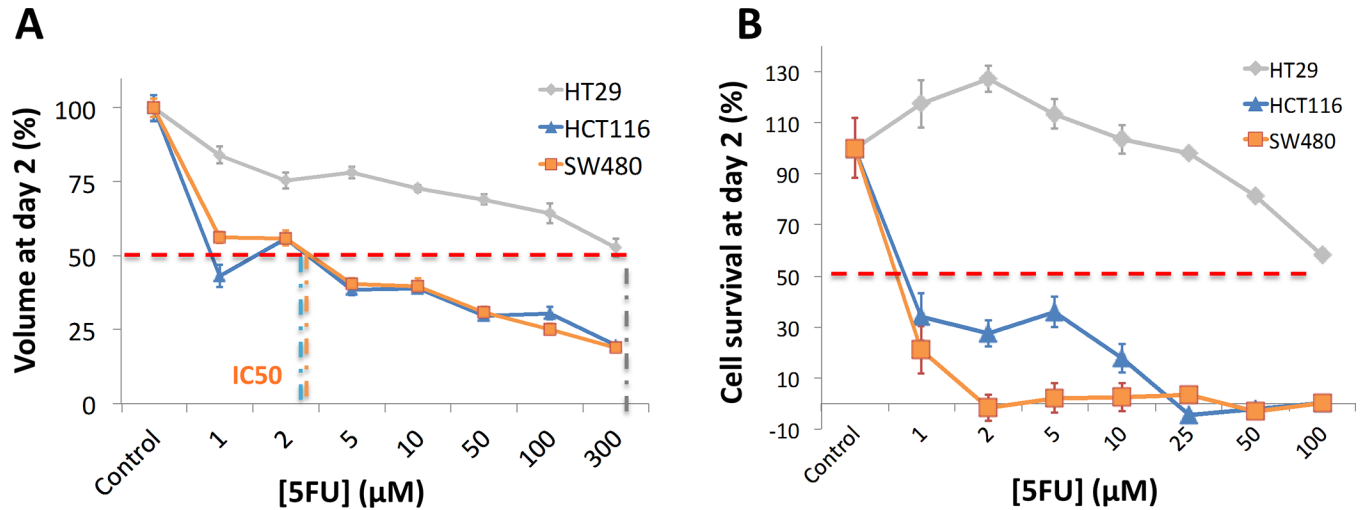


**Fig 6. Volume of the cohesive core of MCTS under drug treatment after transfer to new wells.** Left. Typical images of spheroid cores at 48h for the three CRC cell lines after administration of 0  $\mu\text{M}$  (A, E, I), 10  $\mu\text{M}$  (B, F, J) or 100  $\mu\text{M}$  (C, G, K) 5-FU. The dotted circle represents the size of the control MCTS which is reported as a guide for the eye on other panels. Right. Time series of the mean MCTS volume relative to initial volume at day 0 for each particular condition after 5-FU treatment ranging from 0 to 100  $\mu\text{M}$  (only 3 drug concentrations are reported for clarity). (A-D) HT29, (E-H) HCT116 and (I-L) SW480. Error bars: SEM ( $n = 7-12$  for each cell line). Scale bar: 200  $\mu\text{m}$ .

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saturates and cross the control curve at about 24h for 100  $\mu\text{M}$  and at times larger than 48h for all other concentrations. The HCT116 Cell Index follows qualitatively the same trend but is more sensitive to the drug: it increases faster than the control at short times, it crosses the control curves for times comprised between 18h and 36h depending on the 5-FU concentration and finally decreases quickly to zero by 48h for concentrations larger than 10  $\mu\text{M}$ . We will refer to cell activation in the following the fact that Cell Index (or area) during the first 18-48h after drug administration is larger than the control condition. In presence of drug, the SW480 Cell Index follows the control during the first 12h and drops in a concentration dependent manner abruptly by 24h for all concentrations larger than 5  $\mu\text{M}$ .

To summarize, the response in term of Cell Index or whole cell area is highly cell line dependent. Cells are activated by the drug at short time-points, enter in a quiescent state at intermediate times and decrease at subsequent times due to cell death. Increasing the drug concentration fastens cell death. Except for HT29 with 5FU concentrations lower than 25  $\mu\text{M}$ ,



**Fig 7. 3D versus 2D 5-FU effect at 48h.** (A) 3D cell survival of HCT116, SW480 and HT29 MCTS 48 hours after 5-FU treatment ranging from 0 to 100 μM. Survival represents the ratio of MCTS volume at 48h for a given drug condition to the volume of control MCTS (n = 7–12). (B) 2D cell survival of HCT116, SW480 and HT29 cell lines 48 hours after 5-FU treatment ranging from 0 to 100 μM. Survival represents the ratio of the normalized cell area at 48h for a given drug condition to the control normalized cell area (no drug) (n = 3). Error bars: SD.

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the Cell Index with drug is always lower than the control at 48h. We computed the cell survival at 48h by computing the ratio of the area with drug to the control area at that time (Fig 7B). We next evaluated half maximal inhibitory concentration (IC<sub>50</sub>) values that indicate how much of 5-FU is needed to inhibit the cell proliferation by half (dotted line in Fig 7B). We also evaluated IC<sub>50</sub> at 24h in case of 2D cell culture for comparison with 3D MCTS (Table 1).

### 3D versus 2D drug potency (IC<sub>50</sub>)

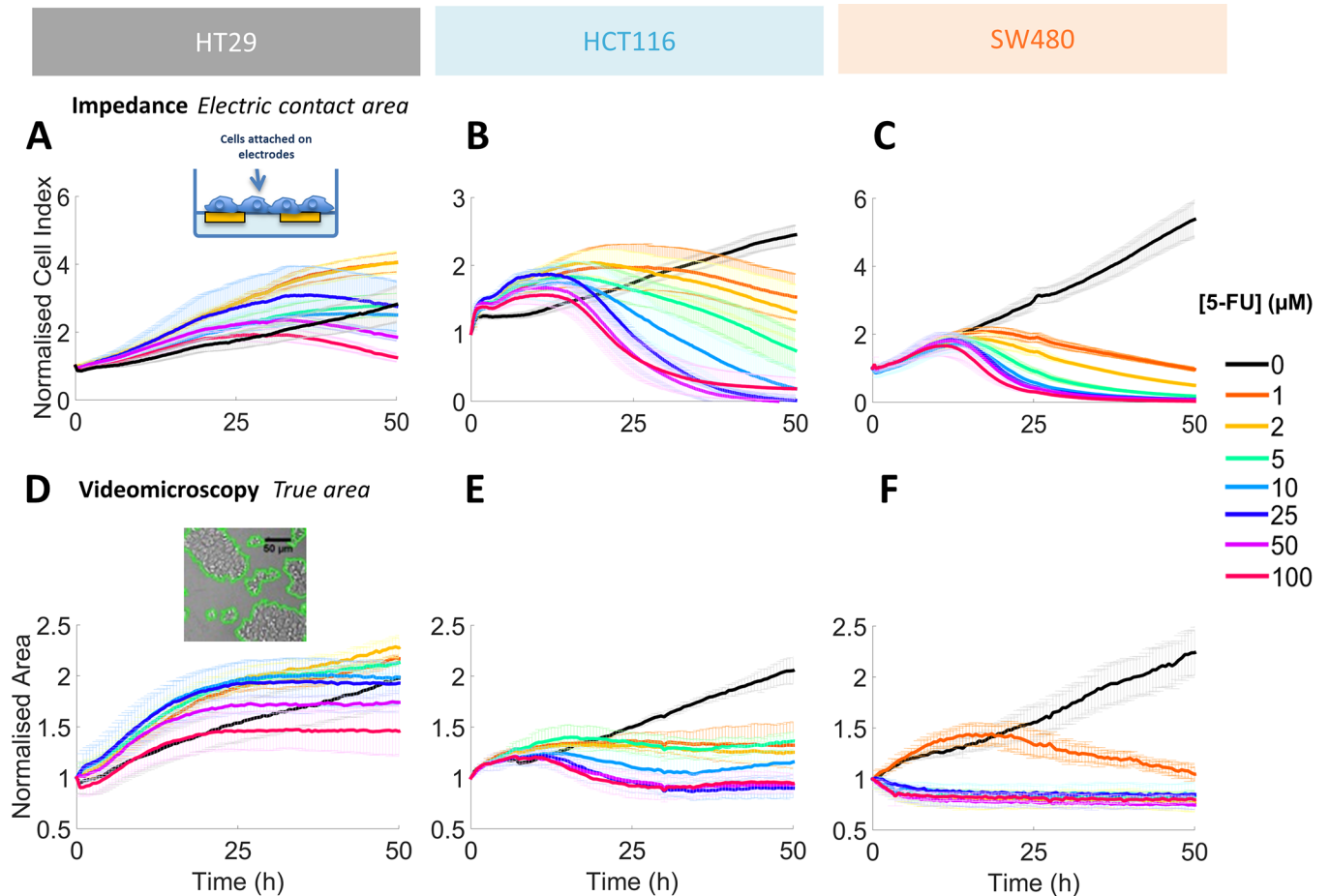
We have found in 2D the same hierarchy than in 3D, with both SW480 and HCT116 much more affected by the drug than HT29 MCTS. The comparison of IC<sub>50</sub> values and cell survival curves at 48h indicates that MCTS are more resistant to 5-FU than 2D cultures. IC<sub>50</sub> values are roughly 2 to 3 times larger in 3D than 2D for the three cell lines (Table 1). At 24 hours post 5-FU, we cannot determine any IC<sub>50</sub> for HT29 both in 2D and 3D. But at that early time, interesting differences appear between the two metastatic cell lines. The IC<sub>50</sub> for SW480 is 1–2 μM and 50–100 μM for HCT116 in 2D while it is not measurable in 3D due to a higher resistance.

As a summary, we found a higher resistance to the drug in 3D than in 2D especially at early times: it is necessary in 3D to wait longer times and administrate higher concentration for a same effect on cell survival. Among the two metastatic cell lines, the SW480 cell line is affected earlier by the drug, a phenomenon seen in 2D on IC<sub>50</sub> values at 24h (IC<sub>50</sub> of 2 μM for SW480 versus 50–100 μM for HCT116) and in 3D from the diffuse outer layer re-apparition at 32h (Fig 4, Supplementary S3 Fig).

**Table 1. IC<sub>50</sub> values for 5-FU for the three cell lines at 24h and 48h in 2D versus 3D spheroids.** A range of IC<sub>50</sub> values (e.g., 1–2 μM) indicates that the precise IC<sub>50</sub> was in between two concentrations tested.

| IC <sub>50</sub> |    | 24h      |           |          | 48h    |        |          |
|------------------|----|----------|-----------|----------|--------|--------|----------|
|                  |    | SW480    | HCT116    | HT29     | SW480  | HCT116 | HT29     |
| IC <sub>50</sub> | 2D | 1-2 μM   | 50-100 μM | > 100 μM | < 1 μM | 2-5 μM | 100 μM   |
|                  | 3D | > 300 μM | > 300 μM  | > 300 μM | 1-5 μM | 1-5 μM | > 300 μM |

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**Fig 8. 2D real time growth analysis.** (A–C) Time series of the Cell Index obtained by RTCA impedance assay for the three CRC cell lines after 5-FU treatment administrated at initial time ([5FU] ranging from 0 to 100  $\mu\text{M}$ , see colour code on the right). (D–F) Time series of the normalized cell area obtained by the videomicroscopy assay for the three CRC cell lines after 5-FU treatment administrated at initial time. Error bars: SD. Results from three distinct experiments in triplicate conditions.

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## Discussion

This study investigated the response to 5-FU treatment of three CRC cell lines. While CRC cell sensitivity to 5-FU has been widely investigated using both 2D [23–26] and 3D models [27–31], to our best knowledge, none of them is comparing the response to 5-FU in terms of kinetics of toxicity and phenotypic analysis for different cell lines presenting different levels of invasive properties. In this study, we found different sensitivity to 5-FU for the three CRC cell lines, ranging from high (SW480), intermediate (HCT116) and low (HT29). We also observe the same hierarchy of CRC cell lines sensitivity between 3D and 2D two days after 5-FU exposure. Moreover, for the three studied cell lines, the 2D  $\text{IC}_{50}$  values are two to three times smaller than in 3D at 48h. Our estimates are comparable to the literature reporting a higher resistance to 5-FU in 3D CRC cell models [13,27–32] and a higher sensitivity for HCT116 compare to HT29 cell lines both in 2D [23–26] and in 3D [7]. As far as the SW480 cell line is concerned, the 5-FU response has never been reported to our best knowledge, despite its relevance as a model of CRC progression [33].

While in 2D the cell survival at 48h is close to zero at a concentration above 5  $\mu\text{M}$  (for SW480) and 25  $\mu\text{M}$  (for HCT116), we show that a core full of living cells is still present in all

MCTS at that time, representing more than 25% of the control core volume. The presence of these living cells may be attributed to (1) a decrease in drug penetration in the center of the spheroid and/or (2) the presence of cells showing an increase resistance to 5-FU treatment in a 3D environment. Numerous studies in various tumoral cell lines have described a good penetration of 5-FU into spheroids compared to other drugs [34–37]. Cell resistance to 5-FU treatment may thus rather be related to a difference in cell-cell and cell-matrix interactions within the spheroid, compared to cells in 2D cultures.

### Spheroid cohesion as an indicator of cell sensitivity to 5-FU

It was reported that tissue cohesion influences the resistance to drugs treatment for HT29 [38]. Several lines of argument indicate that HT29 is much more cohesive than the two metastatic cell lines [39]. HT29 cells are rich in cadherin/catenin cell-to-cell junctions providing a cohesive character implicated in 3D drug resistance [38]. Cadherins are classical cell–cell adhesion molecules and their down-regulation is often associated with increased invasive potential in tumors of epithelial origins [40]. SW480 cells, when confluent, grew in 2D as less cohesive cell clusters, showing only partial membranous localization of E-cadherin and catenins at sites of cell–cell contacts. The membranous localization of E-cadherin and catenins is much stronger for HCT116 and even more for HT29 [39]. To confirm the cohesion degree of our cell lines, we used the method of sequential spheroid trypsinization to obtain qualitatively an index of cell cohesion described before by Carlsson and Nederman [41]. As expected, HT29 MCTS are much more difficult to disrupt than the two other cell types, reflecting their high degree of cohesion. Moreover, increasing cell–cell cohesion decreases spheroid spreading rate on a given substratum [22]. Using such a simple assay of spheroid spreading on a collagen type I, we showed that HT29 cells spread much slower than the two other cell lines HCT116 and SW480 with high spreading potential (S1 Fig). These features are correlated with the cohesion degree of the three cell lines previously described.

It has been shown that adhesion molecules including cadherins, integrins and various components of the ECM can alter tumor mechanical properties [42,43], and hence, cohesion. Cells with an extensive ECM are more susceptible to grow as multicellular spheroids and less subject to disintegration [44]. Moreover, laminin and collagen I, two major components of the ECM and present in large quantities in HT29 spheroids have been reported to be implicated in cell-matrix signaling as a major regulator of various functions of cancer cells [30]. Many studies are now pointing out the role of mechanical cues in tumour progression, and consequently the modification of cell mechanical properties during the metastatic process (reviewed recently in [45], and more specifically for HT29 and SW480 in [46]). In this line, we evidence in this study the presence of a detaching ring of cells for the less cohesive cell lines, allowing us to speculate on an anti-cohesive character to the 5-FU drug treatment, probably due to cell-cell or cell-matrix adhesion properties.

### A novel simple method to quantify cell cohesion and drug resistance

We want to stress out that most of the studies dealing with 3D spheroids are quantifying cell sensitivity to drugs by simply monitoring the volume growth over time, extracted from phase contrast images without considering any gradual change in MCTS density such as the presence of an outer diffuse layer [30]. This approach is working very well in case of a cohesive cancer cell line such as HT29 (grade I) [47]. However, if one wants to focus on drug sensitivity for cells exhibiting higher grades of cancer or for primary cells, the drugs could induce a rapid loss of spheroid cohesion, *i.e.*, a quick disaggregation of the outer cells layer. This was the case in our study for HCT116 and SW480 (both grade IV). Here, the follow-up of the projected area

can lead to completely opposite results. In the present study, by transferring the spheroids at regular time point, we have presented a simple yet robust method to disentangle drug effect on cell proliferation from the effect on cell disaggregation. Note that not only the remaining core, but also some detaching cells are viable cells. These few detaching cells remaining fully active (alive and still capable to spread on collagen type I) are of particular interest as it represents the most resistant subpopulation that should be eliminated to avoid any relapse.

### 5-FU response: 2D cell model reveals cell activation during the first hours of treatment

Surprisingly, we observed an overstimulation of cell growth in 2D the firsts 24 hours after 5-FU mainly for low doses. This interesting effect has been reported recently [26] and can be associated to a mitotic stress after drug addition [23, 48]. It should be pointed out that it is much more difficult to evidence such a dynamic effect in 3D cell model. Indeed, to decipher such increase in mitotic activity and/or mean cell volume within spheroids would require the use of specific optical techniques such as Light Sheet Microscopy or labour intensive immunostaining. As such, this study shows that 2D and 3D cell culture models bring complementary results. 2D models are easy to handle, and even if (1) cells are far from the *in vivo* conditions and (2) the drug dose response is shifted with respect to 3D models, the hierarchy between the resistance of various cell lines to 5-FU is conserved. More importantly, they give information on cellular dynamic that would be difficult to evidence in 3D.

### Conclusions

The identification of active cancer drugs or the combined use of drugs whose action spectrum affects both cohesive and invasive cells is fundamental to increase chemotherapy efficacy [40,49,50]. The proposed methodology yet simple could be a tool readily accessible to analyze the effect of various drugs on spheroids cohesion, a model closer to *in vivo* conditions than traditional 2D cell culture systems. The use of this methodology for CRC cell lines confirms the high resistance to 5-FU treatment of the cohesive HT29 cell line and highlights subtle differences in the sensitivity to 5-FU treatment between the two invasive cell lines SW480 and HCT116. Interestingly, these differences are fully correlated with the MCTS cohesion properties. The early activation that we quantified for HT29 and HCT116 cell lines is very surprising, and we are currently analyzing this effect in more detail. The quantification of the internal structure of spheroids could enable to decipher more precisely the phenotypic changes of cells within the spheroid under 5-FU treatment.

### Supporting information

**S1 Fig. Spreading of the three CRC spheroids on a collagen type I film.** (A-I) Typical images of MCTS taken with the transmission channel of the confocal for the three cell lines at initial time 0h (A, D, G), 24h (B,E,H) and 48h (C,F,I) after depositing the spheroid on the collagen film. The culture medium was not renewed and does not contain 5-FU. Scale bar, 500  $\mu\text{m}$ . (J) Kinetics of spreading (normalized projected cell area Vs. time after deposition). The slower the spreading, the higher is the cell-cell cohesion. Error bars represent SEM (n = 3–5 for each cell line). (TIF)

**S2 Fig. Quantification of live (calcein) and dead (PI) cells in the outer cells layer of SW480 spheroids treated with 0, 2, 10 and 100  $\mu\text{M}$  of 5-FU at day 2.** Left: the disaggregated outer layer is composed of both live and dead cells (50–50%) at all 5-FU concentrations. Right: the

number of dead cells in the outer layer increases gradually with the 5-FU concentration. Because of the difficulty to define the exact boundary between the MCTS core and the outer layer and because some peripheral cells might be lost during the agarose injection step all around the spheroid (see [materials and methods](#)), it is difficult to compare quantitatively the number of cells inside and outside the MCTS core. On the other hand, the transfer technique enables a precise quantification of the number of dead cells in the MCTS core (See [Fig 5](#)). Error bars: SEM. (TIF)

**S3 Fig. Spheroid relative diameter change between 24h (after transfer) and 32h for the two invasive CRC cell lines.** (A) Relative diameter change as a function of the 5-FU concentration. The diameter is evaluated from the spheroid surface area  $A$  including the diffuse outer layer measurement as  $(4A/\pi)^{1/2}$ . Error bars represent SEM ( $n = 7-12$  for each cell line). (B,E) Typical images of MCTS at 24h (after transfer) and 32h for  $10\mu\text{M}$  5-FU. Scale bar,  $200\mu\text{m}$ . (TIF)

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